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Pharmacological profile of methylphenidate-based designer drugs

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Running title: methylphenidate-like designer drugs

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Abstract

Background: Methylphenidate-based designer drugs are new psychoactive substances (NPS) that are used outside medical settings and their pharmacology is largely unexplored. The aim of the present study was to characterize the pharmacology of methylphenidate-based substances in vitro.

Methods: We determined the potencies of the methylphenidate-based NPS N-benzylethylphenidate, 3,4-dichloroethylphenidate, 3,4-dichloromethylphenidate, ethynaphthidate, ethylphenidate, 4-fluoromethylphenidate, isopropylphenidate, 4-methylmethylphenidate, methylmorphenate, and propylphenidate and the potencies of the related compounds cocaine and modafinil with respect to norepinephrine, dopamine, and serotonin transporter inhibition in transporter-transfected human embryonic kidney 293 cells. We also investigated monoamine efflux and monoamine receptor and transporter binding affinities. Furthermore, we assessed the cell integrity under assay conditions.

Results: All methylphenidate-based substances inhibited the norepinephrine and dopamine transporters 4 to >1,000-fold more potently than the serotonin transporter. Similar to methylphenidate and cocaine, methylphenidate-based NPS did not elicit transporter-mediated efflux of monoamines. Besides binding to monoamine transporters, several test drugs had affinity for adrenergic, serotonergic, and rat trace amine-associated receptors but not for dopaminergic or mouse trace amine-associated receptors. No cytotoxicity was observed after drug treatment at assay concentrations.

Conclusion: Methylphenidate-based substances had pharmacological profiles similar to methylphenidate and cocaine. The predominant actions on dopamine transporters vs. serotonin transporters may be relevant when considering abuse liability.
1. Introduction

The psychostimulant methylphenidate (MPH; Ritalin®) is used for the treatment of attention-deficit/hyperactivity disorder and narcolepsy but it also has a history of being misused as a ‘smart drug’ and ‘cognitive enhancer’ (Arria et al., 2008; Liakoni et al., 2015; Maier et al., 2013). In recent years, an increasing number of MPH-based new psychoactive substances (NPS; Fig. 1) (Brandt et al., 2014) have become available as alternatives to MPH (Bailey et al., 2015; European Monitoring Centre for Drugs and Drug Addiction, 2015) and have been associated with several fatalities (Krueger et al., 2014; Maskell et al., 2016; Parks et al., 2015). Characteristic for the NPS phenomenon, many of the currently circulating MPH analogs originated from drug development efforts (Deutsch et al., 1996; Markowitz et al., 2013; Misra et al., 2010), which subsequently appeared on the streets. The pharmacological and subjective-effect profiles of MPH are very similar to cocaine (Simmler et al., 2014; Vogel et al., 2016; Volkow et al., 1999). Furthermore, some of these substances are either sold in their own right or offered in the form of branded products (Bailey et al., 2015; Parks et al., 2015). Methylphenidate predominantly inhibits the norepinephrine (NE) and dopamine (DA) transporters (NET and DAT, respectively), thus, possibly contributing to its abuse
potential (Simmler et al., 2014; Vogel et al., 2016). Correspondingly, questions arise about the extent to which MPH analogs might share MPH-like characteristics. Assessing the pharmacological profile of NPS in vitro is an initial step to gain a better understanding of the potential clinical effects and toxicology of these substances. For this reason, the present study reports on the transporter interaction profiles of the MPH-related NPS N-benzylethylphenidate, 3,4-dichloroethylphenidate, 3,4-dichloromethylphenidate, and isopropylphenidate and the transporter and receptor interaction profiles of ethynaphthidate, ethylphenidate, 4-fluoromethylphenidate, 4-methylmethylphenidate, methylmorphenate, and propylphenidate. Modafinil, a stimulant prescribed for the treatment of narcolepsy, which is frequently offered for sale as a ‘neuroenhancer’ (Ghahremani et al., 2011; Maier et al., 2013; Mereu et al., 2013; Müller et al., 2013), has also been included in this investigation. Stimulants may act as transporter inhibitors or as transporter substrates that cause monoamine efflux into the synaptic cleft (Rothman and Baumann, 2003; Sitte and Freissmuth, 2015). Therefore, additionally to the transporter inhibition potencies of the substances, their mechanism of action (reuptake inhibitor or transporter substrate) was determined.

2. Material and methods

2.1. Drugs

Cocaine, 3,4-methylenedioxymethamphetamine (MDMA) and MPH were purchased from Lipomed (Arlesheim, Switzerland), with high-performance liquid chromatography (HPLC) purity > 98.5%. Modafinil was purchased from Cayman Chemicals (Ann Arbor, MI, USA), with purity > 98%. Methylmorphenate and propylphenidate were obtained from reChem Labs (Ontario, Canada) and afterwards identified and tested for purity using nuclear magnetic resonance (NMR) and HPLC, which revealed purity > 95%. N-Benzylethylphenidate, 3,4-dichloroethylphenidate, 3,4-dichloromethylphenidate,
ethyl-naphthidate, 4-fluoromethylphenidate, isopropylphenidate, and 4-
methylmethylphenidate were part of confiscations by German authorities and test purchases
(Klare et al., 2017). The substances were fully characterized in a previous study (Klare et al.,
2017) and purity values were estimated at > 95% based on spectroscopic and
chromatographic methods of analysis. Ethylphenidate was provided by Dr. Christian Bissig
(Forensic Institute, Zurich, Switzerland) after being confiscated by Swiss authorities and
being tested for purity of > 98%. Modafinil was obtained as racemic base. The other drugs
were obtained as racemic hydrochloride salts. Radiolabeled $^{3}$H-NE (13.1 Ci/mmol) and
$^{3}$H-DA (30.0 Ci/mmol) were obtained from Perkin-Elmer (Schwerzenbach, Switzerland).
Radiolabeled $^{3}$H-5-HT (80 Ci/mmol) was purchased from Anawa (Zürich, Switzerland).

2.2. Monoamine uptake transport inhibition

Monoamine uptake inhibition was assessed using human embryonic kidney (HEK) 293 cells that stably expressed the human SERT, DAT, or NET (Tatsumi et al., 1997) as previously described (Hysek et al., 2012). Briefly, the cells were cultured to 70-90% confluence, detached, and resuspended in Krebs-Ringer Bicarbonate Buffer (Sigma-Aldrich, Buchs, Switzerland). For $^{3}$H-DA uptake experiments, the uptake buffer was supplemented with 1.14 mM ascorbic acid. The cells were then treated with vehicle control and drug in the range of 1 nM to 900 µM for 10 min at room temperature. Additionally, monoamine-specific inhibitors were added (10 µM fluoxetine for SERT, 10 µM mazindol for DAT, and 10 µM nisoxetine for NET). To initiate uptake transport, $^{3}$H-5-HT, $^{3}$H-DA, or $^{3}$H-NE were added at a final concentration of 5 nM for an additional 10 min. The cells were then separated from the uptake buffer by centrifugation through silicone oil, and the tubes were frozen in liquid nitrogen. The cell pellet was cut into scintillation vials and lysed. The samples were shaken for 1 h before scintillation fluid (Ultimagold, Perkin Elmer, Schwerzenbach,
Switzerland) was added. Monoamine uptake was then quantified by liquid scintillation counting on a Packard Tri-Carb Liquid Scintillation Counter 1900 TR. Uptake in the presence of the selective inhibitors was determined to be nonspecific and subtracted from the total counts.

2.3. Transporter-mediated monoamine efflux

The potential of the drugs to initiate transporter-mediated NE, DA, or 5-HT efflux was assessed in HEK 293 cells that overexpressed the respective transporter as previously described (Simmler et al., 2013). Briefly, the cells were first preloaded with [³H]-NE, [³H]-DA, or [³H]-5-HT dissolved in Krebs-HEPES buffer for 20 min at 37°C. The cells were then washed and treated with 100 µM of the drugs for 15 min (DAT and SERT) or 45 min (NET). The treatment durations for [³H]-NE, [³H]-DA, and [³H]-5-HT efflux experiments were based on kinetic evaluation of the efflux-over-time curves of MDMA (Simmler et al., 2014). The cells were washed again, and the remaining radioactivity inside the cells was quantified. The monoamine transporter blockers citalopram (SERT), mazindol (DAT), and nisoxetine (NET) were added as a negative control at a concentration of 10 µM to determine “pseudo-efflux” that was caused by nonspecific monoamine efflux and subsequent reuptake inhibition (Scholze et al., 2000).

2.4. Radioligand receptor and transporter binding assays

The radioligand binding assays were performed as previously described for transporters (Hysek et al., 2012) and receptors (Revel et al., 2011). Briefly, membrane preparations of HEK 293 cells (Invitrogen, Zug, Switzerland) that overexpressed the respective transporters (Tatsumi et al., 1997) or receptors (human genes, with the exception of rat and mouse genes for trace amine-associated receptors [TAARs]) (Revel et al., 2011)
were incubated with the radiolabeled selective ligands at concentrations equal to \( K_d \), and ligand displacement by the compounds was measured. Specific binding of the radioligand to the target receptor was defined as the difference between the total binding and nonspecific binding that was determined in the presence of the selected competitors. The following radioligands and competitors, respectively, were used: \( N \)-methyl-[\(^3\text{H}\)]nisoxetine and 10 \( \mu \text{M} \) indatraline (NET), [\(^3\text{H}\)]citalopram and 10 \( \mu \text{M} \) indatraline (SERT), [\(^3\text{H}\)]WIN35,428 and 10 \( \mu \text{M} \) indatraline (DAT), [\(^3\text{H}\)]8-hydroxy-2-(di-\( n \)-propylamine)tetratin and 10 \( \mu \text{M} \) pindolol (5-HT\(_{1\text{A}}\) receptor), [\(^3\text{H}\)]ketanserin and 10 \( \mu \text{M} \) spiperone (5-HT\(_{2\text{A}}\) receptor), [\(^3\text{H}\)]mesulgerine and 10 \( \mu \text{M} \) mianserin (5-HT\(_{2\text{C}}\) receptor), [\(^3\text{H}\)]prazosin and 10 \( \mu \text{M} \) chlorpromazine (\( \alpha \_1\text{A} \) adrenergic receptor), [\(^3\text{H}\)]rauwolscine and 10 \( \mu \text{M} \) phentolamine (\( \alpha _{2\text{A}} \) adrenergic receptor), [\(^3\text{H}\)]spiperone and 10 \( \mu \text{M} \) spiperone (D\(_2\) receptor), and [\(^3\text{H}\)]RO5166017 and 10 \( \mu \text{M} \) RO5166017 (TAAR\(_{1}\)).

### 2.5. Activity at the serotonin 5-HT\(_{2\text{B}}\) receptor

Activity at the 5-HT\(_{2\text{B}}\) receptor was assessed as previously described (Rickli et al., 2016). Briefly, human 5-HT\(_{2\text{B}}\) receptor-expressing HEK 293 cells were incubated in a cell culture plate overnight. The next day, the growth medium was removed by snap inversion, and calcium indicator Fluo-4 solution (Molecular Probes, Eugene, OR, USA) was added to each well. The plates were then incubated for 45 min at 31\(^\circ\)C. The Fluo-4 solution was removed by snap inversion and then added a second time. The cells were then incubated for another 45 min at 31\(^\circ\)C. Immediately before testing, the cells were washed with HBSS and 20 mM HEPES (assay buffer; Gibco) using an EMBLA cell washer, and assay buffer was added. The plates were placed in a FLIPR. Test substances that were diluted in assay buffer were added online, and the increase in fluorescence was measured.

### 2.6. Cytotoxicity
Cytotoxicity was assessed with the ToxiLight bioassay kit (Lonza, Basel, Switzerland) according to the manufacturer’s protocol. The kit measures adenylate kinase release as a result of cell membrane integrity loss. Human SERT-, DAT-, and NET-transfected HEK 293 cells were treated for 1 h at room temperature with the drugs at the highest assay concentrations.

2.7. Statistical analysis

Calculations were performed using Prism 7.0a software (GraphPad, San Diego, CA, USA). Monoamine transporter inhibition data were fit by nonlinear regression to variable-slope sigmoidal dose-response curves and IC_{50} values were assessed. The DAT/SERT ratio is expressed as 1/DAT IC_{50}:1/SERT IC_{50}. Compound-induced monoamine efflux of five independent experiments was compared with negative controls using analysis of variance followed by the Holm-Sidak test. P values lower than 0.05 were considered significant and substances were considered transporter substrates if they caused significantly higher efflux than the negative controls. IC_{50} values of radioligand binding were determined by calculating nonlinear regression curves for a one-site model using three independent 10-point concentration-response curves for each compound. K_i (affinity) values, which correspond to the dissociation constants, were determined using the Cheng-Prusoff equation. Nonlinear regression concentration-response curves were used to calculate the EC_{50} values for the 5-HT_{2B} receptor activation.

3. Results

3.1. Monoamine uptake transporter inhibition

Monoamine uptake inhibition curves are shown in Fig. 2, and the corresponding IC_{50} values and DAT/SERT inhibition ratios are listed in Table 1. Methylphenidate was a potent
inhibitor of the NET and DAT at submicromolar concentrations and a weak inhibitor of the SERT. 3,4-dichloromethylphenidate inhibited the NET more than 10-fold more potently than MPH, whereas the inhibition potency for the DAT was more than 2-fold increased. The NET and DAT inhibition potencies of 3,4-dichloroethylphenidate, ethynaphthidate, 4-fluoromethylphenidate, and 4-methylmethylphenidate were similar to MPH in the range of 0.04–0.42 µM for the NET and 0.08–0.34 µM for the DAT. N-Benzylethylphenidate, ethylphenidate, isopropylphenidate, methylmorphenate, and propylphenidate inhibited the NET with 6 to 800-fold lower potency compared to MPH and the DAT with 4 to 500-fold lower potency. The SERT inhibition potency for all MPH-based NPS was lower than the NET and DAT inhibition potencies. Ethynaphthidate inhibited the SERT at 1.7 µM with a DAT/SERT ratio of 5. The remaining compounds inhibited the SERT 40 to >1000-fold weaker than the DAT and 26 to >1000-fold weaker than the NET. Modafinil was a weak inhibitor of monoamine transporters with an IC₅₀ value >10 µM for the DAT and no relevant NET or SERT inhibition (IC₅₀ values >100 µM). Unlike the MPH-based substances and modafinil, cocaine inhibited all three transporters with similar potency in the range of 0.5–1.5 µM.

3.2. Monoamine efflux

Similar to cocaine, MPH and the MPH-based NPS and related compounds did not cause monoamine efflux (Fig. 3) and are therefore not transporter substrates.

3.3. Monoamine receptor and transporter binding affinities

The interactions between MPH-based NPS and related compounds with monoamine receptors and transporters are shown in Table 2. All MPH-based NPS bound to the NET and DAT but only N-benzylethylphenidate, 3,4-dichloroethylphenidate, 3,4-
dichloromethylphenidate, and ethylnaphthidate bound to the SERT in the concentration range tested. 3,4-Dichloroethylphenidate, 3,4-dichloromethylphenidate, 4-methylmethylphenidate, and ethylnaphthidate bound to the α_{1A} receptor in the range of 1.7–6.5 μM and additionally to the α_{2A} receptor in the range of 7–10 μM. Ethylnaphthidate and propylphenidate bound to the α_{2A} receptor with 14 μM and 8.7 μM, respectively, but did not bind to the α_{1A} receptor in the investigated concentration range. 3,4-Dichloroethylphenidate, 3,4-dichloromethylphenidate, ethylnaphthidate, 4-methylmethylphenidate, and propylphenidate, had affinities of 1–17 μM for the 5-HT_{1A} receptor. Ethylnaphthidate was the only drug to bind to the 5-HT_{2A} receptor with an IC_{50} value of 4.9 μM and only 3,4-dichloromethylphenidate and ethylnaphthidate bound to the 5-HT_{2C} receptor, both with an IC_{50} of 12 μM. None of the compounds activated the 5-HT_{2B} receptor or bound to the mouse TAAR_{1}, and only 3,4-dichloroethylphenidate, 3,4-dichloromethylphenidate, and isopropylphenidate bound to the rat TAAR_{1} with affinities in the range of 6–13 μM. None of the MPH-based NPS had relevant affinity for D_{2} receptors. Modafinil and cocaine bound to the monoamine transporters but did not interact with monoamine or trace amine receptors.

3.4. Cytotoxicity

Cytotoxicity was not observed for any of the drugs in the functional assays at the concentrations tested, thus confirming cell integrity during the assays.

4. Discussion

We characterized the in vitro pharmacological profiles of MPH-based NPS and compared them with MPH and cocaine. All compounds inhibited the DAT substantially more potently than the SERT, suggesting predominantly stimulant-type effects similar to amphetamine and a high abuse liability (Liechti, 2015; Simmler et al., 2013).
4.1. Monoamine uptake transporter inhibition and monoamine efflux

Methylphenidate and MPH-based NPS, with the exception of N-benzylethylphenidate, isopropylphenidate, and methylmorphenate, inhibited the NET at submicromolar concentrations, suggesting cardiostimulant and psychostimulant properties, similar to amphetamines (Hysek et al., 2011; Simmler et al., 2013). Moreover, the NET and DAT inhibition potencies but not the SERT inhibition potency correlate with the psychotropic effective doses of psychostimulants in human (Simmler et al., 2013).

The MPH-based NPS were only monoamine transporter inhibitors and not monoamine transporter substrates, indicating a mechanism of action similar to cocaine but not amphetamines (Fleckenstein et al., 2007; Torres et al., 2003). Ethylnaphthidate inhibited the SERT at low micromolar concentrations, but the remaining MPH-based NPS displayed a clear preference for DAT over SERT, resulting in high DAT/SERT ratios frequently reported for locomotor stimulants (Simmler et al., 2013). Our results are consistent with other studies that reported potent NET and DAT inhibition for MPH (DAT/SERT ratio = 2207) and triple uptake inhibition for cocaine (DAT/SERT ratio = 3.2) (Han and Gu, 2006). Modafinil was a moderate and relatively selective DAT inhibitor, with an IC_{50} value of 11 µM. This finding is consistent with previous in vitro studies that reported IC_{50} values of 4–13 µM (Karabacak et al., 2015; Loland et al., 2012; Madras et al., 2006; Zolkowska et al., 2009). The interaction between modafinil and DAT is also thought to modulate the pharmacological effects of the drug (Wisor, 2013). The psychopharmacological profiles and cognitive-enhancing properties of MPH and modafinil may be different. Modafinil has been shown to improve attention and wakefulness, whereas MPH has been shown to improve memory (Repantis et al., 2010).

4.2. Transporter and receptor binding profiles
Compared with ethylphenidate, replacement of the benzene ring with naphthalene (ethynaphthidate) increased the potency in inhibiting the SERT and increased the affinity for 5-HT receptors. Many stimulant NPS interact with TAARs (Simmler et al., 2016); however, no potent TAAR interactions were found for MPH-based NPS. 3,4-Dichloromethylphenidate and ethynaphthidate interacted with the $\alpha_{1A}$ and 5-HT$_{1A}$ receptor in the low micromolar range. The remaining MPH-based NPS did not potently interact with monoamine receptors, indicating that they exert their primary effects by inhibiting uptake transporters, similar to MPH and cocaine (Ritz et al., 1987, 1988; Volkow et al., 2002). Consistent with the monoamine uptake data, 3,4-dichloroethylphenidate, 3,4-dichloromethylphenidate, and ethynaphthidate potently bound to the NET and DAT and had affinity for the SERT as well. 4-Fluoromethylphenidate, 4-methylmethylphenidate, and methylphenidate bound potently to the NET and DAT but had no affinity to the SERT in the tested concentration range. N-Benzylethylphenidate, ethylphenidate, isopropylphenidate, and propylphenidate showed high affinity for the DAT but not for the NET or SERT. Methylmorphenate did not potently bind to any transporter. Cocaine potently bound to all transporters but not to receptors. No interaction between modafinil and monoamine receptors was observed. To date, no single site of action for modafinil has been identified (Gerrard and Malcolm, 2007).

4.3. Comparison of transporter binding and transporter inhibition

No drug-mediated monoamine efflux was observed for any of the MPH-based compounds, strengthening the argument that they are pure uptake blockers. For uptake blockers, a correlation between the monoamine uptake and radioligand binding affinities has been previously described for the NET (Cheetham et al., 1996; Lee et al., 1982), the DAT (Javitch et al., 1984; Schoemaker et al., 1985), and the SERT (D'Amato et al., 1987; Langer et al., 1980). However, discrepancies between monoamine uptake inhibition and
radioligand binding have been observed for cocaine-like drugs and proposed for MPH-like drugs, when the conditions for the binding and uptake inhibition assays varied (Reith et al., 2005; Rothman et al., 1993).

Highest NET and DAT binding affinities were observed for the most potent NET and DAT inhibitor 3,4-dichloromethylphenidate. However, the increase in potency compared to MPH was much more pronounced with a 76-fold and 12-fold increase for NET and DAT binding, respectively. 3,4-Dichloroethylphenidate bound more than 10-fold more potently to the NET and DAT whereas the NE and DA uptake inhibition was similar to MPH. Ethynaphthidate, 4-fluoromethylphenidate, and 4-methylmethyphenidate inhibited the NET and DAT with similar potency as MPH. These substances bound to the NET with affinity in the range of 0.22–0.31 µM and to the DAT with affinity in the range of 0.026–0.040 µM. MPH bound to the NET and DAT with 0.50 µM and 0.070 µM, respectively. Thus, unlike for the dichloro substituted compounds, the IC\textsubscript{50} values and the \textit{K}\textsubscript{i} values for ethynaphthidate, 4-fluoromethylphenidate, and 4-methylmethyphenidate correlate well. \textit{N}-Benzylethylphenidate, ethylphenidate, isopropylphenidate, methylmorphenate, and propylphenidate inhibited the NET with 6 to 800-fold lower potency compared to MPH and the DAT with 4 to 500-fold lower potency. While the binding affinities for the NET were 8 to 48-fold decreased, the DAT binding affinities were decreased only for \textit{N}-benzylethylphenidate, methylmorphenate, and propylphenidate (5 to 46-fold) whereas the DAT binding affinities of ethylphenidate and isopropylphenidate were close to MPH. Remarkably, \textit{N}-benzylethylphenidate was by far the weakest transporter inhibitor, it did however not have the lowest NET and DAT binding affinities. In the investigated concentration range, only \textit{N}-benzylethylphenidate, 3,4-dichloroethylphenidate, 3,4-dichloromethylphenidate, and ethynaphthidate bound to the SERT. 3,4-dichloroethylphenidate, 3,4-dichloromethylphenidate, and ethynaphthidate, were the most
potent SERT inhibitors, N-benzylethylphenidate was however the weakest SERT inhibitor. Thus, as observed for the NET and DAT, the SERT binding affinity of N-benzylethylphenidate was much higher than might be expected from the uptake inhibition data.

To conclude, the rank order of potency of the radioligand binding and uptake inhibition was similar with the 3,4-substituted and 4-substitued compounds being among the most potent MPH-based NPS; the relative potencies of the uptake inhibition and transporter binding varied however to a certain extent.

Besides cocaine, ethylaminephthidate was the only compound to have considerable inhibition potencies and affinities for all transporters. The inhibition and binding potencies generally decreased with increasing size of the carbon side chain. Compared to MPH, the steric ring-substitution of N-benzylethylphenidate substantially decreased the inhibition potency for all transporters and the binding to the NET and DAT. However, higher binding affinity for the SERT was observed. Modafinil selectively inhibited and bound to the SERT.

The present study has limitations. Possible potent contaminants could theoretically have influenced the results for some drugs with lower purity. Substance-induced efflux was only tested at a high substance concentration. The absence of monoamine efflux could be the result of bell-shaped concentration-efflux curves as it has been demonstrated for amphetamine analogs with known monoamine releasing properties, including MDMA, in different in vitro assays (Seidel et al., 2005). However, such bell-shaped efflux curves were not observed in the assay used in the present study as previously documented (Hysek et al., 2012), strengthening the argument that the MPH-based NPS are in fact pure uptake inhibitors. Moreover, in this study the focus was laid on the NET, DAT, and SERT, as they are main targets of amphetamines and presumably many stimulant NPS (Sitte and Freissmuth, 2015). Other possible mechanisms that may contribute to the effects of NPS,
such as VMAT2 inhibition (Sulzer et al., 2005), calcium-triggered exocytosis of monoamines (Mundorf et al., 1999; Sulzer et al., 2005), mRNA regulation (Douglass et al., 1995), or ion channel blockage (Bauman and DiDomenico, 2002; O'Leary and Hancox, 2010), were not investigated in this study.

5. Conclusion

Similar to MPH and cocaine, MPH-based NPS are potent inhibitors of the NET and DAT. Furthermore, they are not monoamine transporter substrates and have only minor interactions with monoamine receptors. The high selectivity for the DAT vs. SERT suggests that these emerging drugs may have abuse potential. Modafinil is a weak but selective inhibitor at DAT but does not present monoamine receptor interactions.

Author contributions


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Conflict of Interest

M.C.H. is an employee of F. Hoffmann-La Roche. The other authors do not have any conflicts of interest to declare for this work.
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Figures

![Diagram of chemical structures](image)

**Fig. 1.** Chemical structures of MPH-based NPS and related compounds.
Fig. 2. Monoamine uptake inhibition in stably transfected HEK 293 cells that expressed the human NET, DAT, or SERT. Curves were fitted by non-linear regression, and corresponding IC_{50} values are shown in Table 1. The data are presented as the mean ± SEM and numbers in parentheses indicate the number of individual experiments performed in triplicate (NET/DAT/SERT): N-benzylethylphenidate (4/4/4), 3,4-dichloroethylphenidate (3/3/5), 3,4-dichloromethylphenidate (3/3/4), ethynaphthidate (3/3/3), ethylphenidate (3/3/5), 4-fluoromethylphenidate (4/4/4), isopropylphenidate (3/3/4), 4-methylmethylphenidate (4/3/4), methylmorphenate (3/3/4), methylphenidate (3/3/6), propylphenidate (3/3/4), cocaine (3/4/5), modafinil (6/7/6).
**Fig. 3.** Monoamine efflux induced by 100 µM of the compounds after preloading HEK 293 cells that expressed the human NET, DAT, or SERT with radiolabeled monoamine. The efflux is expressed as percentage of $[^3]$H-NE, $[^3]$H-DA, or $[^3]$H-5-HT decrease in monoamine preloaded cells compared to vehicle control. The dashed line marks nonspecific “pseudo-efflux” that arises from monoamine diffusion and subsequent reuptake inhibition. Substances that caused significantly more monoamine efflux (*$p < 0.05$) than pure uptake inhibitors (open bars) were determined to be monoamine transporter substrates. The data are presented as the mean ± SEM of five independent experiments.
### Tables

**Table 1. Monoamine transport inhibition.**

<table>
<thead>
<tr>
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<th>NET  IC\textsubscript{50} [µM] (95% CI)</th>
<th>DAT  IC\textsubscript{50} [µM] (95% CI)</th>
<th>SERT IC\textsubscript{50} [µM] (95% CI)</th>
<th>DAT/SERT ratio (95% CI)</th>
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<tr>
<td><strong>Methylphenidate-based</strong></td>
<td></td>
<td></td>
<td></td>
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</tr>
<tr>
<td>Methylphenidate</td>
<td>0.12 (0.09–0.16)</td>
<td>0.13 (0.10–0.18)</td>
<td>274 (204–366)</td>
<td>2108 (1133–3660)</td>
</tr>
<tr>
<td>4-Methylmethylphenidate</td>
<td>0.09 (0.07–0.11)</td>
<td>0.15 (0.12–0.18)</td>
<td>164 (132–204)</td>
<td>1093 (733–1700)</td>
</tr>
<tr>
<td>Ethylphenidate</td>
<td>0.81 (0.62–1.06)</td>
<td>0.61 (0.45–0.84)</td>
<td>257 (205–322)</td>
<td>421 (244–716)</td>
</tr>
<tr>
<td>4-Fluoromethylphenidate</td>
<td>0.04 (0.03–0.06)</td>
<td>0.15 (0.12–0.20)</td>
<td>40 (33–48)</td>
<td>267 (165–400)</td>
</tr>
<tr>
<td>3,4-Dichloromethylphenidate</td>
<td>0.01 (0.01–0.02)</td>
<td>0.05 (0.04–0.06)</td>
<td>12 (9–15)</td>
<td>240 (150–375)</td>
</tr>
<tr>
<td>Isopropylphenidate</td>
<td>2.3 (1.8–2.9)</td>
<td>0.82 (0.68–1.00)</td>
<td>147 (112–193)</td>
<td>179 (112–284)</td>
</tr>
<tr>
<td>Methylmorphenate</td>
<td>9.3 (7.0–12.3)</td>
<td>13 (11–16)</td>
<td>1831 (932–3600)</td>
<td>141 (58–327)</td>
</tr>
<tr>
<td>3,4-Dichloroethylphenidate</td>
<td>0.13 (0.10–0.16)</td>
<td>0.08 (0.06–0.09)</td>
<td>8.0 (6.9–9.3)</td>
<td>100 (77–155)</td>
</tr>
<tr>
<td>Propylphenidate</td>
<td>0.94 (0.71–1.25)</td>
<td>1.2 (1.0–1.6)</td>
<td>84 (67–106)</td>
<td>70 (42–106)</td>
</tr>
<tr>
<td>N-Benzylethylphenidate</td>
<td>95 (59–154)</td>
<td>60 (41–86)</td>
<td>2515 (958–6605)</td>
<td>42 (11–161)</td>
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<tr>
<td>Ethynaphthidate</td>
<td>0.42 (0.32–0.54)</td>
<td>0.34 (0.28–0.42)</td>
<td>1.7 (1.3–2.1)</td>
<td>5.0 (3.1–7.5)</td>
</tr>
<tr>
<td><strong>Other</strong></td>
<td></td>
<td></td>
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</tr>
<tr>
<td>Modafinil</td>
<td>231 (177–300)</td>
<td>11 (9–14)</td>
<td>2616 (250–27300)</td>
<td>238 (28–1950)</td>
</tr>
<tr>
<td>Cocaine</td>
<td>0.48 (0.36–0.64)</td>
<td>0.90 (0.75–1.08)</td>
<td>1.5 (1.2–1.9)</td>
<td>1.7 (1.1–2.5)</td>
</tr>
</tbody>
</table>

Values are means and 95% confidence intervals (CI). DAT/SERT ratio = 1/DAT IC\textsubscript{50} : 1/SERT IC\textsubscript{50}. 
Table 2. Monoamine transporter and receptor binding affinities.

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<th>NET</th>
<th>DAT</th>
<th>SERT</th>
<th>D₂</th>
<th>α₁A</th>
<th>α₂A</th>
<th>5-HT₁A</th>
<th>5-HT₂A</th>
<th>5-HT₂B</th>
<th>5-HT₂C</th>
<th>TA₁rat</th>
<th>TA₁mouse</th>
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<tr>
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<td>Ki</td>
<td>Ki</td>
<td>Ki</td>
<td>Ki</td>
<td>Kᵢ</td>
<td>Kᵢ</td>
<td>Kᵢ</td>
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<td>Kᵢ</td>
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<tr>
<td><strong>Methylphenidate-based</strong></td>
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<td></td>
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<tr>
<td>Methylphenidate</td>
<td>0.50±0.17</td>
<td>0.070±0.020</td>
<td>&gt;22</td>
<td>&gt;4.4</td>
<td>&gt;8.9</td>
<td>&gt;15</td>
<td>&gt;17</td>
<td>&gt;13</td>
<td>&gt;10</td>
<td>&gt;15</td>
<td>&gt;5.0</td>
<td>&gt;4.7</td>
</tr>
<tr>
<td>4-Methylmethylphenidate</td>
<td>0.31±0.10</td>
<td>0.033±0.007</td>
<td>&gt;22</td>
<td>&gt;4.4</td>
<td>6.5±0.3</td>
<td>10±1</td>
<td>9.9±0.7</td>
<td>&gt;13</td>
<td>&gt;10</td>
<td>&gt;15</td>
<td>&gt;5.0</td>
<td>&gt;4.7</td>
</tr>
<tr>
<td>Ethylphenidate</td>
<td>4.9±0.7</td>
<td>0.081±0.007</td>
<td>&gt;30</td>
<td>&gt;25</td>
<td>&gt;12</td>
<td>14±1</td>
<td>&gt;25</td>
<td>&gt;12</td>
<td>&gt;20</td>
<td>&gt;15</td>
<td>&gt;15</td>
<td>&gt;15*</td>
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<tr>
<td>4-Fluoromethylphenidate</td>
<td>0.22±0.08</td>
<td>0.040±0.007</td>
<td>&gt;22</td>
<td>&gt;4.4</td>
<td>&gt;8.9</td>
<td>&gt;15</td>
<td>&gt;17</td>
<td>&gt;13</td>
<td>&gt;10</td>
<td>&gt;15</td>
<td>&gt;5.0</td>
<td>&gt;4.7</td>
</tr>
<tr>
<td>3,4-Dichloromethylphenidate</td>
<td>0.0066±0.0006</td>
<td>0.0060±0.0005</td>
<td>3.0±0.1</td>
<td>&gt;4.4</td>
<td>1.7±0.1</td>
<td>7.0±0.6</td>
<td>1.8±0.1</td>
<td>&gt;13</td>
<td>&gt;15</td>
<td>6.2±0.6</td>
<td>&gt;14</td>
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<tr>
<td>Isopropylphenidate</td>
<td>4.2±0.4</td>
<td>0.097±0.014</td>
<td>&gt;23</td>
<td>&gt;4.4</td>
<td>&gt;11</td>
<td>&gt;15</td>
<td>&gt;17</td>
<td>&gt;13</td>
<td>&gt;15</td>
<td>13±2</td>
<td>&gt;14</td>
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<tr>
<td>Methylmorphenate</td>
<td>24±1</td>
<td>3.2±0.3</td>
<td>&gt;22</td>
<td>&gt;4.4</td>
<td>&gt;8.9</td>
<td>&gt;15</td>
<td>&gt;17</td>
<td>&gt;13</td>
<td>&gt;10</td>
<td>&gt;15</td>
<td>&gt;5.0</td>
<td>&gt;4.7</td>
</tr>
<tr>
<td>3,4-Dichloroethylphenidate</td>
<td>0.028±0.003</td>
<td>0.0065±0.0002</td>
<td>1.5±0.2</td>
<td>&gt;4.4</td>
<td>4.3±0.1</td>
<td>7.5±0.3</td>
<td>4.5±0.4</td>
<td>&gt;13</td>
<td>12±0.3</td>
<td>6.6±1.3</td>
<td>&gt;14</td>
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<tr>
<td>Propylphenidate</td>
<td>3.8±1.3</td>
<td>0.33±0.07</td>
<td>&gt;22</td>
<td>&gt;4.4</td>
<td>&gt;8.9</td>
<td>8.7±0.5</td>
<td>17±1</td>
<td>&gt;13</td>
<td>&gt;10</td>
<td>&gt;15</td>
<td>&gt;5.0</td>
<td>&gt;4.7</td>
</tr>
<tr>
<td>N-Benzylethyl phenidate</td>
<td>5.5±0.5</td>
<td>0.33±0.01</td>
<td>8.4±1.0</td>
<td>&gt;4.4</td>
<td>&gt;11</td>
<td>&gt;15</td>
<td>&gt;17</td>
<td>&gt;13</td>
<td>&gt;15</td>
<td>&gt;15</td>
<td>&gt;14</td>
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<tr>
<td>Ethylnaphthidate</td>
<td>0.27±0.06</td>
<td>0.026±0.003</td>
<td>0.58±0.05</td>
<td>&gt;4.4</td>
<td>1.8±0.2</td>
<td>8.6±0.5</td>
<td>1.3±0.2</td>
<td>4.9±0.5</td>
<td>&gt;10</td>
<td>12±3</td>
<td>&gt;5.0</td>
<td>&gt;4.7</td>
</tr>
<tr>
<td><strong>Other</strong></td>
<td></td>
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<td></td>
</tr>
<tr>
<td>Modafinil</td>
<td>&gt;26</td>
<td>4.0±0.7</td>
<td>&gt;22</td>
<td>&gt;4.4</td>
<td>&gt;8.9</td>
<td>&gt;15</td>
<td>&gt;17</td>
<td>&gt;13</td>
<td>&gt;10</td>
<td>&gt;15</td>
<td>&gt;5.0</td>
<td>&gt;4.7</td>
</tr>
<tr>
<td>Cocaine</td>
<td>1.6±0.3</td>
<td>0.20±0.02</td>
<td>0.87±0.04</td>
<td>&gt;4.4</td>
<td>&gt;8.9</td>
<td>&gt;15</td>
<td>&gt;17</td>
<td>&gt;13</td>
<td>&gt;10</td>
<td>&gt;15</td>
<td>&gt;5.0</td>
<td>&gt;4.7</td>
</tr>
</tbody>
</table>

Values are given as µM (mean±SD).

*a*From Simmler et al., 2016