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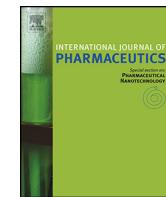
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Investigation on the aerosol performance of dry powder inhalation hypromellose capsules with different lubricant levels

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ABSTRACT

HPMC capsules are made by a dipping process and a surface lubricant for the mould pins is an essential processing aid for removing dried capsules shells. For the purpose of this study, the level was determined by quantifying methyloleate (MO) a component found in the lubricant but not in the hypromellose capsules. Here we investigated the influence of the lubricant, low ($10.81 \mu\text{g}/\text{capsule} = 60 \text{ mg/kg MO}$), medium ($15.97 \mu\text{g}/\text{capsule} = 90 \text{ mg/kg MO}$) and high ($23.23 \mu\text{g}/\text{capsule} = 127 \text{ mg/kg MO}$) content on powder (binary mixture of salbutamol: lactose, 1:50 w/w) aerosolization properties was investigated. Results indicated significantly lower emitted dose from capsules with 60 mg/kg MO. Furthermore, the 90 and 127 mg/kg MO level of lubricant capsules produced almost double the Fine Particle Dose & Fine Particle Fraction compared with the low level of lubricant. The data indicates that lubricant level within capsules has an influence on deposition profiles and amount of drug remaining in capsule and inhaler device after actuation. It is suggested lubricant levels greater than 60 mg/kg MO per capsule are required to minimise powder retention within capsules and maximise deposition profiles. AFM (atomic force microscopy) data suggest that internal surface roughness may be related with this phenomena.

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1. Introduction

Delivery of therapeutic agents *via* the pulmonary route has gained increasing applications for lung diseases such as asthma and COPD. Pulmonary delivery has many advantages including delivery of medication directly to site of action, bypassing first pass metabolism in the liver (Geller, 2009; Labiris and Dolovich, 2003), it is non-invasive and can achieve therapeutic outcome at lower doses than administration *via* the oral route (Smith and Parry-Billings, 2003).

Dry powder inhaler (DPI) are able to deliver low and high doses within the range of 5–500 µg, do not require co-ordination between actuation and inspiration as with pMDI (Kaialy et al., 2012). They have been developed since the 1960's for a range of conditions such as asthma and COPD using short and long acting beta agonists, anti-cholinergic agents and corticosteroids drugs in order to facilitate drug administration to the lungs *via* the inhalation route (Atkins, 2005). Today there are currently more than twenty commercially available DPI, both active and passive (Chan et al., 2014). New active DPI incorporate additional

mechanisms within the device to aid the fluidization of the powder from the device and reduce the reliance on the patient's inspiratory force. These mechanisms include vibration mesh which oscillates upon the patient's inhalation, others include release of the powder formulation only when the patient has achieved the correct inspiratory force (Chan et al., 2014). Passive DPI have unit doses of drug in either blister packs or capsules, which contain the drug and a carrier, e.g. lactose, and drug deposition relies on the patient's inspiratory force to de-aggregate the drug from the carrier (Chan et al., 2014; Kaialy et al., 2012; Zhou and Morton, 2012).

The powder mass in the capsules allows flexibility for the administration of low and high dose drugs within the range of 5 to 500 µg. Examples of capsule based devices include the single unit HandiHaler® (Boehringer-Ingelheim) (Islam and Gladki, 2008), TOBI® Podhaler™ (tobramycin) and Colobreathe® Turbospin® for delivery of large doses (Claus et al., 2014), Breezhaler® (Novartis), (Young et al., 2014) and novel multiple pre-metered unit-dose Flowcaps® (Hovione) that contains up to 20 capsules (Friebel and Steckel, 2010). These devices are simple to use, cost-effective and can administer low and high doses. In addition, the capsule based devices improve patient compliance, as they can provide feedback to the patient in the form of a rattling sound, indicating correct

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49 inhalation flow rate was achieved and passed through the device to
50 **Q4** deliver the correct dose (Behara et al., 2014; Smith et al., 2010).
51 Moreover, the patient can visually check the capsule to determine
52 if the dose has been administered (Smith et al., 2010).

53 Hypromellose (HPMC) is used to make inhalation grade
54 capsules (Quali-V®-I) for use in DPI, as it is unaffected by moisture
55 content changes (Jones, 2008). Hence, it does not become brittle as
56 it loses moisture, a common phenomenon with gelatin capsules as
57 patients do not store them as directed resulting in broken capsules
58 and poor performance in their DPI (Nagata, 2002; Ogura et al.,
59 1998; Renswouw et al., 2010). In addition, it has also been shown
60 that HPMC capsules are less influenced by triboelectrification
61 which is common with gelatin capsules (Nakate et al., 2005). Inhalation
62 grade HPMC capsules are made from different grades of
63 raw material chosen for their puncturing properties (Torrisi et al.,
64 2013) and as a result have a slightly higher moisture content; 4.5–
65 6.5% compared to 4.0–6.0% in oral pharmaceutical grade capsules.
66 The HPMC Capsules are manufactured by dipping stainless steel
67 mould pins at room temperature into a warm solution of
68 hypromellose containing carrageenan as a network former and
69 potassium chloride as a network promoter (Jones, 2004). The
70 change in temperature causes the HPMC solution to gel and form a
71 film on the surface of the mould pins. The films are dried by passing
72 groups of pins through a series of drying kilns in which large
73 volumes of air at controlled temperature and humidity is blown
74 over them. As the films dry they shrink on to the pins. To remove
75 them without damage it is essential for the mould pins to be coated
76 with a surface lubricant to act as a release aid. Capsules cannot be
77 manufactured without this lubricant (Jones, 2008). However, a
78 search of the literature only shows one study investigating the
79 influence of the amount of mould lubricant on the internal surfaces
80 of capsules in relation to the aerosolization properties of powders
81 from a capsule based DPI (Saim and Horhota, 2002). Furthermore,
82 this study relates to gelatin capsules and not HPMC.

83 The lubricant is a mixture of food and pharmaceutical grade
84 materials registered with regulatory authorities and the composition
85 is proprietary for each capsule manufacturer. Hence for
86 quantitative analysis it is necessary to choose a component of the
87 lubricant which is not found in the HPMC capsules. In this study we
88 chose methyloleate (MO) as a marker for lubricant content
89 consisting of free fatty acids together with their esters. A number
90 of sample preparation methods have been proposed in the
91 literature to convert free fatty acids into their esters, such as
92 silylation (Woo and Kim, 1999) or reaction with alkyl chloroformates
93 (Gimeno-Adelantado et al., 2001) as well as for trans-
94 esterification of the triglycerides (Mason and Waller, 1964).

95 The aim of this study was to investigate the aerosolization
96 properties of dry powder formulations composed of inhalation
97 grade lactose and micronized salbutamol, filled in to size 3HPMC
98 inhalation grade capsules manufactured with 3 different lubricant
99 levels via an 8-pin inhaler device. Size 3HPMC capsules was chosen
100 because this is the size used in the pharmaceutical industry for
101 development of capsule-based DPI. For example the last significant
102 developments in inhalation capsule-based devices, Ultibro and
103 Seebri Breezhaler, incorporate their respective dry powder
104 formulation into a size 3 capsule. This size capsule (0.8 mg/mL)
105 has a powder fill weight of 225 mg. Furthermore, to the best of our
106 knowledge, the capsule inner surface lubricant content has not
107 been determined by GCMS or its distribution in HPMC capsules by
108 AFM. Hence, we describe a new technique with results obtained
109 using these methods in this study.

2. Material and methods

2.1. Materials

112 8-pin monodose inhaler was provided by Plastiape S.p.a Italy.
113 Hypromellose (HPMC) inhalation grade capsules, size 3 (Quali-V®-
114 I) for this inhaler, manufactured using three different lubricant
115 levels (satisfactory physical quality capsules were made at each
116 level); low ($10.81 \mu\text{g}/20 \text{ mg}$ of blended powder within capsule =
117 60 mg/kg MO), medium ($15.97 \mu\text{g}/20 \text{ mg}$ of blended powder
118 within capsule = 90 mg/kg MO) and high ($23.23 \mu\text{g}/20 \text{ mg}$ of
119 blended powder within capsule = 127 mg/kg MO) were obtained
120 from Qualicaps® Europe, S.A.U, Spain. Inhalation grade lactose
121 (Respitose) was supplied by DFE Pharma, The Netherlands.
122 Micronized salbutamol was obtained from Lusochimica, Spain.
123 Methanol and 1-heptane sulphonic acid sodium salt were
124 purchased from Sigma, UK. Methyloleate analytical standard,
125 hexane and chloroform were from Sigma-Aldrich (St. Louis USA).
126 1,2,3 Trichlorobenzene was purchased from Fluka and was used as
127 internal standard. Trimethylsulfonium hydroxide solution,
128 (TMSH), 0.25 M in methanol was used for GC derivatization.

2.2. Determination of methyl oleate (MO) in capsules by gas chromatography mass spectrometry

131 Gas chromatography mass spectrometry, GCMS, is the tech-
132 nique most suitable for its qualitative and quantitative determina-
133 tion after derivatization and extraction into an organic solvent
134 (Driscoll et al., 2009; Sutherland, 2007; Zhang et al., 2014).
135 Capsules inner lubricant content was evaluated by determining
136 MO which was taken as a marker of the lubricant content using
137 GCMS. Eleven HPMC capsules were weighed in a glass vial and
138 5 mL of Hexane: chloroform, 60:40 (v:v) extraction solvent
139 containing 10 mg/L of the internal standard was added. The vial
140 was sonicated for 1 h in an ultrasonic bath; then 100 μL of the
141 extract was transferred into a 2 mL vial for derivatization using
142 50 μL of TMSH. The MO was identified by MS (Mass spectrometry)
143 and was quantified using an internal calibration method with six
144 points in the 0.5–20 mg/kg concentration range. 1 μL of the
145 derivatized MO was injected in split less mode in the GCMS
146 instrument.

2.3. Preparation of inhalation grade lactose & powder mix

148 Inhalation grade lactose and powder mix were prepared
149 according to previously published method (Saleem et al., 2008)
150 with slight modifications. Inhalation grade lactose was fractionated
151 by sieving with a sieve stack (250, 125, 90, 63, and 45 μm) using
152 vibration amplitude of 40 for 10 min and collected on a 90 μm
153 sieve to be used in all subsequent studies. Micronized salbutamol
154 sulphate and lactose were mixed in a ratio of 1:50 (w/w) via
155 geometric dilution to obtain a 2% binary blend. The formulations
156 were blended with a Turbula® orbital mixer (Glen Mills, Clifton,
157 New Jersey) for 30 min at 46 rpm. The blend uniformity was
158 determined by randomly selecting five 20 mg samples, and
159 formulations were considered uniform when the coefficient of
160 variation (% CV) was $\leq 6\%$. Samples were analyzed using high-
161 performance liquid chromatography (HPLC) method below (Section
162 2.4). Once blend uniformity was achieved $20 \pm 1 \text{ mg}$ of
163 blended powder was manually loaded into HPMC capsules (size 3)
164 with different lubricant levels (low, medium and high) and stored
165 in a humidity chamber (Sanyo Atmos Chamber) at 22°C and 40%
166 RH for 2 weeks (Nine HPMC capsules were filled for each lubricant
167 level at weeks 1 and 2).

168

2.4. In vitro aerosolization performance

169 For each lubricant level, three capsules (20 ± 1 mg of powder
 170 loaded into HPMC capsules (size 3)) corresponding to a dose of
 171 $408 \mu\text{g}$ were dispersed through a 8-pin DPI inhaler into a next
 172 generation cascade impactor (NGI; MSP Corporation, Shoreview,
 173 MN) at a flow rate of 60 L min^{-1} actuated for 4 s, with 15 mL of
 174 mobile phase added to the pre-separator. This was repeated three
 175 times ($n=3$). Drug depositing in the capsule, inhaler, mouthpiece
 176 adaptor, induction port, pre-separator and NGI stages were
 177 collected by rinsing each component with mobile phase. This
 178 was repeated at week 2 and the drug content was assessed via
 179 HPLC method (Section 2.4).

180 The emitted dose (ED) was calculated as the total mass of drug
 181 depositing in the mouthpiece, induction port, pre-separator, and
 182 NGI stages. The fine particle dose (FPD) was determined as the
 183 mass of drug deposited in the NGI with aerodynamic diameters \leq
 184 $4.46 \mu\text{m}$. The percentage fine particle fraction (% PPF) of each dose
 185 was the ratio of the drug mass depositing in the NGI (aerodynamic
 186 diameter $\leq 4.46 \mu\text{m}$) over the emitted dose. Mass median
 187 aerodynamic diameter (MMAD) was calculated by subjecting
 188 the inertial impaction data to log-probability analysis.

189 2.5. Chemical analysis

190 Capsules internal lubricant analysis were carried out by gas
 191 chromatography coupled to mass spectrometry, GCMS, using a GC
 192 7890 (Agilent Technologies, Palo Alto, CA, USA) and a 5975C
 193 quadrupole mass spectrometer (Agilent Technologies, TX, USA). A
 194 Supelcowax 10 ($30 \text{ m} \times 0.25 \text{ mm} \times 0.25 \mu\text{m}$ film thickness) fused
 195 silica capillary column was used. The injection port temperature
 196 was 240°C and the oven temperature program changed from 40°C
 197 to 240°C at $10^\circ\text{C}/\text{min}$.

198 Salbutamol sulphate was analyzed by HPLC (Agilent Technologies)
 199 using a Kinetex C18 column ($50 \times 4.6 \text{ mm}$ i.d. packed with
 200 $2.6 \mu\text{m}$, phenomenex, UK). The mobile phase consisted of
 201 methanol and 0.25% (w/v) 1-heptane sulphonic acid sodium salt
 202 (45:55 v/v), the flow rate was 1 mL/min , injection volume $10 \mu\text{L}$,
 203 temperature 25°C and wavelength was 200 nm . The retention time

204 for salbutamol sulphate was 3 min and the limits of detection and
 205 quantification were 0.60 and $1.12 \mu\text{g/mL}$ respectively.

206 2.6. Morphology of inner capsule surface

207 Atomic force microscopy, AFM, is a widely used technique for
 208 micro- and nanoscale material characterization creating a three
 209 dimensional image of a physical surface (García and Pérez, 2002).
 210 A commonly employed measurement approach is based on
 211 tapping-mode AFM which involves a short and pulsed contact
 212 between the tip of an oscillating micro-cantilever and the sample
 213 surface. Vibrations of the cantilever tip are induced through dither
 214 piezo oscillations from which heights and phases are monitored for
 215 imaging purposes (García, 2010).

216 The AFM (atomic force microscopy) experiments were made
 217 using a multimode Nanoscope III A (Bruker) in tapping mode in
 218 order to access surface topography. It is equipped with three
 219 scanners of 1, 1.5 and $150 \mu\text{m}$. Small pieces of $5 \times 5 \text{ mm}$ were cut and
 220 placed onto the Nanoscope probe. The instrument standard sample
 221 capsule probe was made of stainless steel but it was not suitable to
 222 handle the curved shape of a capsule; so, a special home-made
 223 support device was designed. Topographic measurements of the
 224 inner capsules surface at a fixed scanning angle equal to zero were
 225 made using tapping mode (intermittent contact mode). The
 226 cantilever/tip assembly was sinusously vibrated by a piezo device
 227 mounted above it, and the oscillating tip slightly taped the surface
 228 at the resonant frequency of the cantilever with constant
 229 oscillating amplitude introduced in the vertical direction with a
 230 feedback loop keeping the average normal force constant.
 231 Measurements were made using a silicon probe (Veeco probe)
 232 with a spring constant of 5 N/m and a resonance frequency of
 233 150 kHz . All experiments were performed in air at ambient
 234 conditions. In order to stabilize thermally the piezo driver, the
 235 machine was turned on two hours before use. During each
 236 measurement a $15 \times 15 \mu\text{m}$ surface was covered using a FESP tip.

237 2.7. Statistical analysis

238 The data obtained were analyzed statistically by one-way
 239 analysis of variance (ANOVA) with the Tukey's comparison using

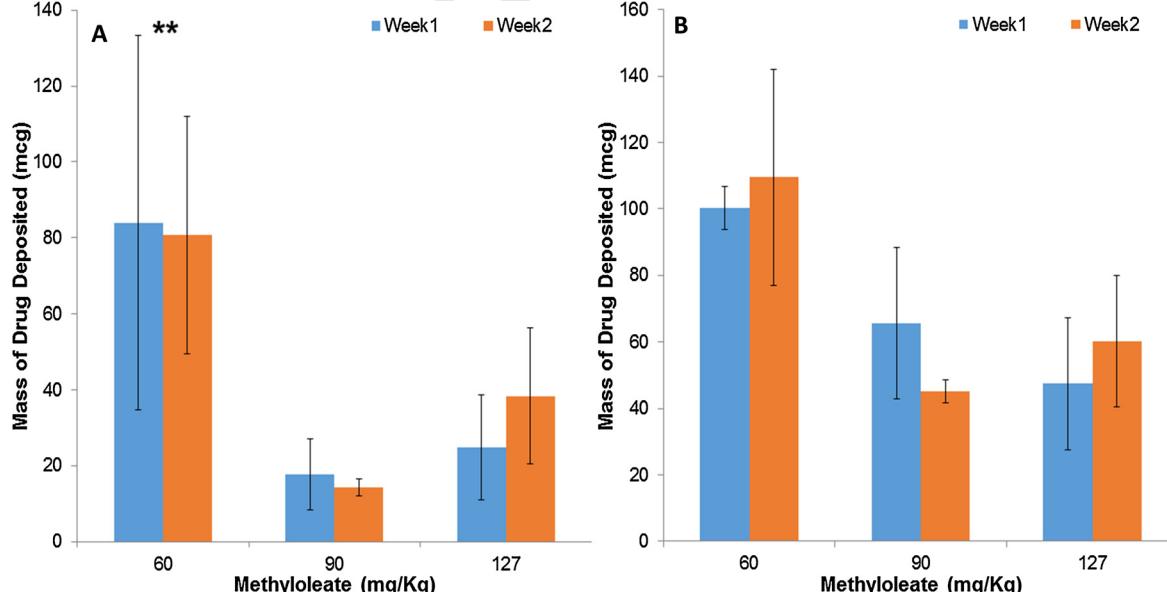


Fig. 1. Deposition of salbutamol sulphate remaining in capsules (A) and device (B) following aerosolisation at 60 L min^{-1} from a 8-pin inhaler (Mean \pm SD, $n=3$) ** $P < 0.05$ (ANOVA/Tukey's) Low (60 mg/kg MO) versus medium (90 mg/kg MO) & high (127 mg/kg MO) lubricant levels at weeks 1 and 2.

240 Minitab 17 Statistical Software® (Minitab Inc., PA, USA). Statistical
 241 significance were considered when $p < 0.05$. All values are
 242 expressed as the mean \pm standard deviation.

243 3. Results and discussion

244 3.1. In vitro aerosolization performance

245 3.1.1. Comparing capsules and device

246 Comparing HPMC capsules (Fig. 1A) the results clearly indicate
 247 a significantly larger salbutamol retention in the low lubricant
 248 capsule (week 1: $84.01 \pm 49.23 \mu\text{g}$, week 2: $80.76 \pm 31.25 \mu\text{g}$)
 249 compared to the medium (week 1: $17.70 \pm 9.34 \mu\text{g}$, week 2:
 250 $14.37 \pm 2.20 \mu\text{g}$) and high lubricant capsules (week 1:
 251 $24.91 \pm 13.79 \mu\text{g}$, week 2: $38.36 \pm 17.89 \mu\text{g}$) ($p < 0.05$, ANOVA/
 252 Tukey's). However, there was no significant difference regarding
 253 salbutamol retention between medium and high lubricant
 254 capsules at week 1 and 2. A similar trend was observed with
 255 drug deposition remaining in the 8-pin inhaler (Fig. 1A). It is
 256 evident from the data that lubricant level within the capsule is
 257 important, with data suggesting lubricant levels between 90 and
 258 127 mg/kg MO per capsule result in significantly lower drug
 259 deposition within the capsule and inhaler device. The high
 260 salbutamol retention in low lubricant capsules occurred, because
 261 during the removal of the capsules from the mould pins there is a
 262 high degree of adhesion that causes roughness, which can be seen
 263 by the mountain and deep valleys, as shown by AFM (see
 264 Section 3.2). Hence during inhalation the salbutamol particles
 265 become entrapped or lodged within these peaks and crevices
 266 (Saim and Horhota, 2002). However, this effect is reduced
 267 significantly as the level of lubricant increases to greater than
 268 90 mg/kg MO lubricant per capsule. This results in a smooth inner
 269 surface of the capsule with less mountain and valleys (see
 270 Section 3.2) as there is less adhesion during the removal from the

271 moulding pins, and hence, reduction in contact between particles
 272 and crevices (Ibrahim et al., 2000; Saim and Horhota, 2002).

273 3.1.2. Comparing emitted dose, fine particle dose, fine particle fraction 274 & MMAD

275 Fig. 2 shows the ED, FPD, FPF and MMAD of salbutamol
 276 aerosolized from an 8-pin inhaler at 60 L/min. The ED is
 277 significantly lower for the low lubricant level capsules (week 1:
 278 $223.73 \pm 42.72 \mu\text{g}$, week 2: $217.69 \pm 63.85 \mu\text{g}$) compared to the
 279 medium (week 1: $324.57 \pm 32.06 \mu\text{g}$, week 2: $2348.42 \pm 1.17 \mu\text{g}$) and
 280 high lubricant capsules (week 1: $335.65 \pm 33.70 \mu\text{g}$, week 2:
 281 $309.35 \pm 37.67 \mu\text{g}$) ($p < 0.05$, ANOVA/Tukey's). These results were
 282 repeated for fine particle dose (μg) (Fig. 2B) and fine particle
 283 fraction (%) (Fig. 2C) where the values are almost twice that
 284 obtained using low lubricant capsules ($p < 0.05$, ANOVA/Tukey's).
 285 This coincides with the high deposition of salbutamol remaining in
 286 the low lubricant capsules (Fig. 1A) and the device (Fig. 1A).
 287 Furthermore, the MMAD (μm) (Fig. 2D) is significantly greater
 288 from low lubricant capsules (week 1: $3.37 \pm 0.78 \mu\text{m}$, week 2:
 289 $2.71 \pm 0.03 \mu\text{m}$) compared to medium (week 1: $2.13 \pm 0.11 \mu\text{m}$,
 290 week 2: $1.78 \pm 0.57 \mu\text{m}$) and high (week 1: $2.12 \pm 0.16 \mu\text{m}$, week 2:
 291 $1.65 \pm 0.17 \mu\text{m}$) ($p < 0.05$, ANOVA/Tukey's). This also confirms the
 292 low FPD and FPF within the lungs, due to large particle size and
 293 hence less drug depositing within the deep lungs.

294 3.2. AFM studies

295 Topographic plots from three capsule inner surfaces are
 296 represented in Fig. 3. They were taken as examples of the twelve
 297 capsules analyzed, which were selected at a low, medium and high
 298 concentrations that were previously determined by GCMS (Driscoll
 299 et al., 2009; Sutherland, 2007; Zhang et al., 2014).

300 As can be seen, the topographic images show a mountain and
 301 deep valley distribution of the lubricant oil in the inner surface of
 302 the capsules. Similar results have been obtained using gelatin

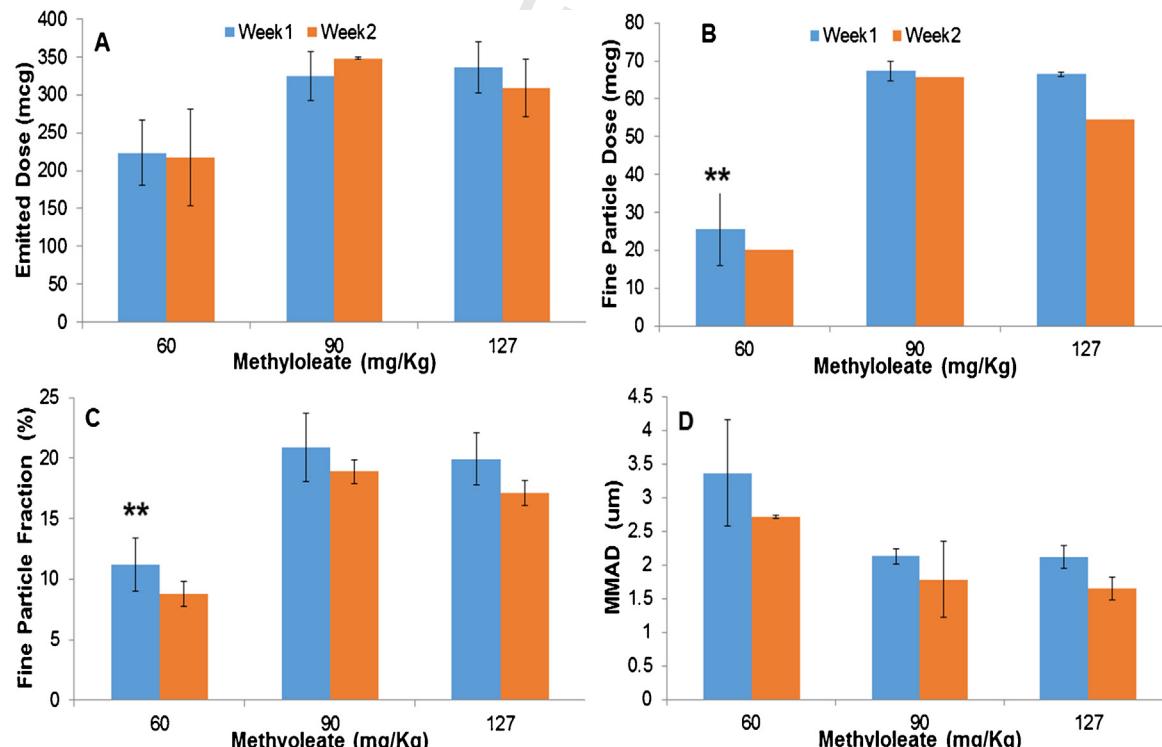


Fig. 2. Emitted dose (μg) (A), Fine particle dose (μg) (B), Fine particle fraction (%) (C), MMAD (μm) (D) of salbutamol sulphate following aerosolisation at 60 L min^{-1} from an 8-pin inhaler (Mean \pm SD, $n = 3$). ** $P < 0.05$ (ANOVA/Tukey's) Low (60 mg/kg MO) versus medium (90 mg/kg MO) & high (127 mg/kg MO) lubricant levels at weeks 1 and 2.

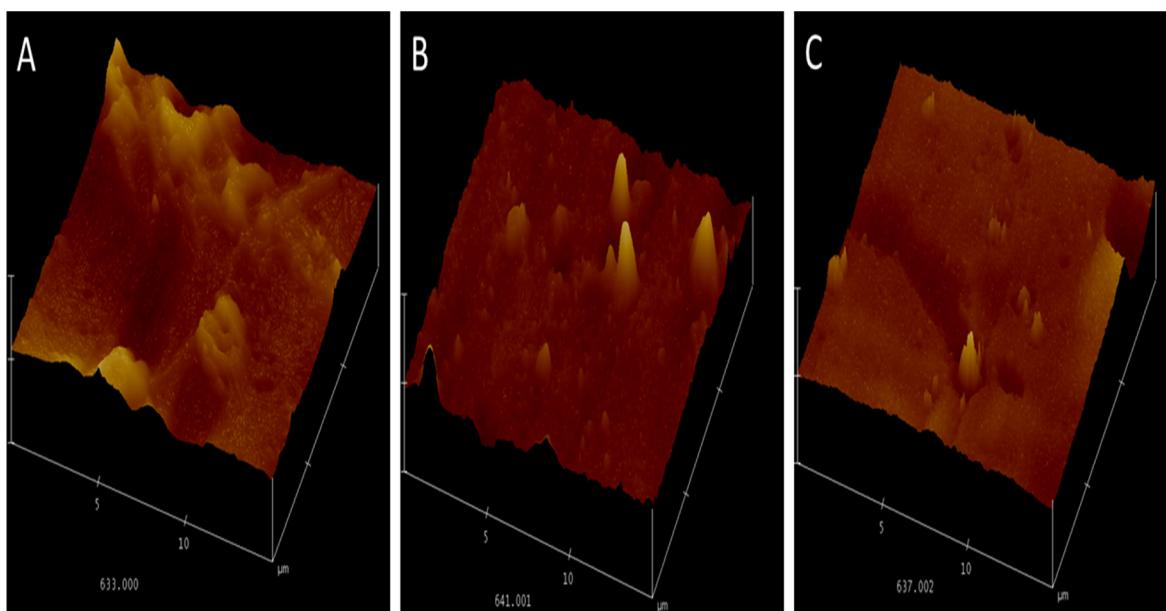


Fig. 3. Topographical image of capsules inner surface containing (A) low (26 mg/kg MO), (B) medium (63 mg/kg MO) and (C) high (137 mg/kg MO) levels of methyloleate.

capsules (Ibrahim et al., 2005). When the MO amount increased, the inner surface appeared to be more homogenous (*i.e.* reduced mountain and deep valley distribution) (Fig. 3A and C).

Four parameters derived from these plots (roughness, depth, particle height and grain height) (Fig. 4) were used to obtain better knowledge regarding inner capsule lubricant distribution. The average from three different points on each capsule surface was used to plot the different parameters versus MO concentration. It is apparent (Fig. 4a) when the MO concentration increases, the roughness, represented by Ra, decreased, indicating that

homogeneity of capsule surface is higher when it is covered more completely with the lubricant.

Reproducibility of the capsule inner surface can be evaluated by paying attention to two capsules groups containing three capsules with similar MO concentration, approximately 59 and 104 mg/kg respectively as indicated by the red circle in Fig. 4A. The reproducibility of the Ra parameter increases when MO concentration is higher. This seems to indicate that homogeneity of the lubricant inner surface is also enhanced when its concentration is higher.

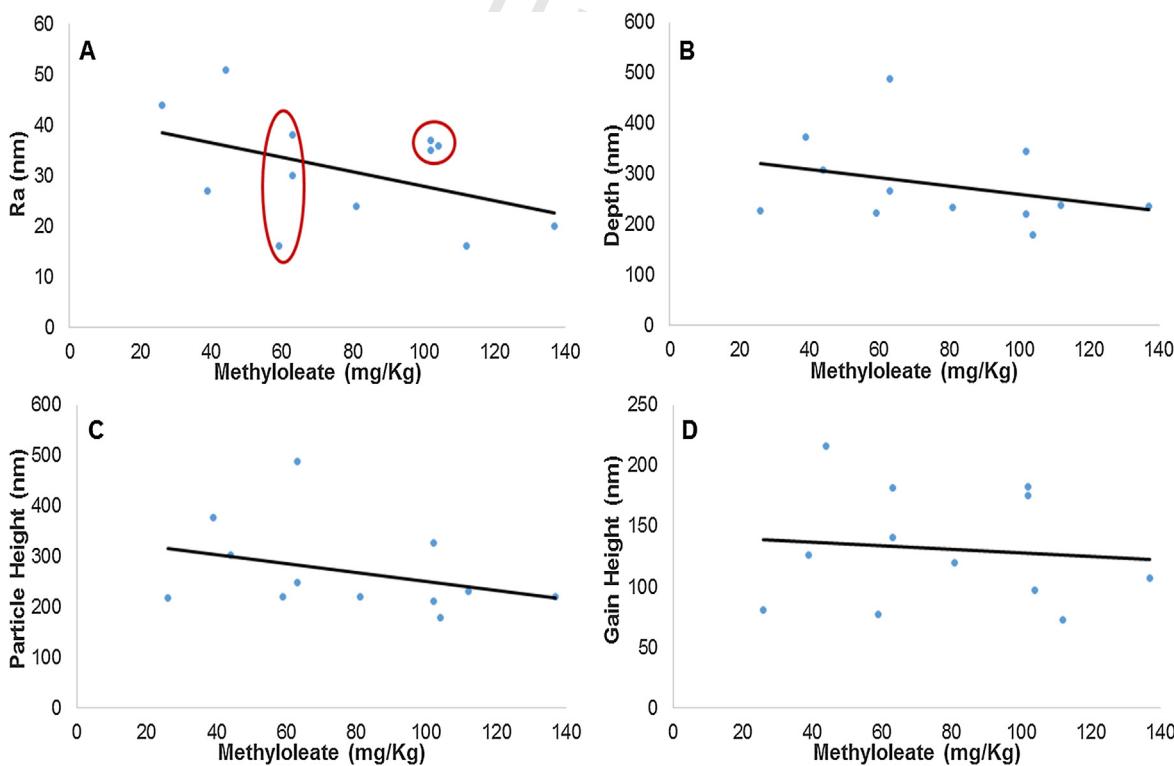


Fig. 4. Methyloleate concentration effect on (A) roughness, (B) Depth, (C) particle height and (D) grain height.

Similarly, the average depth, particle height and height gain also decreased when MO concentration increased (Fig. 4B–D), which confirmed the surface decreased when MO concentration increases. In fact this tendency is also apparent in Fig. 4A–D, in which dispersion of the respective parameters through an average straight line decreases when the inner lubricant concentration increases.

330 4. Conclusion

331 The study clearly indicates that the lubricant level inside
332 capsules has an influence on deposition profiles, amount of drug
333 remaining in capsule and inhaler device after actuation. The results
334 obtained suggest lubricant levels greater than 10.81 µg per capsule
335 (60 mg/kg MO) are beneficial in decreasing drug deposition from
336 capsules in an 8-pin inhaler device, while more than doubling the
337 fine particle dose and fraction. It seems this effect is related to the
338 capsule internal surface roughness. Measurements with AFM
339 indicated that homogeneity of the internal capsule surface is
340 higher when the inner lubricant concentration increases.

341 Conflict of interest and role of funding source

342 Q7 This work was funded by Qualicaps Europe S.A.U., Alcobendas
343 (Madrid), Spain. They were not involved in the study design,
344 collection, analysis or interpretation of data, but did have an input
345 in checking the manuscript before submission.

346 References

- 347 Atkins, P.J., 2005. Dry powder inhalers: an overview. *Respir. Care* 50, 1304–1312.
348 Behara, S.R., Longest, P.W., Farkas, D.R., Hindle, M., 2014. Development and
349 comparison of new high-efficiency dry powder inhalers for carrier-free
350 formulations. *J. Pharm. Sci.* 103, 465–477.
351 Chan, J.G., Wong, J., Zhou, Q.T., Leung, S.S., Chan, H.K., 2014. Advances in device and
352 formulation technologies for pulmonary drug delivery. *AAPS PharmSciTech* 15,
353 882–897.
354 Claus, S., Weiler, C., Schiewe, J., Friess, W., 2014. How can we bring high drug doses to
355 the lung? *Eur. J. Pharm. Biopharm.* 86, 1–6.
356 Driscoll, D.F., Ling, P.R., Bistrian, B.R., 2009. Pharmacopeial compliance of fish oil-
357 containing parenteral lipid emulsion mixtures: Globule size distribution (GSD)
358 and fatty acid analyses. *Int. J. Pharm.* 379, 125–130.
359 Friebel, C., Steckel, H., 2010. Single-use disposable dry powder inhalers for
360 pulmonary drug delivery. *Expert Opin. Drug Deliv.* 7, 1359–1372.
361 García, R., 2010. Theory of Amplitude Modulation AFM, Amplitude Modulation
362 Atomic Force Microscopy. Wiley-VCH Verlag GmbH & Co. KGaA, pp. 41–57.
363 Garcí'a, R., Pérez, R., 2002. Dynamic atomic force microscopy methods. *Surf. Sci. Rep.*
364 47, 197–301.
365 Geller, D.E., 2009. Aerosol antibiotics in cystic fibrosis. *Respir. Care* 54, 658–670.
366 Gimeno-Adelantado, J.V., Mateo-Castro, R., Domenech-Carbo, M.T., Bosch-Reig, F.,
367 Domenech-Carbo, A., Casas-Catalan, M.J., Osete-Cortina, L., 2001. Identification
368 of lipid binders in paintings by gas chromatography. Influence of the pigments. *J.*
369 *Chromatogr. A* 922, 385–390.
370 Ibrahim, T.H., Burk, T.R., Etzler, F.M., Neuman, R.D., 2000. Direct adhesion
371 measurements of pharmaceutical particles to gelatin capsule surfaces. *J. Adhes. Sci. Technol.* 14, 1225–1242.
372 Ibrahim, T.H., Burk, T.R., Etzler, F.M., Neuman, R.D., 2005. Direct adhesion
373 measurements of pharmaceutical particles to gelatin capsules. In: Drellich, J.,
374 Mittal, K.L. (Eds.), *Atomic Force Microscopy in Adhesion Studies*. VSP, Leiden,
375 Boston, pp. 137–154.
376 Islam, N., Gladki, E., 2008. Dry powder inhalers (DPIs)—a review of device reliability
377 and innovation. *Int. J. Pharm.* 360, 1–11.
378 Jones, B.E., 2004. Manufacture and properties of two-piece hard capsules, In:
379 odczeck, F., Jones, B.E. (Eds.), *Pharmaceutical Capsules*. 2nd ed. Pharmaceutical
380 Press, London, pp. 79–100.
381 Jones, B.E., 2008. The evolution of DPI capsules. *Inhalation* 2, 20–23.
382 Kaialy, W., Alhalaweh, A., Velaga, S.P., Nokhodchi, A., 2012. Influence of lactose
383 carrier particle size on the aerosol performance of budesonide from a dry
384 powder inhaler. *Powder Technol.* 227, 74–85.
385 Labiris, N.R., Dolovich, M.B., 2003. Pulmonary drug delivery. Part I: physiological
386 factors affecting therapeutic effectiveness of aerosolized medications. *Br. J. Clin.
387 Pharmacol.* 56, 588–599.
388 Mason, M.E., Waller, G.R., 1964. Dimethoxyp propane induced transesterification of
389 fats and oils in preparation of methyl esters for gas chromatographic. *Anal. Chem.* 36, 583.
390 Nagata, S., 2002. Advantages to HPMC capsules: a new generation's hard capsule.
391 *Drug Deliv. Technol.* 2, 34–39.
392 Nakate, T., Yoshida, H., Ohike, A., Tokunaga, Y., Ibuiki, R., Kawashima, Y., 2005.
393 Formulation development of inhalation powders for FK888 using the E-haler to
394 improve the inhalation performance at a high dose, and its absorption in
395 healthy volunteers. *Eur. J. Pharm. Biopharm.* 59, 25–33.
396 Ogura, T., Furuya, Y., Matsuura, S., 1998. HPMC capsules—an alternative to gelatin.
397 *Pharm. Technol. Eur.* 10, 32–42.
398 Renswouw, D.C., Laarhoven, A.C., Haren, M.J., Bouvy, M.L., Weda, M., 2010. Storage
399 instructions for inhalation capsules: consequences of incorrect storage and
400 adherence in daily practice. *J. Pharm. Pract.* 23, 548–552.
401 Saim, S., Horhota, S.T., 2002. Process for overcoming drug retention in hard gelatin
402 inhalation capsules. *Drug Dev. Ind. Pharm.* 28, 641–654.
403 Saleem, I., Smyth, H., Telko, M., 2008. Prediction of dry powder inhaler formulation
404 performance from surface energetics and blending dynamics. *Drug Dev. Ind.
405 Pharm.* 34, 1002–1010.
406 Smith, I.J., Parry-Billings, M., 2003. The inhalers of the future? A review of dry
407 powder devices on the market today. *Pulm. Pharmacol. Ther.* 16, 79–95.
408 Smith, I.J., Bell, J., Bowman, N., Everard, M., Stein, S., Weer, J.C., 2010. Inhaler device
409 what remainto be done? *J. Aerol. Med. Pulm. Drug Deliv.* 37 (Suppl. 2), S25–S37.
410 Sutherland, K., 2007. Derivatisation using m-(trifluoromethyl)
411 phenyltrimethylammonium hydroxide of organic materials in artworks for
412 analysis by gas chromatography–mass spectrometry: unusual reaction
413 products with alcohols. *J. Chromatogr. A* 1149, 30–37.
414 Torrisi, B.M., Birchall, J.C., Jones, B.E., Diez, F., Coulman, S.A., 2013. The development
415 of a sensitive methodology to characterise hard shell capsule puncture by dry
416 powder inhaler pins. *Int. J. Pharm.* 456, 545–552.
417 Woo, K.L., Kim, J.I., 1999. New hydrolysis method for extremely small amount of
418 lipids and capillary gas chromatographic analysis as N(O)-tert-
419 butyldimethylsilyl fatty acid derivatives compared with methyl ester
420 derivatives. *J. Chromatogr. A* 862, 199–208.
421 Young, D., Wood, L., Singh, D., Dederichs, J., 2014. The history and performance of the
422 Breezhaler device. In: Trifilieff, A. (Ed.), *Indacaterol*. Springer, Basel, pp. 117–128.
423 Zhang, X.J., Huang, L.L., Su, H., Chen, Y.X., Huang, J., He, C., Li, P., Yang, D.Z., Wan, J.B.,
424 2014. Characterizing plasma phospholipid fatty acid profiles of polycystic ovary
425 syndrome patients with and without insulin resistance using GC-MS and
426 chemometrics approach. *J. Pharm. Biomed. Anal.* 95, 85–92.
427 Zhou, Q.T., Morton, D.A., 2012. Drug-lactose binding aspects in adhesive mixtures:
428 controlling performance in dry powder inhaler formulations by altering lactose
429 carrier surfaces. *Adv. Drug Deliv. Rev.* 64, 275–284.