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Rubisco Evolution in C₄ Eudicots: An Analysis of Amaranthaceae *sensu lato*

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**Abstract**

**Background:** Rubisco (ribulose-1,5-bisphosphate carboxylase/oxygenase) catalyses the key reaction in the photosynthetic assimilation of CO₂. In C₄ plants CO₂ is supplied to Rubisco by an auxiliary CO₂-concentrating pathway that helps to maximize the carboxylase activity of the enzyme while suppressing its oxygenase activity. As a consequence, C₄ Rubisco exhibits a higher maximum velocity but lower substrate specificity compared with the C₃ enzyme. Specific amino-acids in Rubisco are associated with C₄ photosynthesis in monocots, but it is not known whether selection has acted on Rubisco in a similar way in eudicots.

**Methodology/Principal Findings:** We investigated Rubisco evolution in Amaranthaceae *sensu lato* (including Chenopodiaceae), the third-largest family of C₄ plants, using phylogeny-based maximum likelihood and Bayesian methods to detect Darwinian selection on the chloroplast rbcL gene in a sample of 179 species. Two Rubisco residues, 281 and 309, were found to be under positive selection in C₄ Amaranthaceae with multiple parallel replacements of alanine by serine at position 281 and methionine by isoleucine at position 309. Remarkably, both amino-acids have been detected in other C₄ plant groups, such as C₄ monocots, illustrating a striking parallelism in molecular evolution.

**Conclusions/Significance:** Our findings illustrate how simple genetic changes can contribute to the evolution of photosynthesis and strengthen the hypothesis that parallel amino-acid replacements are associated with adaptive changes in Rubisco.

**Introduction**

Rubisco (ribulose-1,5-bisphosphate carboxylase/oxygenase, EC 4.1.1.39) serves as the main gateway for inorganic carbon to enter metabolic pathways in most ecosystems and hence is unique in its importance to support life. Observations of significant variation in Rubisco kinetics between plant species [1,2,3], the correlation of Rubisco kinetics with temperature [4] and CO₂ availability [5], and positive selection on Rubisco at the molecular level in all principal lineages of land plants [6] support the hypothesis that all Rubiscos may be well adapted to their subcellular environment [7]. However, the molecular mechanisms responsible for optimizing the relationship between Rubisco specificity and its maximum rate of catalytic turnover in particular conditions are still open to debate [8]. Here we use a phylogeny-based approach to investigate how the occurrence of C₄ photosynthesis has influenced Rubisco evolution at the molecular level in eudicots as represented by the family Amaranthaceae *sensu lato*.

Rubisco discriminates imperfectly between CO₂ and O₂ as substrates, and under present-day atmospheric conditions (385 p.p.m. CO₂), the carboxylase activity of Rubisco is saturated in C₃ plants, and the oxygenase activity gives rise directly to the competing process of photorespiration. Photorespiratory rates in C₃ plants increase steeply with increasing temperature and give rise to a distinct temperature optimum for net photosynthesis, above which plant yields decline steeply. Increased carbon loss via photorespiration at higher temperatures is attributable mainly to the declining specificity of Rubisco for CO₂ relative to O₂ (S_c/o). In fact, it has been proposed that the very slow turnover of Rubisco (k_cat ≈ 3 s⁻¹) is a direct consequence of the enzyme’s particular reaction mechanism, in which S_c/o is maximized by tight binding of the transition-state intermediate [7]. Land plants also depend on the enzyme rubisco activase which removes tightly binding inhibitors at the active site of Rubisco and thus prevents the loss of its catalytic activity. The cascade of side-reactions performed by Rubisco is yet to be fully understood although recent achievements in mathematical modelling of Rubisco reactions offer the theoretical background for predicting ‘side-effects’ by simulating the overall kinetic behaviour [9]. Another corollary of low k_cat and of the large size of the holozyme (560 kDa) is that Rubisco comprises up to 50% of soluble protein in photosynthetic tissues and is probably the most abundant enzyme on Earth [10].

In terrestrial plants with C₄ photosynthesis or crassulacean acid metabolism (CAM), and in many aquatic organisms, photorespiration is partially or completely suppressed by the operation of an auxiliary CO₂-concentrating mechanism. C₄ plants initially fix atmospheric carbon in the mesophyll cells using phosphoenolpyr-
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Materials and Methods

Phylogenetic analysis

We obtained all Amaranthaceae rbcL nucleotide sequences available in GenBank and aligned them. Sequences shorter than 1341 base pairs and sequences with missing data were excluded. The resulting trimmed alignment consisted of 179 rbcL sequences of 1341 base pairs long which represented 94% of the rbcL coding region and corresponded to positions 64 to 1404 of the rbcL sequence of Spinacia oleracea (GenBank AJ400848). The analysed dataset consisted of 95 C3 and 84 C4 species (Table S1). Most of the included sequences came from four studies [19,28,29,30] and evenly represented all main lineages within the family (Fig. 1). Phylogeny was reconstructed using a maximum-likelihood inference (ML) conducted with RAxML version 7.2.6 [31] using the raxmlGUI interface [32]. We conducted five independent runs from different starting points to assess convergence within two likelihood units of the best tree, which was consistently selected. The parameters of partition were allowed to vary independently under the GTR+GAMMA model of evolution as implemented in RAxML. ML nodal support was calculated by analysing 1000 bootstrap replicates. The best-scoring ML tree was used for tests of positive selection (see below).

Tests for positive selection

Positive, neutral, or purifying selection at the molecular level can be inferred by comparing rates of non-synonymous (dNS) and synonymous (dSN) mutations along a phylogenetic tree [33]. Under neutrality, the two rates are expected to be equal (dNS/dSN = 1), while purifying (negative) or adaptive (positive) selection is expected to deflate (dNS/dSN<1) or inflate (dNS/dSN>1) this ratio, respectively. One can use likelihood ratio tests to detect positive selection that affects only a subset of codons in a protein-coding gene, with positive selection indicated by accelerated nonsynonymous substitutions. Models assuming positive selection along all phylogeny or prespecified branches only (e.g. C4 lineages in our case) can be employed within Phylogenetic Analysis by Maximum Likelihood (PAML) framework [33].

We used the codeml program in the PAML v.4.4 package [33] to estimate dNS/dSN ratio in the model M0, that allows for a single dNS/dSN value across the whole phylogenetic tree obtained previously (see Phylogenetic analyses section). Further, codeml was used to perform likelihood ratio tests (LRTs) for positive selection among amino
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acid sites. The tree length value obtained from the model M0 was compared with tree length values obtained from other models to control for consistency among models. We performed two LRTs to compare null models which assume the same selective pressure along all branches of a phylogeny and do not allow positive selection \( d_k/d_s > 1 \) with nested models which do allow it [33]. The first LRT, A1-aM2a, compares the M1a model (Nearly Neutral) which allows \( 0 \leq d_k/d_s \leq 1 \) with the M2a model (Selection model); same as the M1 model plus an extra class under positive selection with \( d_k/d_s > 1 \). The second LRT, M8a-M8, compares the M8a model which assumes a discrete beta distribution for \( d_k/d_s \), which is constrained between 0 and 1, including a class with \( d_k/d_s = 1 \), with the M8 model which allows the same distribution as M8a but an extra class under positive selection with \( d_k/d_s > 1 \).

Finally, we performed two branch-site tests of positive selection along prespecified foreground branches [33, 34, 35]. The first was the A model for basal C4 branches only where positive selection was allowed only on branches leading to C4 clades. The second was the A model for all C4 branches where positive selection was allowed on branches leading to C4 clades and branches within C4 clades. The A1-A LRT compares the null model A1 with the nested model A. Both the A1 and A models allow \( d_k/d_s \) ratios to vary among sites and among lineages. The A1 model allows \( 0 \leq d_k/d_s < 1 \) and \( d_k/d_s = 1 \) for all branches, and also two additional classes of codons with fixed \( d_k/d_s = 1 \) along prespecified foreground branches while restricted as \( 0 < d_k/d_s < 1 \) and \( d_k/d_s = 1 \) on background branches. The alternative A model allows \( 0 < d_k/d_s < 1 \) and \( d_k/d_s = 1 \) for all branches, and also two additional classes of codons under positive selection with \( d_k/d_s > 1 \) along prespecified foreground branches while restricted as \( 0 < d_k/d_s < 1 \) and \( d_k/d_s = 1 \) on background branches. C4 lineages were marked as foreground branches.

For all LRTs, the first model is a simplified version of the second, with fewer parameters, and is thus expected to provide a poorer fit to the data (lower maximum likelihood). The M1a, M8a and A1 models are null models which do not allow codons with \( d_k/d_s > 1 \), whereas the M2a, M8 and A models are alternative models which do allow codons with \( d_k/d_s > 1 \). The significance of the LRTs was calculated assuming that twice the difference in the log of maximum likelihood between the two models was distributed as a chi-square distribution with the degrees of freedom (df) given by the difference in the numbers of parameters in the two nested models [34, 36]. For the M1a-M2a comparison \( \text{df} = 2 \), and for M8a-M8, A1-A and M0 vs 2-rates model comparisons \( \text{df} = 1 \). Each LRT was run two times using different initial \( d_k/d_s \) values (0.1 and 0.4) to test for suboptimal local peaks. To identify amino acid sites potentially under positive selection, the parameter estimates from M2a, M8 and A models were used to calculate the posterior probabilities that an amino acid belongs to a class with \( d_k/d_s > 1 \) using the Bays Empirical Bayes (BEB) approaches implemented in PAML [37]. Independently from codeml we used the SLR program which implements “sitewise likelihood-ratio” (SLR) method for detecting non-neutral evolution, a statistical test

### Table 1. Analysis of the Amaranthaceae rbcl genes for positively selected sites.

<table>
<thead>
<tr>
<th>Model with positive selection *</th>
<th>Positively selected sites</th>
<th>Null model</th>
<th>LRT d</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Analysis for positively selected sites common for C4 and C3 clades</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>M2a</td>
<td>-10711.44</td>
<td>( \chi = 3.00, p_0 = 0.93, ) ( \epsilon_0 = 0.02, ) ( p_1 = 0.01, ) ( \epsilon_1 = 0.26 )</td>
<td>32, 145, 279, 499</td>
</tr>
<tr>
<td>M8</td>
<td>-10705.58</td>
<td>( \chi = 2.94, p_0 = 0.96, ) ( \rho = 0.15, ) ( q = 3.04, ) ( \epsilon = 1.56 )</td>
<td>32, 43, 145, 225, 262, 279, 439, 443</td>
</tr>
<tr>
<td>SLR</td>
<td>NA</td>
<td>( \chi = 2.75, ) ( \epsilon = 0.10 )</td>
<td>32, 145, 225, 279, 439</td>
</tr>
<tr>
<td><strong>Analysis for positively selected sites specific for branches leading to C4 clades</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>A</td>
<td>-10729.13</td>
<td>( \chi = 2.94, p_0 = 0.93, ) ( \epsilon_0 = 0.02, ) ( \epsilon_1 = NA )</td>
<td>no</td>
</tr>
<tr>
<td><strong>Analysis for positively selected sites specific for C4 clades</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>A</td>
<td>-10723.60</td>
<td>( \chi = 2.94, p_0 = 0.92, ) ( \epsilon_0 = 0.02, ) ( p_1 = 0.01, ) ( \epsilon_1 = 3.15 )</td>
<td>281, 309</td>
</tr>
</tbody>
</table>

*M1a (nearly neutral), M2a (positive selection), M8a (beta & \( \epsilon = 1 \)) and M8 (beta & \( \epsilon \)) are PAML site models; A1 and A are PAML branch site models; SLR is “sitewise likelihood-ratio” method.

**K** is transition/transversion rate ratio; \( \epsilon \) is \( d_k/d_s \) ratio; \( \epsilon_0 \) is \( d_k/d_s \) ratio in a class under putative positive selection; \( p_0 \) and \( p_1 \) are proportion of codons with \( \epsilon = 0 \) and \( \epsilon > 1 \), respectively; \( p \) and \( q \) are parameters of beta distribution in the range (0, 1); for the SLR test, the parameter values given are those optimal under M0.

The sites listed are those at which positive selection is detected with a cutoff (significance level or posterior probability, as appropriate to the method used) >95%; those >99% are in italics. For the SLR test, the italic underlined sites are those at which there is still evidence for positive selection after correcting for multiple comparisons.

LRT is likelihood ratio test, \( \text{df} \) is twice the difference of model log-likelihoods.

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Figure 1. Maximum likelihood phylogeny based on *rbcl* sequences of 179 Amaranthaceae species. Numbers above the branches are ML bootstrap support percentages. Filled orange circles of the first, second and third columns after species names indicate presence of C4 photosynthesis, serine at the position 281 and isoleucine at the position 309, respectively. The figure was composed using iTOL program [62]. doi:10.1371/journal.pone.0052974.g001
that can identify sites under positive selection even when the strength of selection is low [30]. The SLR test [30] consists of performing a likelihood-ratio test on a sitewise basis, testing the null model (neutrality, \( d_\alpha/d_\beta = 1 \)) against an alternative model (\( d_\alpha/d_\beta \neq 1 \)). SLR method is a test of whether a given site has undergone selection or not, and the test statistic summarizes the strength of the evidence for selection rather than the strength of the selection itself [38]. The same input files with sequence alignment and species phylogeny were used for both codeml and SLR.

**Analysis of correlated evolution on phylogenies**

Closely related taxa are not independent data points and they consequently violate the assumptions of conventional statistical methods [36]. Thus, we used analysis of correlated evolution on phylogenies to test the significance of correlation between pairs of discrete characters: (1) the presence/absence of C_4 photosynthesis and (2) the presence/absence of particular amino-acid at sites found to be under positive selection along C_4 branches in the A model of codeml. For this purpose, we used the phylogeny obtained using RAxML (see above) and performed Pagel’s test of correlated (discrete) character evolution [40] implemented in the Mesquite package (version 2.72) [41]. Test was performed separately for each Rubisco residue under positive selection along C_4 branches and Bonferroni correction was performed for simultaneous statistical testing.

**Structural analysis of Rubisco**

We used the published Rubisco protein structure from spinach (Spinacia oleracea, Amaranthaceae) from data file 1RBO [42] obtained from the RCSB Protein Data Bank. Throughout the paper, the numbering of Rubisco large subunit residues is based on the spinach sequence. The locations and properties of individual amino acids in the Rubisco structure were analysed using DeepView – Swiss-PdbViewer v.3.7 [43] and by CUPSAT [44].

**Results**

**Phylogenetic analysis**

The ML phylogenetic tree (Fig. 1) for rbcL sequences from 179 Amaranthaceae species was largely congruent with previously obtained phylogenies and accepted taxonomic subdivisions of the family [19,28,29,30,45,46,47,48]; however no statistical tests for topological similarity between our tree and previously published trees were performed because of different sizes and species compositions of datasets. A minimum of 16 independent origins of C_4 photosynthesis were represented in the Amaranthaceae phylogeny if conservative approach for observed polytomies had been taken (Fig. 1), which is consistent with the estimate by Sage et al. [16]. The other assumption of this estimate was that no reversals from C_4 to C_3 were allowed. Predominance of C_4 gains over reversals to C_3 is supported by both empirical data and theoretical work [49].

**Tests for positive selection**

Likelihood ratio tests (LRTs) for variation in \( d_\alpha/d_\beta \) ratios and for positive selection [33] were applied to the dataset of rbcL sequences from 179 C_3 and C_4 Amaranthaceae species. LRTs that were run using two different initial \( d_\alpha/d_\beta \) values (0.1 and 0.4) to test for suboptimal local peaks produced identical results. LRTs for positive selection [33] showed that the models assuming positive selection (M2a and M8) fit the data better than the nested models without positive selection (M1a and M8a; \( p\)-value <0.00001; Table 1). To test whether selection occurs specifically in C_4 clades we used two branch site models (aka model A [33,34]), one of which allowed positive selection only on branches leading to C_4 clades and the other also allowed positive selection within the C_4 clades. Each of these models was compared to an alternative model that allowed for no positive selection and only the latter of the two models demonstrated better fit to data than the model without positive selection (\( p\)-value <0.05; Table 1).

**Sites under positive selection**

Four sites were identified as evolving under positive selection with a posterior probability >0.95 by BEB [37] implemented in the M2a model (residues 32, 145, 279, 439), but eight sites when BEB was implemented in the M8 model (all the same that in M2a plus sites 43, 225, 262, 443). Independent SLR analysis showed five sites evolving under positive selection (32, 145, 225, 279, 439), but only for one of them (site 279) evidence for positive selection remained significant after correcting for multiple comparisons. Two sites (residues 281 and 309) were shown to be under positive selection within C_4 clades while under relaxed or purifying selection within C_3 clades with a posterior probability >0.99 by BEB in the A model for C_4 branches. Both sites had only two alternative amino acids in this dataset (Table 2). One of the two alternative amino acids was more frequent among C_4 species, while the other was more frequent among C_3 species (Table 2), but there were no fixed differences between C_4 and C_3 species. We refer to amino acids more frequently associated with C_4 taxa as the ‘C_4’ amino acids, but only for the sake of brevity, as they are not invariably associated with C_4 photosynthesis. Pagel’s test of correlated character evolution [40] on phylogeny showed significant positive associations (\( p\)-value <0.05) between the presence of C_4 photosynthesis and the presence of ‘C_4’ amino acids at sites 281 and 309, shown to be under positive selection along C_4 branches.

**Discussion**

**Widespread positive selection on Rubisco**

As the performance of Rubisco can directly affect plant growth and crop yields, substantial efforts have been made to study its structure and function, with the ultimate aim of trying to improve Rubisco performance [50]. The last few years have brought new approaches to improving our understanding of Rubisco evolution and its genetic mechanisms. The initial molecular-phylogenetic analysis of rbcL showed that positive selection is widespread among all main lineages of land plants, but is restricted to a relatively small number of Rubisco amino acid residues within functionally important sites [6]. Following studies showed that rbcL is under positive selection in particular taxonomic groups [26,27,51,52,53,54,55,56]. Coevolution of residues is common in Rubisco of land plants as well as positive selection and there is an overlap between coevolving and positively selected residues [57]. Hence, phylogeny-based genetic analyses suggest there has been a constant fine-tuning of Rubisco to optimize its performance in specific conditions, in agreement with empirical observations that Rubisco enzymes from different organisms show diversity of kinetics better related to species ecology than phylogeny [4].

All eight residues shown under selection in Amaranthaceae using SLR and PAML models M2 and M8 were already shown to be under Darwinian selection in other groups of plants [6]. Five of these residues (145, 225, 262, 279 and 439) were among twenty most commonly selected Rubisco large subunit residues [6]. Findings in Amaranthaceae are in agreement with the previously described uneven distribution of putative fine-tuning residues in Rubisco [6]. Residues 43, 145, 225, 262 and 279 had only two
alternative amino acids in the analyzed dataset, while residues 32 and 439 had three and residue 443 had four alternative amino acids. Residue 145 is involved in dimer-dimer interactions, residue 225 is involved in interactions with small subunit, while residue 262 is involved in both [6]. C4 photosynthesis has increased the availability of CO2 for Rubisco. There are no monospecific had ‘C4’ amino acids, while nine C4 clades of which the hypothesis that C3 ancestors of C4 species, C3–C4 intermediate and C4 species at the dawn of their origin have Rubisco with C3 kinetics, but once C4 pump is fully functional it creates a strong accessibility, or location within the tertiary structure of the enzyme.

### Table 2. Characteristics of amino-acid replacements under positive selection in the C4 lineages of Amaranthaceae.

<table>
<thead>
<tr>
<th>AA No.*</th>
<th>AA changes C2 – C4</th>
<th>Type of changes b</th>
<th>ΔHc (kJ/mol)</th>
<th>ΔPd</th>
<th>ΔV* (%)</th>
<th>SAf (%)</th>
<th>RFPS (%)</th>
<th>% C3/% C4 species g</th>
<th>Location of residue</th>
<th>Structural motifs within 5 Å</th>
<th>Interactions j</th>
</tr>
</thead>
<tbody>
<tr>
<td>281</td>
<td>A → S</td>
<td></td>
<td>-2.6</td>
<td>1.1</td>
<td>0.4</td>
<td>0.00</td>
<td>2.7</td>
<td>2.1/34.5</td>
<td></td>
<td>Helix 4</td>
<td>DD</td>
</tr>
<tr>
<td>309</td>
<td>M → I</td>
<td></td>
<td>2.6</td>
<td>-0.5</td>
<td>3.8</td>
<td>8.50</td>
<td>5 (-1.3)</td>
<td>19.6</td>
<td></td>
<td>Strand F</td>
<td>ID</td>
</tr>
</tbody>
</table>

* Amino acid (AA) numbering is based on the spinach sequence after [63].

b Side chain type changes. Types abbreviations: H – hydrophobic; N – non-polar aliphatic; P – polar uncharged; U – hydrophilic (after [64]).

c Hydrophylicity difference [65].

d Van der Waals volume difference [67].

e Solvent accessibility calculated using the spinach structure (pdb file 1RBO) by CUPSAT [44].

f Overall stability of the protein predicted using the spinach structure (pdb file 1RBO) by CUPSAT [44]. DS – destabilizing, S – stabilizing.

g RFPS – relative frequency of the particular residue to be under positive selection in C3 plants. Data from 112 rbcL datasets with detected positive selection from [6].

h Percentage of C3 and C4 species that have ‘C4’ amino acid among the 95 C3 species and 84 C4 species of Amaranthaceae analysed.

i Interactions in which the selected residues and/or residues within 5 Å of them are involved. ID – intradimer interactions; DD – dimer-dimer interactions (after [63]).

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Parallel amino-acid replacements in Rubisco from phylogenetically distant lineages

Bayesian analyses of rbcL sequences in a phylogenetic framework allowed us to identify two residues under directional selection along C4 branches within Amaranthaceae (Table 2). There are no common trends in physicochemical properties of ‘C4’ amino acids with respect to proteins such as residue hydrophobicity, solvent accessibility, or location within the tertiary structure of the enzyme (Table 2). Alanine at the position 281 was replaced by serine at least eleven times within the studied species with nine of replacements taking place within C4 clades and two replacements in C4 species Chenopodium bonus-henricus and Spinacia oleracea (Fig. 1). Methionine at the position 309 was replaced by isoleucine at least four times, all of which within C4 clades (Fig. 1). Only three C4 species, Atriplex spongiosa, A. rosea and Horaninovia ulicina, had both ‘C4’ amino acids simultaneously. Seven C4 clades of which one was monospecific had ‘C4’ amino acids, while nine C4 clades of which six consisted of only one species did not have ‘C4’ amino acids (Fig. 1). More frequent occurrence of ‘C4’ amino acids in clades consisting of many species compared to monospecific clades corresponds to our findings of stronger positive selection within C4 clades (Table 1).

Interestingly, both selected residues in C4 Amaranthaceae are among the eight residues selected in C4 Cyperaceae and Poaceae [26] and the ‘C4’ amino acid 309I is also among selected in C4 Flaveria [27]. None of the ‘C4’ amino acids is fixed among C4 species, but they are more frequent among C4 lineages, ranging from 17 to 35% in C4 Amaranthaceae, and from 14 to 87% in C4 Cyperaceae and Poaceae (Table 2; percentage for C4 Cyperaceae and Poaceae calculated using numbers from [26]). Although ‘C4’ amino acids are not fixed among all C4 species, there is a significant positive association between their presence and C4 photosynthetic type in Amaranthaceae. Given the existence of C4 species without ‘C4’ amino acids, it is likely that other as yet unidentified amino acids replacements may be involved in Rubisco adaptation. The model of sequence evolution used to identify Rubisco residues under positive selection within C4 lineages averages selective pressure among selected branches (C4 branches in our case) and allows detection only of the most typical substitutions, potentially missing ones that are unique for a particular branch. Other possible explanations are variation in Rubisco kinetic properties not only between C4 and C3 groups of species but also within these groups [3,4,5,23] and putative differences in other proteins which form the Rubisco complex (small subunit, Rubisco activase). Although the large subunits contain active sites, changes in small subunits may make significant contribution to kinetic properties of plant and algal Rubiscos [59], including differences observed between C3 and C4 plants [60], and the rbcS genes encoding small subunits have been shown under positive selection in C4 Flaveria [27].

Identical amino-acids in Rubisco of C4 Amaranthaceae and C4 Cyperaceae and Poaceae, representing eudicots and monocots with significantly different anatomy and ecological preferences [22], constitute a remarkable example of parallel molecular evolution in phylogenetically distant groups. This example becomes even more interesting if C3 plants are considered as well. Various groups of C3 plants such as some aquatic species and C3 species from cold habitats have faster but less CO2-specific Rubisco compared with their C4 relatives from terrestrial and warm conditions, respectively [3,23]. Hence, some groups of C3 plants can arrive at the same evolutionary solutions for Rubisco fine-tuning as C4 plants. Indeed, ‘C4’ amino acids are not fixed among all C4 species, but they are more frequent among C4 lineages, ranging from 17 to 35% in C4 Amaranthaceae, and from 14 to 87% in C4 Cyperaceae and Poaceae (Table 2; percentage for C4 Cyperaceae and Poaceae calculated using numbers from [26]). Although ‘C4’ amino acids are not fixed among all C4 species, there is a significant positive association between their presence and C4 photosynthetic type in Amaranthaceae. Given the existence of C4 species without ‘C4’ amino acids, it is likely that other as yet unidentified amino acids replacements may be involved in Rubisco adaptation. The model of sequence evolution used to identify Rubisco residues under positive selection within C4 lineages averages selective pressure among selected branches (C4 branches in our case) and allows detection only of the most typical substitutions, potentially missing ones that are unique for a particular branch. Other possible explanations are variation in Rubisco kinetic properties not only between C4 and C3 groups of species but also within these groups [3,4,5,23] and putative differences in other proteins which form the Rubisco complex (small subunit, Rubisco activase). Although the large subunits contain active sites, changes in small subunits may make significant contribution to kinetic properties of plant and algal Rubiscos [59], including differences observed between C3 and C4 plants [60], and the rbcS genes encoding small subunits have been shown under positive selection in C4 Flaveria [27].
Amaranthaceae in the present study and for C₄ monocots and Flaveria previously [26,27], have been reported to be under positive selection in various groups of C₄ plants by Kapralov and Filatov [6]. Moreover, residue 309 is among the most frequently positively selected sites in land plants, and although residue 281 itself is not, its close neighbours, residues 279 and 282, are among the most often positively selected ones [6]. Thus, we can conclude that both ‘C₄’ amino acids, 281S and 309I, evolved in parallel in various phyllogenetically distant lineages of C₃ and C₄ plants in which faster but less specific Rubisco was needed.

The residue 309 is located on the interface of large subunits within a large subunit dimer, while the residue 281 is involved into dimer-dimer interactions (Table 2). Methionine at position 309 is replaced by the smaller and more hydrophobic isoleucine, which has a stabilising and favourable effect on overall molecule stability according to CSPAT calculations using spinach pdb-structure [44], while A281S replacement decreases hydrophobicity and may be destabilising (Table 2).

Effects of A281S replacement on kinetics of land plants Rubisco has not been studied, while recent study by Whitney et al. [61] using mutagenic approach showed that M309I replacement in Flaveria changed Rubisco kinetics from “C₃-like” to “C₄-like” making the enzyme faster but less CO₂-specific. Importance of M309I replacement for changes in kinetics of Flaveria Rubisco was predicted using in silico approach similar to one used in the present study [27] and confirmed in planta by the study of Whitney et al. [61] making it a good case in support of further application of phylogenetic-based methods for detecting residues under positive selection in Rubisco and elsewhere.

Towards the periodic table of functional amino-acid replacements in Rubisco

Continuing population growth creating increasing demand for food, coupled with future climate change and its potentially dire consequences such as biome collapse and crop failure, both call for an improved understanding of mechanisms allowing plant species to adapt the photosynthetic process to a wide range of conditions. Hence, there is a necessity for more phylogenetically based studies of genes encoding Rubisco from various lineages of phototrophs established in different conditions to better understand Rubisco evolution at the molecular level. The integration of phylogenetic and biochemical research is required to study how Darwinian selection has created a range of enzymes with different kinetic and physical properties tailored to function in virtually all ecosystems on our planet. Knowledge of the role of specific residues in Rubisco adaptation to the particular conditions may provide clues for engineering better enzymes suited to contemporary agricultural needs as well as helping to understand what modifications in the enzyme may have been (and perhaps will be) driven by adaptation to different environmental conditions.

Supporting Information

Table S1 List of studied species.

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Author Contributions

Conceived and designed the experiments: MVK. Performed the experiments: MVK. Analyzed the data: MVK. Wrote the paper: MVK JACS DAF.

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