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1 Title

Landscape resistance affects individual habitat selection but not genetic relatedness in a reintroduced desert
 ungulate

4 Abstract

5 The long-term success of species reintroductions is strongly dependent on the availability of large areas of suitable habitat and the genetic make-up of the population. If available habitat is poorly connected this can 6 7 hinder gene flow and lead to genetic fragmentation of the population, potentially increasing its extinction risk. 8 We employed a conservation genomics approach in which we combined analyses of genetic structure with 9 testing for potential landscape effects on habitat selection and gene flow in reintroduced Asiatic wild ass Equus 10 hemionus ssp. in the Israeli Negev desert. Genetic structure and pairwise relatedness were first investigated 11 followed by examination of landscape effects on individual habitat selection using records of GPS collared 12 individuals. We then built habitat resistance surfaces and used electrical circuit theory to test for landscape 13 effects on genetic relatedness. We detected weak genetic structuring, yet low spatial coherence among 14 individuals from the same genetic cluster. Landscape variables had a significant impact on individual habitat 15 selection, with wild ass avoiding steep slopes and habitats of low suitability as predicted by a species distribution model. However, the landscape genetic analysis revealed no effect of habitat resistance on genetic 16 17 relatedness. These results suggest that gene flow in the reintroduced population is not impacted by landscape resistance. Indeed, the high mobility of the species may increase its resistance to the genetic effects of habitat 18 19 fragmentation, at least over a small number of generations. We discuss other potential causes for the observed

20 genetic structure including a behavioural effect. Our study highlights the importance of understanding species-

21 habitat interactions for the long-term success of reintroductions.

22 Keywords

23 Landscape resistance, habitat selection, genetic structure, reintroduction, *Equus hemionus*, circuit theory

1. Introduction

25 Reintroductions are one of the most powerful conservation tools for reinforcing and re-establishing populations 26 of threatened species, but success rates are often low. The most important determinants of the long-term success 27 of a reintroduction are i) the availability of large areas of suitable habitat and ii) the genetic makeup of the 28 reintroduced population (Wolf et al. 1998; Armstrong & Seddon 2008; Baguette et al. 2013). Genetic makeup is 29 important since many reintroductions are based on a small number of founders. The resulting small population 30 size during the early stages of the reintroduction can lead to increased genetic drift and inbreeding, causing the 31 loss of genetic diversity and adaptive flexibility in the established population (Frankham et al. 2002; Templeton 32 2017). These negative effects are further exacerbated if the reintroduced population is fragmented. Resulting 33 genetic isolation of subpopulations can make these population fragments more vulnerable to extinction due to 34 inbreeding and stochastic genetic and demographic processes (Saccheri et al. 1998; Bozzuto et al. 2019).

35 Large connected areas of suitable habitat are also crucial to facilitate sufficient demographic growth of the

36 reintroduced population (Armstrong & Seddon 2008). In contrast, habitats with low functional connectivity

37 (whereby the landscape impedes individual movement) can hinder range expansion and prevent reintroduced

populations from successfully colonising the available habitat (Templeton et al. 2011; Neuwald & Templeton

2013; Ziółkowska et al. 2016). Furthermore, low connectivity can also limit gene flow between occupied
 patches, resulting in spatial sub-structuring of the population (Manel et al. 2003; Bergl & Vigilant 2007). This

40 patches, resulting in spatial sub-structuring of the population (Waller et al. 2005, Bergi & Vignant 2007). This 41 may explain observations of within-population genetic structure in reintroduced populations, with genetic

42 clusters centring around release sites (Howell et al. 2016; Grauer et al. 2017; Moraes et al. 2017). In order to

43 avoid the problem of genetic isolation, individuals must be able to disperse between occupied patches into new

44 suitable territory at a rate that facilitates sufficient gene flow (Mills & Allendorf 1996).

Gene flow is limited by factors restricting individual dispersal, here defined as the movement between habitat patches or subpopulations (Benton & Bowler 2012). In terrestrial mammals, dispersal ability is usually affected by landscape structure, climatic and anthropogenic factors, or specific combinations of these (Howell et al.

48 2016). Major landscape features (e.g., roads, mountain ridges) may act as physical barriers completely

- 49 preventing movement across them, but areas of less preferred habitat may also reduce gene flow (Storfer et al.
- 50 2007). For example, in female-philopatric mountain goats, male habitat selection best predicted gene flow and
- 51 relatedness across the landscape (Shafer et al. 2012). However, for many reintroduced populations, information
- 52 on habitat use and preference is limited, especially when the species has been absent from the area for a long
- 53 time or when it is replaced by a closely-related group (e.g., a different subspecies) which makes prediction of
- 54 resource use and dispersal more difficult (Seddon & Soorae 1999). Therefore, directly assessing habitat
- 55 connectivity and gene flow and the factors impacting them is an important measure to optimise population
- 56 management to enhance chances of long-term population persistence.
- 57 The Asiatic wild ass *Equus hemionus ssp.* (Pallas, 1775) reintroduced to Israel, presents an ideal opportunity for
- 58 furthering our understanding of the environmental effects on the dispersal and genetic structure in small,
- 59 reintroduced populations. Equus hemionus ssp. was reintroduced in Israel after the local subspecies (Syrian wild
- 60 ass E.h.hemippus) became extinct (Saltz & Rubenstein 1995). A captive breeding facility was established by the 61 Israeli Nature and Parks Authority (INPA) in 1968 from individuals of two subspecies; Iranian onager
- 62 E.h.onager and Turkmen kulan E.h.kulan (Saltz et al. 2000). From this breeding facility, 38 individuals were
- released into the Negev desert between 1982 and 1993 at two release sites (Fig. 1) (Saltz & Rubenstein 1995). 63
- 64 The population has since expanded its spatial distribution across the highly heterogenous landscape and is
- 65 currently estimated at >250 individuals (Gueta et al. 2014; Renan et al. 2018).
- 66 Asiatic wild as have a highly flexible fission-fusion social structure and a resource defence polygyny mating 67 system (Boyd et al. 2016; Renan et al. 2018). Dominant males defend territories near permanent water sources, 68 while females form unstable social groups with individual associations based on reproductive status rather than relatedness (Rubenstein 1994; Saltz et al. 2000; Wallach et al. 2007; Altman 2016). Previous analyses based on 69 70 mitochondrial DNA haplotypes and nuclear microsatellite markers identified a weak spatial genetic structure in 71 the established population (Gueta et al. 2014; Renan 2014). The authors suggested a combined effect of range
- 72 expansion and low habitat connectivity between colonised areas to be the underlying cause (Gueta et al. 2014). 73 This possibility is supported by previous studies which identified resource distribution and topography as the
- 74 main predictors for wild ass presence and pathway usage (Davidson et al. 2013; Nezer et al. 2017). In the
- 75 Negev, patches of suitable habitat appear to be separated by areas of low resource availability and challenging
- 76 topography such as steep cliffs and canyons that could act as barriers to wild ass movement, hence limiting gene
- 77 flow between patches. Since the recently established population in Israel is geographically isolated with no
- 78 opportunity for external migrants from neighbouring countries, it is particularly vulnerable to the negative
- 79 effects of genetic drift (Frankham et al. 2002). Further spatial subdivision would be a severe threat to this
- 80 recently established population and could jeopardise the long-term success of the reintroduction (With & King
- 81 1999; Wang et al. 2017; Pelletier et al. 2019).
- 82 The aim of the present study was to investigate potential landscape effects that may cause genetic structuring of
- 83 the reintroduced population. First, we assessed genetic clustering of the population using a panel of 1645
- 84 genome-wide single nucleotide polymorphisms (SNPs). Then, we analysed individual GPS collar data and
- 85 investigated habitat selection with respect to slope and habitat suitability based on a species distribution model
- 86 (SDM). Finally, we created landscape resistance surfaces from habitat selection data and applied electrical
- 87 circuit theory to test for an effect of habitat resistance on genetic relatedness. Based on wild ass ecology and
- 88 previous studies of the population, we predict: i) the population in Israel is genetically structured into spatially
- distinct clusters ii) individuals avoid areas of low habitat suitability (based on the SDM) and steep slope, as 89 90 reported for wild ass in other populations (e.g. Sharma et al., 2004), and iii) steep cliffs form a complete barrier
- 91 to wild ass movement and hence we predict a stronger effect of slope-based landscape resistance than
- 92 suitability-based landscape resistance on genetic relatedness in the population.

2. Materials and Methods 93

94 2.1 Study site

95 The Negev is a hyper-arid desert that extends throughout Southern Israel. The landscape is defined by a steep 96 gradient in elevation ranging from the Negev Highlands in the Northwest (>1000m above sea level) decreasing

- 97 towards the Arava valley and the Dead Sea in the East (<300m below sea level) (Stern et al. 1986). This
- 98 elevation gradient coincides with a gradient in mean maximum annual temperature and precipitation, ranging
- 99 from 22.6°C and 150mm in the Negev Highlands to 31.1°C and 30mm in the hotter and more arid Arava (Israel 100

- 101 Negev highlands giving way to sand and desert savannoid vegetation types in the Arava (Danin 1999). The
- 102 topography of the Negev is complex and characterised by steep cliffs and levelled floodplains. Vegetation is
- 103 mostly limited to ephemeral streambeds and floodplains. Permanent water sources are scarce, however, flash
- 104 floods occurring after heavy rainfall in the winter fill up natural rock pools which retain water for several
- 105 months (Nezer et al. 2017). In addition, there are three artificial water sources which are maintained throughout
- the year by the INPA to provide wildlife with water, which have also become activity centres of the wild ass
- 107 population (Gueta et al. 2014; Nezer et al. 2017).

108 2.2 DNA sample collection and sequencing

109 DNA samples were collected opportunistically by rangers and veterinarians of the INPA across seasons, 110 between 2010 and 2017. Blood or tissue samples were taken from individuals that were killed in road traffic 111 accidents, from injured individuals receiving veterinary treatment or during the fitting of global positioning system (GPS) collars. Precise geographical locations were available for all samples (Fig. 1). Whole blood 112 113 samples were stored in EDTA tubes (not heparinized; BD Vacutainer K2E 18.0mg, Vacuette K3E 3mg), tissue 114 samples were either stored in 70% ethanol or untreated in paper envelopes. All blood and tissue samples were 115 stored frozen (at -20°C or -80°C). We purified DNA from samples using commercial silica spin column-based extraction kits (QIAGEN DNeasy Blood and Tissue Kit, Thermo Scientific GeneJET Genomic DNA 116 117 Purification Kit), following manufacturers protocol. We sequenced samples using double digest restriction-site 118 associated DNA marker sequencing (ddRADseq) methods and the high-fidelity versions of the enzymes EcoRI 119 and SbfI (R3101S and R3642L, respectively; New England Biolabs). We prepared libraries following a protocol adapted from Peterson et al. (2012) and sequenced them on a single flow cell lane of an Illumina HiSeq 4000 120 121 system. Over 400 million raw paired-end sequence reads were produced with a mean read length of 300bp. We 122 assessed the quality of raw reads using the FastQC tool (Andrews 2010). A mean Phred+33 quality score >30 was recorded for all bases. We processed raw sequences in the STACKS 2.0 pipeline (Catchen et al. 2013) and 123 124 assembled loci de novo using the denovo_map wrapper program in STACKS and identified optimal parameter 125 settings using an approach adapted from Paris et al. (2017) and SNP error rates, calculated using seven replicate 126 pairs of individuals included in the data set. To avoid linkage between markers we retained only the first SNP on 127 a locus using the --write-single-snp function in the population program in STACKS. Subsequently, we filtered 128 called SNPs in 3 steps in the vcftools programme (Danecek et al. 2011) using site and individual filtering 129 options (minimum mean individual coverage \geq 35x, minor allele count \geq 3, SNPs present in minimum of 80% of 130 individuals). Finally, SNPs that deviated from Hardy-Weinberg equilibrium as defined by p-value threshold

131 > 0.05 were removed.

132 2.3 Genetic structure analysis

133 Initially, we explored the data using Principal Component Analysis (PCA), which fits orthogonal Principal

- Components (PCs) that summarise overall variability between individuals. Subsequently, we investigated
 genetic structure in more detail using two different approaches: A discriminant analysis of principal components
 (DAPC; Jombart, 2008) and a Bayesian cluster analysis implemented in the program STRUCTURE (Pritchard et
 al. 2000).
- 138 DAPC is a multivariate approach which performs a PCA in a first step and then subjects the PC scores to a 139 discriminant function analysis (DFA). Unlike PCA, DFA fits orthogonal discriminant functions that maximise 140 between group relative to within-group variation. Therefore, it is suited to differentiating between genetic groups (Jombart et al. 2010). A K-means clustering approach can be applied to assess the number and 141 142 composition of genetic clusters (K) in the data. The best supported model is identified using the Bayesian 143 Information Criterion (BIC), where the lowest BIC, which is often indicated by an elbow in the curve, is 144 preferred. We performed PCA and DAPC in the 'adegenet' package (Jombart 2008) in R (R 3.5.3, R core team 145 2020). In both analyses we retained the first 10 PCs, which explained 54.96% of the total variance.
- 146We ran the program STRUCTURE with the admixture model and correlated allele frequencies, for K=1-10, with14710 repetitions for each K. The runs were performed with 1×10^6 iterations of the Markov Chain Monte Carlo148(MCMC) chain preceded by 1×10^5 burn-in iterations. We assessed STRUCTURE outputs for the optimal value of149K using the log likelihood (Pritchard et al. 2000) and the Evanno method (Evanno et al. 2005) in the web-based150version of STRUCTURE HARVESTER (Earl & vonHoldt 2012). Pritchard et al. (2000) suggest that a value of K
- 151 which maximizes the model log likelihood Ln(PD) is optimal. However, Ln(PD) often plateaus or continues to
- increase after reaching the optimal K-value and so Evanno et al. (2005) proposed an improved method to
- estimating optimal K, based on the second order rate of change of the likelihood function. We produced ancestry

- 154 bar plots in STRUCTURE HARVESTER. As GPS data from collared individuals suggested fidelity to a smaller range
- during the breeding season (Supplementary material, Fig. A1), population genetic differentiation may be easier 155
- 156 to detect at this time. We therefore repeated our analyses on the 15 individuals sampled during the breeding
- 157 season.

158 2.4 Individual habitat selection

159 To investigate a potential effect of landscape characteristics on gene flow we used individual movement as a

- 160 proxy for gene flow and investigated factors that restrict individual movement using high resolution movement
- data from GPS collars. Previous studies have highlighted two habitat characteristics impacting wild ass 161
- distribution: Species-specific habitat suitability and topography (Davidson et al. 2013; Gueta et al. 2014; Nezer 162 163
- et al. 2017). To verify the impact of these characteristics on wild ass movement, we first performed a 164 compositional analysis of habitat selection (Aebischer et al. 1993). The analysis compares the relative
- abundance of a specific habitat type with its relative use by individuals. This way, habitat types that are avoided 165
- 166 by individuals and potentially represent barrier to movement can be identified.
- 167 We investigated habitat selection with respect to habitat suitability, based on a previously developed species distribution model (SDM, Nezer et al., 2017). The model, which covered most of the area of the present study 168
- 169 (Fig.1), was created using indirect observations and a data set of nine habitat variables from different categories
- 170 relevant to wild ass biology (water, topography, biotic conditions, climate, anthropogenic disturbance). Since
- 171 the model was based on faecal mount surveys rather than direct observations of wild ass, we tested the
- 172 predictive power of the model using the high resolutions GPS-collar data. We used the model output, a
- 173 probabilistic distribution map that represents the probability of wild ass distribution in the area with values
- 174 ranging from 0 (low probability) to 99 (high probability), as an indicator for habitat suitability (habitat 175 suitability index). Specifically, low probability values indicate low habitat suitability and high probability values
- 176 indicate high habitat suitability. Since the SDM from which habitat suitability was derived did not cover the
- 177 entire study area, the analysis was restricted to the part of the study area covered by the SDM. The SDM did not
- 178 consider seasonal differences in habitat suitability, and potential seasonal patterns in wild ass natal dispersal are
- 179 unknown. Hence, the analysis did not account for seasonality.
- 180 Previous studies have reported topography as one of the most important physical constraints to wild ass
- 181 movement, with steep slopes (> 30°) being avoided entirely (Sharma et al. 2004; Henley et al. 2007; Davidson
- 182 et al. 2013; Nezer et al. 2017). Therefore, we decided to also investigate habitat selection with respect to
- topography as a habitat measure directly linked to movement ability. The same slope layer from the SDM was 183 used, which was generated from a contour dataset retrieved from the Survey of Israel (MAPI; for further details
- 184
- 185 see Nezer et al. 2017).
- 186 To investigate individual habitat selection with respect to habitat suitability and slope, we used movement
- 187 records from 7 GPS collared individuals. Between 2012 and 2013, five individuals (4 males, 1 female) of the
- 188 reintroduced population were fitted with GPS collars (African Wildlife Tracking company) (Giotto et al. 2015)
- 189 and an additional 2 females were collared in 2015. Collars were set to record the location every hour and
- animals wore collars between 10-25 months resulting in a minimum of 2937 records per individual within the 190
- 191 reduced study area (Supplementary material, Table A1).
- 192 Slope and habitat suitability raster layers had a resolution of 100m and we extracted the grid cell values for each
- 193 GPS record from the collared individuals using the extract values to points function in ARCGIS (ESRI 2011).
- 194 Subsequently, we divided extracted data for each variable into categories, to investigate proportional habitat use.
- 195 For habitat suitability index we divided the range from 0-99 equally, rendering a low (0-33), intermediate (34-
- 196 66) and high (67–99) suitability category. Based on previous studies (Sharma et al. 2004; Davidson et al. 2013)
- 197 we set a threshold for steep slopes at 30 degrees and divided the slope into low $(0-15^\circ)$, intermediate $(16-30^\circ)$
- 198 and steep slope, containing all records $>30^{\circ}$.
- 199 We performed a compositional analysis of habitat selection on the defined habitat categories using the compana 200 function in the 'adehabitatHS' package in R (Calenge 2006). The analysis first tests for significance of habitat
- 201 selection using Wilks lambda and subsequently produces a ranking matrix indicating whether a specific habitat
- 202 type is used significantly more or less than another. P-values were estimated by randomisation tests (999
- 203 permutations of the data). Aebischer et al. (1993) recommend using a minimum of 6 individuals, therefore, we
- 204 pooled males and females for the analysis. We analysed habitat use relative to habitat availability within the
- 205 entire habitat area, corresponding to third order selection as described by Johnson et al. (1980). We did not limit

206 the analysis to habitat available within an individual's home range, since gene flow is mediated by long-distance

207 dispersal movements extending beyond home range boundaries. Finally, compositional analysis assumes no

208 individual variation in habitat selection, and we tested this assumption by eigenanalysis of selection ratios with 209 the eisra function.

210 2.5 Landscape genetic analysis

211 2.5.1 Resistance surfaces

212 After assessing the impact of habitat characteristics on individual movement, we created habitat resistance 213 surfaces. This approach assigns resistance values to each cell in a habitat grid, reflecting the relative cost inflicted on an individual moving through it (Spear et al. 2010). We created three habitat resistance surfaces 214 based on habitat suitability index, slope, and geographic distance. First, we inverted the habitat suitability map 215 216 using the raster calculator in ARCGIS. To grid cells with a probability of 0 we assigned a marginally positive 217 value 0.01 to comply with input requirements of downstream analysis. The resulting habitat resistance map 218 based on habitat suitability ranged from 0.01 (low resistance) to 99 (high resistance). We parameterised the 219 second resistance surface based on slope, so that grid cells with a slope of $1^{\circ}-30^{\circ}$ were assigned a resistance 220 value of 1–30 respectively. We set a threshold by assigning grid cells with a slope $>30^{\circ}$ a resistance value of 99. 221 Grid cells with a slope of 0° we assigned a resistance of 0.01. Additionally, we created a control resistance 222 surface based solely on geographic distance, by assigning all grid cells of the resistance surface a value of 1. All resistance surfaces had a spatial resolution of 100m and were produced with ARCGIS (Fig. 2). 223

224 2.5.2 Pairwise distances

225 Since the landscape genetic analysis was restricted to the part of the study area covered by the SDM, we 226 excluded three individuals which fell outside the SDM area from the analysis (Fig. 1). We used the programme 227 CIRCUITSCAPE (version 4.0, McRae et al. 2013) to calculate pairwise resistance distances for the remaining 27 228 individuals for the three resistance surfaces. Circuitscape applies algorithms from electronic circuit theory to 229 estimate resistances to current flow between nodes. The program was run in pairwise mode with individuals set 230 as nodes, connected to all eight neighbouring cells surrounding a node. Pairwise genetic distance was expressed 231 through a relatedness coefficient, which is effectively a measure of the genetic distance between two 232 individuals. We estimated pairwise relatedness coefficients in the 'related' R package (Pew et al. 2015) using the 233 corrected Wang (2002) estimator, which has been shown to achieve high accuracy with small sample sizes 234 (Wang, 2017).

235 2.5.3 Distance-based redundancy analysis

To test for a potential relationship between habitat resistance distance and genetic distance we performed a 236 237 distance-based redundancy analysis (dbRDA) using the *capsscale* function in the 'vegan' R package (Oksanen et 238 al. 2010). dbRDA is an extension of multivariate regression which accepts distance matrices as response 239 variables. The response matrix is transformed into synthetic variables which are then regressed on multiple 240 explanatory variables (Legendre et al. 1999; Buttigieg & Ramette 2014). First, we transformed the pairwise 241 habitat resistance matrices to generate one-dimensional explanatory variables for the dbRDA. For this purpose, 242 we performed principal coordinate analyses using the pcoa function in the 'ape' R package (Paradis & Schliep 243 2018) with a Lingoes correction for negative eigenvalues to preserve all variation of the landscape resistance 244matrices. Subsequently, we used a Broken Stick model to estimate the number of significant principal 245 coordinates (PCos) (MacArthur, 1957; Supplementary material, Fig. A2). For all three resistance variables only 246 the first or first and second PCos explained more variation than expected under the Broken Stick model. 247 However, since this accounted for only ~35% of variation in each variable, we repeated the analysis with the

248 first 4 PCos retained which accounted for >50% of variation (Supplementary material, Table A2).

249 We tested a total of seven models, once with the first 4 PCos and once with only the first PCo retained (Table

1). We ran three models that tested for landscape resistance effects on gene flow by setting the pairwise

relatedness matrix as the response variable and one of the three transformed habitat resistance matrices (based

on either geographic distance, slope, SDM) as explanatory variables. Additionally, we tested four partial models

that controlled for an effect of geographic distance on habitat resistance and the reciprocal. We tested for

significance with permutation tests using the *anova.cca* function with 9999 permutations. Since GPS data from

collared individuals suggested fidelity to a smaller range during the breeding season (Supplementary material,

- Fig. A1), a potential landscape genetic effect may be easier to detect during the breeding season. Hence, we
- repeated our analyses on the 14 individuals sampled during the breeding season.

3. Results

259 3.1 Sequencing and summary statistics

260Illumina sequencing produced a total of 803,092,446 raw sequence reads. The de novo assembly with optimal261parameter settings (m3N0M4n4) produced 2,639 polymorphic loci with an average of 2.27 SNPs per locus and a262mean(\pm SD) SNP error rate of 1.08 \pm 0.31%. After SNP filtering the final data set contained 1496 SNPs and 30263individuals. Initial population genetic analysis revealed no significant difference between the mean (\pm SD)264expected heterozygosity (He=0.344 \pm 0.128) and observed heterozygosity (Ho=0.345 \pm 0.144; Paired Student's t-265test: t(1495)=-0.440, p-value=0.660) of the population. The inbreeding coefficient indicated no population level266inbreeding (Fis=-0.002).

267 3.2 Genetic structure analyses

268 The variation explained by the first two principle components of the PCA was low (PC1 9.86%, PC2 7.65%) 269 and no prominent genetic clusters could be identified (Fig. 3a). Also, the BIC plot of the DAPC indicated K=1 as optimal (Fig. 3c). This suggested no meaningful genetic clustering in the population. In contrast, for the 270 271 STRUCTURE analysis, the Evanno method identified a clear peak of delta(K) for K=4 (delta(K)=57.07; Fig. 3d). 272 However, the Evanno method cannot identify an optimum of K=1 and may indicate peaks at higher values of K 273 even in the absence of any genetic structure (Evanno et al. 2005). The mean Ln P(D) across different values of 274 K remains consistent with no distinct maximum value or plateau (Fig. 3d), suggesting that there may be only a 275 very weak signal of genetic structure. The STRUCTURE ancestry plot highlights 4 clusters with high admixture

276 levels in some individuals (Fig. 4b).

277 Since the two approaches gave slightly different results, we assessed their robustness by comparing the

278 individual assignments to the four clusters between the multivariate and the Bayesian approach. Based on the 279 results of the Evanno method, we ran the DAPC with predefined K=4. Three out of the four described clusters

were differentiated along the first PC while the fourth cluster was differentiated more strongly by the second PC

281 (Supplementary material, Fig. A3). Subsequently, we compared individual assignments from DAPC and

282 STRUCTURE. In the DAPC analysis all individuals had assignment probabilities of 1, whereas in STRUCTURE, 12

 $283 \qquad \text{individuals could not be assigned clearly to a single ancestral population (q-values < 0.7) and these individuals}$

were excluded from the comparison. Of the 18 remaining individuals, 16 clustered together in groups consistent

- between STRUCTURE and DAPC analyses (Fig. 4). However, these clusters were geographically dispersed. Six
- individuals assigned to the same cluster were located in close proximity near an artificial water source (Fig. 4).
 However, most individuals were dispersed across the study area with no clear spatial segregation between
- 287 However, most mutviduals were dispersed across the study area with no clear spatial segregation between 288 genetic clusters. Repeating the genetic structure analysis using only samples collected during the breeding
- season did not impact these findings (Supplementary material, B1).

290 3.3 Individual habitat selection

291 Compositional analysis of habitat categories revealed that individual habitat selection differed significantly from

random with respect to habitat suitability index (Lambda=0.013, p=0.012, by randomisation) and slope

293 (Lambda=0.064, p=0.021, by randomisation). The ranking matrix highlighted a clear preference for low slope

and high suitability habitats (Supplementary material, Table A3). Wild ass used more low slope and more

- intermediate and high suitability habitat than proportionally available (Fig. 5). The analysis using GPS-collar
- 296 data therefore indicated that both habitat suitability index and slope are relevant variables affecting habitat 297 selection in wild ass. Eigenanalysis of selection ratios indicated no difference in habitat selection between
- 298 individuals (Supplementary material, Fig. A4).

299 3.4 Landscape genetic analysis

300 None of the tested models of the distance-based redundancy analysis returned significant results and the

301 explained variance was close to zero for all predictor variables (Table 1). Habitat suitability and slope explained

302 negligible variation in genetic relatedness between individuals of the population. This was also true for models

303 controlling for geographic distance and resistance distances, respectively (Table 1). The results were consistent

- across models that retained only the first PCo or the first 4 PCos of the explanatory variables, hence, the models
- appear to be insensitive to these minor variations, indicating robustness of the results. Repeating the landscape
- 306 genetic analysis using only samples collected during the breeding season did not affect the results
- 307 (Supplementary material, B2).

308 4. Discussion

309 The analysis revealed some genetic structuring in the reintroduced population of wild ass in Israel. However,

310 inconsistencies in the optimal number of clusters and individual assignment between the different methods

- highlight that the genetic differentiation is weak. These results are consistent with a previous study on the same
- 312 population using lower resolution genetic data (eight microsatellite markers) which demonstrated weak yet
- significant genetic differentiation between four a priori defined subpopulations (Renan 2014). Taken together,
 these results suggest a weak genetic structure within the reintroduced population. Nevertheless, our new
- analyses of landscape resistance to individual movement does not support this as being a cause. The analysis of
- 316 GPS data showed that landscape resistance affected wild ass habitat selection, with individuals clearly avoiding
- 317 low suitability habitats and steep slopes. However, the landscape genetic analysis gave no support for an effect
- 318 of landscape resistance on genetic relatedness.

319 The present study demonstrates that the Asiatic wild ass clearly avoid certain habitats, yet functional

- 320 connectivity across the study area appears to be retained. Although large proportions of the habitat have low
- 321 suitability, these are interwoven by a network of low resistance paths, which likely facilitate individual
- movement across the study area (Fig. 2). In contrast to our expectations, habitat resistance based on slope was
- found to have no negative association with relatedness. Slopes above 30° account for only a very small
- 324 proportion of the habitat in the Negev, nonetheless, they occur in the form of steep cliffs extending over large
- 325 geographical areas and are expected to form true barriers to wild ass movement. However, wild ass are large-
- bodied, highly mobile mammals which have been reported to range long distances, and it is likely that even if
- 327 individuals are unable to climb these cliffs, they can circumvent them (Nandintsetseg et al. 2016; Nezer et al.
- 2017). In fact, the GPS data showed some long-distance movements by two females, which support the findings
 that even high resistance habitat does not prevent movement across the landscape in the Negev population
- (Supplementary material, Fig. A1). Therefore, despite being important for individual habitat selection, it
- currently appears that slope and habitat suitability have no to little effect on gene flow. These are promising
- 332 findings for the reintroduced population of wild ass in Israel and potentially for other equid populations in
- 333 heterogenous habitats.

334 The results contradict our expectations and underlines the importance of testing believed landscape barriers to 335 gene flow, as assumptions based on movement behaviour or habitat selection may be misleading. Similarly, 336 other studies have reported differential effects of roads on gene flow, even in cooccurring mammals of similar size and mobility (Frantz et al. 2012). In this study we investigated generic movement from GPS records and 337 338 found that wild ass appeared to avoid low suitability habitats. However, we did not detect any dispersal 339 movements and it is possible that dispersing individuals may be willing to cross low suitability habitats which 340 are otherwise avoided during routine movements (Fey et al. 2016; Keeley et al. 2017). Other studies have found such patterns, for example radio-tracking of red squirrels identified that dispersing individuals frequently 341 342 crossed roads, which were otherwise avoided during routine movements (Fey et al. 2016). Finally, little is 343 known about the natal dispersal of Asiatic wild ass with regard to seasonality or sex bias. Consequently, in our 344 habitat selection analysis we did not differentiate between sexes nor test for seasonal effects. However, if such a 345 bias in natal dispersal existed, it is possible that an existing landscape genetic effect was obscured (Shafer et al. 2012). Long-term data sets from GPS movement records could provide information on wild ass natal dispersal, 346

347 which could be used to parameterise dispersal-specific resistance layers and improve landscape genetic analysis.

348 Despite the apparent lack of a landscape effect on gene flow, the present study identified a weak genetic 349 structure in the population, which is likely caused by factors that have not been measured here. Three potential 350 causes for genetic structuring are related to the population's demographic history. First, at the onset of the 351 reintroduction, a captive breeding core was created from individuals of two different subspecies (Saltz & 352 Rubenstein 1995). Differences in the effective niche of these two subspecies may result in divergent habitat 353 preferences and lead to spatial separation and limited interbreeding, ultimately promoting the rise of genetic 354 substructure (McDonald et al. 2019). However, an analysis investigating spatial autocorrelation based on 355 individual hybrid indices found no support for spatial segregation based on subspecies ancestry (unpublished results). A second possible reason for genetic structuring in our study population is that it could be the signature 356 357 of the multiple release events during establishment of the wild population. Individuals were released at two reintroduction sites, from which they dispersed across the habitat. Founder effects and genetic drift experienced 358 359 by the population during early stages of establishment could have resulted in the weak genetic differentiation. 360 Other studies have described a genetic signature of release events in translocated populations (Williams et al.

- 2000; Biebach & Keller 2009; Puckett et al. 2014; Moraes et al. 2017). For example, Grauer et al. (2017)
- reported unique patterns of genetic structure caused by serial release events of individuals from different
 sources, in a reintroduced population of American Marten. Finally, a behavioural effect related to the resource-
- defence-polygyny of the Asiatic wild ass could be the cause for the observed genetic clustering (Renan 2014).
- 365 Male wild ass defend territories around permanent water sources. Increased resource requirements restrict
- 366 females to the vicinity of these permanent water sources during the foaling and breeding season in the summer
- 367 (Saltz et al. 2000; Wallach et al. 2007; Boyd et al. 2016). The GPS records of radio collared individuals
- 368 reflected these behavioural patterns: Males remained close to a water source all year round, while females
- 369 extended their movement range in the winter, yet returned to the same area of the permanent water source in the
- 370 summer when mating occurs (Supplementary material, Fig. A1). This seasonal range contraction and the
- resulting highly localised breeding activity could result in a genetic differentiation between individuals from
 different activity centres (Renan 2014; Giotto et al. 2015). This could explain the presence of a fine-scale weak
- 372 unrefent activity centres (Kenan 2014; Gloub et al. 2015). This could explain the presence of a fine-scale we 373 genetic structure despite high mobility of the species. A similar effect has been observed in feral horses in
- 374 Nevada: during the hot summer, when most of the mating occurred, herds were unable to disperse from the
- 375 limited water sources, which resulted in a weak genetic differentiation between subgroups from different water
- 376 sources, despite their overlapping winter ranges (Ashley 2004).
- While the current analysis failed to identify an effect of habitat on gene flow, it is important to consider the short lag time since the initial release of individuals which was less than five generations ago (given a generation time
- of 7.5 years; Ransom et al. 2016). Landscape resistance may have an impact on gene flow, however, not enough
- time has passed for the signal to become established (Landguth et al. 2010). At this point it is not possible to
- 381 determine with certainty what is causing the observed weak genetic differentiation. If it is due to the release
- 382 events and range expansion combined with genetic drift during the establishment phase, it is expected to
- diminish over time due to continued gene flow (Short & Petren 2011). However, if it is caused by a behavioural
- 384 or a (not yet detectable) landscape effect, then it is likely to persist or even intensify over time.
- 385 Some restriction to gene flow can increase the potential for retaining genetic diversity and is therefore beneficial
- (Chesser 1991; Chesser et al. 1993). However, intensification of the genetic structure may lead to population
 fragmentation and genetic isolation of subpopulations, which could increase the populations extinction risk
- fragmentation and genetic isolation of subpopulations, which could increase the populations extinction ris
 (With & King 1999; Wang et al. 2017). In an isolated population of woodland caribou, Rangifer tarandus
- 389 caribou, reduced gene flow has caused the rise of genetic substructure over a short time period (15 years)
- 390 (Pelletier et al. 2019). The authors believe that this fragmentation is severely threatening the populations long-
- term persistence, as a 53% reduction in the population's inbreeding effective size has been recorded over a
- timespan of only two generations. To avoid the risks of genetic isolation, management of the Asiatic wild ass
- population should aim to prevent any further reinforcement of the observed structure. Specifically, creating
- additional permanent water sources is expected to increase the number of activity centres, minimise distances
- between these and possibly encourage more dispersal movements. Furthermore, additional permanent water
 sources provide more high-quality territories for Asiatic wild ass, thereby enabling a greater number of males to
- 397 contribute to the gene pool (Greenbaum et al. 2018; Renan et al. 2018).

398 5. Conclusions

399 Here we presented an investigation into landscape barriers to gene flow in a reintroduced population by 400 combining GPS movement records and genetic samples. The results demonstrate the importance of genetic 401 analysis to test presumed landscape barriers to gene flow. Particularly, large-bodied highly mobile species may likely be able to maintain gene flow even across unsuitable habitat. Further, the present study highlights the 402 403 importance for long-term genetic monitoring of reintroduced populations. Genetic structure may develop even 404 after successful establishment of a growing population (Neuwald & Templeton 2013), and in the absence of 405 obvious landscape barriers. While this may be simply a transient phenomenon caused by a founder effect, it may have other underlying causes. If a genetic differentiation persists and intensifies, it can reduce reintroduction 406 407 success even long after initial release of individuals and hence should be considered in conservation 408 management protocols (Kramer-Schadt et al. 2004).

410 References

- Aebischer NJ, Robertson PA, Kenward RE. 1993. Compositional analysis of habitat use from animal radio tracking data. Ecology **74**:1313–1325.
- 413 Altman A. 2016. Female group size in Asiatic wild ass. Ben-Gurion University of the Negev, Israel.
- Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. Available from http://www.bioinformatics.babraham.ac.uk/projects/fastqc.
- 416 Armstrong DP, Seddon PJ. 2008. Directions in reintroduction biology. Trends in Ecology and Evolution 23:20–
 417 25.
- Ashley MC. 2004. Population genetics of feral horses: Implications of behavioral isolation. Journal of
 Mammalogy 85:611–617.
- Baguette M, Blanchet S, Legrand D, Stevens VM, Turlure C. 2013. Individual dispersal, landscape connectivity
 and ecological networks. Biological Reviews 88:310–326.
- Benton TG, Bowler DE. 2012. Dispersal in invertebrates: influences on individual decisions. Pages 41–49 in J.
 Colbert, M. Baguette, T. G. Benton, and J. M. Bullock, editors. Dispersal ecology and evolution. Oxford
 University Press, Oxford.
- Bergl RA, Vigilant L. 2007. Genetic analysis reveals population structure and recent migration within the highly
 fragmented range of the Cross River gorilla (Gorilla gorilla diehli). Molecular Ecology 16:501–516.
- Biebach I, Keller LF. 2009. A strong genetic footprint of the re-introduction history of Alpine ibex (Capra ibex
 ibex). Molecular Ecology 18:5046–5058.
- Boyd L, Scorolli A, Nowzari H, Bouskila A. 2016. Social Organization of Wild Equids. Pages 7–22 in P.
 Kaczensky and J. I. Ransom, editors. Wild Equids: Ecology, Management and Conservation. Johns
 Hopkins University Press, Baltimore, MD.
- Bozzuto C, Biebach I, Muff S, Ives AR, Keller LF. 2019. Inbreeding reduces long-term growth of Alpine ibex
 populations. Nature Ecology & Evolution 3:1359–1364. Available from https://doi.org/10.1038/s41559019-0968-1.
- Buttigieg PL, Ramette A. 2014. A guide to statistical analysis in microbial ecology: a community-focused,
 living review of multivariate data analyses. FEMS microbiology ecology 90:543–550.
- Calenge C. 2006. The package "adehabitat" for the R software: A tool for the analysis of space and habitat use
 by animals. Ecological Modelling **197**:516–519.
- Catchen JM, Hohenlohe PA, Bassham S, Amores A, Cresko WA. 2013. Stacks: An analysis tool set for
 population genomics. Molecular Ecology 22:3124–3140.
- Chesser RK. 1991. Influence of gene flow and breeding tactics on gene diversity within populations. Genetics
 129:573–583.
- Chesser RK, Rhodes OE, Sugg DW, Schnabel A. 1993. Effective sizes for subdivided populations. Genetics
 135:1221–1232.
- Danecek P et al. 2011. The variant call format and VCFtools. Bioinformatics 27:2156–2158. Oxford University
 Press. Available from https://academic.oup.com/bioinformatics/article-
- 447 lookup/doi/10.1093/bioinformatics/btr330 (accessed August 15, 2018).
- 448 Danin A. 1999. Desert rocks as plant refugia in the Near East. Botanical Review 65:93–170.
- 449 Davidson A, Carmel Y, Bar-David S. 2013. Characterizing wild ass pathways using a non-invasive approach:
 450 Applying least-cost path modelling to guide field surveys and a model selection analysis. Landscape
 451 Ecology 28:1465–1478.
- 452 Earl DA, vonHoldt BM. 2012. STRUCTURE HARVESTER: A website and program for visualizing
- 453 STRUCTURE output and implementing the Evanno method. Conservation Genetics Resources 4:359–
 454 361.

- 455 ESRI. 2011. ArcGIS Desktop: Release 10. Environmental Systems Research Institute, Redlands, California.
- Evanno G, Regnaut S, Goudet J. 2005. Detecting the number of clusters of individuals using the software
 STRUCTURE: a simulation study. Molecular ecology 14:2611–20. Available from
 http://www.ncbi.nlm.nih.gov/pubmed/15969739.
- Fey K, Hämäläinen S, Selonen V. 2016. Roads are no barrier for dispersing red squirrels in an urban
 environment. Behavioral Ecology 27:741–747.
- 461 Frankham R, Ballou JD, Briscoe DA, McInnes KH. 2002. Effects of Population Size Reduction. Pages 225–226
 462 in D. A. Briscoe, J. D. Ballou, and R. Frankham, editors. Introduction to Conservation Genetics.
 463 Cambridge University Press, Cambridge.
- Frantz AC, Bertouille S, Eloy MC, Licoppe A, Chaumont F, Flamand MC. 2012. Comparative landscape
 genetic analyses show a Belgian motorway to be a gene flow barrier for red deer (Cervus elaphus), but not
 wild boars (Sus scrofa). Molecular Ecology 21:3445–3457.
- Giotto N, Gerard JF, Ziv A, Bouskila A, Bar-David S. 2015. Space-use patterns of the Asiatic Wild Ass (Equus hemionus): Complementary insights from displacement, recursion movement and habitat selection analyses. PLoS ONE 10:1–21. Available from http://dx.doi.org/10.1371/journal.pone.0143279.
- Grauer JA, Gilbert JH, Woodford JE, Eklund D, Anderson S, Pauli JN. 2017. Unexpected genetic composition
 of a reintroduced carnivore population. Biological Conservation 215:246–253. Elsevier. Available from
 http://dx.doi.org/10.1016/j.biocon.2017.09.016.
- Greenbaum G, Renan S, Templeton AR, Bouskila A, Saltz D, Rubenstein DI, Bar-David S. 2018. Revealing
 life-history traits by contrasting genetic estimations with predictions of effective population size.
 Conservation Biology 32:817–827. Available from http://doi.wiley.com/10.1111/cobi.13068.
- Gueta T, Templeton AR, Bar-David S. 2014. Development of genetic structure in a heterogeneous landscape
 over a short time frame: the reintroduced Asiatic wild ass. Conservation Genetics 15:1231–1242.
- Henley SR, Ward D, Schmidt I. 2007. Habitat selection by two desert-adapted ungulates. Journal of Arid
 Environments 70:39–48.
- Howell PE, Koen EL, Williams BW, Roloff GJ, Scribner KT. 2016. Contiguity of landscape features pose
 barriers to gene flow among American marten (Martes americana) genetic clusters in the Upper Peninsula
 of Michigan. Landscape Ecology 31:1051–1062.
- Johnson DH. 1980. The comparison of usage and availability measurements for evaluating resource preference.
 Ecology 61:65–71. Wiley Online Library.
- Jombart T. 2008. Adegenet: A R package for the multivariate analysis of genetic markers. Bioinformatics
 24:1403–1405.
- Jombart T, Devillard S, Balloux F. 2010. Discriminant analysis of principal components: A new method for the
 analysis of genetically structured populations. BMC Genetics 11:94–109.
- Keeley ATH, Beier P, Keeley BW, Fagan ME. 2017. Habitat suitability is a poor proxy for landscape
 connectivity during dispersal and mating movements. Landscape and Urban Planning 161:90–102.
 Elsevier B.V. Available from http://dx.doi.org/10.1016/j.landurbplan.2017.01.007.
- 492 Kramer-Schadt S, Revilla E, Wiegand T, Breitenmoser U. 2004. Fragmented landscapes, road mortality and
 493 patch connectivity: Modelling influences on the dispersal of Eurasian lynx. Journal of Applied Ecology
 494 41:711–723.
- Landguth EL, Cushman SA, Schwartz MK, McKelvey KS, Murphy M, Luikart G. 2010. Quantifying the lag
 time to detect barriers in landscape genetics. Molecular Ecology 19:4179–4191.
- Legendre PP, Andersson MJ, Anderson M. 1999. Distance-based redundancy analysis: Testing multispecies
 responses in multifactorial ecological experiments. Ecological Monographs 69:1–24.
- MacArthur RH. 1957. On the relative abundance of bird species. Proceedings of the National Academy of
 Sciences of the United States of America 43:293–295.
- 501 Manel S, Schwartz MK, Luikart G, Taberlet P, Manel, Stephanie, Schwartz MK, Luikart G, Taberlet P. 2003.

- 502 Landscape genetics: combining landscape ecology and population genetics. Trends in Ecology and 503 Evolution **18**:8–9.
- McDonald MM, Johnson SM, Henry ER, Cunneyworth PMK. 2019. Differences between ecological niches in northern and southern populations of Angolan black and white colobus monkeys (Colobus angolensis palliatus and Colobus angolensis sharpei) throughout Kenya and Tanzania. American Journal of Primatology:e22975.
- McRae B, Shah V, Mohapatra T. 2013. CIRCUITSCAPE User Guide. Available from
 http://www.circuitscape.org.
- Mills LS, Allendorf FW. 1996. The one-migrant-per-generation rule in conservation and management.
 Conservation Biology 10:1509–1518. Available from http://doi.wiley.com/10.1046/j.1523 1739.1996.10061509.x.
- Moraes AM, Ruiz-Miranda CR, Ribeiro MC, Grativol AD, da S. Carvalho C, Dietz JM, Kierulff MCM, Freitas
 LA, Galetti PM. 2017. Temporal genetic dynamics of reintroduced and translocated populations of the
 endangered golden lion tamarin (Leontopithecus rosalia). Conservation Genetics 18:995–1009.
- Nandintsetseg D, Kaczensky P, Ganbaatar O, Leimgruber P, Mueller T. 2016. Spatiotemporal habitat dynamics
 of ungulates in unpredictable environments: The khulan (Equus hemionus) in the Mongolian Gobi desert
 as a case study. Biological Conservation 204:313–321. Elsevier Ltd. Available from
 http://dx.doi.org/10.1016/j.biocon.2016.10.021.
- Neuwald JL, Templeton AR. 2013. Genetic restoration in the eastern collared lizard under prescribed woodland
 burning. Molecular Ecology 22:3666–3679.
- Nezer O, Bar-David S, Gueta T, Carmel Y. 2017. High-resolution species-distribution model based on
 systematic sampling and indirect observations. Biodiversity and Conservation 26:421–437. Springer
 Netherlands.
- Oksanen J, Blanchet FG, Kindt R, Legendre P, O'hara RB, Simpson GL, Solymos P, Stevens MHH, Wagner H.
 2010. Vegan: community ecology package. R package version 1.17-4. http://cran. r-project. org>. Acesso
 em 23:2010.
- Paradis E, Schliep K. 2018. ape 5.0: An environment for modern phylogenetics and evolutionary analyses in R.
 Bioinformatics 35:526–528. Oxford University Press.
- Paris JR, Stevens JR, Catchen JM. 2017. Lost in parameter space: A road map for stacks. Methods in Ecology
 and Evolution 8:1360–1373.
- Pelletier F, Turgeon G, Bourret A, Garant D, St-Laurent MH. 2019. Genetic structure and effective size of an
 endangered population of woodland caribou. Conservation Genetics 20:203–213. Springer Netherlands.
 Available from http://dx.doi.org/10.1007/s10592-018-1124-1.
- Peterson BK, Weber JN, Kay EH, Fisher HS, Hoekstra HE. 2012. Double digest RADseq: An inexpensive
 method for de novo SNP discovery and genotyping in model and non-model species. PLoS ONE 7.
- Pew J, Muir PH, Wang J, Frasier TR. 2015. related: an R package for analysing pairwise relatedness from
 codominant molecular markers. Molecular Ecology Resources 15:557–561. Wiley Online Library.
- Pritchard JK, Stephens M, Donnelly P. 2000. Inference of population structure using multilocus genotype data.
 Genetics 155:945–959.
- Puckett EE et al. 2014. Influence of drift and admixture on population structure of American black bears (Ursus americanus) in the Central Interior Highlands, USA, 50 years after translocation. Molecular Ecology
 23:2414–2427.
- R core team. 2020. R: A language and environment for statistical computing. R Foundation for Statistical
 Computing, Vienna, Austria. Available from https://www.r-project.org/.%0A.
- Ransom JI, Lagos L, Hrabar H, Nowzari H, Usukhjargal D, Spasskaya N. 2016. Wild and feral equid population
 dynamics. Pages 87–104 in J. I. Ransom and P. Kaczensky, editors. Wild equids: Ecology, management,
 and conservation. Johns Hopkins University Press, Baltimore.
- 549 Renan S. 2014. From behavioral patterns to genetic structure: The reintroduced Asiatic Wild Ass (Equus

- 550 hemionus) in the Negev Desert. Ben-Gurion University of the Negev, Israel.
- Renan S, Speyer E, Ben-Nun T, Ziv A, Greenbaum G, Templeton AR, Bar-David S, Bouskila A. 2018. Fission fusion social structure of a reintroduced ungulate: Implications for conservation. Biological Conservation
 222:261–267. Available from https://doi.org/10.1016/j.biocon.2018.04.013.
- Rubenstein DI. 1994. The Ecology of female social behaivour in Horses Zebras and Asses. Pages 13–28 Animal
 Societies: individuals, interactions and organization.
- Saccheri I, Kuussaari M, Kankare M, Vikman P, Fortelius W, Hanski I. 1998. Inbreeding and extinction in a
 butterfly metapopulation. Nature **392**:491–494. Springer Netherlands. Available from
 http://dx.doi.org/10.1016/j.biocon.2011.12.034 (accessed November 19, 2018).
- Saltz D, Rowen M, Rubenstein DI. 2000. The effect of space-use patterns of reintroduced Asiatic wild ass on
 effective population size. Conservation Biology 14:1852–1861. Available from
 http://dx.doi.org/10.1111/j.1523-1739.2000.99227.x.
- Saltz D, Rubenstein DI. 1995. Population dynamics of a reintroduced Asiatic wild ass (Equus hemionus) herd.
 Ecological Applications 5:327–335.
- Seddon PJ, Soorae PS. 1999. Guidelines for subspecific substitutions in wildlife restoration projects.
 Conservation Biology 13:177–184.
- Shafer ABA, Northrup JM, White KS, Boyce MS, Côté SD, Coltman DW. 2012. Habitat selection predicts
 genetic relatedness in an alpine ungulate. Ecology 93:1317–1329. John Wiley & Sons, Ltd. Available
 from http://doi.wiley.com/10.1890/11-0815.1 (accessed March 8, 2019).
- Sharma BD, Clevers J, De Graaf R, Nawa R. 2004. Mapping Equus kiang (Tibetan Wild Ass) habitat in
 Surkhang, Upper Mustang, Nepal. International Mountain Society 24:149–156.
- Short KH, Petren K. 2011. Fine-scale genetic structure arises during range expansion of an invasive gecko.
 PLoS ONE 6.
- Spear SF, Balkenhol N, Fortin MMJ, McRae BH, Scribner K. 2010. Use of resistance surfaces for landscape
 genetic studies: Considerations for parameterization and analysis. Molecular Ecology 19:3576–3591.
- 575 Stern E, Gradus Y, Meir A, Krakover S, Tsoar H. 1986. Atlas of the Negev. Keter Publishing House, Jerusalem.
- Storfer A, Murphy MA, Evans JS, Goldberg CS, Robinson S, Spear SF, Dezzani R, Delmelle E, Vierling L,
 Waits LP. 2007. Putting the "landscape" in landscape genetics. Heredity 98:128–142.
- Templeton AR. 2017. Measuring biodiversity and monitoring ecological and evolutionary processes with
 genetic and genomic tools. Pages 251–265 in J. Garson, A. Plutynski, and S. Sarkar, editors. The
 Routledge handbook of philosophy of biodiversity. Routledge, New York.
- Templeton AR, Brazeal H, Neuwald JL. 2011. The transition from isolated patches to a metapopulation in the
 eastern collared lizard in response to prescribed fires. Ecology 92:1736–1747.
- Wallach AD, Inbar M, Scantlebury M, Speakman JR, Shanas U. 2007. Water requirements as a bottleneck in the
 reintroduction of European roe deer to the southern edge of its range. Canadian Journal of Zoology
 85:1182–1192. Available from http://www.nrcresearchpress.com/doi/10.1139/Z07-098.
- Wang J. 2002. An estimator for pairwise relatedness using molecular markers. Genetics 160:1203–1215.
 Genetics. Available from http://www.ncbi.nlm.nih.gov/pubmed/10924488 (accessed August 7, 2018).
- Wang J. 2017. Estimating pairwise relatedness in a small sample of individuals. Heredity 119:302–313. Nature
 Publishing Group. Available from http://dx.doi.org/10.1038/hdy.2017.52.
- Wang W, Qiao Y, Li S, Pan W, Yao M. 2017. Low genetic diversity and strong population structure shaped by
 anthropogenic habitat fragmentation in a critically endangered primate, Trachypithecus leucocephalus.
 Heredity 118:542–553.
- Williams RN, Rhodes OE, Serfass TL. 2000. Assessment of genetic variance among source and reintroduced
 fisher populations. Journal of Mammalogy 81:895–907.
- 595 With KA, King AW. 1999. Extinction thresholds for species in fractal landscapes. Conservation Biology

- :314–326.
- Wolf CM, Garland T, Griffith B. 1998. Predictors of avian and mammalian translocation success: Reanalysis
 with phylogenetically independent contrasts. Biological Conservation 86:243–255.
- Ziółkowska E, Perzanowski K, Bleyhl B, Ostapowicz K, Kuemmerle T. 2016. Understanding unexpected
 reintroduction outcomes: Why aren't European bison colonizing suitable habitat in the Carpathians?
 Biological Conservation 195:106–117.

604 Tables and Figures

Table 1 Distance based redundancy analysis models tested and their total variance (Inertia), the % variation607explained (R2) and adjusted % variation explained (adjusted R2), the degrees of freedom (df), F-statistic (F) and608p-value of the permutation tests (9999 permutations). Partial models controlling for a third variable are indicated609with |.

Inertia	% Variation (constrained Inertia or R2)	Adjusted % variation explained	df	F	р
		(adjustedR2)			
4.47	3.74%	<1%	1	0.972	0.610
4.47	3.86%	<1%	1	1.003	0.497
4.47	3.64%	<1%	1	0.943	0.698
4.47	3.96%	<1%	1	1.028	0.407
4.47	3.23%	<1%	1	0.833	0.915
4.47	3.85%	<1%	1	1.000	0.495
4.47	3.01%	<1%	1	0.776	0.961
_					
_					
4.47	15.05%	<1%	4	0.975	0.682
4.47	15.98%	<1%	4	1.046	0.204
4.47	14.48%	<1%	4	0.932	0.881
4.47	15.84%	<1%	4	1.023	0.415
4.47	15.12%	<1%	4	0.967	0.655
4.47	15.27%	<1%	4	0.986	0.577
4.47	13.63%	<1%	4	0.871	0.918
	4.47 4.47 4.47 4.47 4.47 4.47 4.47 4.47	Inertia % variation (constrained Inertia or R2) 4.47 3.74% 4.47 3.86% 4.47 3.64% 4.47 3.96% 4.47 3.96% 4.47 3.01% 4.47 15.05% 4.47 15.98% 4.47 15.84% 4.47 15.12% 4.47 13.63%	Inertia% variation (constrained Inertia or R2)Adjusted % variation explained (adjustedR2) 4.47 3.74% $(adjustedR2)$ 4.47 3.86% (1%) 4.47 3.64% (1%) 4.47 3.96% (1%) 4.47 3.23% (1%) 4.47 3.23% (1%) 4.47 3.01% (1%) 4.47 15.05% (1%) 4.47 15.98% (1%) 4.47 15.98% (1%) 4.47 15.84% (1%) 4.47 15.12% (1%) 4.47 15.27% (1%) 4.47 15.27% (1%) 4.47 15.27% (1%) 4.47 13.63% (1%)	Inertia% variation (constrained Inertia or R2)Adjusted % variation explained (adjustedR2) 4.47 3.74% $(1\%$ 4.47 3.74% $(1\%$ 4.47 3.66% $(1\%$ 4.47 3.64% $(1\%$ 4.47 3.96% $(1\%$ 4.47 3.23% $(1\%$ 4.47 3.23% $(1\%$ 4.47 3.01% $(1\%$ 4.47 15.05% $(1\%$ 4.47 15.98% $(1\%$ 4.47 15.84% $(1\%$ 4.47 15.12% $(1\%$ 4.47 15.27% $(1\%$ 4.47 13.63% $(1\%$	Inertia% variation (constrained Inertia or R2)Adjusted % variation explained (adjustedR2) 4.47 3.74% $(1\%$ 4.47 3.74% $(1\%$ 4.47 3.86% $(1\%$ 4.47 3.64% $(1\%$ 4.47 3.64% $(1\%$ 4.47 3.96% $(1\%$ 4.47 3.23% $(1\%$ 4.47 3.23% $(1\%$ 4.47 3.01% $(1\%$ 4.47 3.01% $(1\%$ 4.47 15.05% $(1\%$ 4.47 15.98% $(1\%$ 4.47 15.98% $(1\%$ 4.47 15.12% $(1\%$ 4.47 15.12% $(1\%$ 4.47 15.27% $(1\%$ 4.47 13.63% $(1\%$ 4 0.986 4.47 13.63% $(1\%$ 4 0.986





Fig. 1 Map of the study area in Southern Israel, depicting locations of *Equus hemionus ssp.* DNA sample
collection (n=30), release sites of the reintroduction and location of three artificial water sources. The orange
outline indicates the area of the species distribution model created by Nezer et al. (2017) and the study area for

- 617 the landscape genetic analysis
- 618
- 619



Fig. 2 Habitat resistance surfaces for the study area in Southern Israel, based on (a) habitat suitability index and
(b) slope. Shading indicates resistance value.





Fig. 3 Optimal number of genetic clusters in the reintroduced population of *Equus hemionus ssp.* in Southern
Israel. Initial exploration using principle component analysis indicates no distinct clustering along (a) the first
and second and (b) along the first and third principle components. (c) The Bayesian information criterion
(DAPC analysis) is lowest for K=1 indicating no genetic clustering. (d) The Evanno method (STRUCTURE
analysis) indicates a clear peak in Delta(K) for K=4, while the mean Ln P(D), in dots, does not reach a plateau.



Fig. 4 Genetic structure analysis of reintroduced *Equus hemionus ssp.* in Southern Israel. (a) Spatial distribution
of sampling locations for individuals consistently assigned to the same cluster by both STRUCTURE and a
discriminant analysis of principle components (DAPC). Only individuals with a high assignment probability
(>=0.7) to a single genetic cluster are displayed. Colours indicate 4 genetic clusters (blue, green, red, orange).
White points indicate individuals not assigned consistently by the two analyses. (b) Proportional ancestry of all
individuals (n=30) for K=4 as estimated by STRUCTURE (top) and DAPC (bottom).



638 Fig. 5 Proportional habitat use by 7 *E.hemionus ssp.* individuals in Southern Israel between 2013-2017 based on

639 GPS record data. Habitat is classified based on (a) slope and (b) suitability index. Dark bars indicate mean (+-

SD) proportional usage by individuals and light bars indicate proportional availability in the study area of each
 habitat class. "*" indicates significance by permutation of differences in mean proportional habitat use between
 categories.

643

644 Supplementary material

645 Appendix A

Table A1 Location records collected for different time intervals for 7 individuals equipped with GPS collars
 recording at hourly intervals

ID	Name	Sex	Start date	End date	Total time	Total number of records	Number of records within reduced study area
6000	Ktsoutsy	Male	16.10.2012	05.12.2014	25 months	15712	5323
6001	Short tail	Male	08.06.2013	18.04.2014	10 months	7786	4011
6002	Nahum	Male	12.07.2013	31.12.2014	17 months	14101	7898
	Tacum						
6003	Gila	Female	07.08.2013	08.02.2015	18 months	14980	5547
6004	Idan	Male	08.08.2013	18.01.2015	17 months	14901	2937
6005	Alona	Female	08.07.2015	22.06.2017	24 months	16700	10718
6006	Ariela	Female	17.07.2015	18.02.2017	19 months	14442	3827

Table A2 Percentage of variation explained by the retained principle coordinates (PCos) of different habitat
 resistance variables

Variable	Variance explained by
	retained principle
	coordinate

only first PCo retained	
Habitat suitability resistance	34.92%
Slope resistance	37.07%
Geographic distance	35.18%

first 4 PCos retained	
Habitat suitability resistance	61.96%
Slope resistance	63.29%
Geographic distance	59.41%

656 **Table A3** Simplified ranking matrix comparing proportional habitat use with overall habitat availability in the

657 study area for a) different categories of habitat slope and b) different categories of habitat suitability. "+"

658 indicates the habitat in the row is used more than the habitat in the column, "-" indicates the opposite. "+++"

and "---" indicate that the difference is significant at p<0.05.

a)	Habitat slope						
	0°-15°	16° -30°	>30° slope	Rank			
	slope	slope	1				
0°-15° slope		+++	+++	2			
16°-30° slope			+++	1			
>30° slope				0			
b)	Habitat suitability						
	TT: 1	Intermodiate	Low	Donl			
	suitability	suitability	suitability	Kalik			
High suitability	suitability	suitability +++	suitability +++	2			
High suitability Intermediate suitability	Hign suitability	suitability +++	suitability +++ +++	2 1			
High suitability Intermediate suitability Low	suitability	suitability +++	suitability +++ +++	2 1 0			

661



Fig. A1 Individual movement tracks for four males (left) and three females (right). Data represent hourly records obtained from GPS collars over a minimum period of 10

664 months. Left panels indicated movements recorded during the breeding season (June-August), right panels represent movements during non-breeding season (October-May).

665 For three individuals (Nahum Tacum, Alona, Ariela) data were obtained for two consecutive breeding seasons. Two females (Alona, Ariela) which displayed long-distance

666 movements during the non-breeding seasons, returned to the area near the permanent water source during breeding season in two consecutive years.





- variation explained as expected under a broken stick model. Only the first (b, c) or first and second (a) principle
 coordinates explain more variation than expected



Fig. A3 DAPC plot of the reintroduced *Equus hemionus ssp.* population in Southern Israel for predefined K=4.
 Right inset shows a bar chart of eigenvalues of the PCA with dark retained eigenvalues. Left inset shows a bar

676 chart of DA eigenvalues with dark corresponding components.





684

Fig. A4 Results of the eigenanalysis of selection ratios to evaluate habitat selection by 7 GPS-collared Asiatic wild ass *E.h.ssp* with respect to a) habitat suitability and b) habitat slope. Top figures show the habitat types,

683 bottom figures show habitat preference of each individual.

- 685 Appendix B
- 686 **B1.** Repeated genetic clustering analysis using only samples (N=15) collected during the breeding season (June 687 August)

The variation explained by the first two principle components of the PCA was low (PC1 7.62%, PC2 6.54%)

and no prominent genetic clusters could be identified (Fig. B1.1a, B1.1b). Also, the BIC plot of the DAPC did

690 not display a clear minimum value after which the BIC rises again, which would indicate the optimal number of

- 691 clusters (Fig. B1.1c). This suggested no meaningful genetic clustering in the population. In contrast, for the 692 STRUCTURE analysis, the Evanno method identified a clear peak of delta(K) for K=2 (delta(K)=118.78; Fig.
- 592 STRUCTURE analysis, the Evanno method identified a clear peak of delta(K) for K=2 (delta(K)=118.78; Fig. B1.1d). However, the Evanno method cannot identify an optimum of K=1 and may indicate peaks at higher
- values of K even in the absence of any genetic structure (Evanno et al. 2005). The mean Ln P(D) across
- 695 different values of K displays a plateau between K=2 K=4 (Fig. B1.1d), supporting the results of the Evanno
- 696 method.
- 697 To conclude, the genetic structure analysis based on the reduced data set also offers support for the presence of a
- 698 weak genetic structure in the populations. The genetic cluster analysis in Structure suggested that the population
- 699 may be differentiated into fewer genetic clusters (Best K by Evanno, K=2). However, this is somewhat 700 expected, given the reduced number of samples.





Fig. B1.1 Optimal number of genetic clusters in the reintroduced population of *Equus hemionus ssp.* in Southern
Israel based on the reduced data (N=15) set including only samples collected during the breeding season (JuneAugust). Initial exploration using principle component analysis indicates no distinct clustering along (a) the first
and second and (b) along the first and third principle components. (c) The Bayesian information criterion
(DAPC analysis) does not clearly identify an optimal number of clusters. (d) The Evanno method
(STRUCTURE analysis) indicates a clear peak in Delta(K) for K=2, which is supported by the mean Ln P(D), in

- dots, which reaches a plateau between K=2-K=4.
- 709
- 710
- 711
- 712

- 713 **B2.** Repeated landscape genetic analysis using only samples (N=14) collected during the breeding season (June-
- 714 August)
- 715

For all three resistance variables only the first or first and second PCos explained more variation than expected

under the Broken Stick model (Fig B2.1). However, since this accounted for only ~40% of variation in each

- variable, we repeated the analysis with the first 4 PCos retained which accounted for ~70% of variation (Table
- 719 B2.1). None of the tested models of the distance-based redundancy analysis returned significant results and the
- explained variance was very low (<3%) for all predictor variables (Table B2.2). Habitat suitability and slope

b

- explained negligible variation in genetic relatedness between individuals of the population. This was also true
- for models controlling for geographic distance and resistance distances, respectively (Table B2.2).
- 723
- 724





731 coordinates explain more variation than expected

Table B2.1 Percentage of variation explained by the retained principle coordinates (PCos) of different habitat

resistance variables

Variable	Variance explained by retained principle coordinate
only first PCo retained	
Habitat suitability resistance	40.88%
Slope resistance	48.11%
Geographic distance	39.65%
first 4 PCos retained	
Habitat suitability resistance	72.01%
Slope resistance	81.92%
Geographic distance	69.03%

737	Table B2.2 Distance based redundant	cy analysis models tested	d for samples (N=14) collected	l during the breeding
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season (June-August). Total variance (Inertia), the % variation explained (R2) and adjusted % variation
 explained (adjusted R2), the degrees of freedom (df), F-statistic (F) and p-value of the permutation tests (9999
 permutations). Partial models controlling for a third variable are indicated with 1

permutations). Partial models controlling for a third variable are indicated with |.

Variable	Inertia	%Variation (constrained Inertia or R2)	Adjusted % variation explained (adjustedR2)	df	F	р
only first PCo retained						
SDM resistance	0.74	9.58%	2.04%	1	1.271	0.175
Slope resistance	0.74	8.71%	1.10%	1	1.145	0.290
Geographic distance	0.74	9.53%	1.99%	1	1.264	0.179
SDM resistance geographic distance	0.74	7.98%	<1%	1	1.063	0.389
Slope resistance geographic distance	0.74	6.01%	<1%	1	0.782	0.773
Geographic distance SDM resistance	0.74	7.93%	<1%	1	1.057	0.401
Geographic distance slope resistance	0.74	6.83%	<1%	1	0.889	0.624
first 4 PCos retained	_					
SDM resistance	0.74	28.87%	<1%	4	0.913	0.726
Slope resistance	0.74	31.59%	1.19%	4	1.039	0.393
Geographic distance	0.74	29.14%	<1%	4	0.926	0.690
SDM resistance geographic distance	0.74	31.23%	<1%	4	0.985	0.520
Slope resistance geographic distance	0.74	32.47%	2.56%	4	1.058	0.407
Geographic distance SDM resistance	0.74	31.51%	<1%	4	0.994	0.514
Geographic distance slope resistance	0.74	30.03%	<1%	4	0.978	0.526