



LJMU Research Online

Turton, N, Bowers, N, Khajeh, S, Hargreaves, IP and Heaton, RA

Coenzyme Q10 and the exclusive club of diseases that show a limited response to treatment

<http://researchonline.ljmu.ac.uk/id/eprint/15108/>

Article

Citation (please note it is advisable to refer to the publisher's version if you intend to cite from this work)

Turton, N, Bowers, N, Khajeh, S, Hargreaves, IP and Heaton, RA (2021) Coenzyme Q10 and the exclusive club of diseases that show a limited response to treatment. Expert Opinion on Orphan Drugs. ISSN 2167-8707

LJMU has developed **LJMU Research Online** for users to access the research output of the University more effectively. Copyright © and Moral Rights for the papers on this site are retained by the individual authors and/or other copyright owners. Users may download and/or print one copy of any article(s) in LJMU Research Online to facilitate their private study or for non-commercial research. You may not engage in further distribution of the material or use it for any profit-making activities or any commercial gain.

The version presented here may differ from the published version or from the version of the record. Please see the repository URL above for details on accessing the published version and note that access may require a subscription.

For more information please contact researchonline@ljmu.ac.uk

<http://researchonline.ljmu.ac.uk/>



Coenzyme Q₁₀ and the exclusive club of diseases that show a limited response to treatment

Nadia Turton, Nathan Bowers, Sam Khajeh, Iain P Hargreaves & Robert A Heaton

To cite this article: Nadia Turton, Nathan Bowers, Sam Khajeh, Iain P Hargreaves & Robert A Heaton (2021): Coenzyme Q₁₀ and the exclusive club of diseases that show a limited response to treatment, Expert Opinion on Orphan Drugs, DOI: [10.1080/21678707.2021.1932459](https://doi.org/10.1080/21678707.2021.1932459)

To link to this article: <https://doi.org/10.1080/21678707.2021.1932459>



© 2021 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.



Published online: 03 Jun 2021.



Submit your article to this journal [↗](#)



Article views: 2



View related articles [↗](#)



View Crossmark data [↗](#)

Coenzyme Q₁₀ and the exclusive club of diseases that show a limited response to treatment

Nadia Turton, Nathan Bowers, Sam Khajeh, Iain P Hargreaves and Robert A Heaton

School of Pharmacy and Biomolecular Sciences, Liverpool John Moores University, Liverpool UK

ABSTRACT

Introduction: Coenzyme Q₁₀ (CoQ₁₀) is a ubiquitous organic molecule with a significant role in the mitochondrial electron transport chain (ETC). As a result of its role in such an important biological process, CoQ₁₀ deficiency has been implicated in the pathogenesis of numerous diseases such as Parkinson's disease (PD) and multiple sclerosis (MS). This has led to multiple attempts to use CoQ₁₀ supplementation as a treatment or pre-treatment with varying degrees of success.

Areas covered: The present review will identify evidence of mitochondrial dysfunction in MS, PD and mitochondrial ETC disorders. In addition, the inability of Co₁₀ supplementation to elicit significant clinical outcome in these disorders and possible flaws in these studies will be discussed. The databases utilized for this review were the Web of science and PubMed, with inclusive dates (1957–2021).

Expert opinion: A lack of improved neurological outcome in these disorders post treatment with CoQ₁₀ may be attributable to the limited ability of CoQ₁₀ to cross the blood–brain barrier. Thus, CoQ₁₀ alternatives should also be considered. Other factors including time of disease diagnosis, dosage, administration, and duration of CoQ₁₀ therapy may have a significant influence on the efficacy of this treatment.

ARTICLE HISTORY

Received 15 February 2021
Accepted 18 May 2021

KEYWORDS

Coenzyme Q₁₀;
mitochondrial electron
transport chain disorders;
multiple sclerosis;
Parkinson's disease

1. Introduction

Coenzyme Q₁₀ (CoQ₁₀) is the dominant form of ubiquinone found in humans and can be found in almost all cellular membranes [1]. This lipid-soluble molecule is made up of a benzoquinone moiety, with a 10-subunit polyisoprenoid lipid tail [2] (Figure 1). The central benzoquinone ring includes two redox active sites, essential for its cellular function, whereas the polyisoprenoid tail is responsible for the correct positioning in a variety of membranes [2].

The synthesis of CoQ₁₀ can be divided into three distinct steps. Firstly, the synthesis of the benzoquinone ring from 4-hydroxybenzoate, followed by the synthesis of the polyisoprenoid tail from acetyl-coenzyme A (CoA) utilizing the mevalonate pathway and finally, the condensation of the two structures to form CoQ₁₀ [3]. The condensation step in CoQ₁₀ synthesis takes place within the mitochondria and has been suggested to be the rate limiting step in the CoQ₁₀ biosynthetic pathway [4].

CoQ₁₀ plays a significant role in a number of biochemical pathways within the mitochondrion [2]. However, CoQ₁₀'s principal function is as an electron carrier, exchanging electrons derived from both complex I and II and transferring them to complex III facilitating electron transfer in the mitochondrial electron transport chain (ETC; Figure 2). In addition, CoQ₁₀ also functions as an essential lipid-soluble antioxidant within the cell, and because of CoQ₁₀'s ubiquitous nature it not only protects the mitochondrial membrane from free radical induced oxidation, but also that of other organelle

membranes together with the plasma membrane [3]. Additionally, CoQ₁₀ is also involved in the regeneration of other cellular antioxidants such as vitamin C and E [5]. In a recent study by Heaton, Heales [6] evidence showed that cellular CoQ₁₀ concentration can even influence the lysosomal lumen pH, although the mechanism by which this occurs has yet to be elucidated.

In view of CoQ₁₀'s functional role, CoQ₁₀ has been supplemented in response to three disorders of the central nervous system (CNS), of which they have displayed evidence of mitochondrial dysfunction and oxidative stress [3,7,8]. These disorders include multiple sclerosis (MS), mitochondrial electron transport chain (ETC) disorders and Parkinson's disease (PD). The present review will evaluate the inability of Co₁₀ supplementation to elicit significant clinical outcome in these disorders and possible flaws in these studies will be discussed.

1.1. CoQ₁₀ Deficiency

CoQ₁₀ deficiencies can be divided into two categories, primary and secondary deficiencies [2]. Primary CoQ₁₀ deficiencies are caused by mutations in the genes directly involved in the biosynthesis of CoQ₁₀ [9,10]; whereas secondary deficiencies have been associated with mutations in genes that are indirectly related to CoQ₁₀ biosynthesis, or to other non-genetic factors [11].

Due to the plethora of different biochemical functions of CoQ₁₀, the clinical consequences resulting from a CoQ₁₀ deficiency can have a wide spectrum of presentations. In

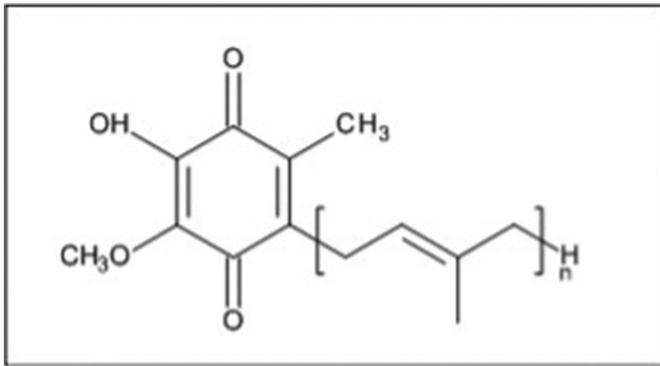


Figure 1. Chemical structure of coenzyme Q₁₀ (CoQ₁₀) N = 10.

particular, a CoQ₁₀ deficiency results in the impairment of the METC, leading to a reduced cellular ATP production [12]. Primary CoQ₁₀ deficiencies are most often reported in tissues with high energy demand such as the CNS [12]. The clinical presentation and severity of primary CoQ₁₀ deficiencies can vary drastically ranging from a mild clinical phenotype to a severe infantile presentation which can often be fatal [13].

Secondary CoQ₁₀ deficiency typically presents in diseases such as mitochondrial myopathies [14,15], cardiovascular disease [16], type II diabetes [17], chronic kidney disease [18], liver disease and critical illness [19]. In common with a primary CoQ₁₀ deficiency the deficit in CoQ₁₀ contributes to disease pathophysiology in these disorders by impairing energy metabolism and compromising cellular antioxidant status. As discussed previously, a secondary CoQ₁₀ deficiency can be caused by non-genetic factors such as the pharmacological agents, statins [20]. Statins reduce the circulatory levels of low-density lipoprotein (LDL) through the competitive inhibition of the enzyme, 3-hydroxy-3-methyl-glutaryl-coenzyme

A reductase (HMG-CoA reductase) (Figure 3) which serves as a major regulatory role in both cholesterol and CoQ₁₀ biosynthesis [21]. A number of studies have reported reduced CoQ₁₀ status following statin treatment [21]. However, the majority of these studies assessed endogenous CoQ₁₀ status in plasma/serum, which may only reflect the decrease in LDL, as LDL is the principal carrier of CoQ₁₀ in the circulation [3]. Only a small number of studies have directly measured CoQ₁₀ muscle status which is thought to be a more accurate reflection of endogenous CoQ₁₀ status [22].

2. Multiple sclerosis

MS is a chronic inflammatory disease of the CNS and currently the most common cause of non-traumatic neurological disability in young adults [23]. While the etiology and pathogenesis of MS still remain to be fully elucidated, research suggests that genetic predisposition and environmental factors trigger the permeation of myelin-specific autoreactive lymphocytes into the CNS, an important causative factor for MS [24–26]. The entry of these lymphocytes into the CNS results in multifocal demyelination, neural and axonal damage, oligodendrocyte loss, blood–brain barrier (BBB) breakdown and oxidative stress [23,27–29]. The clinical presentation of MS includes optic neuritis, diplopia, fatigue, paralysis, paresthesias and cognitive dysfunction [30]. MS can be categorized into four disease states including relapsing-remitting multiple sclerosis (RRMS), secondary progressive multiple sclerosis (SPSM), primary progressive multiple sclerosis (PPMS), and progressive-relapsing multiple sclerosis (RPMS) [31,32]. The majority of MS patients experience RRMS, during which an episode of neurological impairment is followed by partial or complete remission [33,34].

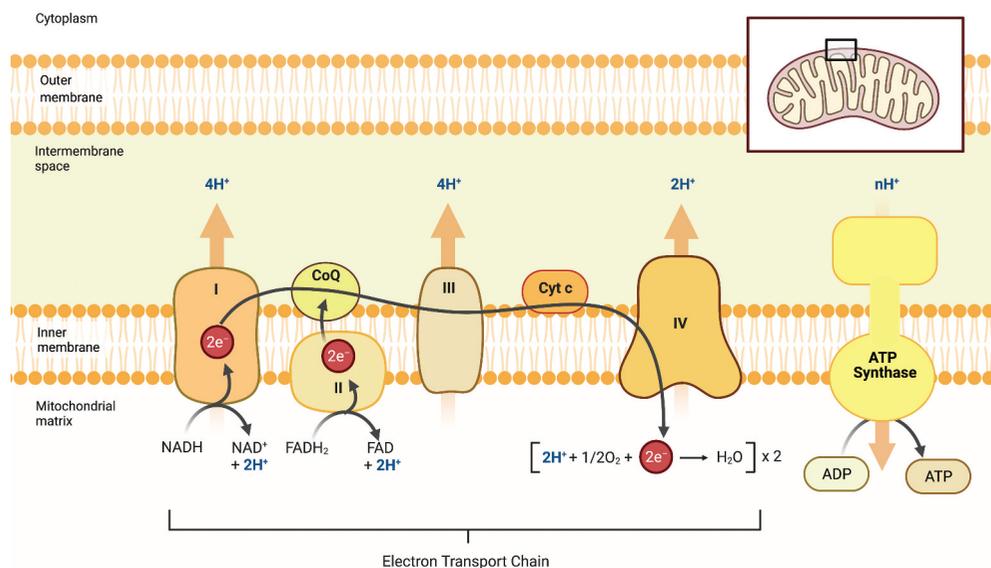


Figure 2. Diagram of electron transport chain (ETC) and ATP synthase illustrating the pumping of H⁺ ions into the intermembrane space, and how CoQ₁₀ augments the passage of electrons. Created using BioRender.com (template provided by Biorender).

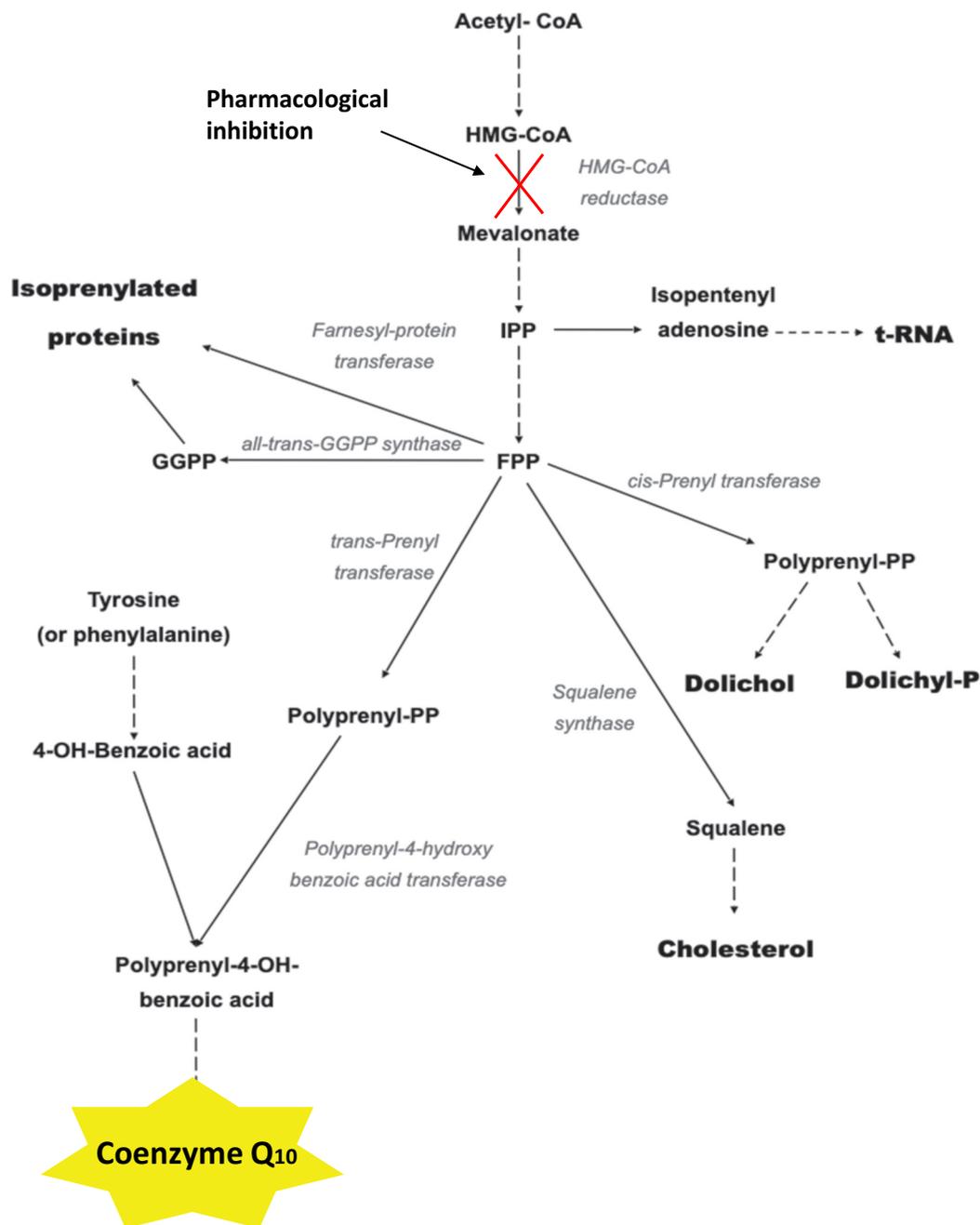


Figure 3. Schematic diagram of the CoQ₁₀ biosynthetic pathway, demonstrating how pharmacological inhibition, such as statin therapy may also influence the synthesis of CoQ₁₀. Adapted from Hargreaves, Duncan [20].

2.1. Mitochondrial dysfunction and oxidative stress in MS

It is becoming increasingly apparent that oxidative stress accompanies neuro-inflammation observed in MS, which can result in secondary mitochondrial dysfunction [35]. This injurious effect upon the mitochondria can result in further reactive oxygen species (ROS) production continuing the cycle of free radical generation, eventually resulting in decreased neuronal ATP production [36,37]. These factors ultimately lead to activation of apoptosis and progressive axon degeneration, which can extenuate to the neuronal body and dendrites and eventually the presynaptic and postsynaptic neurons

[38]. Mitochondrial dysfunction has been associated with the disease pathogenesis of MS, since deficiencies in ETC complex IV, citrate synthase (mitochondrial matrix marker) and complex I (NADH dehydrogenase: ubiquinone reductase) activities have been detected in the active lesions of postmortem brain samples taken from chronic MS patients [39]. Similarly, Sadeghian et al. [40] reported that the activity of ETC complex I was selectively compromised in the spinal tissue in a mouse model of the disease (experimental autoimmune encephalomyelitis; EAE), together with an increase in astrocytic phosphofructokinase 2 (PFK2) expression and a shift from oxidative to glycolytic metabolism. These findings accompanied depolarization of the axonal mitochondria and axons themselves,

thus, correlating with neurological deficits. Evidence of mitochondrial dysfunction is relatively unexplored in other tissues; however, Hargreaves et al. [41] reported statistically significant alterations to ETC complex I and complex II/III (succinate: ubiquinone reductase) activities at the pre-symptomatic and asymptomatic stages in spinal cord, jaw muscle and liver tissues of EAE mouse models of MS. ETC complex I has been reported to be the main site of ROS production in the ETC [42]; thus, increased levels of oxidative stress reported in MS patients [43] may be attributable to a decrease observed in complex I activity [39,40]. Additionally, lymphocytes isolated from MS patients were found to have a significant reduction in mitochondrial membrane potential compared to healthy controls [44]. The differences in mitochondrial function identified in this study were proportional to the degree of MS severity, again suggesting mitochondrial dysfunction may contribute to the pathogenesis of disease progression in MS and is not limited to the CNS.

2.2. CoQ₁₀ limitations in MS

In view of the evidence of mitochondrial dysfunction and oxidative stress reported in the pre-symptomatic and symptomatic stages of MS, the ETC electron carrier and lipid-soluble antioxidant, CoQ₁₀ may be considered as a potential adjunct therapy to control disease progression [15,45]. However, although proven to be beneficial in some cases, several studies have suggested that CoQ₁₀ still has some therapeutic limitations in the treatment of MS and this will be outlined in the following section.

Sanoobar et al. [46] demonstrated that CoQ₁₀ supplementation had no effect on the activity of the antioxidant enzyme, glutathione peroxidase (GPx) in RRMS patients. In this study, fasting blood samples were taken from 45 subjects with MS before and after a 12-week intervention with CoQ₁₀ and a placebo control. However, despite an increase in superoxide dismutase activity (SOD), a decrease in malondialdehyde (MDA; marker of lipid peroxidation) levels and reduced total antioxidant capacity (TAC) levels detected in the CoQ₁₀ treated MS patient group, no significant difference in GPx activity was identified between the control and treated groups. Previously, reduced levels of peripheral GPx activity have been reported in MS patients compared to healthy controls [47]. It has been suggested that GPx may make more of a contribution than the antioxidant enzyme, catalase in detoxifying cellular hydrogen peroxide [48]. Thus, in view of the susceptibility of the CNS to ROS induced oxidative damage, CoQ₁₀ supplementation does not appear to have the capacity to up-regulate the activity of the vital antioxidant enzyme, GPx in MS, which may therefore limit the ability of CoQ₁₀ to delay disease progression [46].

One potential therapeutic mechanism of CoQ₁₀ is its ability to adjust the immune response [49]. Inflammation is a key contributor to axonal injury in MS, of which anti-inflammatory cytokines are decreased and pro-inflammatory cytokines are increased in the CSF [50,51]. A study by Sanoobar et al. [7] reported that CoQ₁₀ supplementation for 12 weeks (500 mg/day) did not alter levels of anti-inflammatory cytokines transforming growth factor- β (TGF- β) and interleukin (IL)-4 in the serum of MS patients compared to the placebo group.

Similarly, in an EAE C57BL/6 female adult mouse model study of MS, Soleimani et al. [52] observed no alterations in the levels of IL-4 in the CoQ₁₀ treated group compared to the untreated group. TGF- β can suppress T-cell proliferation, macrophage activity and cytotoxic lymphocyte maturation [53]. Similarly, IL-4 inhibits the ability of macrophages to produce TNF- α , IL-1 and IL-6 [54,55]. Both of which have demonstrated protective properties in EAE models [56,57]. However, CoQ₁₀ may utilize its anti-inflammatory effects via the inhibition of pro-inflammatory cytokines, as opposed to increasing anti-inflammatory cytokines [7,52]. On the contrary, 12-week CoQ₁₀ (200 mg/day) supplementation in RRMS patients treated with interferon beta1a 44 μ g (IFN- β 1a) demonstrated higher levels of IL-4 in the peripheral blood [58]. However, at present we cannot exclude the possibility that the increase in circulatory IL-4 was the result of IFN- β 1a treatment rather than that of CoQ₁₀. The studies discussed also required patient 'follow up reports' that exceeded a 3-month intervention to identify meaningful changes over longer periods of time. Additionally, in view of the fact that MS is an inflammatory disease of the CNS, supplementary markers of MS severity should be included e.g magnetic resonance imaging data allow an assessment of the effect of CoQ₁₀ supplementation on cerebral function.

Depression and fatigue are common symptoms of MS [59], and therefore, a randomized, double-blinded, placebo-controlled trial was undertaken to assess the effect of a CoQ₁₀ supplementation (500 mg/day) on MS-related fatigue over a 12 week period [60]. In the study fatigue symptoms were measured using a fatigue severity scale (FSS). In the CoQ₁₀ (500 mg/day) treated patient group a significant decrease in FSS was observed. Similarly, previous studies have observed improved fatigue sensation and physical performance during fatigue-inducing workload trials at high-dose CoQ₁₀ (300 mg/day) supplementation [61]. However, these levels of CoQ₁₀ may be higher than that achievable with current oral CoQ₁₀ formulation and lower doses of CoQ₁₀ (100 mg/day) have been shown to have no effect on fatigue during physical workload trials in healthy volunteers, which may translate into the effects of fatigue in MS patients [61]. Furthermore, it should also be considered that larger and long-term follow-up studies are required to confirm the anti-fatigue effects of CoQ₁₀ in MS as well as developing more accurate methods of quantifying fatigue severity in MS following CoQ₁₀ supplementation. However, it should be noted that the etiology of MS-related fatigue is unknown and may be multifactorial and therefore may require more than one candidate therapy

3. Parkinson's disease

PD is a neurodegenerative disorder that is pathologically characterized by Lewy body formation and striatal dopamine depletion in the Substantia Nigra [8]. Abnormal accumulation of ROS in the neuronal cells of PD patients leads to a reduction in the antioxidant, glutathione. As a result, increased oxidative stress leads to mitochondrial respiratory chain dysfunction, the release of pro-apoptotic mitochondrial proteins and ultimately, cell death [62]. As well as CoQ₁₀'s role in the METC

(Figure 2), reduced CoQ₁₀ is also an important antioxidant, protecting the membrane and lipoproteins from ROS [63]. Consequently, the therapeutic efficacy of CoQ₁₀ in the treatment of PD has been tested in numerous studies. The following section evaluates the results obtained from some of the key trials that have been conducted, as well as discussing the potential future applications of CoQ₁₀ in the treatment of PD.

3.1. CoQ₁₀ limitation in PD

Results obtained from a phase II trial indicated that the progression of early-stage PD could be stabilized using oral CoQ₁₀ supplementation [64]. In this trial, patients were randomly assigned to a placebo group or CoQ₁₀ treatment at dosages of 300, 600, or 1200 mg/d. Disease progression was then measured at one, four, eight, twelve and sixteen months using the Unified Parkinson's Disease Rating Scale (UPDRS). This scale measures the mental and motor capacity of patients, as well as their ability to complete daily living activities and in doing so considers the various clinical features of PD. After eight months, the mean total UPDRS score of patients receiving 300 mg/d and 600 mg/d dosages was similar, but importantly, lower than that of the placebo group, suggesting that CoQ₁₀ was effective in slowing disease progression. However, the UPDRS scores of the 1200 mg/d group were not significantly lower than both aforementioned groups, indicating that there is a dosage threshold required for maximum drug efficacy. Following these promising findings, a phase III clinical trial involving 600 patients was conducted using largely the same method [65]. In this trial, treated patients received CoQ₁₀ dosages of 1200 mg/d or 2400 mg/d. Despite 1200 mg/d being the highest dosage used in the previous study, the mean change in UPDRS score of treated patients was not significantly lower than that of the placebo group after 16 months. The researchers therefore concluded that CoQ₁₀ showed no benefit and should not be used in the treatment of early-stage PD.

CoQ₁₀ was unable to stabilize PD progression in the study described by Beal, Oakes [65], but further assessment of this potential treatment is not futile. Half of dopaminergic neurons within the substantia nigra of PD patients are damaged before clinical diagnosis [66]. As the average age of participants in this study was 65 years old, it is possible that the neurones of these patients were already significantly damaged prior to treatment with CoQ₁₀, meaning its neuroprotective effects would be limited [67]. The answer to improving its therapeutic efficacy could therefore lie in using novel methods to diagnose the disease earlier. In addition, the loss of glutathione during presymptomatic Parkinson's disease pathogenesis implies that supplementary antioxidants, such as CoQ₁₀, could be advantageous in its treatment [68].

In both of the trials described, the studied populations consisted of a broad range of sporadic PD patients, this means their contrasting findings could have been the result of diverse gene mutations within their respective populations. A number of mutant genes have been linked to familial PD [69], for example, a mutation to the PINK1 gene prevents PINK1 from transferring electrons between complex 1 and CoQ₁₀ in the MECT [70]. This mutation would significantly

reduce the effectiveness of CoQ₁₀ supplementation on disease progression, however, screening for it did not occur during patient selection, meaning the reported null effects of CoQ₁₀ in PD may have been skewed. The capacity of CoQ₁₀ to reduce oxidative stress could still prove beneficial in treating patients suffering from other forms of the disease and should be investigated further. Both studies reported an increase in mean CoQ₁₀ plasma levels from baseline to the last assessment; however, as previously discussed, this is not an entirely accurate way to estimate tissue CoQ₁₀ concentrations. In addition, neuronal cells have yet another limiting factor in the BBB permeability restricting the passage of CoQ₁₀ into the cells [71]. Whilst creating compounds that can penetrate the BBB still proves difficult to this day, CoQ₁₀ analogues that have better water solubility have been produced. Studies by Muthukumar, Leahy [72] and Sikorska, Lanthier [73] have shown that water-soluble Ubisul-Q10 can stabilize the progression of PD in animal models; however, the positive results obtained from Sikorska, Lanthier [73] are caveated by reports that treatment disruption resulted in continued neuronal degradation. Analogues of CoQ₁₀ which possess improved mitochondrial membrane penetration have also been produced [74]. MitoQ consists of CoQ₁₀ bound to triphenyl alkyl phosphonium cations, the cations interact with negative mitochondrial membranes and in doing so improve CoQ₁₀ uptake [75]. The therapeutic efficacy of MitoQ was tested in a study conducted by Snow, Rolfe [67] where treated patients were given 40 mg or 80 mg dosages of the compound each day for 16 months. Whilst the mean UPDRS score of treated patients did not significantly differ from that of the placebo group, it is possible that late diagnosis of the disease, as well as struggles in getting the compound to cross the BBB hindered the results. If the above-mentioned obstacles can be overcome, water-soluble analogues of CoQ₁₀ could still prove beneficial in treating PD. Proposals suggested by Park, Park [76] indicate that the answer to the BBB conundrum is continuous CoQ₁₀ in traditional delivery. In their study, PD was induced in rat models using 6-hydroxydopamine. They were then injected with various dosages of CoQ₁₀ solution, and their neurodegeneration measured. Measurements used in this study included behavioral and dopaminergic evaluation, as well as neurogenesis, angiogenesis and neuroinflammation analysis. Compared to the placebo group, models treated with high dosage CoQ₁₀ solution showed significant improvements in every parameter. These results suggest that convection-enhanced delivery could improve CoQ₁₀ therapeutic efficacy and therefore provide the basis for more research into what seems a promising avenue for PD treatment.

4. Mitochondrial electron transport chain disorders

The clinical heterogeneity and unpredictable nature of ETC disorders have not been conducive to the development of effective therapeutic strategies for these diseases. Treatment of these disorders is extremely challenging with no cure as yet in sight. Most patients with ETC disorders are afforded symptomatic treatment to manage the various clinical presentations of the disease. Treatment can also involve therapy aimed at enhancing ETC function as well as reducing

Table 1. Summary of the current clinical trials which have assessed the therapeutic efficacy of CoQ₁₀ in the treatment of ETC disorders. Abbreviations: MERRF: mitochondrial encephalopathy and ragged red fibers, MELAS: myoclonic epilepsy and lactic acidosis, CPEO: chronic progressive external ophthalmoplegia, KSS: Kearns Sayre syndrome, NARP: Neuropathy, ataxia, and retinitis pigmentosa. Clinical trials which have assessed the efficacy of CoQ₁₀ in the treatment of ETC disorders.

Study	Possible flaws in study
1. In the study by Bresolin, et al. [92] 44 mitochondrial patients (CPEO, KSS and MERRF) were treated CoQ ₁₀ (2 mg/kg/day) for 6 months. 54% of patients following CoQ ₁₀ treatment were found to have a 35% decrease in plasma lactate levels post exercise. These were deemed 'responders' however following a further 3 months of CoQ ₁₀ treatment no differences were found between the 'responders' and the placebo group.	In the study, serum CoQ ₁₀ levels were reported as: 0.93 nmol/L (pre CoQ ₁₀ supplementation) and 1.33 nmol/L (post CoQ ₁₀ supplementation). The general serum/plasma ref range is reported as: 0.48–1.8 PM [13,931]. The level of CoQ ₁₀ only increased by approximately 1.4-fold following CoQ ₁₀ supplementation. Possibly due to the poor bioavailability of the CoQ ₁₀ formulation used. Low dosage of CoQ ₁₀ used (2 mg/kg/day).
2. In the study by Chen, et al. [94] patients with mitochondrial encephalomyopathies (MERRF, MELAS, CPEO) were treated with CoQ ₁₀ (160 mg/day) for 3 months. A significant improvement in the global MRC score which is used for grading the muscle strength was noted in patients together with a nonsignificant decrease in plasma lactate levels post exercise. Patient serum CoQ ₁₀ levels were decreased below control levels, 0.6–0.98 PM (patient); 0.57–1.83 gM (control range). Following 3 months of CoQ ₁₀ therapy the serum CoQ ₁₀ level of patients increased to 3.46 PM (3.5–5fold increase, approximately).	Low dosage of CoQ ₁₀ used (160 mg/day). No details of type of CoQ ₁₀ formulation were provided. Short duration trial (3 months). Patient fatigability was not significantly improved.
3. In the study by Glover, et al. [95] 30 patients with MRC diseases (MELAS, CPEO, NARP, LHON) were treated with CoQ ₁₀ (1200 mg/day) for 60 days. CoQ ₁₀ used in the study was a Q gel; Tishcon, New York. Following CoQ ₁₀ treatment, only a significant decrease in plasma lactate levels were observed post exercise in patients, no other clinical benefit noted.	The plasma CoQ ₁₀ status of the MRC patients showed no evidence of a CoQ ₁₀ deficiency (mean plasma CoQ ₁₀ status of patients: 1.15 gM). Since these patients may not have an underlying CoQ ₁₀ deficiency, they may not be expected to show any clinical improvement following CoQ ₁₀ supplementation [96]. Short duration trial (60 days).

increased ROS generation typically associated with ETC impairment. In light of the prospect that supplementation with CoQ₁₀ may increase the efficiency of electron transfer through the ETC in patients with disorders of the ETC, CoQ₁₀ supplementation is one of the regular therapeutic strategies used in specialist clinical centers [3]. However, at present there is a paucity of high-level evidence supporting the use of CoQ₁₀ in the treatment of patients with ETC disorders. However, a number of case studies have reported evidence of an increase in strength, accelerated post exercise recovery and an improvement in oxygen consumption in patients following CoQ₁₀ supplementation [77]. The most consistent finding, however, following Co₁₀ therapy appears to be at the

biochemical level, with a decrease in plasma lactate levels being reported following supplementation [78].

4.1. CoQ₁₀ limitations in ETC disorders

Presently, only a limited number of clinical trials have assessed the efficacy of CoQ₁₀ as a treatment for mitochondrial disease (Table 1). The results of these studies have demonstrated no overall clinical benefit and mostly included patients with ETC disorders with variable clinical phenotypes and genotypes. This makes it difficult to draw definite conclusions on the responsiveness of CoQ₁₀ therapy on particular patient groups. Although no benefit in any of the outcomes was observed in two trials [79] evidence of improvement in muscle strength [80] and following exercise, a decrease in plasma lactate levels was reported [81] was reported. It has been suggested that the duration of the study and the dose of CoQ₁₀ administered are both factors that may compromise the therapeutic efficacy of CoQ₁₀ in clinical trials [82,83]. The lack of clinical trials that have assessed the effectiveness of CoQ₁₀ for the treatment of mitochondrial disease can be attributed to the difficulty in recruiting a sufficient number of patients and the relatively high cost of conducting the trials themselves. Furthermore, the reasons for the refractory nature of the neurological sequelae associated with ETC dysfunction to CoQ₁₀ supplementation may be a consequence of the relatively poor ability of CoQ₁₀ to cross the BBB. Animal studies have suggested that CoQ₁₀ supplementation can be transported across the BBB [A, B] (whats A and B). However, it is yet to be established whether the degree of CoQ₁₀ uptake observed in these animal studies would be sufficient to restore cerebral ETC function.

The low bioavailability of some of the CoQ₁₀ formulations used in these trials together with their low treatment doses or short durations may have contributed to the ambiguity in the results of the clinical trials (Table 1) [80,84,85]. CoQ₁₀ doses typically in the range of 5–50 mg/kg/day are recommended in the treatment of patients with primary CoQ₁₀ deficiency and soluble forms are recommended instead of tablet forms, due to their higher bioavailability [86]. At present, however, there is no consensus on what dosage of CoQ₁₀ is appropriate to administer to patients with other forms of ETC dysfunction. However, a dosage of at least 5 mg/kg/day is suggested [87]. Importantly, there is evidence of secondary CoQ₁₀ deficiency associated with patients with ETC disorders, linked to a mutation in a gene not specifically involved in the CoQ₁₀ biosynthetic pathway; in some cases, these patients have been shown to be more responsive to CoQ₁₀ therapy and have shown significant clinical improvement post CoQ₁₀ supplementation [88]. Moreover, this demonstrates the importance of determining the CoQ₁₀ status prior to treatment, in order to identify any subgroups of patients with ETC disorders who may be more responsive to therapy [89]. It should also be noted that additionally to its supposed ability to improve electron transport in the ETC, the therapeutic efficacy associated with CoQ₁₀ in the treatment of mitochondrial disease may also be the result of its potent lipid soluble antioxidant capacity [78].

In light of their increased absorption, the use of both gel and oil-based formulations has been recommended over the use of conventional tablet formulations in patients with ETC disorders [90]. In a study by Martinefski, Samassa [91] it was reported that liquid emulsion improved the bioavailability of CoQ₁₀ with respect to solid formulations. Currently, there is considerable debate as to whether formulations of ubiquinol rather than the ubiquinone form of CoQ₁₀ have a better absorption from the gastrointestinal tract (GI). It has been reported that absorption of ubiquinol is three to four times higher than CoQ₁₀ in the GI [91,92]. However, because during absorption in the GI CoQ₁₀ undergoes reduction to ubiquinol, the suggested increased bioavailability of the ubiquinol formulations may be due to the matrix in which the ubiquinol is held in rather than its redox state.

5. Conclusion

A number of studies have highlighted evidence of mitochondrial dysfunction in the pathophysiology of MS, PD and ETC disorders. Thus, in view of CoQ₁₀'s significant role in the ETC, it may be deemed a potential therapeutic strategy in response to the aforementioned disorders. However, in some cases CoQ₁₀ has demonstrated an inability to elicit significant neurological outcome in MS, PD and ETC disorders and this has been discussed in the present review.

Although there is an abundance of evidence indicating the ability of CoQ₁₀ to slow and even limit disease pathogenesis, there are certain diseases in which CoQ₁₀ supplementation has only demonstrated a limited therapeutic benefit to patients. The factors that may contribute to the limited therapeutic efficacy of CoQ₁₀ are at present uncertain, but the paucity of hospital centers that are able to assess the endogenous CoQ₁₀ status of patients and therefore indicate evidence of an underlying CoQ₁₀ deficiency should be taken into consideration. Furthermore, CoQ₁₀ assessments are most often determined based on serum or plasma surrogates, which may not reflect endogenous levels. This may influence the ability of CoQ₁₀ supplementation to impact the progression of the disease, because the severity of the CoQ₁₀ deficiency may be masked by the use of inappropriate surrogates for endogenous CoQ₁₀ determination. In addition, a late diagnosis of a CoQ₁₀ deficiency may result in irreversible disease progression and a limited response to CoQ₁₀ supplementation. Other important factors to consider which may influence the clinical response to CoQ₁₀ supplementation are the type of CoQ₁₀ formulation employed and the dosage used, as these factors will influence the absorption and bioavailability of CoQ₁₀. Furthermore, the use of synthetic analogues of CoQ₁₀ capable of passing through the BBB may also be a consideration.

In conclusion, it is currently uncertain why CoQ₁₀ supplementation may be ineffective in the treatment of the diseases discussed in this review and further studies may be required in order to establish appropriate quinone treatment protocols.

6. Expert opinion

In the present review, the inability of CoQ₁₀ to augment significant improvements in the pathophysiology and clinical

outcome of three disorders of CNS was discussed. In some cases, CoQ₁₀ treatment appeared to elicit little or no improvement on disease pathophysiology; however, this in part may reflect the paucity of information currently available and the limited number of studies assessing the therapeutic efficacy of CoQ₁₀ in the treatment of CNS disorders which is certainly the case for MS [93]. Furthermore, the inclusion of supplementary markers of disease severity should also be considered in clinical studies such as, magnetic resonance imaging data to enable an accurate assessment of the effect of CoQ₁₀ supplementation on cerebral activity in these CNS disorders.

The aforementioned diseases are accompanied by evidence of mitochondrial dysfunction and oxidative stress [39,43]. Additionally, some studies have reported evidence of tangible improvements in mitochondrial functioning post treatment with CoQ₁₀ [46]. Thus, in view of the important role CoQ₁₀ plays as an electron carrier in the ETC and as a potent antioxidant, we cannot at present exclude the potential for CoQ₁₀ to elicit significant enhancement of mitochondrial functioning in the CNS disorders discussed in this review [15,45]. However, a lack of improved neurological outcome post treatment with CoQ₁₀ may be attributable to the limited ability of CoQ₁₀ to cross the BBB [94]. There is still uncertainty about the transports of CoQ₁₀ across the BBB in humans and to date, no studies to date have yet assessed the transport of CoQ₁₀ across the human BBB. Animal studies, however, have shown a degree of CoQ₁₀ transport across the BBB. In a study by Matthews, Yang [95] a 30% increase in cerebral cortex CoQ₁₀ and coenzyme Q9 (CoQ₉; predominant ubiquinone species in rat) was reported following oral supplementation of 12-month-old Sprague-Dawley rats with CoQ₁₀ (200 mg/kg) for 2 months. Additionally, in a study by Smith, Matson [96] it was reported high-dose supplementation (1000–5000 mg/kg) led to a significant increase in CoQ₁₀ concentrations in the brain of mice models. Yet, it is unclear whether this increase in CoQ₁₀ status following supplementation would be sufficient to replenish an underlying cerebral CoQ₁₀ deficiency. An *in vitro* study assessing the transport of CoQ₁₀ across the BBB has reported that the uptake of CoQ₁₀ via the Receptor for Advanced glycation end products (RAGE) and SR-B1 (scavenger Receptor) correlates with an efflux via the low-density lipoprotein receptor (LDLR) transporters, demonstrating that CoQ₁₀ transcytosis occurs in both directions, resulting in no 'net' transport across the BBB [94]. In view of this, administration of LDLR inhibitors or the stimulation of the luminal activity of SR-B1 transporters in conjunction with CoQ₁₀ supplementation may be an appropriate method to enhance the CNS uptake of exogenous CoQ₁₀. Alternatively, the short-chain analog of CoQ₁₀, idebenone which has been reported to readily cross the BBB [97] should also be considered as an alternative means of targeting the ETC dysfunction and oxidative stress associated with the CNS disorder discussed in this review. Idebenone administration has been reported to improve oxidative metabolism in patients with the CNS mitochondrial syndrome, MELAS [98]. Therefore, idebenone may be an appropriate alternative to CoQ₁₀ in the treatment of MS, PD and ETC disorders, although it should be noted that this therapy may be disease dependent. Furthermore, idebenone treatment has been reported to induce oxidative stress in

in vitro studies and exacerbate disease presentation in a patient with a primary deficiency in CoQ₁₀ biosynthesis and therefore caution should always be exercised before replacing CoQ₁₀ with idebenone in therapeutic regime of patients [15].

Factors such as the time of disease diagnosis, dosage, administration, and duration of CoQ₁₀ therapy may have a significant influence on the efficacy of this treatment. For example, the studies discussed in this review generally lacked long-term follow-up reports that exceeded 3-months to identify significant changes with CoQ₁₀ intervention over longer periods of time [58]. Thus, we cannot at present exclude the possibility that there may be possible long-term beneficial effects associated with CoQ₁₀ therapy. In addition, the studies discussed used a range of CoQ₁₀ dosages; however, at present there is no consensus on the appropriate dosage of CoQ₁₀ required to elicit significant clinical benefit to the patient. Doses as high as 2400 mg/day are being administered in the treatment of PD [99]. However, doses exceeding this level may result in a block of GI absorption and split dosing is recommended for patient administration [100,101]. Additionally, primary CoQ₁₀ deficiencies if diagnosed at the early stage of the disease have been widely reported to be responsive to CoQ₁₀ supplementation with accompanying clinical improvement being observed [102]. However, the stage of the disease may be important when CoQ₁₀ supplementation is initiated, as if this occurs after irreversible organ damage has occurred no therapeutic efficacy may be elicited even in cases of primary CoQ₁₀ deficiency [10].

It should also be noted that evidence of an underlying deficit in CoQ₁₀ status has been observed among some patients with ETC disorders and PD [103,104]. Thus, it would be anticipated that these patients may be more responsive to CoQ₁₀ supplementation, and this should be considered upon administration. In view of this, the level of CoQ₁₀ in patients should be assessed prior to administration to determine which patients may benefit from CoQ₁₀ intervention. Interestingly, a deficit in serum COQ₁₀ levels was not reported in MS patients [105]. This may justify why MS patients in some cases do not respond to this therapy [7]. In addition to this, the stage of the disease progression when CoQ₁₀ is administered may be of importance, as patients with early-stage disease (prior to irreversible CNS damage) may be more responsive to CoQ₁₀.

Other dietary preferences and supplementation taken in response to MS, PD and ETC disorders may influence the absorption of CoQ₁₀. For example, high-dose vitamin E administration may compete with CoQ₁₀ for GI absorption [106], resulting in lower plasma CoQ₁₀ levels and these factors should all be taken into consideration prior to supplementing CoQ₁₀ in these disorders. The beneficial effect of CoQ₁₀ on these disorders although not clearly demonstrated cannot at present be excluded at this juncture. Further studies are required addressing the points raised in this section before the therapeutic potential of CoQ₁₀ in the treatment of MS, PD and ETC disorders can be finally discounted.

Funding

This paper was not funded.

Declaration of interest

N Turton has received a PhD research grant funded by VitaFlo Ltd UK. Apart from those disclosed, the authors have no other relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript.

Reviewer disclosure

Peer reviewers on this manuscript have no relevant financial or other relationships to disclose.

References

1. Crane FL. Isolation of a quinone from beef heart mitochondria. *Biochim Biophys Acta*. 1957;25:220–221.
2. Crane FL. Biochemical functions of coenzyme Q10. *J Am Coll Nutr*. 2001;20(6):591–598.
3. Hargreaves IP. Ubiquinone: cholesterol's reclusive cousin. *Ann Clin Biochem*. 2003;40(3):207–218.
4. Acosta MJ, Fonseca LV, Desbats MA, et al. Coenzyme Q biosynthesis in health and disease. *Biochimica et Biophysica Acta (BBA)-Bioenergetics*. 2016;1857(8):1079–1085. .
5. Rodick TC, Seibels DR, Babu JR, et al. Potential role of coenzyme Q 10 in health and disease conditions. *Nutrit Diet Suppl*. 2018;10:1–11.
6. Heaton RA, Heales S, Rahman K, et al. The Effect of Cellular Coenzyme Q(10) Deficiency on Lysosomal Acidification. *J Clin Med*. 2020 Jun 19;9(6):6.
7. Sanoobar M, Eghtesadi S, Azimi A, et al. Coenzyme Q10 supplementation ameliorates inflammatory markers in patients with multiple sclerosis: a double blind, placebo, controlled randomized clinical trial. *Nutr Neurosci*. 2015 May;18(4):169–176. .
8. Chai C, Lim K-L. Genetic insights into sporadic Parkinson's disease pathogenesis. *Curr Genomics*. 2013;14(8):486–501.
9. Quinzii C, Naini A, Salviati L, et al. A mutation in para-hydroxybenzoate-polyprenyl transferase (COQ2) causes primary coenzyme Q10 deficiency. *Am J Hum Genet*. 2006;78(2):345–349. .
10. Duncan AJ, Bitner-Glindzicz M, Meunier B, et al. A nonsense mutation in COQ9 causes autosomal-recessive neonatal-onset primary coenzyme Q10 deficiency: a potentially treatable form of mitochondrial disease. *Am J Hum Genet*. 2009;84(5):558–566. .
11. Neergheen V, Hargreaves I. Secondary coenzyme Q10 deficiency: causes and consequence. Coenzyme Q10-uses, health effects and role in disease. Nova Science Publishers. New York; 2018. p. 89–111.
12. Potgieter M, Pretorius E, Pepper MS. Primary and secondary coenzyme Q10 deficiency: the role of therapeutic supplementation. *Nutr Rev*. 2013 Mar;71(3):180–188.
13. Salviati L, Trevisson E, Doimo M, et al. Primary coenzyme Q10 deficiency. *GeneReviews*[®]. University of Washington, Seattle, Seattle (WA); 2017.
14. Yubero D, Montero R, Martín MA, et al. Secondary coenzyme Q10 deficiencies in oxidative phosphorylation (OXPHOS) and non-OXPHOS disorders. *Mitochondrion* 2016;30:51–58.
15. Hargreaves IP. Coenzyme Q10 as a therapy for mitochondrial disease. *Int J Biochem Cell Biol*. 2014 Apr;49:105–111.
16. Coenzyme MD. Q10 and cardiovascular disease: an overview. *Br J Cardiol*. 2015;22(4):1–7.
17. Mantle D. Coenzyme Q10 supplementation for diabetes and its complications: an overview. *Brit J Diabet*. 2017;17(4):145–148.
18. Hargreaves I, Mantle D, Milford D. Chronic kidney disease and coenzyme Q10 supplementation. *Journal of Kidney Care*. 2019;4(2):82–90.
19. Mantle D, Hargreaves I. Coenzyme Q10 and degenerative disorders affecting longevity: an overview. *Antioxidants*. 2019;8(2):44.

20. Hargreaves IP, Duncan AJ, Heales SJ, et al. The Effect of HMG-CoA Reductase Inhibitors on Coenzyme Q 10. *Drug Saf.* 2005;28(8):659–676. .
21. Hargreaves I, Heaton RA, Mantle D. Disorders of human coenzyme q10 metabolism: an overview. *Int J Mol Sci.* 2020;21(18):6695.
22. Hargreaves IP, Duncan AJ, Heales SJ, et al. The effect of HMG-CoA reductase inhibitors on coenzyme Q10: possible biochemical/clinical implications. *Drug Saf.* 2005;28(8):659–676.
23. Dutta R, Trapp BD. Mechanisms of neuronal dysfunction and degeneration in multiple sclerosis. *Prog Neurobiol.* 2011;93(1):1–12.
24. Korn T, Reddy J, Gao W, et al. Myelin-specific regulatory T cells accumulate in the CNS but fail to control autoimmune inflammation. *Nat Med.* 2007;13(4):423–431. .
25. Engelhardt B. Molecular mechanisms involved in T cell migration across the blood–brain barrier. *J Neural Transm.* 2006 [2006 April 01];113(4):477–485. .
26. Bălașa R, Barcutean L, Balasa A, et al. THE action of TH17 cells on blood brain barrier in multiple sclerosis and experimental autoimmune encephalomyelitis. *Human Immunol.* 2020 February 01;81(5):81.
27. Oliveira SR, Kallaur AP, Simão AN, et al. Oxidative stress in multiple sclerosis patients in clinical remission: association with the expanded disability status scale. *J Neurol Sci.* 2012 Oct 15;321(1–2):49–53.
28. Ferguson B, Matyszak MK, Esiri MM, et al. Axonal damage in acute multiple sclerosis lesions. *Brain.* 1997;120(3):393–399. .
29. Lassmann H. Multiple Sclerosis Pathology. *Cold Spring Harb Perspect Med.* 2018;8(3):a028936.
30. Kurnellas MP, Donahue KC, Elkabes S. Mechanisms of neuronal damage in multiple sclerosis and its animal models: role of calcium pumps and exchangers. *Biochem Soc Trans.* 2007;35(Pt 5):923–926.
31. Lublin FD, Reingold SC. Defining the clinical course of multiple sclerosis. *Resul Int Survey.* 1996;46(4):907–911.
32. Michailidou I, De Vries HE, Hol EM, et al. Activation of endogenous neural stem cells for multiple sclerosis therapy. *Front Neurosci.* 2015;8:454.
33. Lublin FD, Reingold SC, Cohen JA, et al. Defining the clinical course of multiple sclerosis. *The 2013 Rev.* 2014;83(3):278–286.
34. Polman CH, Reingold SC, Banwell B, et al. Diagnostic criteria for multiple sclerosis: 2010 Revisions to the McDonald criteria. *Ann Neurol.* 2011;69(2):292–302. .
35. Mir F, Lee D, Ray H, et al. CSF isoprostane levels are a biomarker of oxidative stress in multiple sclerosis. *Neurol Neuroimmunol Neuroinflamm.* 2014;1(2):e21. .
36. Dutta R, McDonough J, Yin X, et al. Mitochondrial dysfunction as a cause of axonal degeneration in multiple sclerosis patients. *Ann Neurol.* 2006 Mar;59(3):478–489. .
37. Lazzarino G, Amorini A, Eikelenboom M, et al. Cerebrospinal fluid ATP metabolites in multiple sclerosis. *Mult Scler J.* 2010;16(5):549–554. .
38. Dendrou CA, Fugger L, Friese MA. Immunopathology of multiple sclerosis. *Nat Rev Immunol.* 2015 Sep 15;15(9):545–558.
39. Lu F, Selak M, O'Connor J, et al. Oxidative damage to mitochondrial DNA and activity of mitochondrial enzymes in chronic active lesions of multiple sclerosis. *J Neurol Sci.* 2000 [2000 August 15];177(2):95–103. .
40. Sadeghian M, Mastrolia V, Rezaei Haddad A, et al. Mitochondrial dysfunction is an important cause of neurological deficits in an inflammatory model of multiple sclerosis. *Sci Rep.* 2016 [2016 September 14];6(1):33249. .
41. Ng X, Sadeghian M, Heales S, et al. Assessment of Mitochondrial Dysfunction in Experimental Autoimmune Encephalomyelitis (EAE) Models of Multiple Sclerosis. *International Journal of Molecular Sciences.* 2019 Oct 9;20(20):20.
42. Kushnareva Y, Murphy AN, Andreyev A. Complex I-mediated reactive oxygen species generation: modulation by cytochrome c and NAD(P)⁺ oxidation-reduction state. *Biochem J.* 2002;368(Pt 2):545–553.
43. Choi IY, Lee P, Adany P, et al. In vivo evidence of oxidative stress in brains of patients with progressive multiple sclerosis. *Mult Scler.* 2018 Jul;24(8):1029–1038. .
44. Armon-Omer A, Neuman H, Sharabi-Nov A, et al. Mitochondrial activity is impaired in lymphocytes of MS patients in correlation with disease severity. *Mult Scler Relat Disord.* 2020 [2020 June 01];41:102025.
45. Littarru GP, Tiano L. Bioenergetic and antioxidant properties of coenzyme Q10: recent developments. *Mol Biotechnol.* 2007 Sep;37(1):31–37.
46. Sanoobar M, Eghtesadi S, Azimi A, et al. Coenzyme Q10 supplementation reduces oxidative stress and increases antioxidant enzyme activity in patients with relapsing-remitting multiple sclerosis. *Int J Neurosci.* 2013 Nov;123(11):776–782. .
47. Tasset I, Agüera E, Sánchez-López F, et al. Peripheral oxidative stress in relapsing–remitting multiple sclerosis. *Clin Biochem.* 2012 [2012 April 01];45(6):440–444. .
48. Baud O, Greene AE, Li J, et al. Glutathione Peroxidase-Catalase Cooperativity Is Required for Resistance to Hydrogen Peroxide by Mature Rat Oligodendrocytes. *J Neurosci.* 2004;24(7):1531–1540. .
49. Kumar A, Kaur H, Devi P, et al. Role of coenzyme Q10 (CoQ10) in cardiac disease, hypertension and Meniere-like syndrome. *Pharmacol Ther.* 2009 [2009 December 01];124(3):259–268. .
50. Farrokhi M, Jahanbani-Ardakani H, Eskandari N, et al. Cerebrospinal Fluid and Serum Markers of Inflammation in Patients with Multiple Sclerosis. *Adv Neuroimmun Biol.* 2017;6(3–4):149–152. .
51. Göbel K, Ruck T, Meuth SG. Cytokine signaling in multiple sclerosis: lost in translation. *Mult Scler J.* 2018;24(4):432–439.
52. Soleimani M, Jameie SB, Barati M, et al. Effects of coenzyme Q10 on the ratio of TH1/TH2 in experimental autoimmune encephalomyelitis model of multiple sclerosis in C57BL/6. *Iran Biomed J.* 2014;18(4):203–211. .
53. Wahl SM, Chen W. Transforming growth factor-beta-induced regulatory T cells referee inflammatory and autoimmune diseases. *Arthritis Res Ther.* 2005;7(2):62–68.
54. Souza PPC, Brechter AB, Reis RI, et al. IL-4 and IL-13 inhibit IL-1β and TNF-α induced kinin B1 and B2 receptors through a STAT6-dependent mechanism. *Br J Pharmacol.* 2013;169(2):400–412. .
55. Hart PH, Vitti GF, Burgess DR, et al. Potential antiinflammatory effects of interleukin 4: suppression of human monocyte tumor necrosis factor alpha, interleukin 1, and prostaglandin E2. *Proc Natl Acad Sci U S A.* 1989 May;86(10):3803–3807. .
56. Xu J, Wang Y, Jiang H, et al. TGF-β in Mice Ameliorates Experimental Autoimmune Encephalomyelitis in Regulating NK Cell Activity. *Cell Transplant.* 2019 Sep-Oct;28(9–10):1155–1160. .
57. Casella G, Colombo F, Finardi A, et al. Extracellular Vesicles Containing IL-4 Modulate Neuroinflammation in a Mouse Model of Multiple Sclerosis. *Mol Ther.* 2018 Sep 5;26(9):2107–2118.
58. Moccia M, Capacchione A, Lanzillo R, et al. Coenzyme Q10 supplementation reduces peripheral oxidative stress and inflammation in interferon-β1a-treated multiple sclerosis. *Ther Adv Neurol Disord.* 2019;12:1756286418819074.
59. Greeke EE, Chua AS, Healy BC, et al. Depression and fatigue in patients with multiple sclerosis. *J Neurol Sci.* 2017 Sep ;15(380):236–241. .
60. Sanoobar M, Dehghan P, Khalili M, et al. Coenzyme Q10 as a treatment for fatigue and depression in multiple sclerosis patients: a double blind randomized clinical trial. *Nutr Neurosci.* 2016 [2016 March 15];19(3):138–143. .
61. Mizuno K, Tanaka M, Nozaki S, et al. Antifatigue effects of coenzyme Q10 during physical fatigue. *Nutrition.* 2008 [2008 April 01];24(4):293–299. .
62. Al Shahrani M, Heales S, Hargreaves I, et al. Oxidative stress: mechanistic insights into inherited mitochondrial disorders and Parkinson's disease. *J Clin Med.* 2017;6(11):100. .
63. Villalba JM, Parrado C, Santos-Gonzalez M, et al. Therapeutic use of coenzyme Q10 and coenzyme Q10-related compounds and formulations. *Expert Opin Investig Drugs.* 2010;19(4):535–554. .

64. Shults CW, Oakes D, Kiebertz K, et al. Effects of coenzyme Q10 in early Parkinson disease: evidence of slowing of the functional decline. *Arch Neurol.* 2002;59(10):1541–1550. .
65. Beal MF, Oakes D, Shoulson I, et al. A randomized clinical trial of high-dosage coenzyme Q10 in early Parkinson disease: no evidence of benefit. *JAMA Neurol.* 2014;71(5):543–552. .
66. Damier P, Hirsch E, Agid Y, et al. The substantia nigra of the human brain: II. Patterns of Loss of Dopamine-containing Neurons in Parkinson's Disease. *Brain* 1999;122(8):1437–1448.
67. Snow BJ, Rolfe FL, Lockhart MM, et al. A double-blind, placebo-controlled study to assess the mitochondria-targeted antioxidant MitoQ as a disease-modifying therapy in Parkinson's disease. *Mov Disord.* 2010;25(11):1670–1674. .
68. Jenner P, Dexter D, Sian J, et al. Oxidative stress as a cause of nigral cell death in Parkinson's disease and incidental Lewy body disease. *Ann Neurol.* 1992;32(5):S82–S87. .
69. Dauer W, Przedborski S. Parkinson's disease: mechanisms and models. *Neuron.* 2003;39(6):889–909.
70. Morais VA, Haddad D, Craessaerts K, et al. PINK1 loss-of-function mutations affect mitochondrial complex I activity via Ndufa10 ubiquinone uncoupling. *Science.* 2014;344(6180):203–207. .
71. Duberley KE, Abramov AY, Chalasani A, et al. Human neuronal coenzyme Q10 deficiency results in global loss of mitochondrial respiratory chain activity, increased mitochondrial oxidative stress and reversal of ATP synthase activity: implications for pathogenesis and treatment. *J Inherit Metab Dis.* 2013;36(1):63–73. .
72. Muthukumar K, Leahy S, Harrison K, et al. Orally delivered water soluble Coenzyme Q 10 (Ubisol-Q 10) blocks on-going neurodegeneration in rats exposed to paraquat: potential for therapeutic application in Parkinson's disease. *BMC Neurosci.* 2014;15(1):1–11. .
73. Sikorska M, Lanthier P, Miller H, et al. Nanomicellar formulation of coenzyme Q10 (Ubisol-Q10) effectively blocks ongoing neurodegeneration in the mouse 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine model: potential use as an adjuvant treatment in Parkinson's disease. *Neurobiol Aging.* 2014;35(10):2329–2346. .
74. Murphy MP, Smith RA. Targeting antioxidants to mitochondria by conjugation to lipophilic cations. *Annu Rev Pharmacol Toxicol.* 2007;47(1):629–656.
75. Zaki NM. Strategies for oral delivery and mitochondrial targeting of CoQ10. *Drug Deliv.* 2016;23(6):1868–1881.
76. Park HW, Park CG, Park M, et al. Intrastriatal administration of coenzyme Q10 enhances neuroprotection in a Parkinson's disease rat model. *Sci Rep.* 2020;10(1):1–12. .
77. Matalonga L, Arias A, Coll MJ, et al. Treatment effect of coenzyme Q (10) and an antioxidant cocktail in fibroblasts of patients with Sanfilippo disease. *J Inherit Metab Dis.* 2014 May;37(3):439–446.
78. Neerghen V, Chalasani A, Wainwright L, et al. Coenzyme Q10 in the treatment of mitochondrial disease. *J Inborn Errors Metabol Screen.* 2017;5:2326409817707771.
79. Haack TB, Danhauser K, Haberberger B, et al. Exome sequencing identifies ACAD9 mutations as a cause of complex I deficiency. *Nat Genet.* 2010;42(12):1131. .
80. Chen R-S, Chin-Chang H, Chu N-S. Coenzyme Q10 treatment in mitochondrial encephalomyopathies. *Eur Neurol.* 1997;37(4):212.
81. Kerr DS. Review of clinical trials for mitochondrial disorders: 1997–2012. *Neurotherapeutics.* 2013;10(2):307–319.
82. Lin C-M TP. Valproic acid aggravates epilepsy due to MELAS in a patient with an A3243G mutation of mitochondrial DNA. *Metab Brain Dis.* 2007;22(1):105–109.
83. Maloney PC, Kashket E, Wilson TH. A proton motive force drives ATP synthesis in bacteria. *Proc Nat Acad Sci.* 1974;71(10):3896–3900.
84. Bresolin N, Doriguzzi C, Ponzetto C, et al. Ubidecarenone in the treatment of mitochondrial myopathies: a multi-center double-blind trial. *J Neurol Sci.* 1990;100(1–2):70–78. .
85. Glover EI, Martin J, Maher A, et al. A randomized trial of coenzyme Q10 in mitochondrial disorders. *Muscle Nerve.* 2010;42(5):739–748.
86. Žmitek J, Šmidovnik A, Fir M, et al. Relative bioavailability of two forms of a novel water-soluble coenzyme Q10. *Ann Nutr Metab.* 2008;52(4):281–287. .
87. Stacpoole PW, deGrauw TJ, Feigenbaum AS, et al. Design and implementation of the first randomized controlled trial of coenzyme CoQ₁₀ in children with primary mitochondrial diseases. *Mitochondrion* 2012 Nov;12(6):623–629. .
88. Sacconi S, Trevisson E, Salviati L, et al. Coenzyme Q10 is frequently reduced in muscle of patients with mitochondrial myopathy. *Neuromuscul Disord.* 2010;20(1):44–48. .
89. Sato Y, Kobayashi H, Higuchi T, et al. Metabolomic Profiling of Pompe Disease-Induced Pluripotent Stem Cell-Derived Cardiomyocytes Reveals That Oxidative Stress Is Associated with Cardiac and Skeletal Muscle Pathology. *Stem Cells Transl Med.* 2017 Jan;6(1):31–39. .
90. Bhagavan HN, Chopra RK. Plasma coenzyme Q10 response to oral ingestion of coenzyme Q10 formulations. *Mitochondrion* 2007;7:578–588.
91. Martinefski M, Samassa P, Buontempo F, et al. Relative bioavailability of coenzyme Q10 formulation for paediatric individualized therapy. *J Pharm Pharmacol.* 2017;69(5):567–573. .
92. García-Corzo L, Luna-Sánchez M, Doerrier C, et al. Ubiquinol-10 ameliorates mitochondrial encephalopathy associated with CoQ deficiency. *Biochim Biophys Acta Mol Basis Dis.* 2014;1842(7):893–901. .
93. Ömerhoça S, Akkaş SY, İçen NK. Multiple Sclerosis: diagnosis and Differential Diagnosis. *Noro Psikiyatı Ars.* 2018;55(1):S1–S9.
94. Wainwright L, Hargreaves IP, Georgian AR, et al. CoQ10 Deficient Endothelial Cell Culture Model for the Investigation of CoQ10 Blood–Brain Barrier Transport. *J Clin Med.* 2012;321(1–2):3236. .
95. Matthews RT, Yang L, Browne S, et al. Coenzyme Q10 administration increases brain mitochondrial concentrations and exerts neuroprotective effects. *Proc Nat Acad Sci.* 1998;95(15):8892–8897. .
96. Smith KM, Matson S, Matson WR, et al. Dose ranging and efficacy study of high-dose coenzyme Q10 formulations in Huntington's disease mice. *Biochim Biophys Acta Mol Basis Dis.* 2006;1762(6):616–626. .
97. Erb M, Hoffmann-Enger B, Deppe H, et al. Features of idebenone and related short-chain quinones that rescue ATP levels under conditions of impaired mitochondrial complex I. *PLoS One.* 2012;7(4):e36153–e36153. .
98. Ikejiri Y, Mori E, Ishii K, et al. Idebenone improves cerebral mitochondrial oxidative metabolism in a patient with MELAS. *Neurology* 1996 Aug;47(2):583–585. .
99. Shults CW, Flint Beal M, Song D, et al. Pilot trial of high dosages of coenzyme Q10 in patients with Parkinson's disease. *Exp Neurol.* 2004 Aug;188(2):491–494. .
100. Bhagavan HN, Chopra RK. Plasma coenzyme Q10 response to oral ingestion of coenzyme Q10 formulations. *Mitochondrion.* 2007 [2007/06];7:578–88.
101. Miles MV, Patterson BJ, Schapiro MB, et al. Coenzyme Q10 Absorption and Tolerance in children with down syndrome: a Dose-Ranging Trial. *Pediatr Neurol.* 2006 [2006 July 01];35(1):30–37. .
102. Hargreaves I. Coenzyme Q10 as a therapy for mitochondrial disease. *Int J Biochem Cell Biol.* 2014;49:105–111.
103. Moreno-Fernández AM, Cordero MD, Garrido-Maraver J, et al. Oral treatment with amitriptyline induces coenzyme Q deficiency and oxidative stress in psychiatric patients. *J Psychiatr Res.* 2012 [2012 March 01];46(3):341–345. .
104. Hargreaves IP, Lane A, Sleiman PMA. The coenzyme Q10 status of the brain regions of Parkinson's disease patients. *Neurosci Lett.* 2008 [2008 December 05];447(1):17–19. .
105. De Bustos F, Jiménez-Jiménez FJ, Molina JA, et al. Serum levels of coenzyme Q10 in patients with multiple sclerosis. *Acta Neurol Scand.* 2000;101(3):209–211. .
106. Kaikkonen J, Nyssönen K, Tomasi A, et al. Antioxidative efficacy of parallel and combined supplementation with coenzyme Q10 and d- α -Tocopherol in mildly hypercholesterolemic subjects: a randomized placebo-controlled clinical study. *Free Radic Res.* 2000 [2000 January 01];33(3):329–340. .