

# LJMU Research Online

Tryfonos, A, Green, DJ and Dawson, EA

Effects of Catheterization on Artery Function and Health: When Should Patients Start Exercising Following Their Coronary Intervention?

http://researchonline.ljmu.ac.uk/id/eprint/10198/

Article

**Citation** (please note it is advisable to refer to the publisher's version if you intend to cite from this work)

Tryfonos, A, Green, DJ and Dawson, EA (2019) Effects of Catheterization on Artery Function and Health: When Should Patients Start Exercising Following Their Coronary Intervention? Sports Medicine. ISSN 0112-1642

LJMU has developed LJMU Research Online for users to access the research output of the University more effectively. Copyright © and Moral Rights for the papers on this site are retained by the individual authors and/or other copyright owners. Users may download and/or print one copy of any article(s) in LJMU Research Online to facilitate their private study or for non-commercial research. You may not engage in further distribution of the material or use it for any profit-making activities or any commercial gain.

The version presented here may differ from the published version or from the version of the record. Please see the repository URL above for details on accessing the published version and note that access may require a subscription.

For more information please contact <a href="mailto:researchonline@ljmu.ac.uk">researchonline@ljmu.ac.uk</a>

http://researchonline.ljmu.ac.uk/

Andrea Tryfonos<sup>1</sup>, Daniel J Green<sup>2</sup> & Ellen A Dawson<sup>1</sup>

# Effects of catheterization on artery function and health: When should patients start exercising following their coronary intervention?

<sup>1</sup>Research Institute for Sport and Exercise Science, Liverpool John Moores University, Liverpool L3 3AF, United Kingdom

<sup>2</sup>School of Human Sciences (Exercise and Sport Science), The University of Western Australia, Crawley, Western Australia, 6009

Running Title. Catheters, vascular function and exercise rehabilitation

<u>Author for correspondence.</u> Dr Ellen A Dawson, Research Institute for Sport and Exercise Science, Liverpool John Moores University, Liverpool L3 3AF, United Kingdom, Tel: +44 (0)151 904 6264, Fax: +44 (0)151 094 6284, E-mail: e.dawson@ljmu.ac.uk

# ORCID

Andrea Tryfonos. 0000-0002-8226-0724 Prof Daniel J Green. 0000-0003-3226-2921 Dr Ellen A Dawson. 0000-0002-5958-267X

# Key points:

- Coronary interventions can result in artery dysfunction and acute injury, potentially leading to thrombosis and stenosis.
- Due to the arterial injury, a period of relative arterial vulnerability post-coronary interventions may exist.
- Acute exercise during this period, particularly at higher intensities, may result to an increase in oxidative stress, inflammation, transient endothelial dysfunction and a pro-thrombotic milieu, contributing to elevated event susceptibility.
- Patients following coronary interventions can start an exercise-training program between 2 and 4 weeks post-PTCA and/or PCI, recognizing that there may be a "grey area" for functional recovery between 2-12 weeks post-catheterization.
- Individual characteristics of patients (age, severity of disease, comorbidities) and parameters of exercise (intensity, duration, frequency) should all be considered when prescribing exercise following coronary interventions.

#### Abstract:

Coronary artery disease (CAD) is a leading cause of death worldwide, and percutaneous transluminal coronary angiography (PTCA) and/or percutaneous coronary intervention (PCI; angioplasty) are commonly used to diagnose and/or treat the obstructed coronaries. Exercise-based rehabilitation is recommended for all CAD patients; however, most guidelines do not specify when exercise training should commence following PTCA and/or PCI. Catheterization can result in arterial dysfunction and acute injury, and given that fact that exercise, particularly at higher intensities, is associated with elevated inflammatory and oxidative stress, endothelial dysfunction and a pro-thrombotic milieu, performing exercise post-PTCA/PCI may transiently elevate the risk of cardiac events. This review aims to summarize extant literature relating to the impacts of coronary interventions on arterial function, including the time-course of recovery and the potential deleterious and/or beneficial impacts of acute versus long-term exercise. The current literature suggests that arterial dysfunction induced by catheterization recovers 4-12 weeks following catheterization. This review proposes that a period of relative arterial vulnerability may exist and exercise during this period may contribute to elevated event susceptibility. We therefore suggest that CAD patients start an exercise-training program between 2 and 4 weeks post-PCI, recognizing that the literature suggest there is a "grey area" for functional recovery between 2-12 weeks post catheterization. The timing of exercise onset should take into consideration the individual characteristics of patients (age, severity of disease, comorbidities) and the intensity, frequency and duration of the exercise prescription.

#### 1.0 Introduction:

Cardiovascular disease (CVD) is the leading cause of death worldwide with 17.8 million deaths per year [1]. Atherosclerosis, the inflammatory process which underlies most CVD, results in thickening of the artery wall, plaque development and, sometimes, plaque rupture. Endothelial dysfunction is an early atherogenic event, with subsequent transmigration and accumulation of white blood cells and vascular smooth muscle cells (VSMC) into the intima [2-4], resulting in plaque formation and stenosis.

Coronary artery disease (CAD) is the most common cause of cardiovascular deaths [5, 6]. The presence and extent of coronary artery plaque and associated stenosis is commonly determined using percutaneous transluminal coronary angiography (PTCA), a technique that utilizes contrast agent and X-ray based detection to visualize the vessel lumen [7]. Treatment for an obstructed artery can include angioplasty (PCI; percutaneous coronary intervention); where a catheter with balloon tip is inserted and advanced to the stenosed coronary artery. A balloon is inflated to restore the blood flow and a stent (expandable tube-shaped device), is commonly deployed inside the artery to keep it open. PCI is the most common treatment of CAD [8] and one of most widely performed medical procedures in Western world; with 97,376 PCI reported in UK in 2015 [9], although it is worth noting that critical appraisal of the benefits of PCI, relative to conservative medical management, has only recently begun to emerge [10, 11].

Guidelines released by representative bodies recommend that all patients with CAD should receive a program of structured exercise-based cardiac rehabilitation (CR) [8, 12, 13, 6]. However, these guidelines do not typically specify a time for the onset of rehabilitation following PCI, reflecting the relatively weak and conflicting data in regards to the effects of acute exercise in patients following coronary interventions. The UK National Institute for Health and Care Excellence (NICE) is the only body that provides guidance, recommending that programs start 2 weeks post-PCI, and 6 weeks after coronary artery bypass graft (CABG) surgery [14]. The basis for this recommendation regarding timing is, however, unclear and PCI can result in artery dysfunction and/or acute injury [15-17], the extent of which may vary according to procedure and patient characteristics. The time-course of recovery from the "insult" of catheterization, and whether exercise during this period presents a risk, has not been fully considered. The purpose of this review is to summarize extant literature relating to the impacts of coronary interventions on arterial function, including the time-course of the recovery and the potential deleterious and/or beneficial impacts of acute versus long-term exercise. Our aim is to raise consciousness among clinicians and exercise scientists regarding the acute impacts of catheterization on arterial function and to develop an evidence-based approach to recommendations pertaining to the optimal timing to safely begin rehabilitative exercise-based programs following PCI.

### 2.0 Do coronary interventions cause arterial injury?

PTCA/PCI are commonly utilized in the management of CAD, resulting in reduced angina [18], and are considered safe and effective [19, 20]. However, vascular dysfunction and other complications such as re-stenosis and thrombosis are well recognized [21]. There is clear evidence that invasive procedures mechanically disrupt and can damage the vascular endothelium. Acutely, endothelial dysfunction or damage can convert dilator responses to increased flow and shear stress into constriction and possibly even spasm. Chronically, an injured or denudated endothelium permits the development of atherosclerosis, as it no longer provides an anti-thrombotic surface, and VSMC are exposed to the circuiting blood, resulting in platelet binding and activation of the clotting cascade,

possibly leading to thrombosis [21-23]. In addition, loss of endothelial function and cytokine release by platelets and macrophages further stimulate migration and proliferation of VSMC to the intima, contributing to the formation of neo-intima layer and restenosis [24] (Figure 2).

# **2.1** What are the effects of catheter insertion, balloon inflation and stent implantation on arterial structure and function?

# 2.1.1 Structural changes.

Whilst catheterization on its own (i.e. PTCA) may damage the artery, the addition of balloon inflation and stent deployment may cause a greater degree of injury, resulting in neo-intima thickening [25]. Neo-intima thickening appears to depend on the degree of endothelial denudation [26], a small denuded area results in little intimal hyperplasia [27], while larger denuded areas lead to greater intima thickening [28]. Newer, 2<sup>nd</sup> generation bioresorbable drug-eluting stents (DES), with antiinflammatory and anti-proliferative properties, have shown lower neo-intima thickening [29], and superior clinical outcomes [30] including reduced mortality, morbidity [31], and revascularization rates [32]. However, the incidence of in-stent restenosis [33, 34] and late thrombosis [35, 36] is still an issue. Recently, stents with seeding cells, endothelial progenitor cells (EPC) or endothelial cells (EC), have been studied utilizing *in vitro* and *in vivo* models, with promising outcomes regarding in-stent restenosis [37, 38]. Further research and clinical trials are needed in this area.

Transradial PTCA/PCI has become increasingly popular [39], and the radial artery represents a useful surrogate for epicardial arteries as they are comparable in histopathology and size [40]. Radial artery injury has been identified in approximately one-fifth of patients following transradial procedures, one-third of which have significant dissection extending into the media layer [17]. Moreover, greater vascular injury is associated with repeated catheterization [15]. The radial artery can therefore provide insight into acute and long-term structural changes, post-catheterization.

Intima media thickness (IMT), a non-invasive assessment of atherosclerosis, has been shown to increase at 1- [41], 3- [42] and 4-months [43] following radial procedures, potentially indicating a chronic impact on the structure of the artery wall. Interestingly, IMT has also been shown to increase after only 1 day [41], which likely reflects the acute impact on smooth muscle contraction, which itself can increase wall thickness measures [44]. Previous catheterizations may result in long-term structural remodeling, either due to an increased IMT or a decreased diameter [41, 42, 45, 46], which may have consequences for the long-term patency of the vessel. If the radial artery is removed and used as a graft for CABG, the stenosis-free graft patency is higher if the artery had not been previously catheterized for angiogram/PCI [46].

To summarize, there is evidence for an initial increase in artery diameter due to catheter insertion. Long term, further IMT thickening and decreases in diameter may evolve. In additional to structural changes, PCI may also affect arterial function. Structure and function interact [47], as reduced endothelial function post-injury has been associated with the degree of intimal thickening [48, 49].

# 2.1.2 Functional changes.

Numerous methods can be used to assess vascular function, either invasively or non-invasively, in both coronary and peripheral arteries. The most common method to evaluate coronary artery function to date has involved intracoronary infusion of endothelium-dependent and –independent

dilators and quantitative angiography. More recently, non-invasive techniques (cardiac magnetic resonance imaging, positron emission tomography, and computed tomography) have been used to assess changes in coronary diameter.

Impaired endothelium-dependent and -independent dilation, and paradoxical constriction are major functional complications following PTCA and/or PCI in coronary arteries [50, 48, 25, 51-54] and in other catheterized arteries [55-58, 16, 59, 17, 60]. The grade of endothelium-dependent dysfunction may depend on the severity of denudation [56], whereas VSMC function appears not to be related with the extent of denudation [48]. Plain-balloon injury, bare-metal stents (BMS) and drug-eluting stents (DES) all result in acute endothelial and VSMC dysfunction [25, 61]. In addition to the directly stented section, proximal and distal segments may experience impaired vascular responses [62-64]. Greater dysfunction occurs distal than proximal to stented area [65-68], resulting in greater vasoconstriction [53] and distal coronary vasospasm [69], potentially increasing thrombotic risk. Moreover, endothelial dysfunction assessed in this distal section of the vessel has been associated with poor endothelial coverage in this area [67].

Direct measurement of coronary artery function can be difficult and is highly invasive. Over the past few decades, a non-invasive assessment, called flow-mediated dilation (FMD), has emerged to assess endothelium-dependent function in humans, usually in the brachial artery. The change in artery diameter in response to sublingual-glyceryl-trinitrate spray (GTN) is also commonly used to evaluate endothelial-independent, but nitric oxide (NO)-mediated, VSMC function. FMD has been shown to be acutely lower in catheterized radial arteries compared to the non-catheterized arteries [70, 59, 60, 17, 16]. In general, equivocal findings regarding radial artery function post-catheterization are reported, with a reduction noted at 4- [17], 9- [60] and 12-weeks [70] in some studies, whereas others suggest that vascular function returns to baseline at 3- [59, 17, 16] and 12-months [71]. This may mirror coronary function as patients who experienced coronary artery stenosis post-PCI had lower coronary blood flow at 3 months [72]. In addition to dilatory function, the ability of the artery to constrict is also reduced 24h and 7-weeks post-catheterization [73]. However, VSMC function appears to recover earlier than endothelial function, with reduced function at 24h [17, 70, 59, 60], and 1-week [17] but not 1 and 3 months [17] (Figure 1).

Whilst change in function is likely due to direct damage, there may also be a systemic effect. Lower brachial artery function following PCI, assessed in the non-catheterized vessel, was associated with late in-stent restenosis at 1-month [74], 6-month [49] and 12-month [75, 76]. In contrast, VSMC function was unrelated to the incidence of in-stent restenosis [74]. Such vascular responses were independent of stent type [76]. Given that preserved function is often reported in a non-catheterized control limb [16, 17], further work is needed to determine if there is a systemic effect of PCI and whether inflammatory or oxidative stress mechanisms are involved.

In general, evidence suggests that endothelial denudation following endovascular procedures impairs vasomotor function, while VSMC function is often relatively preserved. It remains unclear whether such interventions result in systemic vasomotor dysfunction, but some indicative evidence suggests that this may be the case. Endothelial dysfunction in coronary arteries, assessed by ACh infusion has been associated with 9-13% cardiac event risk and 21% revascularization requirement in 8-year follow-up [77]. Similarly, a 1% decrease in peripheral endothelial function (FMD) has been associated with 8-

22% increase in risk of future CVD events [78], and risk prediction appears to be stronger in diseased than healthy individuals [79]. Nonetheless, FMD post-PCI might also be considered an independent marker to predict late restenosis or revascularization rates induced by coronary stenting [80].

### 2.2 Does contrast media using during PTCA/PCI cause vascular dysfunction?

Contrast media (CM) is a chemical substance used to enhance visibility of structures or fluids within the body, improving the medical imaging and allowing diagnosis of stenosis. However, the intravascular administration of CM can be associated with vasoconstriction [81] and nephrotoxic effects (contrast-induced nephropathy; CIN). Interestingly, CM may change vascular tone differently, depending on the health of the artery. CM normally induces dilation in healthy subjects [82, 83], however, in CAD patients who typically exhibit endothelial dysfunction, coronary vasoconstriction or even vasospasm may occur [83, 84]. *In vitro* studies have demonstrated that CM may exert negative effects on both EC [85, 86] and VSMC [87, 88]. CM is more likely to exert dose-dependent [89] vasoconstriction in segments close to a stenotic lesion [83, 90], emphasizing a possible involvement of endothelial dysfunction, and constriction occurs more frequently following ionic CM than the currently used non-ionic CM [91]. Ionic CM has greater cytotoxic effects on VSMC, compared with non-ionic CM as it is likely to release of tissue plasminogen activator inhibitor (PAI) from platelets and ECs, contributing to intimal hyperplasia following coronary interventions [92].

CM volume is negatively correlated with subsequent FMD, but positively correlated with von willebrand factor (vWF), thiobarbituric acid reactive substances (TBARS), C-reactive protein (C-RP), interleukin (IL)-6), inferring increased oxidative stress and inflammation induced by CM [93]. CIN may be associated with increased oxidative stress and inflammation [94, 93, 95] and lower level of circulating EPC [96], which may underlie the increased major adverse cardiac events [96, 97]. Collectively, these studies suggest that PCTA with CM alone may negatively affect vascular function, potentially due to increased oxidative stress.

# 2.3 Mechanisms leading to neo-intima formation and vasomotor dysfunction post PTCA and PCI

Increased oxidative stress and inflammation post PTCA/PCI has been associated with restenosis and vascular dysfunction [98-105]. Neo-intima formation (and VSMC proliferation) within the first week [106, 27] may be a consequence of the release or synthesis of growth factors by VSMC, including PDGF (platelet-derived growth factor; a potent VSMC mitogen), while the elevated VSMC proliferation in the following weeks [24, 27] may be explained by reduced NO production due to endothelial damage [107]. Loss of eNOS and reduced NO production post-injury initially occurs due to the actual damage/death of EC during the procedure, and subsequently due to increased production of reactive oxygen species (ROS) [100, 99, 108-110], which appears to follow a similar time course to proliferation post-injury [111]. ROS production and inflammation increase immediately after vascular injury (following both PCI and PTCA [102]) and remain elevated for hours [100] or even days (1-15 days) [112, 113, 104, 98], returning close to baseline levels at 1-month [114]. Inflammation, oxidative stress and platelet adhesion are associated with endothelial dysfunction in CAD patients [115] and increased radial IMT in post-PCI patients [43]. In further support of this, some studies indicate that antioxidant treatments in both animals and patients are associated with reduced neo-intima formation [116, 112, 117, 104, 118, 119, 114], greater luminal diameter, and lower risk of major adverse cardiac events [118]. Finally, lower antioxidant capacity in post-PCI patients is a predictor for cardiac event risk [120].

### 2.4 Recovery of function and structure following catheterization.

In animal models, impaired endothelium-dependent dilation of catheterized arteries is evident in the first days to 4 weeks [25, 48, 54, 58] and tends to recover by 4-8 weeks [57, 51]. In contrast, reduced VSMC function is apparent only the first week post-denudation [54, 25, 51]. Similarly, reduced endothelium-dependent constriction observed the first 1-2 weeks post-injury [56, 25], recovers at 1 month [25, 57].

In humans, reduced endothelium-dependent function at 4 weeks [17, 60, 70] typically returns to baseline at 3-12 months [59, 17, 71], whereas VSMC function is generally preserved or appears to recover more quickly, with improved function at 1 month post-catheterization [65, 17, 16]. In addition, marked endothelium-dependent constriction has been reported in denuded radial arteries at 6 months post-PCI, compared to the control vessel. Structural remodeling seems to be more complicated and persistent; with increased IMT still reported 1-4 months post-PCI [41-43] (Figure 1).

EPC derived from bone marrow have been proposed as a source in endothelial regeneration following vascular injury [121-124]. EPC have been shown to decrease in count [125] and function [126] following PCI. However, EPC cannot predict recovery of vessels [17] and do not enhance reendothelialization or reduce restenosis [127, 128]. Therefore, further research is needed to define the role of EPC in vascular recovery post coronary intervention. Alternatively, following endothelial denudation, re-endothelialization begins to cover and repair the denuded area. Endothelial repair is predominantly a result of replication and migration of intact adjacent non-injured endothelial cells, close to the denuded area. In support of this, endothelial repair tends to begin from the edges of the denuded zone (close to intact endothelium) and then converge on the center [129-132].

Regenerating endothelium appears to re-establish the capacity for arterial constriction before the ability to induce dilation [57]. This may result in a greater long-term contractile capacity, increasing the risk for vasospasm and thrombotic events. Indeed, paradoxical vasoconstriction in coronary arteries (treated and non-treated) has been reported [53, 133] at 1 to 6 months post-PCI. Interestingly, incomplete re-endothelialization is apparent at 1-month post-PCI only in DES-treated arteries, while complete recovery is reported in arteries treated with balloon-inflation only or BMS [25]. DES induces non-specific anti-proliferative effects; not only reducing the VSMC proliferation rate but also influencing the growth of other cell types, including EC. It may negatively affect the recovery of the endothelium post-denudation.

Endothelial regeneration post-injury is associated with the incidence of thrombotic events [21], supporting the importance of early re-endothelialization. Delayed recovery may be associated with endothelial dysfunction [134, 135], explaining in-stent restenosis [33, 34] or late thrombosis [35, 36], post-PCI. However, it is important that the endothelial lining function well in order to reduce thrombosis and restenosis risk [136, 137, 67, 128]. Even in arteries with intact endothelial coverage [68], functional abnormalities may occur and are related to intimal thickening [48, 138, 51, 139]. Promoting functional re-endothelialization may be an important strategy to eliminate adverse complications following coronary interventions; however further research is required to understand the cellular and molecular mechanisms that drive EC to induce endothelial regeneration.

Tradionally, the impact of exercise training in CAD patients has been focussed on improvements in fitness levels [140-143] and cardiovascular risk factors, such as hypertension and lipid profiles [143, 144]. However, this cannot fully explain the exercise-induced cardiovascular risk reduction [145, 144]. It has been suggested that this 'risk-factor gap', can be explained through exercise-mediated improvemnts in factors such as endothelial function [47, 146]. Indeed, there is compelling evidence that exercise in CAD patients directly improves vascular function [147-153, 140, 154-156, 143], in exercised limbs [149], non-exercised limbs [143, 157, 158] and in coronary arteries [159], all of which contribute to reduced ischaemic events [160]. This is found in traditional aerobic-based training sessions, combined circuit training with resistance exercise [161, 157] and high intensity exercise training (HIIT) [143].

The impact of exercise training can be profound. Exercise training has been shown to have superior outcomes at 1 year (event-free survival, re-hospitalization, and repeat revascularizations) in CAD patients, compared with those who had PCI [162] with reduced inflammation and ischaemic events still apparent at 2 years [163]. In contrast, no difference in the progression of *de novo* lesions [164] and plaque formation [165] have been found among exercised and non-exercised groups, although this may be related to efficacy of drug treatment and/or the short time-course for the development of atherosclerosis. However, exercised patients exhibited greater improvements in recurrent angina and maximum exercise tolerance than non-exercised patients [166]. More importantly, exercise training has been associated with lower 5-year all-cause mortality in post-PCI patients [167] and is recommended as a key treatment in CAD [8].

It is worthy of note that CAD patients are typically prescribed a range of CVD medications, including statins [168, 169], beta-blockers [170], angiotensin converting enzyme inhibitors and/or angiotensin receptor blockers [171-174], and sometimes calcium channel blockers [175], all of which may positively impact upon the endothelial function. Few studies have directly assessed the combined impacts of exercise and CV medications on endothelial function, but Walsh *et al.* (2003) showed that basal NO bioactivity was increased following exercise in both treated (with statins) and untreated subjects, suggesting that exercise may benefit vascular function independently of the impact of statins per se [157]. To further support this, large-scale meta-analysis comparing medical treatment with and without exercise training, reported that the combination contributes to significantly lower cardiovascular risk and mortality [176, 177], indicating that some of the direct effects of exercise may be due to the impacts of shear stress and hemodynamics [146]. In conclusion, CVD medication and exercise may have independent effects on vascular function, mediated through distinct pathways that may culminate in synergistic enhancement in endothelial function.

A prominent mechanism through which exercise training leads to vascular adaptation is increase in shear stress. Shear stress is the tangential force of the flowing blood on the endothelial surface of the blood vessel, and it is known that high shear stress promotes EC survival, enhancing vasodilation. Conversely, low shear stress will result in EC apoptosis and to vasoconstriction, along with VSMC proliferation and platelet aggregation [178]. Hambrecht *et al.* demonstrated, in CAD patients, that exercise training results in upregulation of eNOS Ser1177, which is known to be related to shear stress transduction [179]. Upregulation of eNOS increases NO production and bioavailability, resulting in

vasodilation. To illustrate the relevance of shear stress in terms of exercise-mediated vascular adaptation, studies were performed in healthy people using an inflated cuff on one exercised limb, in order to blunt the shear stress that accompanied exercise compared to the contralateral unimpeded limb. Following 8 weeks of exercise training, the uncuffed (shear stressed) exercised limb exhibited significant improvement of endothelial function when compared with the cuffed limb [180, 47], supporting the proposal, that shear stress is the key stimuli during exercise to improve endothelial function (see recent review [47]). This was further supported by similar studies in which shear stress increases induced by passive heating (i.e. in the absence of exercise) induced identical adaptation, with similar levels of attenuation in the cuffed side [181].

Mechanisms other than shear stress, such as reduced inflammation and oxidative stress following exercise training, will all contribute to improved endothelial function [182, 152, 183]. This may also be associated with reduced platelet aggregation [184, 185], contributing further to decrease in the risk of thrombotic events [186] and reduced restenosis risk at 6- [152] and 9-month angiographic follow-up [187, 188]. In summary, improved endothelial function following exercise training may be a result of increased eNOS and NO production [189, 190, 151] and decreased oxidative stress [151] and NO scavenging, such that regular physical activity restores the balance between NO production and NO inactivation in CAD [156]. In addition, increased numbers of EPC following exercise training may be another mechanism by which exercise ameliorates endothelial function in CAD [191]. This supported by an association in EPC count and improved FMD and NO synthesis [154].

The studies outlined above indicate that there are profound direct and indirect effects of repeated exercise stimulation on the function, structure and health of conduit arteries in humans. Such benefits have been summarized in a recent review [47]. These effects would be expected to enhance the recovery from catheter related injury, but there is scant evidence regarding the most appropriate time to begin a preventive exercise program, or indeed whether pre-rehabilitation prior to catheterization maybe be as beneficial as post hoc training to enhance recovery [192]. Despite the evidence supporting that exercise-based CR benefits the event-free survival of patients post coronary interventions, the participation rate in both Europe and United States is far lower than desirable; approximately one-third of patients participate in cardiac rehab programs after a cardiac event [193-196]. In the United States, CR referral was remarkably lower in post-PCI than post-CABG patients; 48% and 91%, while the hospital performing the procedure was the strongest predictor of referral [197]. Therefore, an increase in referrals should be considered as a priority in CAD management.

#### 4.0 What are the vascular effects of acute exercise after PTCA/PCI?

Whilst regular exercise has clearly associated with improved endothelial function and reduction of cardiac events, the acute response to a single bout of exercise remains a controversial issue, particularly exercise in patients following coronary interventions. It is proposed that strenuous exercise might acutely enhance the risk of events in cardiac patients, including thromboembolism and myocardial infarction [198]. Although such events are extremely rare; stent thrombosis risk 0-0.02% [199] and 1% [200], some isolated case-reports of sub-total or total occlusion of coronary arteries [198], and fatal acute stent-thrombosis [201-203], have been related with acute exercise following coronary interventions. The dichotomy between the increased acute risk of exercise and the well-established sustained benefits of prolonged exercise training is commonly known as the 'exercise paradox', whereby reduced endothelial function [204] and increased platelet aggregation [205] have

been shown in acute strenuous exercise, whereas improved endothelial function has been reported as a result of prolonged exercise training in CAD patients [143].

4.1 Endothelial function in healthy and CAD population. The impact of acute exercise on endothelial function (as assessed using FMD immediately following exercise) in healthy individuals is equivocal; studies showing an increase [206-208], a decrease [209-211], or no change [212-214]. This variation may relate to exercise intensity, type, duration and the subject's fitness level [215]. In particular, prolonged exercise has been shown to reduce endothelial function in healthy individuals [216] and result in negative effects on vascular stiffness in CAD patients [217]. Despite the duration, exercise intensity (typically defined as the % of maximal heart rate), has been considered as the 'key-factor' driving the vascular responses. In general, high intensity exercise (HIE) causes greater acute endothelial dysfunction than moderate intensity exercise (MIE) [215, 211, 218]. However, the role of exercise intensity on vascular responses in CAD population is unclear, with studies showing greater dysfunction in HIE [219], and others no difference compared to MIE [220-222]. The above controversy may be explained, in part, by the fact that each study had different exclusion criteria to define CAD patients; i.e. including or excluding patients with previous coronary interventions within 3 months. Differences in baseline FMD appears to affect the vascular responses acutely post-exercise; lower baseline FMD may result in increase in endothelial function post-exercise, while higher baseline-FMD may be associated with a decrease [222], further clouding the understanding of acute effects of exercise on endothelial function. Apart from non-invasive FMD data collected post-exercise, more invasive studies in animals have demonstrated an impairment in endothelial function to increased blood flow during exercise, paradoxical vasoconstriction to acetylcholine (ACh), but preserved VSMC dilation in atherosclerotic animals, whilst arteries from healthy animals dilate [223]. Similarly, Gordon et al. [224] suggested that vasoconstriction in response to ACh and exercise is apparent only in patients with atherosclerosis, while patients with angiographically smooth vessels appeared to preserve endothelial vasodilation.

Circulating EC, soluble E-selectin and vWf, (markers of endothelial dysfunction), are all elevated acutely post-exercise stress tests in CAD patients [225], suggesting that exercise bouts may impact on endothelial function in these patients. To our knowledge, only one study evaluated VSMC function immediately post a single bout of submaximal cycling exercise in CAD patients, showing a decrease at 15min post-exercise [222]. Whilst the above studies have examined peripheral arteries, changes in coronary artery function with isometric handgrip exercise has also been examined in patients with CAD [226, 227]. Isometric handgrip exercise resulted in abnormal coronary responses with reduced vasodilation and blood flow [226], supporting the incidence of coronary endothelial-dependent dysfunction [227].

<u>4.2 Endothelial function post-denudation.</u> Even less evidence exists regarding vascular responses of acute exercise in vessels that have been catheterized (Figure 1&2). Pohl *et al.* 1986 first illustrated i*n vivo* paradoxical vasoconstriction post-denudation in response to increased blood flow and ACh (endothelial-mediated function) in femoral arteries, whereas dilation appeared to be preserved in response to nitroglycerin (VSMC function) [58]. Following on from this, Berdeaux *et al.* 1994 reported marked vasoconstriction in response to exercise in canine epicardial arteries post endothelial denudation. This 'paradoxical' vasoconstriction in denuded arteries has also been observed following administration of ACh (endothelial-mediated function) and nitroglycerin (VSMC function). VSMC

function was restored at 3 days and endothelial-mediated function in response to exercise and ACh at 9 days [54].

More recently, studies in patients performing supine bicycle exercise during coronary catheterization reported an exercise-induced paradoxical vasoconstriction in coronary-treated artery, at 6 months post-PCI with 1<sup>st</sup> generation [228], and at 16 months with 2<sup>nd</sup> generation DES [229], with normal vasodilation in the non-catheterized vessel [229]. VSMC-induced dilation was abolished in the stented area, whereas vasodilation was still apparent proximally and distally to the stent [229]. Coronary vasoconstriction during exercise, 6-months post-PCI, has been implicated in chest-pain with the absence of significant stenosis in a recent case report [135]. Overall, these data suggest that coronary interventions result in impaired vascular responses to acute exercise, mainly in endothelium-dependent function, which may increase the risk for vasoconstriction, spasm and possibly cardiac events.

<u>4.3 EPC.</u> The role of EPC in post-catheterization injury and repair is unclear. Nonetheless, EPC count is increased immediately post exercise stress test in revascularized patients [230, 160], although this may only be apparent in patients who experienced exercise-induced myocardial ischemia [160]. In addition, the increase in EPC number appears to be delayed in CAD patients, compared to healthy controls, suggesting a delayed exercise-induced EPC mobilization in CAD patients [231]. More studies need to be done, to confirm whatever circulating EPC contribute the vascular remodeling post-PCI and to investigate their role (and time-course) post-acute exercise.

4.4 <u>Platelet activation.</u> In addition to endothelial dysfunction induced by acute exercise, platelet aggregation may also be increased, contributing to increased thrombotic risk. HIE results in platelet activation and aggregation [205, 232], coagulation [233-235], platelet thrombus formation [236] and platelet-derived microparticles [237] in healthy and CAD populations, and this increase may be more pronounced in CAD patients [238]. Interesting, these effects may not fully reverse with aspirin treatment [239, 240], suggesting that post-PCI patients with prescribed anti-platelet therapy may continue to be at thrombotic risk post-exercise. Tokuue *et al.* (1996) suggested that platelet activation acutely post-exercise may be a result of high shear stress [241], while MIE appears to have some cardioprotective effects; resulting in decreased platelet activation, platelet adhesion [242], and platelet aggregability [232]. This increase in platelet activation following exercise is in line with the exercise paradox and could explain the increased risk of thrombotic events associated predominantly with acute high intensity exercise [198].

According to the most recent ACC/AHA guidelines [243], dual anti-platelet therapy (aspirin and P2Y<sub>12</sub> inhibitor) for 3-12 months (the duration depends on the severity of disease and other comorbidities, risk of bleeding etc.) has been recommended to CAD patients following catheterization. This decreases thrombotic risk and consequently adverse cardiac events following such procedures [244]. Given the evidence that acute exercise, especially at higher intensities, may induce platelet activation, it is important to emphasize that patients who aim to start exercise training following catheterization, should adherence to anti-platelet therapy.

<u>4.5 Potential mechanisms.</u> There are a number of different mechanisms underlying both the acute decrease in vascular function and increased platelet aggregation, explaining the risk of exercise-

related cardiac events (Figure 2). Changes in inflammation and oxidative stress can affect endothelial and platelet function. Inflammatory markers (C-RP, IL-6, IL-8, tumor necrosis factor alpha; TNF-a) are increased immediately following HIE, in healthy subjects [205] and CAD patients [245, 246], while no significant inflammation response appears following lower intensity exercise [205]. It is suggested that this acute immune response may stimulate thrombosis by enhancing both platelet activation and endothelial damage [205, 247]. Furthermore, a negative correlation between vascular function and platelet aggregation post-exercise has been reported in CAD [238], but not in healthy subjects [238] or pre-clinical populations [248], suggesting that existing endothelial dysfunction may be associated with attenuated platelet aggregation, resulting in increased thrombotic risk post-exercise. In addition, strenuous exercise leads to an immediate increase in oxidative stress in CAD patients [249], which appears to promote platelet responsiveness [250] and endothelial dysfunction by reducing NO bioavailability [205]. Vitamin C (an antioxidant) abolished the increased oxidative stress and endothelial dysfunction (FMD) post-exercise in CVD patients [251]. Interestingly, some in vivo studies have maintained that anti-oxidant therapy (ascorbic acid or glutathione) results in reduced coronary artery spasm [252], and platelet aggregation [253]. All of these mechanisms appear to follow a similar time-course, with an increase immediately post-strenuous exercise, followed by a normalization.

#### 6.0 Conclusion:

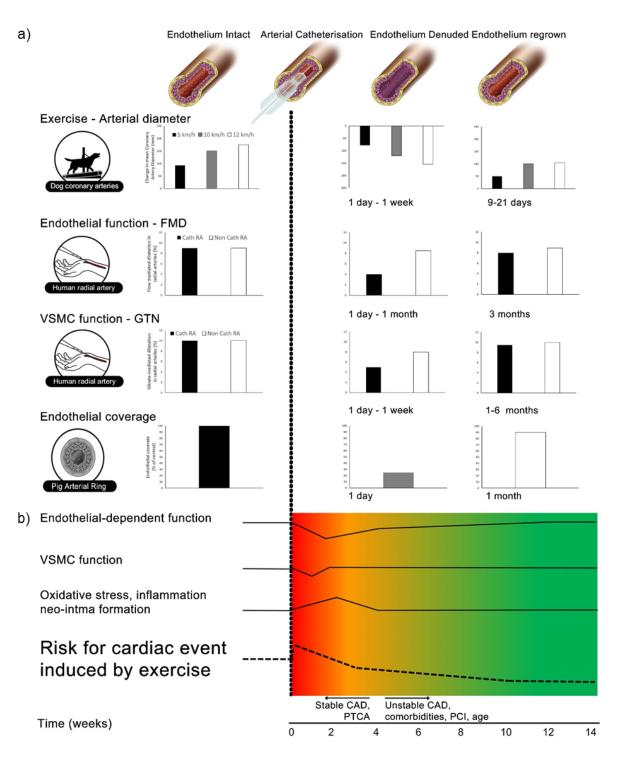
Whilst coronary catheterization is a routine procedure widely used to manage obstructed coronary arteries, insertion of a catheter, CM, balloon inflation and stents, all lead to vascular injury and endothelial dysfunction. Endothelial denudation may result in impaired vascular responses and neointima formation, with further cardiac complications such as restenosis and thrombosis. Endotheliumdependent dysfunction is typically evident, while VSMC function is usually preserved. In general, PCI appears to cause greater damage than diagnostic PTCA, and therefore more time may be needed for vascular responses to recover [25]. Newer generations of DES are superior to 1<sup>st</sup> generation DES and BMS regarding in-stent restenosis, whereas the possibility exists that re-endothelialization may be delayed in DES, with an associated increased risk of late thrombosis.

Exercise training is an important intervention for CAD patients, resulting in improved exercise capacity, quality of life, reduced angina, improved vascular function and a lower risk of cardiac events and/or repeated revascularisation [166, 167, 162.]. More importantly, exercise training is generally considered safe for CAD patients with/without coronary interventions [143]. Exercise-based CR beginning at 8-10 days post-PCI has not been associated with adverse complications [254]. Similarly, a large-scale study, including more than 13,000 patients post-PCI, reported that stent-treated patients should start submaximal exercise at 7 days post-PCI and maximal exercise testing at 14 days [199], while another study with 1000 patients demonstrated that exercise was not associated with any cardiac events, even when exercise stress tests were applied the day after PCI with stenting [200]. Moreover, in a recent study, patients performed maximal exercise test immediately post-PCI, and demonstrated that PCI results in an immediate improvement of exercise tolerance, and increases epicardial and coronary microvascular responses to exercise [255]. Nonetheless, it is important to emphasize that the above studies were performed in a hospital environment, under close monitoring and supervision. Large-scale studies are required to examine the safety and benefits of exercise in a more 'free-living' environment, in order to establish the impact of exercise at different exercise intensities following catheterization. In the near future, the development of devices that track physical activity levels and also some cardiovascular parameters (i.e. heart rate and rhythm) may allow such a real world experiment to be ethically completed.

It is also clear that acute exercise, particularly at higher intensities, has been associated with elevated inflammatory and oxidative stress, transient endothelial dysfunction and a pro-thrombotic milieu [215, 205]. Therefore, caution should be applied when generalizing regarding the safety of exercise immediately post-catheterization. Studies suggest that impaired vascular function immediately following coronary intervenions [50, 55, 58, 54, 59, 17, 16], returns towards pre-catheterization levels 4 to 12 weeks post-PCI [59, 17, 71]. There is also evidence that these detrimental acute effects of exercise may be exacerbated in clinical populations.

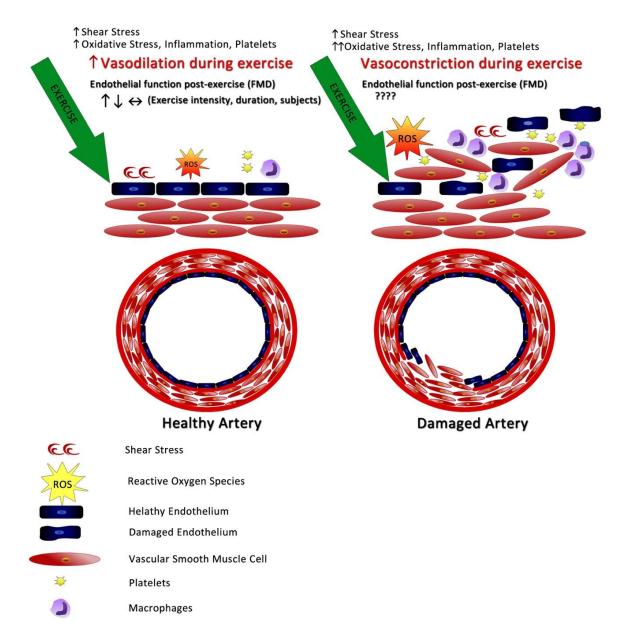
Taking into account the above evidence, we propose that a period of relative arterial vulnerability may exist in the post-catheterization period and that increases in shear stress, oxidative stress and inflammation during this period may contribute to elevated event susceptibility. We suggest that patients should start an exercise-training program between 2 and 4 weeks post-PCI, recognizing that there is a grey area between 2-12 weeks post-catheterization and that discretionary judgement is called for in those at higher risk, related to the individual characteristics of the patients such as age, severity and sub-type of CAD (stable CAD, unstable CAD, non-ST-segment elevation myocardial infarction, ST-segment elevation myocardial infarction), comorbidities, controlled/uncontrolled medication (Figure 1). Consideration should be given to conducting exercise training, particularly that at higher intensity or load, in a supervised and controlled environment in the initial phases, especially in unstable patients and/or those with complicated PCI and/or patients with poor medication compliance. Even though some studies suggest that is safe to start higher-intensity exercise early post-PCI, we believe that the traditional prescriptive approach of beginning with lower and/or lowmoderate intensity exercise, and gradually increasing the duration and/or frequency, followed by intensity, as tolerated by the patient under initial supervision, remains a valid and appropriate approach to progression and clinical exercise prescription.

To conclude, although there is an extensive literature in regard to exercise training in CAD, there is a lack of quality evidence currently available with regard to the effects of acute exercise in arteries following catheterisation. Therefore, further studies are needed to investigate the mechanisms and time-course of vascular repair following catheterization, particularly the mechanisms associated with endothelial dysfunction which may occur following a single bout of exercise in denuded arteries. Further research is also required to characterize differences in the impacts of catheterization between sub-classes of CAD. We believe that the most recent and relatively less invasive techniques, such as MRI and PET, will be helpful in addressing vascular responses to exercise following invasive procedures in future. The guiding principal of "primum non nocere" should be observed when it comes to exercise prescription in high-risk patients and imprudent adoption of the notion that more pain (e.g. a higher intensity) equates with more "gain", threatens this abiding tenet.



**Figure 1:** Time-course of arterial recovery following coronary interventions. Adapted from Figure 4 (Green *et al.* 2017) [47]. a) Summary of the outcomes of studies that investigated the effects of catheterization in arterial function and structure. Paradoxical vasoconstriction reported in *canine* denuded coronaries in response to exercise 3 days post-catheterization, and recovered at 9 days [54]. Endothelial dysfunction, assessed by flow mediated dilation (FMD), reported in patients' catheterized arteries 1 day to 1 month post-catheterization and tend to recover at 3 months [59, 17, 71]. Vascular smooth muscle cell (VSMC) function generally recovers more quickly; in humans reduced VSMC function is apparent only the first week post-denudation [65, 17, 16]. Endothelial coverage in pig's arterial rings was 25% at 1 day post-catheterization, when compared to the control-uninjured vessel,

and recovered up to 80% at 1 month [51]. b) Optimal time to onset exercise training post coronary interventions (PTCA: percutaneous transluminal coronary angiography, PCI: percutaneous coronary intervention). We proposed that patients who undertake PTCA and/or PCI should safely begin exercising at 4 weeks. Stable coronary artery disease (CAD) patients who undergo only PTCA may be able to start exercise training between 2-4 weeks post-PTCA, at a hospital setting, under supervision. Unstable CAD patients, including non ST-segment elevation myocardial infarction (NSTEMI) and ST-segment elevation myocardial infarction (STEMI), with/without comorbidities, severe disease and/or those who undergo PCI should consider starting exercise training at somewhat later, under supervision if possible. Worth noting that oxidative stress, inflammation markers and neo-intima formation which related to the denudation [98-105], follows the same time pattern; increase immediately post denudation [102], remaining elevated for hours [100] or even days (1-15 days) [112, 113, 104, 98], and returning close to baseline levels at 1-month [114].



**Figure 2**: Mechanisms of artery responses to exercise with and without endothelial denudation. Exercise typically induced vasodilation (increase arterial diameter) in healthy arteries (left). Shear stress is elevated during exercise, which will result in increased dilation due to an increase in NO production [179]. Oxidative stress, inflammation and platelets (count/agreeability) may be increased during exercise, particularly following strenuous exercise. Endothelial function immediately post-exercise, assessed by flow mediated dilation (FMD), is equivocal and has been shown to increase, decrease or not change. This variation may relate to exercise intensity, type, duration and the subject's fitness level [215]. The damaged artery, following catheterization (right) will result in paradoxical vasoconstriction [54]. Even though there is an increase in blood flow and shear stress during exercise, the absence of endothelium in the damaged artery can abolish the dilatory response of artery to an exercise [58]. Higher levels of oxidative stress, inflammation and platelets are typically presented in damaged arteries post-catheterization. Endothelial denudation will also result in vascular smooth muscle cell (VSMC) proliferation and migration into the intima, leading to neo-intima formation. There is no information yet in regards to endothelial function post-exercise in damaged arteries.

# Compliance with Ethical Standards:

<u>Conflict of interest.</u> Andrea Tryfonos, Daniel J Green and Ellen A Dawson declare that they have no conflicts of interest.

<u>Funding.</u> Prof. Green is funded by the National Health and Medical Research Council of Australia Principal Research Fellowship (1080914).

#### References

1. Norrving; SMPPB. Global Atlas on Cardiovascular Disease Prevention and Control. 1 ed. 2011.

 Wu MY, Li CJ, Hou MF, Chu PY. New Insights into the Role of Inflammation in the Pathogenesis of Atherosclerosis. International journal of molecular sciences. 2017;18(10). doi:10.3390/ijms18102034.
 Falk E. Pathogenesis of atherosclerosis. Journal of the American College of Cardiology. 2006;47(8 Suppl):C7-12. doi:10.1016/j.jacc.2005.09.068.

4. Ross R. The pathogenesis of atherosclerosis: a perspective for the 1990s. Nature. 1993;362(6423):801-9. doi:10.1038/362801a0.

5. Herrington W, Lacey B, Sherliker P, Armitage J, Lewington S. Epidemiology of Atherosclerosis and the Potential to Reduce the Global Burden of Atherothrombotic Disease. Circulation research. 2016;118(4):535-46. doi:10.1161/circresaha.115.307611.

6. Mozaffarian D, Benjamin EJ, Go AS, Arnett DK, Blaha MJ, Cushman M et al. Executive Summary: Heart Disease and Stroke Statistics--2016 Update: A Report From the American Heart Association. Circulation. 2016;133(4):447-54. doi:10.1161/cir.00000000000366.

7. Campeau L. Percutaneous radial artery approach for coronary angiography. Catheterization and cardiovascular diagnosis. 1989;16(1):3-7.

8. Montalescot G, Sechtem U, Achenbach S, Andreotti F, Arden C, Budaj A et al. 2013 ESC guidelines on the management of stable coronary artery disease: the Task Force on the management of stable coronary artery disease of the European Society of Cardiology. European heart journal. 2013;34(38):2949-3003. doi:10.1093/eurheartj/eht296.

9. Kunadian V, Qiu W, Lagerqvist B, Johnston N, Sinclair H, Tan Y et al. Gender Differences in Outcomes and Predictors of All-Cause Mortality After Percutaneous Coronary Intervention (Data from United Kingdom and Sweden). The American journal of cardiology. 2017;119(2):210-6. doi:10.1016/j.amjcard.2016.09.052.

10. Boden WE, O'Rourke RA, Teo KK, Hartigan PM, Maron DJ, Kostuk WJ et al. Optimal medical therapy with or without PCI for stable coronary disease. The New England journal of medicine. 2007;356(15):1503-16. doi:10.1056/NEJMoa070829.

11. Al-Lamee R, Thompson D, Dehbi HM, Sen S, Tang K, Davies J et al. Percutaneous coronary intervention in stable angina (ORBITA): a double-blind, randomised controlled trial. Lancet (London, England). 2018;391(10115):31-40. doi:10.1016/s0140-6736(17)32714-9.

12. Amsterdam EA, Wenger NK, Brindis RG, Casey DE, Jr., Ganiats TG, Holmes DR, Jr. et al. 2014 AHA/ACC Guideline for the Management of Patients with Non-ST-Elevation Acute Coronary Syndromes: a report of the American College of Cardiology/American Heart Association Task Force on Practice Guidelines. Journal of the American College of Cardiology. 2014;64(24):e139-228. doi:10.1016/j.jacc.2014.09.017.

13. O'Gara PT, Kushner FG, Ascheim DD, Casey DE, Jr., Chung MK, de Lemos JA et al. 2013 ACCF/AHA guideline for the management of ST-elevation myocardial infarction: executive summary: a report of the American College of Cardiology Foundation/American Heart Association Task Force on Practice Guidelines: developed in collaboration with the American College of Emergency Physicians and Society for Cardiovascular Angiography and Interventions. Catheterization and cardiovascular interventions : official journal of the Society for Cardiac Angiography & Interventions. 2013;82(1):E1-27. doi:10.1002/ccd.24776.

14. NICE Guidelines CG 172. November 2013. MI – secondary prevention: Secondary prevention in primary and secondary care for patients following a myocardial infarction <u>https://www.nice.org.uk/guidance/cg48</u>

15. Yonetsu T, Kakuta T, Lee T, Takayama K, Kakita K, Iwamoto T et al. Assessment of acute injuries and chronic intimal thickening of the radial artery after transradial coronary intervention by optical coherence tomography. European heart journal. 2010;31(13):1608-15. doi:10.1093/eurheartj/ehq102.

16. Dawson EA, Rathore S, Cable NT, Wright DJ, Morris JL, Green DJ. Impact of introducer sheath coating on endothelial function in humans after transradial coronary procedures. Circulation Cardiovascular interventions. 2010;3(2):148-56. doi:10.1161/circinterventions.109.912022.

17. Mitchell A, Fujisawa T, Mills NL, Brittan M, Newby DE, Cruden NLM. Endothelial Progenitor Cell Biology and Vascular Recovery Following Transradial Cardiac Catheterization. Journal of the American Heart Association. 2017;6(11). doi:10.1161/jaha.117.006610.

18. Armstrong PW. A comparison of pharmacologic therapy with/without timely coronary intervention vs. primary percutaneous intervention early after ST-elevation myocardial infarction: the WEST (Which Early ST-elevation myocardial infarction Therapy) study. European heart journal. 2006;27(13):1530-8. doi:10.1093/eurheartj/ehl088.

19. Palmerini T, Serruys P, Kappetein AP, Genereux P, Riva DD, Reggiani LB et al. Clinical outcomes with percutaneous coronary revascularization vs coronary artery bypass grafting surgery in patients with unprotected left main coronary artery disease: A meta-analysis of 6 randomized trials and 4,686 patients. American heart journal. 2017;190:54-63. doi:10.1016/j.ahj.2017.05.005.

20. Kerr A, Williams MJ, White H, Grey C, Jiang Y, Nunn C. 30-day mortality after percutaneous coronary intervention in New Zealand public hospitals (ANZACS-QI 18). The New Zealand medical journal. 2017;130(1459):54-63.

21. McDonald AI, Iruela-Arispe ML. Healing arterial ulcers: Endothelial lining regeneration upon vascular denudation injury. Vascular pharmacology. 2015;72:9-15. doi:10.1016/j.vph.2015.06.007.

22. Otsuka F, Finn AV, Yazdani SK, Nakano M, Kolodgie FD, Virmani R. The importance of the endothelium in atherothrombosis and coronary stenting. Nature reviews Cardiology. 2012;9(8):439-53. doi:10.1038/nrcardio.2012.64.

23. Kipshidze N, Dangas G, Tsapenko M, Moses J, Leon MB, Kutryk M et al. Role of the endothelium in modulating neointimal formation: vasculoprotective approaches to attenuate restenosis after percutaneous coronary interventions. Journal of the American College of Cardiology. 2004;44(4):733-9. doi:10.1016/j.jacc.2004.04.048.

24. Hadoke P, Wainwright CL, Wadsworth RM, Butler K, Giddings MJ. Characterization of the morphological and functional alterations in rabbit subclavian artery subjected to balloon angioplasty. Coronary artery disease. 1995;6(5):403-15.

25. Plass CA, Sabdyusheva-Litschauer I, Bernhart A, Samaha E, Petnehazy O, Szentirmai E et al. Time course of endothelium-dependent and -independent coronary vasomotor response to coronary balloons and stents. Comparison of plain and drug-eluting balloons and stents. JACC Cardiovascular interventions. 2012;5(7):741-51. doi:10.1016/j.jcin.2012.03.021.

26. Ip JH, Fuster V, Badimon L, Badimon J, Taubman MB, Chesebro JH. Syndromes of accelerated atherosclerosis: role of vascular injury and smooth muscle cell proliferation. Journal of the American College of Cardiology. 1990;15(7):1667-87.

 27. Fingerle J, Au YP, Clowes AW, Reidy MA. Intimal lesion formation in rat carotid arteries after endothelial denudation in absence of medial injury. Arteriosclerosis (Dallas, Tex). 1990;10(6):1082-7.
 28. Bjorkerud S, Bondjers G. Arterial repair and atherosclerosis after mechanical injury. 5. Tissue response after induction of a large superficial transverse injury. Atherosclerosis. 1973;18(2):235-55.

29. Pendyala L, Yin X, Li J, Shinke T, Xu Y, Chen JP et al. Polymer-free cerivastatin-eluting stent shows superior neointimal inhibition with preserved vasomotor function compared to polymer-based paclitaxel-eluting stent in rabbit iliac arteries. EuroIntervention : journal of EuroPCR in collaboration with the Working Group on Interventional Cardiology of the European Society of Cardiology. 2010;6(1):126-33. doi:10.4244/.

30. Sarno G, Lagerqvist B, Frobert O, Nilsson J, Olivecrona G, Omerovic E et al. Lower risk of stent thrombosis and restenosis with unrestricted use of 'new-generation' drug-eluting stents: a report from the nationwide Swedish Coronary Angiography and Angioplasty Registry (SCAAR). European heart journal. 2012;33(5):606-13. doi:10.1093/eurheartj/ehr479.

31. Qian F, Zhong Y, Hannan EL. Four-year comparative effectiveness of bare-metal and everolimuseluting stents in New York. Catheterization and cardiovascular interventions : official journal of the Society for Cardiac Angiography & Interventions. 2017. doi:10.1002/ccd.27144.

32. Kirtane AJ, Gupta A, Iyengar S, Moses JW, Leon MB, Applegate R et al. Safety and efficacy of drugeluting and bare metal stents: comprehensive meta-analysis of randomized trials and observational studies. Circulation. 2009;119(25):3198-206. doi:10.1161/circulationaha.108.826479.

33. Sabbah M, Kadota K, El-Eraky A, Kamal HM, Abdellah AT, El Hawary A. Comparison of in-stent neoatherosclerosis and tissue characteristics between early and late in-stent restenosis in second-generation drug-eluting stents: an optical coherence tomography study. The international journal of cardiovascular imaging. 2017. doi:10.1007/s10554-017-1146-7.

34. Farooq V, Gogas BD, Serruys PW. Restenosis: delineating the numerous causes of drug-eluting stent restenosis. Circulation Cardiovascular interventions. 2011;4(2):195-205. doi:10.1161/circinterventions.110.959882.

35. Stettler C, Wandel S, Allemann S, Kastrati A, Morice MC, Schomig A et al. Outcomes associated with drug-eluting and bare-metal stents: a collaborative network meta-analysis. Lancet (London, England). 2007;370(9591):937-48. doi:10.1016/s0140-6736(07)61444-5.

36. Mauri L, Hsieh WH, Massaro JM, Ho KK, D'Agostino R, Cutlip DE. Stent thrombosis in randomized clinical trials of drug-eluting stents. The New England journal of medicine. 2007;356(10):1020-9. doi:10.1056/NEJMoa067731.

37. Wu X, Zhao Y, Tang C, Yin T, Du R, Tian J et al. Re-Endothelialization Study on Endovascular Stents Seeded by Endothelial Cells through Up- or Downregulation of VEGF. ACS applied materials & interfaces. 2016;8(11):7578-89. doi:10.1021/acsami.6b00152.

38. Choi WG, Kim SH, Yoon HS, Lee EJ, Kim DW. Impact of an endothelial progenitor cell capturing stent on coronary microvascular function: comparison with drug-eluting stents. The Korean journal of internal medicine. 2015;30(1):42-8. doi:10.3904/kjim.2015.30.1.42.

39. Sandhu K, Butler R, Nolan J. Expert Opinion: Transradial Coronary Artery Procedures: Tips for Success. Interventional cardiology (London, England). 2017;12(1):18-24. doi:10.15420/icr.2017:2:2.

40. Barry MM, Foulon P, Touati G, Ledoux B, Sevestre H, Carmi D et al. Comparative histological and biometric study of the coronary, radial and left internal thoracic arteries. Surgical and radiologic anatomy : SRA. 2003;25(3-4):284-9. doi:10.1007/s00276-003-0142-x.

41. Zhenxian Y, Yujie Z, Yingxin Z, Zhiming Z, Shiwei Y, Zhijian W. Impact of transradial coronary procedures on radial artery. Angiology. 2010;61(1):8-13. doi:10.1177/0003319709348293.

42. Nagai S, Abe S, Sato T, Hozawa K, Yuki K, Hanashima K et al. Ultrasonic assessment of vascular complications in coronary angiography and angioplasty after transradial approach. The American journal of cardiology. 1999;83(2):180-6.

43. Adingupu DD, Westergren HU, Dahgam S, Jonsson-Rylander AC, Blomster J, Albertsson P et al. Radial artery intima-media thickness regresses after secondary prevention interventions in patients' post-acute coronary syndrome and is associated with cardiac and kidney biomarkers. Oncotarget. 2017;8(32):53419-31. doi:10.18632/oncotarget.18511.

44. Thijssen DH, Scholten RR, van den Munckhof IC, Benda N, Green DJ, Hopman MT. Acute change in vascular tone alters intima-media thickness. Hypertension (Dallas, Tex : 1979). 2011;58(2):240-6. doi:10.1161/hypertensionaha.111.173583.

45. Wakeyama T, Ogawa H, lida H, Takaki A, Iwami T, Mochizuki M et al. Intima-media thickening of the radial artery after transradial intervention. An intravascular ultrasound study. Journal of the American College of Cardiology. 2003;41(7):1109-14.

46. Kamiya H, Ushijima T, Kanamori T, Ikeda C, Nakagaki C, Ueyama K et al. Use of the radial artery graft after transradial catheterization: is it suitable as a bypass conduit? The Annals of thoracic surgery. 2003;76(5):1505-9.

47. Green DJ, Hopman MT, Padilla J, Laughlin MH, Thijssen DH. Vascular Adaptation to Exercise in Humans: Role of Hemodynamic Stimuli. Physiological reviews. 2017;97(2):495-528. doi:10.1152/physrev.00014.2016.

48. Weidinger FF, McLenachan JM, Cybulsky MI, Gordon JB, Rennke HG, Hollenberg NK et al. Persistent dysfunction of regenerated endothelium after balloon angioplasty of rabbit iliac artery. Circulation. 1990;81(5):1667-79.

49. Kitta Y, Nakamura T, Kodama Y, Takano H, Umetani K, Fujioka D et al. Endothelial vasomotor dysfunction in the brachial artery is associated with late in-stent coronary restenosis. Journal of the American College of Cardiology. 2005;46(4):648-55. doi:10.1016/j.jacc.2005.04.055.

50. Strotmann JM, Bauersachs J, Fraccarollo D, Kirchengast M, Schnabel PA, Sykora J et al. Trauma induced by nontraumatic coronary devices and its impact on vascular reactivity and morphology. American journal of physiology Heart and circulatory physiology. 2002;283(6):H2356-62. doi:10.1152/ajpheart.00402.2002.

51. Fonseca FA, Izar MC, Fuster V, Gallo R, Padurean A, Fallon JT et al. Chronic endothelial dysfunction after oversized coronary balloon angioplasty in pigs: a 12-week follow-up of coronary vasoreactivity in vivo and in vitro. Atherosclerosis. 2001;154(1):61-9.

52. Lamping KG, Marcus ML, Dole WP. Removal of the endothelium potentiates canine large coronary artery constrictor responses to 5-hydroxytryptamine in vivo. Circulation research. 1985;57(1):46-54.

53. Mc Fadden EP, Bauters C, Lablanche JM, Quandalle P, Leroy F, Bertrand ME. Response of human coronary arteries to serotonin after injury by coronary angioplasty. Circulation. 1993;88(5 Pt 1):2076-85.

54. Berdeaux A, Ghaleh B, Dubois-Rande JL, Vigue B, Drieu La Rochelle C, Hittinger L et al. Role of vascular endothelium in exercise-induced dilation of large epicardial coronary arteries in conscious dogs. Circulation. 1994;89(6):2799-808.

55. Jerius H, Bagwell D, Beall A, Brophy C. The impact of balloon embolectomy on the function and morphology of the endothelium. The Journal of surgical research. 1997;67(1):9-13. doi:10.1006/jsre.1996.4908.

56. Saitoh S, Saito T, Ohwada T, Ohtake A, Onogi F, Aikawa K et al. Morphological and functional changes in coronary vessel evoked by repeated endothelial injury in pigs. Cardiovascular research. 1998;38(3):772-81.

57. Cartier R, Pearson PJ, Lin PJ, Schaff HV. Time course and extent of recovery of endotheliumdependent contractions and relaxations after direct arterial injury. The Journal of thoracic and cardiovascular surgery. 1991;102(3):371-7.

58. Pohl U, Holtz J, Busse R, Bassenge E. Crucial role of endothelium in the vasodilator response to increased flow in vivo. Hypertension (Dallas, Tex : 1979). 1986;8(1):37-44.

59. Dawson EA, Rathore S, Cable NT, Wright DJ, Morris JL, Green DJ. Impact of catheter insertion using the radial approach on vasodilatation in humans. Clinical science (London, England : 1979). 2010;118(10):633-40. doi:10.1042/cs20090548.

60. Burstein JM, Gidrewicz D, Hutchison SJ, Holmes K, Jolly S, Cantor WJ. Impact of radial artery cannulation for coronary angiography and angioplasty on radial artery function. The American journal of cardiology. 2007;99(4):457-9. doi:10.1016/j.amjcard.2006.08.055.

61. Horigome M, Kumazaki S, Hattori N, Kasai H, Horigome M, Aizawa K et al. Noninvasive evaluation of coronary endothelial function following sirolimus-eluting stent implantation by using positron emission tomography. Cardiology. 2009;114(3):157-63. doi:10.1159/000226093.

62. Pendyala LK, Matsumoto D, Shinke T, Iwasaki T, Sugimoto R, Hou D et al. Nobori stent shows less vascular inflammation and early recovery of endothelial function compared with Cypher stent. JACC Cardiovascular interventions. 2012;5(4):436-44. doi:10.1016/j.jcin.2011.11.013.

63. Gogas BD, Benham JJ, Hsu S, Sheehy A, Lefer DJ, Goodchild TT et al. Vasomotor Function Comparative Assessment at 1 and 2 Years Following Implantation of the Absorb Everolimus-Eluting Bioresorbable Vascular Scaffold and the Xience V Everolimus-Eluting Metallic Stent in Porcine Coronary Arteries: Insights From In Vivo Angiography, Ex Vivo Assessment, and Gene Analysis at the Stented/Scaffolded Segments and the Proximal and Distal Edges. JACC Cardiovascular interventions. 2016;9(7):728-41. doi:10.1016/j.jcin.2015.12.018.

64. Hamilos M, Ribichini F, Ostojic MC, Ferrero V, Orlic D, Vassanelli C et al. Coronary vasomotion one year after drug-eluting stent implantation: comparison of everolimus-eluting and paclitaxel-eluting coronary stents. Journal of cardiovascular translational research. 2014;7(4):406-12. doi:10.1007/s12265-014-9568-2.

65. Kim JW, Seo HS, Park JH, Na JO, Choi CU, Lim HE et al. A prospective, randomized, 6-month comparison of the coronary vasomotor response associated with a zotarolimus- versus a sirolimus-eluting stent: differential recovery of coronary endothelial dysfunction. Journal of the American College of Cardiology. 2009;53(18):1653-9. doi:10.1016/j.jacc.2009.01.051.

66. Hamilos MI, Ostojic M, Beleslin B, Sagic D, Mangovski L, Stojkovic S et al. Differential effects of drug-eluting stents on local endothelium-dependent coronary vasomotion. Journal of the American College of Cardiology. 2008;51(22):2123-9. doi:10.1016/j.jacc.2007.12.059.

67. Mitsutake Y, Ueno T, Yokoyama S, Sasaki K, Sugi Y, Toyama Y et al. Coronary endothelial dysfunction distal to stent of first-generation drug-eluting stents. JACC Cardiovascular interventions. 2012;5(9):966-73. doi:10.1016/j.jcin.2012.06.010.

68. Mitsutake Y, Ueno T, Ikeno F, Yokoyama S, Sasaki K, Nakayoshi T et al. Serial changes of coronary endothelial function and arterial healing after paclitaxel-eluting stent implantation. Cardiovascular intervention and therapeutics. 2016;31(1):21-8. doi:10.1007/s12928-015-0341-5.

69. Li J, Jabara R, Pendyala L, Otsuka Y, Shinke T, Hou D et al. Abnormal vasomotor function of porcine coronary arteries distal to sirolimus-eluting stents. JACC Cardiovascular interventions. 2008;1(3):279-85. doi:10.1016/j.jcin.2008.01.009.

70. Yan Z, Zhou Y, Zhao Y, Zhou Z, Yang S, Wang Z. Impact of transradial coronary procedures on radial artery function. Angiology. 2014;65(2):104-7. doi:10.1177/0003319713479650.

71. Madssen E, Haere P, Wiseth R. Radial artery diameter and vasodilatory properties after transradial coronary angiography. The Annals of thoracic surgery. 2006;82(5):1698-702. doi:10.1016/j.athoracsur.2006.06.017.

72. De Vita A, Milo M, Sestito A, Lamendola P, Lanza GA, Crea F. Association of coronary microvascular dysfunction with restenosis of left anterior descending coronary artery disease treated by percutaneous intervention. International journal of cardiology. 2016;219:322-5. doi:10.1016/j.ijcard.2016.06.031.

73. Dawson EA, Alkarmi A, Thijssen DH, Rathore S, Marsman DE, Cable NT et al. Low-flow mediated constriction is endothelium-dependent: effects of exercise training after radial artery catheterization. Circulation Cardiovascular interventions. 2012;5(5):713-9. doi:10.1161/circinterventions.112.971556. 74. Patti G, Pasceri V, Melfi R, Goffredo C, Chello M, D'Ambrosio A et al. Impaired flow-mediated dilation and risk of restenosis in patients undergoing coronary stent implantation. Circulation. 2005;111(1):70-5. doi:10.1161/01.cir.0000151308.06673.d2.

75. Akcakoyun M, Kargin R, Tanalp AC, Pala S, Ozveren O, Akcay M et al. Predictive value of noninvasively determined endothelial dysfunction for long-term cardiovascular events and restenosis in patients undergoing coronary stent implantation: a prospective study. Coronary artery disease. 2008;19(5):337-43. doi:10.1097/MCA.0b013e328301ba8e.

76. Mizia-Stec K, Gasior Z, Haberka M, Mizia M, Chmiel A, Janowska J et al. In-stent coronary restenosis, but not the type of stent, is associated with impaired endothelial-dependent vasodilatation. Kardiologia polska. 2009;67(1):9-17; discussion 8.

77. Halcox JP, Schenke WH, Zalos G, Mincemoyer R, Prasad A, Waclawiw MA et al. Prognostic value of coronary vascular endothelial dysfunction. Circulation. 2002;106(6):653-8.

78. Inaba Y, Chen JA, Bergmann SR. Prediction of future cardiovascular outcomes by flow-mediated vasodilatation of brachial artery: a meta-analysis. The international journal of cardiovascular imaging. 2010;26(6):631-40. doi:10.1007/s10554-010-9616-1.

79. Ras RT, Streppel MT, Draijer R, Zock PL. Flow-mediated dilation and cardiovascular risk prediction: a systematic review with meta-analysis. International journal of cardiology. 2013;168(1):344-51. doi:10.1016/j.ijcard.2012.09.047.

80. Kubo M, Miyoshi T, Oe H, Ohno Y, Nakamura K, Ito H. Prognostic significance of endothelial dysfunction in patients undergoing percutaneous coronary intervention in the era of drug-eluting stents. BMC cardiovascular disorders. 2015;15:102. doi:10.1186/s12872-015-0096-z.

81. Ribeiro L, de Assuncao e Silva F, Kurihara RS, Schor N, Mieko E, Higa S. Evaluation of the nitric oxide production in rat renal artery smooth muscle cells culture exposed to radiocontrast agents. Kidney international. 2004;65(2):589-96. doi:10.1111/j.1523-1755.2004.00408.x.

82. Zelis R, Caudill CC, Baggette K, Mason DT. Reflex vasodilation induced by coronary angiography in human subjects. Circulation. 1976;53(3):490-3.

83. Limbruno U, Petronio AS, Amoroso G, Baglini R, Paterni G, Merelli A et al. The impact of coronary artery disease on the coronary vasomotor response to nonionic contrast media. Circulation. 2000;101(5):491-7.

84. Satoh A, Matsuda Y, Sakai H, Nakatsuka M, Ogawa H, Katayama K et al. Coronary artery spasm during cardiac angiography. Clinical cardiology. 1990;13(1):55-8.

85. Barstad RM, Buchmann MS, Hamers MJ, Orning L, Orvim U, Stormorken H et al. Effects of ionic and nonionic contrast media on endothelium and on arterial thrombus formation. Acta radiologica (Stockholm, Sweden : 1987). 1996;37(6):954-61. doi:10.1177/02841851960373p2102.

86. Kaessmeyer S, Sehl J, Khiao In M, Hiebl B, Merle R, Jung F et al. Organotypic soft-tissue co-cultures: Morphological changes in microvascular endothelial tubes after incubation with iodinated contrast media. Clinical hemorheology and microcirculation. 2016;64(3):391-402. doi:10.3233/ch-168119.

87. Wang YX, Chan P, Morcos SK. The effect of radiographic contrast media on human vascular smooth muscle cells. The British journal of radiology. 1998;71(844):376-80. doi:10.1259/bjr.71.844.9659129.

88. Takatsuki H, Furukawa T, Liu Y, Hirano K, Yoshikoshi A, Sakanishi A. Effect of contrast media on vascular smooth muscle cells. Acta radiologica (Stockholm, Sweden : 1987). 2004;45(6):635-40.

89. Gomi N. Vasoconstriction by angiographic contrast media in isolated canine arteries. The British journal of radiology. 1992;65(779):961-7. doi:10.1259/0007-1285-65-779-961.

90. Kelly RV, Gillespie MJ, Cohen MG, McLaughlin DP, Magnus Ohman E, Stouffer GA. The contrast media iohexol causes vasoconstriction of the proximal left anterior descending coronary artery: implications for appropriate stent sizing. Angiology. 2008;59(5):574-80. doi:10.1177/0003319708318375.

91. Karstoft J, Baath L, Jansen I, Edvinsson L. Vasoconstriction of isolated arteries induced by angiographic contrast media. A comparison of ionic and non-ionic contrast media iso-osmolar with plasma. Acta radiologica (Stockholm, Sweden : 1987). 1995;36(3):312-6.

92. Giedrojc J, Radziwon P, Krupinski K, Kielpinska K, Galar M, Bielawiec M. Effect of nonionic and ionic contrast media on fibrinolysis in patients undergoing angiography. Polish journal of pharmacology. 1996;48(3):323-6.

93. Xiang L, Xiang G, Zhang J, Yue L, Zhao L. Contrast agent suppresses endothelium-dependent arterial dilation after digital subtraction angiography procedure in patients with diabetic foot. Endocrine. 2014;46(3):505-11. doi:10.1007/s12020-013-0095-8.

94. Genovesi E, Romanello M, De Caterina R. [Contrast-induced acute kidney injury in cardiology]. Giornale italiano di cardiologia (2006). 2016;17(12):984-1000. doi:10.1714/2612.26891.

95. Cao S, Wang P, Cui K, Zhang L, Hou Y. [Atorvastatin prevents contrast agent-induced renal injury in patients undergoing coronary angiography by inhibiting oxidative stress]. Nan fang yi ke da xue xue bao = Journal of Southern Medical University. 2012;32(11):1600-2.

96. Chiang CH, Huang PH, Chiu CC, Hsu CY, Leu HB, Huang CC et al. Reduction of circulating endothelial progenitor cell level is associated with contrast-induced nephropathy in patients undergoing percutaneous coronary and peripheral interventions. PloS one. 2014;9(3):e89942. doi:10.1371/journal.pone.0089942.

97. Mehran R, Aymong ED, Nikolsky E, Lasic Z, lakovou I, Fahy M et al. A simple risk score for prediction of contrast-induced nephropathy after percutaneous coronary intervention: development and initial validation. Journal of the American College of Cardiology. 2004;44(7):1393-9. doi:10.1016/j.jacc.2004.06.068.

98. Azevedo LC, Pedro MA, Souza LC, de Souza HP, Janiszewski M, da Luz PL et al. Oxidative stress as a signaling mechanism of the vascular response to injury: the redox hypothesis of restenosis. Cardiovascular research. 2000;47(3):436-45.

99. Juni RP, Duckers HJ, Vanhoutte PM, Virmani R, Moens AL. Oxidative stress and pathological changes after coronary artery interventions. Journal of the American College of Cardiology. 2013;61(14):1471-81. doi:10.1016/j.jacc.2012.11.068.

100. Shi Y, Niculescu R, Wang D, Patel S, Davenpeck KL, Zalewski A. Increased NAD(P)H oxidase and reactive oxygen species in coronary arteries after balloon injury. Arteriosclerosis, thrombosis, and vascular biology. 2001;21(5):739-45.

101. Tsimikas S, Lau HK, Han KR, Shortal B, Miller ER, Segev A et al. Percutaneous coronary intervention results in acute increases in oxidized phospholipids and lipoprotein(a): short-term and long-term immunologic responses to oxidized low-density lipoprotein. Circulation. 2004;109(25):3164-70. doi:10.1161/01.cir.0000130844.01174.55.

102. Berg K, Wiseth R, Bjerve K, Brurok H, Gunnes S, Skarra S et al. Oxidative stress and myocardial damage during elective percutaneous coronary interventions and coronary angiography. A comparison of blood-borne isoprostane and troponin release. Free radical research. 2004;38(5):517-25.

103. Pendyala LK, Li J, Shinke T, Geva S, Yin X, Chen JP et al. Endothelium-dependent vasomotor dysfunction in pig coronary arteries with Paclitaxel-eluting stents is associated with inflammation and oxidative stress. JACC Cardiovascular interventions. 2009;2(3):253-62. doi:10.1016/j.jcin.2008.11.009. 104. Nunes GL, Robinson K, Kalynych A, King SB, 3rd, Sgoutas DS, Berk BC. Vitamins C and E inhibit O2-production in the pig coronary artery. Circulation. 1997;96(10):3593-601.

105. Blum A, Schneider DJ, Sobel BE, Dauerman HL. Endothelial dysfunction and inflammation after percutaneous coronary intervention. The American journal of cardiology. 2004;94(11):1420-3. doi:10.1016/j.amjcard.2004.07.146.

106. Majesky MW, Reidy MA, Bowen-Pope DF, Hart CE, Wilcox JN, Schwartz SM. PDGF ligand and receptor gene expression during repair of arterial injury. The Journal of cell biology. 1990;111(5 Pt 1):2149-58.

107. Barbato JE, Tzeng E. Nitric oxide and arterial disease. Journal of vascular surgery. 2004;40(1):187-93. doi:10.1016/j.jvs.2004.03.043.

108. Pasternak RC, Baughman KL, Fallon JT, Block PC. Scanning electron microscopy after coronary transluminal angioplasty of normal canine coronary arteries. The American journal of cardiology. 1980;45(3):591-8.

109. Jeremy JY, Rowe D, Emsley AM, Newby AC. Nitric oxide and the proliferation of vascular smooth muscle cells. Cardiovascular research. 1999;43(3):580-94.

110. Sundaresan M, Yu ZX, Ferrans VJ, Irani K, Finkel T. Requirement for generation of H2O2 for platelet-derived growth factor signal transduction. Science (New York, NY). 1995;270(5234):296-9.

111. Bai H, Masuda J, Sawa Y, Nakano S, Shirakura R, Shimazaki Y et al. Neointima formation after vascular stent implantation. Spatial and chronological distribution of smooth muscle cell proliferation and phenotypic modulation. Arteriosclerosis and thrombosis : a journal of vascular biology. 1994;14(11):1846-53.

112. Gong KW, Zhu GY, Wang LH, Tang CS. Effect of active oxygen species on intimal proliferation in rat aorta after arterial injury. Journal of vascular research. 1996;33(1):42-6.

113. Szocs K, Lassegue B, Sorescu D, Hilenski LL, Valppu L, Couse TL et al. Upregulation of Nox-based NAD(P)H oxidases in restenosis after carotid injury. Arteriosclerosis, thrombosis, and vascular biology. 2002;22(1):21-7.

114. Kochiadakis GE, Arfanakis DA, Marketou ME, Skalidis EI, Igoumenidis NE, Nikitovic D et al. Oxidative stress changes after stent implantation: a randomized comparative study of sirolimuseluting and bare metal stents. International journal of cardiology. 2010;142(1):33-7. doi:10.1016/j.ijcard.2008.12.105. 115. Di Serafino L, Sarma J, Dierickx K, Ntarladimas I, Pyxaras SA, Delrue L et al. Monocyte-platelets aggregates as cellular biomarker of endothelium-dependent coronary vasomotor dysfunction in patients with coronary artery disease. Journal of cardiovascular translational research. 2014;7(1):1-8. doi:10.1007/s12265-013-9520-x.

116. Liang D, Zhang Q, Yang H, Zhang R, Yan W, Gao H et al. Anti-oxidative stress effect of loadingdose rosuvastatin prior to percutaneous coronary intervention in patients with acute coronary syndrome: a prospective randomized controlled clinical trial. Clinical drug investigation. 2014;34(11):773-81. doi:10.1007/s40261-014-0231-0.

117. Freyschuss A, Stiko-Rahm A, Swedenborg J, Henriksson P, Bjorkhem I, Berglund L et al. Antioxidant treatment inhibits the development of intimal thickening after balloon injury of the aorta in hypercholesterolemic rabbits. The Journal of clinical investigation. 1993;91(4):1282-8. doi:10.1172/jci116326.

118. Liu J, Li M, Lu H, Qiao W, Xi D, Luo T et al. Effects of probucol on restenosis after percutaneous coronary intervention: a systematic review and meta-analysis. PloS one. 2015;10(4):e0124021. doi:10.1371/journal.pone.0124021.

119. Tardif JC, Gregoire J, Schwartz L, Title L, Laramee L, Reeves F et al. Effects of AGI-1067 and probucol after percutaneous coronary interventions. Circulation. 2003;107(4):552-8.

120. Abe N, Kashima Y, Izawa A, Motoki H, Ebisawa S, Miyashita Y et al. A 2-year follow-up of oxidative stress levels in patients with ST-segment elevation myocardial infarction: a subanalysis of the ALPS-AMI study. Angiology. 2015;66(3):271-7. doi:10.1177/0003319714525656.

121. Walter DH, Rittig K, Bahlmann FH, Kirchmair R, Silver M, Murayama T et al. Statin therapy accelerates reendothelialization: a novel effect involving mobilization and incorporation of bone marrow-derived endothelial progenitor cells. Circulation. 2002;105(25):3017-24.

122. Iwakura A, Luedemann C, Shastry S, Hanley A, Kearney M, Aikawa R et al. Estrogen-mediated, endothelial nitric oxide synthase-dependent mobilization of bone marrow-derived endothelial progenitor cells contributes to reendothelialization after arterial injury. Circulation. 2003;108(25):3115-21. doi:10.1161/01.cir.0000106906.56972.83.

123. Lin HH, Chen YH, Yet SF, Chau LY. After vascular injury, heme oxygenase-1/carbon monoxide enhances re-endothelialization via promoting mobilization of circulating endothelial progenitor cells. Journal of thrombosis and haemostasis : JTH. 2009;7(8):1401-8. doi:10.1111/j.1538-7836.2009.03478.x.

124. Werner N, Priller J, Laufs U, Endres M, Bohm M, Dirnagl U et al. Bone marrow-derived progenitor cells modulate vascular reendothelialization and neointimal formation: effect of 3-hydroxy-3-methylglutaryl coenzyme a reductase inhibition. Arteriosclerosis, thrombosis, and vascular biology. 2002;22(10):1567-72.

125. Hill JM, Zalos G, Halcox JP, Schenke WH, Waclawiw MA, Quyyumi AA et al. Circulating endothelial progenitor cells, vascular function, and cardiovascular risk. The New England journal of medicine. 2003;348(7):593-600. doi:10.1056/NEJMoa022287.

126. Aicher A, Heeschen C, Mildner-Rihm C, Urbich C, Ihling C, Technau-Ihling K et al. Essential role of endothelial nitric oxide synthase for mobilization of stem and progenitor cells. Nature medicine. 2003;9(11):1370-6. doi:10.1038/nm948.

127. Conte MS, Choudhury RP, Shirakowa M, Fallon JT, Birinyi LK, Choudhry RP. Endothelial cell seeding fails to attenuate intimal thickening in balloon-injured rabbit arteries. Journal of vascular surgery. 1995;21(3):413-21.

128. Lan H, Wang Y, Yin T, Wang Y, Liu W, Zhang X et al. Progress and prospects of endothelial progenitor cell therapy in coronary stent implantation. Journal of biomedical materials research Part B, Applied biomaterials. 2016;104(6):1237-47. doi:10.1002/jbm.b.33398.

129. Schwartz SM, Haudenschild CC, Eddy EM. Endothelial regneration. I. Quantitative analysis of initial stages of endothelial regeneration in rat aortic intima. Laboratory investigation; a journal of technical methods and pathology. 1978;38(5):568-80.

130. Itoh Y, Toriumi H, Yamada S, Hoshino H, Suzuki N. Resident endothelial cells surrounding damaged arterial endothelium reendothelialize the lesion. Arteriosclerosis, thrombosis, and vascular biology. 2010;30(9):1725-32. doi:10.1161/atvbaha.110.207365.

131. Hagensen MK, Raarup MK, Mortensen MB, Thim T, Nyengaard JR, Falk E et al. Circulating endothelial progenitor cells do not contribute to regeneration of endothelium after murine arterial injury. Cardiovascular research. 2012;93(2):223-31. doi:10.1093/cvr/cvr278.

132. Fishman JA, Ryan GB, Karnovsky MJ. Endothelial regeneration in the rat carotid artery and the significance of endothelial denudation in the pathogenesis of myointimal thickening. Laboratory investigation; a journal of technical methods and pathology. 1975;32(3):339-51.

133. Kirigaya H, Aizawa T, Ogasawara K, Sato H, Nagashima K, Onoda M et al. Incidence of acetylcholine-induced spasm of coronary arteries subjected to balloon angioplasty. Japanese circulation journal. 1993;57(9):883-90.

134. Finn AV, Nakazawa G, Joner M, Kolodgie FD, Mont EK, Gold HK et al. Vascular responses to drug eluting stents: importance of delayed healing. Arteriosclerosis, thrombosis, and vascular biology. 2007;27(7):1500-10. doi:10.1161/atvbaha.107.144220.

135. Hiasa K, Takemoto M, Matsukawa R, Matoba T, Kuga T, Sunagawa K. Chest pain without significant coronary stenosis after implantation of sirolimus-eluting stents. Internal medicine (Tokyo, Japan). 2009;48(4):213-7.

136. Won H, Kim JS, Shin DH, Kim BK, Ko YG, Choi D et al. Relationship between endothelial vasomotor function and strut coverage after implantation of drug-eluting stent assessed by optical coherence tomography. The international journal of cardiovascular imaging. 2014;30(2):263-70. doi:10.1007/s10554-013-0325-4.

137. Nakata T, Fujii K, Fukunaga M, Shibuya M, Kawai K, Kawasaki D et al. Morphological, Functional, and Biological Vascular Healing Response 6 Months After Drug-Eluting Stent Implantation: A Randomized Comparison of Three Drug-Eluting Stents. Catheterization and cardiovascular interventions : official journal of the Society for Cardiac Angiography & Interventions. 2016;88(3):350-7. doi:10.1002/ccd.26273.

138. Shimokawa H, Aarhus LL, Vanhoutte PM. Porcine coronary arteries with regenerated endothelium have a reduced endothelium-dependent responsiveness to aggregating platelets and serotonin. Circulation research. 1987;61(2):256-70.

139. Bosmans JM, Bult H, Vrints CJ, Kockx MM, Herman AG. Balloon angioplasty and induction of nonendothelial nitric oxide synthase in rabbit carotid arteries. European journal of pharmacology. 1996;310(2-3):163-74.

140. Luk TH, Dai YL, Siu CW, Yiu KH, Chan HT, Lee SW et al. Effect of exercise training on vascular endothelial function in patients with stable coronary artery disease: a randomized controlled trial. European journal of preventive cardiology. 2012;19(4):830-9. doi:10.1177/1741826711415679.

141. Van Craenenbroeck EM, Frederix G, Pattyn N, Beckers P, Van Craenenbroeck AH, Gevaert A et al. Effects of aerobic interval training and continuous training on cellular markers of endothelial integrity in coronary artery disease: a SAINTEX-CAD substudy. American journal of physiology Heart and circulatory physiology. 2015;309(11):H1876-82. doi:10.1152/ajpheart.00341.2015.

142. Bacon SL, Sherwood A, Hinderliter A, Plourde A, Pierson L, Blumenthal JA. The influence of endothelial function and myocardial ischemia on peak oxygen consumption in patients with coronary artery disease. International journal of vascular medicine. 2012;2012:274381. doi:10.1155/2012/274381.

143. Conraads VM, Pattyn N, De Maeyer C, Beckers PJ, Coeckelberghs E, Cornelissen VA et al. Aerobic interval training and continuous training equally improve aerobic exercise capacity in patients with coronary artery disease: the SAINTEX-CAD study. International journal of cardiology. 2015;179:203-10. doi:10.1016/j.ijcard.2014.10.155.

144. Mora S, Cook N, Buring JE, Ridker PM, Lee IM. Physical activity and reduced risk of cardiovascular events: potential mediating mechanisms. Circulation. 2007;116(19):2110-8. doi:10.1161/circulationaha.107.729939.

145. Green DJ, Walsh JH, Maiorana A, Best MJ, Taylor RR, O'Driscoll JG. Exercise-induced improvement in endothelial dysfunction is not mediated by changes in CV risk factors: pooled analysis of diverse patient populations. American journal of physiology Heart and circulatory physiology. 2003;285(6):H2679-87. doi:10.1152/ajpheart.00519.2003.

146. Joyner MJ, Green DJ. Exercise protects the cardiovascular system: effects beyond traditional risk factors. The Journal of physiology. 2009;587(Pt 23):5551-8. doi:10.1113/jphysiol.2009.179432.

147. Cornish AK, Broadbent S, Cheema BS. Interval training for patients with coronary artery disease: a systematic review. European journal of applied physiology. 2011;111(4):579-89. doi:10.1007/s00421-010-1682-5.

148. Cornelissen VA, Onkelinx S, Goetschalckx K, Thomaes T, Janssens S, Fagard R et al. Exercise-based cardiac rehabilitation improves endothelial function assessed by flow-mediated dilation but not by pulse amplitude tonometry. European journal of preventive cardiology. 2014;21(1):39-48. doi:10.1177/2047487312460516.

149. Gokce N, Vita JA, Bader DS, Sherman DL, Hunter LM, Holbrook M et al. Effect of exercise on upper and lower extremity endothelial function in patients with coronary artery disease. The American journal of cardiology. 2002;90(2):124-7.

150. Ades PA, Savage PD, Lischke S, Toth MJ, Harvey-Berino J, Bunn JY et al. The effect of weight loss and exercise training on flow-mediated dilatation in coronary heart disease: a randomized trial. Chest. 2011;140(6):1420-7. doi:10.1378/chest.10-3289.

151. Edwards DG, Schofield RS, Lennon SL, Pierce GL, Nichols WW, Braith RW. Effect of exercise training on endothelial function in men with coronary artery disease. The American journal of cardiology. 2004;93(5):617-20. doi:10.1016/j.amjcard.2003.11.032.

152. Munk PS, Staal EM, Butt N, Isaksen K, Larsen AI. High-intensity interval training may reduce instent restenosis following percutaneous coronary intervention with stent implantation A randomized controlled trial evaluating the relationship to endothelial function and inflammation. American heart journal. 2009;158(5):734-41. doi:10.1016/j.ahj.2009.08.021.

153. Kim C, Choi HE, Jung H, Kang SH, Kim JH, Byun YS. Impact of aerobic exercise training on endothelial function in acute coronary syndrome. Annals of rehabilitation medicine. 2014;38(3):388-95. doi:10.5535/arm.2014.38.3.388.

154. Steiner S, Niessner A, Ziegler S, Richter B, Seidinger D, Pleiner J et al. Endurance training increases the number of endothelial progenitor cells in patients with cardiovascular risk and coronary artery disease. Atherosclerosis. 2005;181(2):305-10. doi:10.1016/j.atherosclerosis.2005.01.006.

155. Currie KD, Dubberley JB, McKelvie RS, MacDonald MJ. Low-volume, high-intensity interval training in patients with CAD. Medicine and science in sports and exercise. 2013;45(8):1436-42. doi:10.1249/MSS.0b013e31828bbbd4.

156. Linke A, Erbs S, Hambrecht R. Exercise and the coronary circulation-alterations and adaptations in coronary artery disease. Progress in cardiovascular diseases. 2006;48(4):270-84. doi:10.1016/j.pcad.2005.10.001.

157. Walsh JH, Bilsborough W, Maiorana A, Best M, O'Driscoll GJ, Taylor RR et al. Exercise training improves conduit vessel function in patients with coronary artery disease. Journal of applied physiology (Bethesda, Md : 1985). 2003;95(1):20-5. doi:10.1152/japplphysiol.00012.2003.

158. Green DJ, Walsh JH, Maiorana A, Burke V, Taylor RR, O'Driscoll JG. Comparison of resistance and conduit vessel nitric oxide-mediated vascular function in vivo: effects of exercise training. Journal of applied physiology (Bethesda, Md : 1985). 2004;97(2):749-55; discussion 8. doi:10.1152/japplphysiol.00109.2004.

159. Hambrecht R, Wolf A, Gielen S, Linke A, Hofer J, Erbs S et al. Effect of exercise on coronary endothelial function in patients with coronary artery disease. The New England journal of medicine. 2000;342(7):454-60. doi:10.1056/nejm200002173420702.

160. Adams V, Lenk K, Linke A, Lenz D, Erbs S, Sandri M et al. Increase of circulating endothelial progenitor cells in patients with coronary artery disease after exercise-induced ischemia.

 Arteriosclerosis,
 thrombosis,
 and
 vascular
 biology.
 2004;24(4):684-90.

 doi:10.1161/01.ATV.0000124104.23702.a0.

161. Williams MA, Haskell WL, Ades PA, Amsterdam EA, Bittner V, Franklin BA et al. Resistance exercise in individuals with and without cardiovascular disease: 2007 update: a scientific statement from the American Heart Association Council on Clinical Cardiology and Council on Nutrition, Physical Activity, and Metabolism. Circulation. 2007;116(5):572-84. doi:10.1161/circulationaha.107.185214.

162. Hambrecht R, Walther C, Mobius-Winkler S, Gielen S, Linke A, Conradi K et al. Percutaneous coronary angioplasty compared with exercise training in patients with stable coronary artery disease: a randomized trial. Circulation. 2004;109(11):1371-8. doi:10.1161/01.cir.0000121360.31954.1f.

163. Walther C, Mobius-Winkler S, Linke A, Bruegel M, Thiery J, Schuler G et al. Regular exercise training compared with percutaneous intervention leads to a reduction of inflammatory markers and cardiovascular events in patients with coronary artery disease. European journal of cardiovascular prevention and rehabilitation : official journal of the European Society of Cardiology, Working Groups on Epidemiology & Prevention and Cardiac Rehabilitation and Exercise Physiology. 2008;15(1):107-12. doi:10.1097/HJR.0b013e3282f29aa6.

164. Choi HE, Lee BJ, Kim C. Impact of exercise-based cardiac rehabilitation on de novo coronary lesion in patients with drug eluting stent. Annals of rehabilitation medicine. 2014;38(2):256-62. doi:10.5535/arm.2014.38.2.256.

165. van Oort G, Gross DR, Spiekerman AM. Effects of eight weeks of physical conditioning on atherosclerotic plaque in swine. American journal of veterinary research. 1987;48(1):51-5.

166. Yang X, Li Y, Ren X, Xiong X, Wu L, Li J et al. Effects of exercise-based cardiac rehabilitation in patients after percutaneous coronary intervention: A meta-analysis of randomized controlled trials. Scientific reports. 2017;7:44789. doi:10.1038/srep44789.

167. Goel K, Lennon RJ, Tilbury RT, Squires RW, Thomas RJ. Impact of cardiac rehabilitation on mortality and cardiovascular events after percutaneous coronary intervention in the community. Circulation. 2011;123(21):2344-52. doi:10.1161/circulationaha.110.983536.

168. Altun I, Oz F, Arkaya SC, Altun I, Bilge AK, Umman B et al. Effect of statins on endothelial function in patients with acute coronary syndrome: a prospective study using adhesion molecules and flow-mediated dilatation. Journal of clinical medicine research. 2014;6(5):354-61. doi:10.14740/jocmr1863w.

169. O'Driscoll G, Green D, Taylor RR. Simvastatin, an HMG-coenzyme A reductase inhibitor, improves endothelial function within 1 month. Circulation. 1997;95(5):1126-31.

170. Peller M, Ozieranski K, Balsam P, Grabowski M, Filipiak KJ, Opolski G. Influence of beta-blockers on endothelial function: A meta-analysis of randomized controlled trials. Cardiology journal. 2015;22(6):708-16. doi:10.5603/CJ.a2015.0042.

171. Cheetham C, Collis J, O'Driscoll G, Stanton K, Taylor R, Green D. Losartan, an angiotensin type 1 receptor antagonist, improves endothelial function in non-insulin-dependent diabetes. Journal of the American College of Cardiology. 2000;36(5):1461-6.

172. Cheetham C, O'Driscoll G, Stanton K, Taylor R, Green D. Losartan, an angiotensin type I receptor antagonist, improves conduit vessel endothelial function in Type II diabetes. Clinical science (London, England : 1979). 2001;100(1):13-7.

173. O'Driscoll G, Green D, Maiorana A, Stanton K, Colreavy F, Taylor R. Improvement in endothelial function by angiotensin-converting enzyme inhibition in non-insulin-dependent diabetes mellitus. Journal of the American College of Cardiology. 1999;33(6):1506-11.

174. Collis J, Cheetham C, Dembo L, O'Driscoll J, Stanton K, Taylor R et al. Losartan, an angiotensin type 1 receptor inhibitor, and endothelial vasodilator function in Type 1 diabetes mellitus. Diabetic medicine : a journal of the British Diabetic Association. 2000;17(7):553-4.

175. Radenkovic M, Stojanovic M, Prostran M. Calcium Channel Blockers in Restoration of Endothelial Function: Systematic Review and Meta-Analysis of Randomized Controlled Trials. Current medicinal chemistry. 2018. doi:10.2174/0929867325666180713144806.

176. Kokkinos PF, Faselis C, Myers J, Panagiotakos D, Doumas M. Interactive effects of fitness and statin treatment on mortality risk in veterans with dyslipidaemia: a cohort study. Lancet (London, England). 2013;381(9864):394-9. doi:10.1016/s0140-6736(12)61426-3.

177. Naci H, Ioannidis JP. Comparative effectiveness of exercise and drug interventions on mortality outcomes: metaepidemiological study. British journal of sports medicine. 2015;49(21):1414-22. doi:10.1136/bjsports-2015-f5577rep.

178. Paszkowiak JJ, Dardik A. Arterial wall shear stress: observations from the bench to the bedside. Vascular and endovascular surgery. 2003;37(1):47-57. doi:10.1177/153857440303700107.

179. Hambrecht R, Adams V, Erbs S, Linke A, Krankel N, Shu Y et al. Regular physical activity improves endothelial function in patients with coronary artery disease by increasing phosphorylation of endothelial nitric oxide synthase. Circulation. 2003;107(25):3152-8. doi:10.1161/01.cir.0000074229.93804.5c.

180. Birk GK, Dawson EA, Atkinson C, Haynes A, Cable NT, Thijssen DH et al. Brachial artery adaptation to lower limb exercise training: role of shear stress. Journal of applied physiology (Bethesda, Md : 1985). 2012;112(10):1653-8. doi:10.1152/japplphysiol.01489.2011.

181. Tinken TM, Thijssen DH, Hopkins N, Black MA, Dawson EA, Minson CT et al. Impact of shear rate modulation on vascular function in humans. Hypertension (Dallas, Tex : 1979). 2009;54(2):278-85. doi:10.1161/hypertensionaha.109.134361.

182. Cesari F, Sofi F, Caporale R, Capalbo A, Marcucci R, Macchi C et al. Relationship between exercise capacity, endothelial progenitor cells and cytochemokines in patients undergoing cardiac rehabilitation. Thrombosis and haemostasis. 2009;101(3):521-6.

183. Munk PS, Breland UM, Aukrust P, Ueland T, Kvaloy JT, Larsen AI. High intensity interval training reduces systemic inflammation in post-PCI patients. European journal of cardiovascular prevention and rehabilitation : official journal of the European Society of Cardiology, Working Groups on Epidemiology & Prevention and Cardiac Rehabilitation and Exercise Physiology. 2011;18(6):850-7. doi:10.1177/1741826710397600.

184. Haynes A, Linden MD, Robey E, Naylor LH, Ainslie PN, Cox KL et al. Beneficial impacts of regular exercise on platelet function in sedentary older adults: Evidence from a randomized 6-month walking trial. Journal of applied physiology (Bethesda, Md : 1985). 2018. doi:10.1152/japplphysiol.00079.2018. 185. de Meirelles LR, Matsuura C, Resende Ade C, Salgado AA, Pereira NR, Coscarelli PG et al. Chronic exercise leads to antiaggregant, antioxidant and anti-inflammatory effects in heart failure patients. European journal of preventive cardiology. 2014;21(10):1225-32. doi:10.1177/2047487313491662.

186. Sasaki Y, Morimoto A, Ishii I, Morita S, Tsukahara M, Yamamoto J. Preventive effect of long-term aerobic exercise on thrombus formation in rat cerebral vessels. Haemostasis. 1995;25(5):212-7.

187. Lee JY, Yun SC, Ahn JM, Park DW, Kang SJ, Lee SW et al. Impact of cardiac rehabilitation on angiographic outcomes after drug-eluting stents in patients with de novo long coronary artery lesions. The American journal of cardiology. 2014;113(12):1977-85. doi:10.1016/j.amjcard.2014.03.037.

188. Lee HY, Kim JH, Kim BO, Byun YS, Cho S, Goh CW et al. Regular exercise training reduces coronary restenosis after percutaneous coronary intervention in patients with acute myocardial infarction. International journal of cardiology. 2013;167(6):2617-22. doi:10.1016/j.ijcard.2012.06.122.

189. Gielen S, Schuler G, Hambrecht R. Exercise training in coronary artery disease and coronary vasomotion. Circulation. 2001;103(1):E1-6.

190. Laurent M, Daline T, Malika B, Fawzi O, Philippe V, Benoit D et al. Training-induced increase in nitric oxide metabolites in chronic heart failure and coronary artery disease: an extra benefit of waterbased exercises? European journal of cardiovascular prevention and rehabilitation : official journal of the European Society of Cardiology, Working Groups on Epidemiology & Prevention and Cardiac Rehabilitation and Exercise Physiology. 2009;16(2):215-21. doi:10.1097/HJR.0b013e3283292fcf.

191. Cesari F, Marcucci R, Gori AM, Burgisser C, Francini S, Sofi F et al. Impact of a cardiac rehabilitation program and inflammatory state on endothelial progenitor cells in acute coronary syndrome patients. International journal of cardiology. 2013;167(5):1854-9. doi:10.1016/j.ijcard.2012.04.157.

192. Alkarmi A, Thijssen DH, Albouaini K, Cable NT, Wright DJ, Green DJ et al. Arterial prehabilitation: can exercise induce changes in artery size and function that decrease complications of catheterization? Sports medicine (Auckland, NZ). 2010;40(6):481-92. doi:10.2165/11531950-000000000-00000.

193. Aragam KG, Dai D, Neely ML, Bhatt DL, Roe MT, Rumsfeld JS et al. Gaps in referral to cardiac rehabilitation of patients undergoing percutaneous coronary intervention in the United States. Journal of the American College of Cardiology. 2015;65(19):2079-88. doi:10.1016/j.jacc.2015.02.063.

194. Kotseva K, Wood D, De Backer G, De Bacquer D, Pyorala K, Keil U. Cardiovascular prevention guidelines in daily practice: a comparison of EUROASPIRE I, II, and III surveys in eight European countries. Lancet (London, England). 2009;373(9667):929-40. doi:10.1016/s0140-6736(09)60330-5.

195. de Belder MA, Ludman PF, McLenachan JM, Weston CF, Cunningham D, Lazaridis EN et al. The national infarct angioplasty project: UK experience and subsequent developments. EuroIntervention : journal of EuroPCR in collaboration with the Working Group on Interventional Cardiology of the European Society of Cardiology. 2014;10 Suppl T:T96-t104. doi:10.4244/eijv10sta15.

196. Pack QR, Squires RW, Lopez-Jimenez F, Lichtman SW, Rodriguez-Escudero JP, Lindenauer PK et al. Participation Rates, Process Monitoring, and Quality Improvement Among Cardiac Rehabilitation Programs in the United States: A NATIONAL SURVEY. Journal of cardiopulmonary rehabilitation and prevention. 2015;35(3):173-80. doi:10.1097/hcr.000000000000108.

197. Beatty AL, Bradley SM, Maynard C, McCabe JM. Referral to Cardiac Rehabilitation After Percutaneous Coronary Intervention, Coronary Artery Bypass Surgery, and Valve Surgery: Data From the Clinical Outcomes Assessment Program. Circulation Cardiovascular quality and outcomes. 2017;10(6). doi:10.1161/circoutcomes.116.003364.

198. Ciampricotti R, el Gamal MI. Unstable angina, myocardial infarction and sudden death after an exercise stress test. International journal of cardiology. 1989;24(2):211-8.

199. Goto Y, Sumida H, Ueshima K, Adachi H, Nohara R, Itoh H. Safety and implementation of exercise testing and training after coronary stenting in patients with acute myocardial infarction. Circulation journal : official journal of the Japanese Circulation Society. 2002;66(10):930-6.

200. Roffi M, Wenaweser P, Windecker S, Mehta H, Eberli FR, Seiler C et al. Early exercise after coronary stenting is safe. Journal of the American College of Cardiology. 2003;42(9):1569-73.

201. Samuels B, Schumann J, Kiat H, Friedman J, Berman DS. Acute stent thrombosis associated with exercise testing after successful percutaneous transluminal coronary angioplasty. American heart journal. 1995;130(5):1120-2.

202. Kim HS, Kim SY, Lee UJ, Kim W. Terrible stent thrombosis induced by a treadmill test performed three days after percutaneous coronary intervention. Chonnam medical journal. 2014;50(1):23-6. doi:10.4068/cmj.2014.50.1.23.

203. Nygaard TW, Beller GA, Mentzer RM, Gibson RS, Moeller CM, Burwell LR. Acute coronary occlusion with exercise testing after initially successful coronary angioplasty for acute myocardial infarction. The American journal of cardiology. 1986;57(8):687-8.

204. Choi Y, Akazawa N, Zempo-Miyaki A, Ra SG, Shiraki H, Ajisaka R et al. Acute Effect of High-Intensity Eccentric Exercise on Vascular Endothelial Function in Young Men. Journal of strength and conditioning research. 2016;30(8):2279-85. doi:10.1519/jsc.00000000000536.

205. Chen YW, Apostolakis S, Lip GY. Exercise-induced changes in inflammatory processes: Implications for thrombogenesis in cardiovascular disease. Annals of medicine. 2014;46(7):439-55. doi:10.3109/07853890.2014.927713.

206. Atkinson CL, Carter HH, Dawson EA, Naylor LH, Thijssen DH, Green DJ. Impact of handgrip exercise intensity on brachial artery flow-mediated dilation. European journal of applied physiology. 2015;115(8):1705-13. doi:10.1007/s00421-015-3157-1.

207. Johnson BD, Mather KJ, Newcomer SC, Mickleborough TD, Wallace JP. Brachial artery flowmediated dilation following exercise with augmented oscillatory and retrograde shear rate. Cardiovascular ultrasound. 2012;10:34. doi:10.1186/1476-7120-10-34.

208. Rakobowchuk M, Tanguay S, Burgomaster KA, Howarth KR, Gibala MJ, MacDonald MJ. Sprint interval and traditional endurance training induce similar improvements in peripheral arterial stiffness

and flow-mediated dilation in healthy humans. American journal of physiology Regulatory, integrative and comparative physiology. 2008;295(1):R236-42. doi:10.1152/ajpregu.00069.2008.

209. Dawson EA, Whyte GP, Black MA, Jones H, Hopkins N, Oxborough D et al. Changes in vascular and cardiac function after prolonged strenuous exercise in humans. Journal of applied physiology (Bethesda, Md : 1985). 2008;105(5):1562-8. doi:10.1152/japplphysiol.90837.2008.

210. Llewellyn TL, Chaffin ME, Berg KE, Meendering JR. The relationship between shear rate and flowmediated dilation is altered by acute exercise. Acta physiologica (Oxford, England). 2012;205(3):394-402. doi:10.1111/j.1748-1716.2012.02417.x.

211. Bond B, Hind S, Williams CA, Barker AR. The Acute Effect of Exercise Intensity on Vascular Function in Adolescents. Medicine and science in sports and exercise. 2015;47(12):2628-35. doi:10.1249/mss.000000000000715.

212. Shenouda N, Skelly LE, Gibala MJ, MacDonald MJ. Brachial artery endothelial function is unchanged after acute sprint interval exercise in sedentary men and women. Experimental physiology. 2018. doi:10.1113/ep086677.

213. Rognmo O, Bjornstad TH, Kahrs C, Tjonna AE, Bye A, Haram PM et al. Endothelial function in highly endurance-trained men: effects of acute exercise. Journal of strength and conditioning research. 2008;22(2):535-42. doi:10.1519/JSC.0b013e31816354b1.

214. McClean C, Harris RA, Brown M, Brown JC, Davison GW. Effects of Exercise Intensity on Postexercise Endothelial Function and Oxidative Stress. Oxidative medicine and cellular longevity. 2015;2015;723679. doi:10.1155/2015/723679.

215. Dawson EA, Green DJ, Cable NT, Thijssen DH. Effects of acute exercise on flow-mediated dilatation in healthy humans. Journal of applied physiology (Bethesda, Md : 1985). 2013;115(11):1589-98. doi:10.1152/japplphysiol.00450.2013.

216. Johnson BD, Padilla J, Wallace JP. The exercise dose affects oxidative stress and brachial artery flow-mediated dilation in trained men. European journal of applied physiology. 2012;112(1):33-42. doi:10.1007/s00421-011-1946-8.

217. Michaelides AP, Soulis D, Antoniades C, Antonopoulos AS, Miliou A, Ioakeimidis N et al. Exercise duration as a determinant of vascular function and antioxidant balance in patients with coronary artery disease. Heart (British Cardiac Society). 2011;97(10):832-7. doi:10.1136/hrt.2010.209080.

218. Bailey TG, Perissiou M, Windsor M, Russell F, Golledge J, Green DJ et al. Cardiorespiratory fitness modulates the acute flow-mediated dilation response following high-intensity but not moderateintensity exercise in elderly men. Journal of applied physiology (Bethesda, Md : 1985). 2017;122(5):1238-48. doi:10.1152/japplphysiol.00935.2016.

219. Farsidfar F, Kasikcioglu E, Oflaz H, Kasikcioglu D, Meric M, Umman S. Effects of different intensities of acute exercise on flow-mediated dilatation in patients with coronary heart disease. International journal of cardiology. 2008;124(3):372-4. doi:10.1016/j.ijcard.2006.11.243.

220. Tanasescu M, Leitzmann MF, Rimm EB, Willett WC, Stampfer MJ, Hu FB. Exercise type and intensity in relation to coronary heart disease in men. Jama. 2002;288(16):1994-2000.

221. Currie KD, McKelvie RS, Macdonald MJ. Flow-mediated dilation is acutely improved after highintensity interval exercise. Medicine and science in sports and exercise. 2012;44(11):2057-64. doi:10.1249/MSS.0b013e318260ff92.

222. Currie KD, McKelvie RS, Macdonald MJ. Brachial artery endothelial responses during early recovery from an exercise bout in patients with coronary artery disease. BioMed research international. 2014;2014:591918. doi:10.1155/2014/591918.

223. McLenachan JM, Williams JK, Fish RD, Ganz P, Selwyn AP. Loss of flow-mediated endotheliumdependent dilation occurs early in the development of atherosclerosis. Circulation. 1991;84(3):1273-8.

224. Gordon JB, Ganz P, Nabel EG, Fish RD, Zebede J, Mudge GH et al. Atherosclerosis influences the vasomotor response of epicardial coronary arteries to exercise. The Journal of clinical investigation. 1989;83(6):1946-52. doi:10.1172/jci114103.

225. Boos CJ, Balakrishnan B, Lip GY. The effects of exercise stress testing on soluble E-selectin, von Willebrand factor, and circulating endothelial cells as indices of endothelial damage/dysfunction. Annals of medicine. 2008;40(1):66-73. doi:10.1080/07853890701652833.

226. Hays AG, Stuber M, Hirsch GA, Yu J, Schar M, Weiss RG et al. Non-invasive detection of coronary endothelial response to sequential handgrip exercise in coronary artery disease patients and healthy adults. PloS one. 2013;8(3):e58047. doi:10.1371/journal.pone.0058047.

227. Hays AG, Iantorno M, Soleimanifard S, Steinberg A, Schar M, Gerstenblith G et al. Coronary vasomotor responses to isometric handgrip exercise are primarily mediated by nitric oxide: a noninvasive MRI test of coronary endothelial function. American journal of physiology Heart and circulatory physiology. 2015;308(11):H1343-50. doi:10.1152/ajpheart.00023.2015.

228. Togni M, Windecker S, Cocchia R, Wenaweser P, Cook S, Billinger M et al. Sirolimus-eluting stents associated with paradoxic coronary vasoconstriction. Journal of the American College of Cardiology. 2005;46(2):231-6. doi:10.1016/j.jacc.2005.01.062.

229. Puricel S, Kallinikou Z, Espinola J, Arroyo D, Goy JJ, Stauffer JC et al. Comparison of endotheliumdependent and -independent vasomotor response after abluminal biodegradable polymer biolimuseluting stent and persistent polymer everolimus-eluting stent implantation (COMPARE-IT). International journal of cardiology. 2016;202:525-31. doi:10.1016/j.ijcard.2015.09.085.

230. Rummens JL, Daniels A, Dendale P, Hensen K, Hendrikx M, Berger J et al. Suppressed increase in blood endothelial progenitor cell content as result of single exhaustive exercise bout in male revascularised coronary artery disease patients. Acta clinica Belgica. 2012;67(4):262-9. doi:10.2143/acb.67.4.2062670.

231. Kazmierski M, Wojakowski W, Michalewska-Wludarczyk A, Podolecka E, Kotowski M, Machalinski B et al. Exercise-induced mobilisation of endothelial progenitor cells in patients with premature coronary heart disease. Kardiologia polska. 2015;73(6):411-8. doi:10.5603/KP.a2014.0248.

232. Wang JS, Jen CJ, Kung HC, Lin LJ, Hsiue TR, Chen HI. Different effects of strenuous exercise and moderate exercise on platelet function in men. Circulation. 1994;90(6):2877-85.

233. Ikarugi H, Shibata M, Shibata S, Ishii H, Taka T, Yamamoto J. High intensity exercise enhances platelet reactivity to shear stress and coagulation during and after exercise. Pathophysiology of haemostasis and thrombosis. 2003;33(3):127-33. doi:77820.

234. Ikarugi H, Yamamoto J. The exercise paradox may be solved by measuring the overall thrombotic state using native blood. Drug discoveries & therapeutics. 2017;11(1):15-9. doi:10.5582/ddt.2016.01077.

235. Posthuma JJ, van der Meijden PE, Ten Cate H, Spronk HM. Short- and Long-term exercise induced alterations in haemostasis: a review of the literature. Blood reviews. 2015;29(3):171-8. doi:10.1016/j.blre.2014.10.005.

236. Cadroy Y, Pillard F, Sakariassen KS, Thalamas C, Boneu B, Riviere D. Strenuous but not moderate exercise increases the thrombotic tendency in healthy sedentary male volunteers. Journal of applied physiology (Bethesda, Md : 1985). 2002;93(3):829-33. doi:10.1152/japplphysiol.00206.2002.

237. Chen YW, Chen JK, Wang JS. Strenuous exercise promotes shear-induced thrombin generation by increasing the shedding of procoagulant microparticles from platelets. Thrombosis and haemostasis. 2010;104(2):293-301. doi:10.1160/th09-09-0633.

238. Petidis K, Douma S, Doumas M, Basagiannis I, Vogiatzis K, Zamboulis C. The interaction of vasoactive substances during exercise modulates platelet aggregation in hypertension and coronary artery disease. BMC cardiovascular disorders. 2008;8:11. doi:10.1186/1471-2261-8-11.

239. Li N, Wallen NH, Hjemdahl P. Evidence for prothrombotic effects of exercise and limited protection by aspirin. Circulation. 1999;100(13):1374-9.

240. Kobusiak-Prokopowicz M, Kuliczkowski W, Karolko B, Prajs I, Mazurek W. Platelet aggregation and P-selectin levels during exercise treadmill test in patients with ischaemic heart disease. Kardiologia polska. 2006;64(10):1094-100; discussion 101. 241. Tokuue J, Hayashi J, Hata Y, Nakahara K, Ikeda Y. Enhanced platelet aggregability under high shear stress after treadmill exercise in patients with effort angina. Thrombosis and haemostasis. 1996;75(5):833-7.

242. Wang JS, Liao CH. Moderate-intensity exercise suppresses platelet activation and polymorphonuclear leukocyte interaction with surface-adherent platelets under shear flow in men. Thrombosis and haemostasis. 2004;91(3):587-94. doi:10.1160/th03-10-0644.

243. Levine GN, Bates ER, Bittl JA, Brindis RG, Fihn SD, Fleisher LA et al. 2016 ACC/AHA guideline focused update on duration of dual antiplatelet therapy in patients with coronary artery disease: A report of the American College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines. The Journal of thoracic and cardiovascular surgery. 2016;152(5):1243-75. doi:10.1016/j.jtcvs.2016.07.044.

244. Mehta SR, Yusuf S, Peters RJ, Bertrand ME, Lewis BS, Natarajan MK et al. Effects of pretreatment with clopidogrel and aspirin followed by long-term therapy in patients undergoing percutaneous coronary intervention: the PCI-CURE study. Lancet (London, England). 2001;358(9281):527-33.

245. Kop WJ, Weissman NJ, Zhu J, Bonsall RW, Doyle M, Stretch MR et al. Effects of acute mental stress and exercise on inflammatory markers in patients with coronary artery disease and healthy controls. The American journal of cardiology. 2008;101(6):767-73. doi:10.1016/j.amjcard.2007.11.006.

246. Lara Fernandes J, Serrano CV, Jr., Toledo F, Hunziker MF, Zamperini A, Teo FH et al. Acute and chronic effects of exercise on inflammatory markers and B-type natriuretic peptide in patients with coronary artery disease. Clinical research in cardiology : official journal of the German Cardiac Society. 2011;100(1):77-84. doi:10.1007/s00392-010-0215-x.

247. Lominadze D, Dean WL, Tyagi SC, Roberts AM. Mechanisms of fibrinogen-induced microvascular dysfunction during cardiovascular disease. Acta physiologica (Oxford, England). 2010;198(1):1-13. doi:10.1111/j.1748-1716.2009.02037.x.

248. Haynes A, Linden MD, Robey E, Naylor LH, Cox KL, Lautenschlager NT et al. Relationship between monocyte-platelet aggregation and endothelial function in middle-aged and elderly adults. Physiological reports. 2017;5(10). doi:10.14814/phy2.13189.

249. Guarnieri C, Melandri G, Caldarera I, Cervi V, Semprini F, Branzi A. Spontaneous superoxide generation by polymorphonuclear leukocytes isolated from patients with stable angina after physical exercise. International journal of cardiology. 1992;37(3):301-7.

250. Tozzi-Ciancarelli MG, Penco M, Di Massimo C. Influence of acute exercise on human platelet responsiveness: possible involvement of exercise-induced oxidative stress. European journal of applied physiology. 2002;86(3):266-72.

251. Silvestro A, Scopacasa F, Oliva G, de Cristofaro T, Iuliano L, Brevetti G. Vitamin C prevents endothelial dysfunction induced by acute exercise in patients with intermittent claudication. Atherosclerosis. 2002;165(2):277-83.

252. Onogi F, Saitoh S, Aikawa K, Ishibashi T, Maruyama Y. Antioxidant is a useful supportive agent for the treatment of coronary vasospasm with endothelial dysfunction in pig. Coronary artery disease. 2007;18(2):133-40. doi:10.1097/MCA.0b013e328010a48b.

253. Aikawa K, Saitoh S, Muto M, Osugi T, Matsumoto K, Onogi F et al. Effects of antioxidants on coronary microvascular spasm induced by epicardial coronary artery endothelial injury in pigs. Coronary artery disease. 2004;15(1):21-30.

254. Lyamina NP, Razborov IB, Karpov ES. [Clinical and Economic Aspects of Meldonium as Part of Physical Rehabilitation Programs in Patients With Coronary Heart Disease After Percutaneous Coronary Interventions]. Kardiologiia. 2016;56(8):13-8.

255. Cook CM, Ahmad Y, Howard JP, Shun-Shin MJ, Sethi A, Clesham GJ et al. Impact of Percutaneous Revascularization on Exercise Hemodynamics in Patients With Stable Coronary Disease. Journal of the American College of Cardiology. 2018;72(9):970-83. doi:10.1016/j.jacc.2018.06.033.