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Using Malaise traps to assess aculeate Hymenoptera associated with farmland linear habitats across a range of farming intensities

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1 **Title**

2 Using Malaise traps to assess aculeate Hymenoptera associated with farmland linear habitats
3 across a range of farming intensities

4

5 **Running Title**

6 Malaise trap to assess habitats quality

7

8 **Abstract**

- 9 1. The intensification of farming practices, along with the loss and fragmentation of semi-
10 natural habitats within agricultural areas, has contributed significantly to insect decline
11 worldwide including flower-visiting aculeate Hymenoptera.
- 12 2. In this study aculeate Hymenoptera were collected using bi-directional Malaise traps
13 placed along farmland linear habitats across a range of farming intensities. The aim was
14 to further our understanding of the value of farmland linear habitats to this insect group
15 and in particular the Vespinae, an understudied family.
- 16 3. Overall, significantly greater aculeate Hymenoptera species richness was found on
17 extensive than on intermediate and intensive farms. Significantly more species and
18 specimens were collected on the side of the traps adjacent to the linear habitats
19 compared to the side which opened onto the fields. Aculeate Hymenoptera species
20 richness was also significantly greater in dense hedgerows than in open hedgerows.
21 Furthermore two out of six Vespinae species, *Vespula rufa* and *Vespula vulgaris*, had
22 significantly more individuals on extensive than intensive farms.
- 23 4. This study highlights that low-intensity farming practices and farmland linear habitats,
24 especially dense hedgerows, may enhance aculeate Hymenoptera occurrence in
25 agricultural areas. It also demonstrates that Malaise traps set up along linear habitats

26 across a range of farming intensities can make a significant contribution to knowledge
27 regarding the biodiversity value. Given that selected Vespinae species follow similar
28 trends to aculeate Hymenoptera, the possibility of using them as simple biodiversity
29 indicators is worthy of further exploration.

30

31 **Key words**

32 Bi-directional Malaise traps, farm hedgerows, farm watercourses, farming intensity, social
33 wasps (Vespinae), wild bees (Apidae), biodiversity

34

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55

56 **Introduction**

57 During the last few decades agricultural production has undergone significant intensification
58 (Robinson & Sutherland, 2002). The intensification of farming practices through the utilisation
59 of high agrochemical inputs and monocultural cropping systems, in addition to the loss and
60 fragmentation of semi-natural habitats, are the primary causes of the rapid decrease of farmland
61 biodiversity (Stoate *et al.*, 2001; Benton *et al.*, 2003; Fahrig, 2003; Kleijn *et al.*, 2009).
62 Furthermore, it is one of the major causes of insect decline worldwide over the past sixty years,
63 including flower-visiting aculeate Hymenoptera (Sánchez-Bayo & Wyckhuys, 2019).

64 The ecological consequences of aculeate Hymenoptera decline is a current topic because they
65 affect important ecosystem services such as pollination of crops and wild plants (Biesmeijer *et*
66 *al.*, 2006; Klein *et al.*, 2007; Potts *et al.*, 2010; Vanbergen & Insect Pollinators Initiative, 2013),
67 which are closely linked to food production and human well-being (Zhang *et al.*, 2007; Haines-
68 Young & Potschin, 2010). The conservation and/or restoration of semi-natural habitats in
69 agricultural areas are known to positively influence aculeate Hymenoptera counteracting their
70 overall decline (Garibaldi *et al.*, 2011; Kennedy *et al.*, 2013). Farmland linear habitats (e.g.
71 hedgerows/watercourses), particularly those in agriculturally productive agricultural areas
72 (Morandin & Kremen, 2013; Garratt *et al.*, 2017), are recognised as valuable habitats providing
73 essential resources for flower-visiting insects (Pollard & Holland, 2006; Herzon & Helenius,

74 2008; Hannon & Sisk, 2009). Furthermore these linear habitats have been reported to function
75 as biological corridors facilitating flower-visiting insect movements (Cranmer *et al.*, 2012).
76 While the ecological value of farmland linear habitats for wild bees has been investigated in
77 great detail in recent years, very little is known about the ecological interactions between these
78 habitats and social wasps within agricultural areas. The study of social wasps has been much
79 neglected worldwide largely due to their negative image (Sumner *et al.*, 2018). The exceptions
80 to this are countries such as New Zealand, Tasmania and Hawaii where social wasps are
81 accidentally introduced pests with negative impacts on native species (Harris, 1991; Richter,
82 2000; Hanna *et al.*, 2012; Potter-Craven *et al.*, 2018). Studies elsewhere are mainly limited to
83 tropical ecosystems where they have been shown to provide fundamental ecosystem services
84 such as pest control (Pereira *et al.*, 2007a, b; Picanço *et al.*, 2011) and pollination (Heithaus,
85 1979; Hermes & Köhler, 2006; Clemente *et al.*, 2012). However, little is known about the
86 ecological interactions of social wasps within agricultural systems in temperate regions.
87 The decline of flower-visiting insects in general in recent years has led to the need to monitor
88 their status in agricultural areas using appropriate sampling methods (Westphal *et al.*, 2008;
89 Grundel *et al.*, 2011). Different sampling methods have shown to effectively collect flower-
90 visiting insects, including coloured pan traps and Malaise traps (Ozanne 2005; Campbell &
91 Hanula 2007; Westphal *et al.*, 2008; Devigne *et al.*, 2014). Although these two methods have
92 demonstrated to deliver reliable indications of insect assemblages in agricultural areas, the
93 coloured pan trap has been shown to provide valuable inventories of bees while Malaise trap
94 catches reflect multiple groups of insects (Bartholomew & Powell, 2005).
95 In this study Malaise traps were set up with the aim of collecting as much information as
96 possible on the ecological value of farmland linear habitats through the capture of a wide range
97 of insect groups. Aculeate Hymenoptera were chosen for the purpose of this paper because
98 they are an important ecological group in terrestrial ecosystems with a key role in providing

99 fundamental ecosystem services such as pollination and pest control (LaSalle & Gauld, 1993).
100 Furthermore, they have been proven to be good indicators of habitat quality and environmental
101 change in agricultural areas (Tscharntke *et al.*, 1998). The aims of this study were, therefore,
102 to:

- 103 1. Describe aculeate Hymenoptera assemblages associated with farmland linear habitats
104 across a range of farming intensities;
- 105 2. Compare the value of farmland linear habitats to the more abundant Apidae and
106 Vespinae species;
- 107 3. Consider how aculeate Hymenoptera collected using Malaise traps contributes to our
108 understanding of the ecological value of farmland linear habitats.

109

110 **Materials and Methods**

111 *Study sites*

112 The study was carried out in County Sligo, in the north-west of Ireland on farmlands dominated
113 by cattle and sheep grazing. Fields were classified as extensive, intermediate and intensive
114 using the HNV index after Boyle, Hayes *et al.*, (2015). The HNV index was calculated by
115 incorporating different parameters such as the Livestock Units per hectare (LU/ha), the area of
116 improved grasslands, the areas owned and farmed, and the size of fields and boundaries. Two
117 Malaise traps of Townes design (Townes, 1972) were placed along linear habitats (hedgerows
118 and/or watercourses) across five fields within each farm category (30 traps in total). One set of
119 two traps was set up in each field at least 200 m apart to ensure that the adjacent set was
120 independent (Gittings *et al.*, 2006). Linear habitats within each intensity category were
121 classified as “dense hedgerow” (< 50% gaps) or “open hedgerow” (> 50% gaps), with each
122 hedgerow type consisting of a hedgerow with/without stonewall/bank and a hedgerow with an
123 adjacent watercourse (ditch/stream). Gaps were defined as those spaces occupied by fences,

124 brambles or non-structural hedgerow species, walls and dead sections of hedgerow (Defra,
125 2007). A third linear habitat (watercourse only) was selected according to Williams *et al.*,
126 (2004) based on the presence of ditches/streams and the absence of hedgerows. Dense
127 hedgerows, open hedgerows and watercourses are, hereafter, referred to as DH, OH and W
128 respectively (Appendix 1).

129

130 *Sampling protocol*

131 Aculeate Hymenoptera specimens were captured using Malaise traps. At each field two
132 Malaise traps were positioned 20 m apart after Carey *et al.*, (2017), and 2 m away from the
133 linear habitat to prevent sampling bias after Wolton *et al.*, (2014). Each bi-directional Malaise
134 trap was placed parallel to the linear habitat on the southern side, thus separating invertebrates
135 caught on the field side of the trap from those caught on the linear habitat side of the trap.
136 Collection bottles were oriented in an easterly direction and filled with 70% ethanol solution
137 to kill and preserve the catch. A portable electric fence was placed around each trap to prevent
138 damage by livestock. Fortnightly sampling commenced when Malaise traps were set up on
139 May 24th and ended on September 13th (2018), resulting in a total of eight collections during the
140 whole sampling period. All samples collected were returned to the laboratory for identification.
141 Aculeate Hymenoptera were identified to species level using Dvořák & Roberts (2006),
142 Richards (1980), Yeo & Corbet (1983), and Falk & Lewington (2017). Due to the difficulties
143 involved in separating workers of *Bombus lucorum* L. and *Bombus terrestris* L. (Prys-Jones &
144 Corbet, 1991; Saville *et al.*, 1997; Pywell *et al.*, 2005; Öckinger & Smith, 2007), the specimens
145 were collectively referred to as *B. lucorum* and treated as a single species due to the higher
146 abundances of queens of *B. lucorum* collected compared to queens of *B. terrestris*.
147 Furthermore, in order to have a formal rank name for all bees (Anthophila) and a more
148 compatible classification with the higher-level system used for the aculeate Hymenoptera, all

149 the bees collected in this study were included in only one family, the Apidae, as suggested by
150 previous authors (Melo & Goncalves, 2005). Although a few individuals of the European
151 honey bee (*Apis mellifera* L.) were present, they were not included in the analysis since they
152 depend primarily on the management of hives rather than purely ecological factors (Kremen *et*
153 *al.*, 2004; Winfree *et al.*, 2007). In addition, the family Formicidae, which also belongs to the
154 aculeate Hymenoptera (Brothers, 1999), was not included in this study because of the small
155 number of individuals collected. Hereafter, where the term “aculeate Hymenoptera” is
156 mentioned, it is inferred that ants and the European honey bee are not included.

157

158 *Data analysis*

159 The data were analysed statistically for the above aculeate Hymenoptera species captured and
160 then separately for the dominant Apidae and Vespinae species. Statistical analyses were
161 performed using the SPSS v25 software (IBM SPSS Statistics 2017). Aculeate Hymenoptera
162 species richness was shown to display a Poisson distribution using the non-parametric
163 Kolmogorov-Smirnov test and was therefore analysed using Poisson error distribution and log
164 link function. Aculeate Hymenoptera abundance which did not display a Poisson distribution,
165 was log-transformed ($\ln(x + 0.1)$) before analysis to achieve normally distributed residuals and
166 tested using General Linear Mixed Models with normal error distribution. Farming intensity,
167 farmland linear habitat and trap side were included as fixed factors in the models. To account
168 for the hierarchical study design, trap identity was included as random factor. Residual analyses
169 were performed to assess model appropriateness and whether the models fitted the data. Post-
170 hoc pairwise comparison among the levels of a factor was used to test the effects of farming
171 intensity, linear habitat and trap side types on aculeate Hymenoptera species richness and
172 abundance. For the analysis the effects significance was set at $P < 0.05$. Since the data of the
173 dominant Apidae and Vespinae species were not normally distributed after logarithmic

174 transformation, Mann-Whitney *U* tests were performed to test the effects of farming intensity
175 and linear habitat types. PC-Ord version 6 (MjM Software Design) was also used to construct
176 species-area curves and assess the adequacy of sampling across all selected fields for aculeate
177 Hymenoptera.

178

179 **Results**

180 A total of 32 species (1334 individuals) of aculeate Hymenoptera were collected during the
181 sampling period in the bi-directional Malaise traps placed between farm fields and linear
182 habitats. Overall, more than twice the number of individuals (903) was collected on the linear
183 habitat side of the Malaise traps compared to the field side (431). The number of individuals
184 collected fortnightly in each of the eight collections ranged from a minimum of 57 specimens
185 in the last collection (September 13th) to a maximum of 286 in the first collection (June 7th)
186 (Table 1). Species-area curves demonstrate sufficient sampling for the collection of total
187 aculeate Hymenoptera using bi-directional Malaise traps (Fig. 1).

188 Of all the specimens collected, the family Apidae was the most abundant group in terms of
189 species richness (17) and abundance (954), with the highest numbers in terms of species and
190 individuals belonging to the genus *Bombus* (Appendix 2). The three most abundant Apidae
191 species, representing almost 90% of the total Apidae individuals collected, were *Bombus*
192 *pascuorum* Scopoli (52.1%), *B. lucorum* (31.5%) and *Bombus pratorum* L. (5.8%). The sub-
193 family Vespinae was the next most abundant group, with 6 species and 328 individuals. The
194 three most abundant Vespinae species recorded, i.e. *Vespula vulgaris* L. (36.6%), *Vespula*
195 *germanica* Fabricius (31.7%) and *Vespula rufa* L. (20.4%), constituted almost 90% of all
196 Vespinae collected. All Vespinae species collected in this study represent the full spectrum of
197 Vespinae species recorded in Ireland to date (Else et al., 2016). The (sub-) families Crabronidae
198 (6 species) and Eumeninae (3 species) were less abundant with 36 and 16 individuals

199 respectively.

200 Aculeate Hymenoptera species richness and abundance differed significantly among farming
201 intensities, linear habitats, and trap side (see Table 2 for P values). Pairwise comparison
202 indicated significantly greater species richness on extensive compared to intermediate ($P =$
203 0.015) and intensive farms ($P = 0.004$) (Fig. 2). Overall, significantly greater species richness
204 ($P = 0.011$) and abundance ($P < 0.001$) were also found in the Malaise trap collecting bottles
205 connected to nets which opened onto the side adjacent to the linear farm habitat compared to
206 the side which opened onto the field (Fig. 3). A comparison of the different linear habitats
207 across all farming intensities indicates that aculeate Hymenoptera species richness was
208 significantly greater in dense hedgerows compared to open hedgerows ($P = 0.012$) (Fig. 4).

209 Analyses of the dominant aculeate hymenopteran species showed different patterns for Apidae
210 and Vespinae species. Although some of the three most abundant Apidae species showed
211 decreasing abundances with increases in farming intensity and with increasing openness of the
212 linear habitats, the differences were not significant (Fig. 5: Appendix 3). Similarly, dominant
213 Vespinae species showed no significant differences in abundance across the different linear
214 habitat types (Fig. 6: Appendix 3). However, *V. rufa* abundance was significantly greater on
215 extensive farms compared to intermediate ($P < 0.001$) and intensive ($P < 0.001$) farms, and
216 significantly more *V. vulgaris* individuals were captured on extensive compared to intensive
217 farms ($P = 0.005$) (Fig. 6).

218

219 **Discussion**

220 Species-area curves show adequate sampling for the collection of aculeate Hymenoptera using
221 Malaise traps demonstrating the robustness of the sampling method employed in this study.
222 Significantly greater species richness of aculeate Hymenoptera was found on extensive farms
223 compared to intermediate and intensive farms. In addition, our results indicate the importance

224 of farmland linear habitats for aculeate Hymenoptera where significantly greater species
225 richness and abundance were recorded on the linear habitat side of the traps than on the field
226 side. Further examination of linear habitat type demonstrates that dense hedgerows harboured
227 significantly greater species richness of aculeate Hymenoptera than open hedgerows. Although
228 there were no significant differences for aculeate Hymenoptera abundances across farming
229 intensity and linear habitats types, more specimens were found on extensive farms and in dense
230 hedgerows.

231 In this study farms were classified using the HNV index (see Appendix 1 for details). The main
232 factors which determined the intensity of farming were the stocking rates and the total area of
233 improved grasslands that had been ploughed and reseeded. Previous studies have shown that
234 management practices such as increased stocking rates, ploughing and reseeded with
235 agricultural grasses, in addition to the application of nitrogenous fertiliser, result in a reduction
236 in grassland biodiversity (Plantureux *et al.*, 2005). Heavy grazing associated with higher
237 stocking rates have been reported to negatively affect field plant species richness in grasslands,
238 thereby reducing valuable resources for many invertebrates (McMahon *et al.*, 2012). Previous
239 studies have shown that the reduction of plant diversity as a consequence of intensive grazing
240 negatively influence invertebrate diversity and abundance (Vickery *et al.*, 2001; Kruess &
241 Tschardtke, 2002). Similarly, an increase of nutrient input levels has been found to influence
242 plant and arthropod communities, causing a decrease of insect species richness (Haddad *et al.*,
243 2000; Vickery *et al.*, 2001). A study of 117 European grasslands by Klimek *et al.*, (2007), has
244 also shown that the reduction of both stocking rates and nitrogenous fertiliser input can
245 contribute significantly to the conservation of biodiversity in agricultural grasslands, as
246 supported by the current study with greater species richness and abundances of aculeate
247 Hymenoptera in more extensively managed farms. The application of herbicides, which are
248 commonly used on intensive farms to facilitate reseeded, control weeds and maintain grass

249 growth, may also decrease plant diversity in grasslands (Plantureux *et al.*, 2005), while its
250 reduction has been shown to favour a richer flora within and around the farm fields providing
251 more forage resources for invertebrates (Hyvönen *et al.*, 2003). Although insecticides in
252 grasslands are generally applied in lower amounts and frequency than in cultivated fields
253 (Plantureux *et al.*, 2005), they may also negatively affect aculeate Hymenoptera communities
254 in intensively managed farms through direct lethal or sub-lethal effects and the modification of
255 the habitat quality (Goulson *et al.*, 2015). Given that low intensity grasslands have been
256 demonstrated to be important for many invertebrate groups, including wild bees and solitary
257 wasps (Carvell, 2002; Kruess & Tschardtke, 2002; Steffan-Dewenter & Leschke, 2003), it is
258 not surprising that our results reveal significantly greater aculeate Hymenoptera species
259 richness on extensive compared to intermediate and intensive farms.

260 Our results also indicate that farmland linear habitats are valuable habitats in agricultural areas
261 with significantly more species and specimens found in the linear habitat side of the traps
262 compared to those in the field side. This is supported by previous studies which demonstrate
263 that farmland linear habitats such as hedgerows and ditches provide invertebrate species in
264 general with resources for foraging, shelter from adverse conditions in addition to
265 overwintering and nesting sites (Pollard & Holland, 2006; Herzon & Helenius, 2008; Hannon
266 & Sisk, 2009). The positive effects of farmland linear habitats are probably because aculeate
267 Hymenoptera find the above resources primarily in the farmland linear habitats and not in the
268 surrounding agriculturally productive grasslands. This is supported by Garratt *et al.*, (2017)
269 who suggest that wild bees in agricultural areas are likely to find essential resources for their
270 occurrences predominantly in hedgerows. Even watercourses in agricultural areas have been
271 shown to supply valuable resources for many invertebrate taxa otherwise absent in intensively
272 managed areas (Herzon & Helenius, 2008).

273 Although we found that farmland linear habitats regardless of their type or quality were more
274 valuable habitats than the fields, our results demonstrate that their value for aculeate
275 Hymenoptera as a whole depends on their quality, with significantly greater species richness
276 associated with dense hedgerows. Dense hedgerows seem, therefore, to be farmland linear
277 habitats of greater value, providing more valuable resources to aculeate Hymenoptera
278 compared to open hedgerows or watercourses only. This conclusion is supported by previous
279 studies in which dense continuous hedgerows with a high diversity of structural hedge species
280 and vegetation layers have been shown to provide essential resources to many invertebrate
281 species (Graham *et al.*, 2018), including wild bees (Garratt *et al.*, 2017). In addition to
282 increasing the provision of food resources, these complex hedgerows may also deliver a greater
283 number of refuge sites against predators and adverse weather conditions (Dainese *et al.*, 2015).
284 Likewise, Amy *et al.*, (2015) demonstrated that dense continuous hedgerows with higher
285 foliage density positively influenced insect fauna, while the increase in hedge gap size was
286 negatively correlated with invertebrate diversity and abundance.

287 The value of farmland linear habitats overall across a range of farming intensities to the
288 dominant Apidae and Vespinae species in this study varies according to species. While two
289 Vespinae species (i.e. *V. rufa* and *V. vulgaris*) show significantly greater abundances on
290 extensive than on intensive farms, the three most abundant Apidae bumblebee species show no
291 significant differences across farming intensities and linear habitat types. This may be
292 explained by the fact that many bee species, and in particular large body sized bees such as
293 bumblebees, seem to be more affected by factors at a broader scale, i.e. landscape scale than at
294 the local scale (Happe *et al.*, 2018). Similarly, other studies have shown that large body sized
295 bees have larger foraging ranges than small sized bees, suggesting that they may exploit
296 resources at a bigger scale and therefore be less influenced by local factors (Steffan-Dewenter
297 *et al.*, 2002). Unlike *B. lucorum* and *B. pratorum*, we found more *B. pascuorum* specimens on

298 extensive farms compared to intermediate and intensive farms. This may be related with the
299 habitat preferences of workers of *B. pascuorum* which seem to prefer farm fields including
300 grasslands (Falk & Lewington, 2017). It is, therefore, likely that less disturbed agricultural
301 areas such as those under extensive management may support greater densities of *B. pascuorum*
302 workers compared to more disturbed areas under intensive management. This may explain why
303 more *B. pascuorum* specimens were found on extensive farms, while more generalist species,
304 in terms of habitat preferences, such as *B. lucorum* and *B. pratorum* (Falk & Lewington, 2017)
305 showed less pronounced preferences for farming intensity types.

306 Similar to farming intensity, linear habitat types did not significantly influence the most
307 dominant Apidae species, although more individuals of *B. lucorum* and *B. pascuorum* were
308 found in dense hedgerows. Greater abundances of these species in dense hedgerows can be
309 explained by the fact that these linear habitats may provide more valuable resources such as
310 food resources and nesting opportunities. Rollin *et al.*, (2013) demonstrated that in agricultural
311 areas, wild bees prefer to forage mainly in woody habitats, including farmland linear habitats
312 such as hedgerows because these habitats seem to provide more food resources and nesting
313 sites for many species. Dense continuous hedgerows, in particular, have been shown to provide
314 essential food resources such as flowering plants for bumblebees (Garratt *et al.*, 2017). In
315 addition to foraging resources, which have been shown to positively influence bee communities
316 (Kleijn & van Langevelde, 2006), another important resource that may explain greater *B.*
317 *pascuorum* abundances in dense hedgerows is the presence of more suitable nesting sites. Nest-
318 site preferences are site-specific and queens of *B. pascuorum* seem to display a preference for
319 nesting along sheltered boundaries running between agricultural fields and woody landscape
320 elements such as hedgerows (Svensson *et al.*, 2000; Kells & Goulson, 2003).

321 Vespinae species *V. rufa* and *V. vulgaris*, on the other hand, were captured in significantly
322 greater abundances on extensive than on intensive farms. These contrasting responses to

323 farming intensity between Apidae and Vespinae species may be explained by different feeding
324 behaviours. Unlike bees, which depend primarily on floral resources such as pollen and nectar,
325 social wasps have a more varied diet, ranging from nectar and pollen to invertebrate prey
326 (Richter, 2000). Invertebrate prey, which include serious crop pests such as aphids and
327 caterpillars, are found mainly in farm fields where they colonize and feed on crop plants (Hill,
328 1987). Low intensity managed grasslands are known to support greater plant species richness
329 and therefore better foraging opportunities for many invertebrate species (McMahon *et al.*,
330 2012) which are potential prey for social wasps. It is therefore likely that extensive farms in
331 this study may provide more food resources for social wasps, including a wider variety of prey,
332 thus explaining greater Vespinae abundances on extensive compared to intermediate and
333 intensive farms. It is also possible that social wasps may be more influenced by factors at a
334 smaller scale than the Apidae. However, while the influence of landscape on bees has been
335 well studied (Steffan-Dewenter *et al.*, 2002), little is currently known about this in relation to
336 Vespinae species.

337 Although linear habitat types did not significantly influence the occurrences of the dominant
338 Vespinae species, *V. germanica* and *V. vulgaris* were more abundant in dense hedgerows and
339 on watercourses. High abundances of these two species on watercourses overall could have
340 been due to the presence, in the vicinity of one of the watercourse sites in particular, of a range
341 of habitats including a wet grassland and a dense, continuous hedgerow. The complex and
342 heterogeneous landscape at this site may have provided wasp communities with a large amount
343 of valuable resources resulting in large colonies with numerous individuals. Indeed, *V. vulgaris*
344 and *V. germanica* are known to generally form large colonies with several thousand workers
345 when the ecological conditions for the colony growth are optimal (Wenseleers *et al.*, 2005).
346 However, further research is required to determine the influence of landscape features on such
347 Vespinae populations.

348

349 **Conclusions**

350 This study demonstrates that extensive farms and farmland linear habitats, particularly dense
351 hedgerows, represent important management conditions and valuable habitats for aculeate
352 Hymenoptera. Furthermore, we have highlighted that farmland linear habitats, regardless of
353 the type or quality, are of importance to aculeate Hymenoptera. While aculeate Hymenoptera,
354 in general, reflect farming intensity and habitat quality in agricultural landscapes, certain
355 Vespinae species may be used as possible indicators of farming intensity in temperate regions.
356 The results of this study also demonstrate that Malaise traps set up along linear habitats in a
357 range of farming intensities can make a significant contribution to knowledge regarding the
358 biodiversity value. In conclusion, our results indicate that both extensive management and
359 farmland linear habitats, especially high quality habitats such as dense hedgerows, can be used
360 as tools to enhance aculeate Hymenoptera occurrence in agricultural areas. Therefore, the
361 introduction of low-intensity farming practices and the conservation or restoration of farmland
362 linear habitats are highly recommended to promote invertebrate diversity and counteract the
363 worldwide insect decline.

364

365

366

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374

375 **References**

376 Amy, S.R., Heard, M.S., Hartley, S.E., George, C.T., Pywell, R.F., & Staley, J.T. (2015)
377 Hedgerow rejuvenation management affects invertebrate communities through changes
378 to habitat structure. *Basic and applied ecology*, **16**, 443-451.

379 Bartholomew, C.S., & Prowell, D. (2005) Pan compared to malaise trapping for bees
380 (Hymenoptera: Apoidea) in a longleaf pine savanna. *Journal of the Kansas*
381 *Entomological Society*, **78**, 390-393.

382 Batáry, P., Baldi, A., Kleijn, D., & Tschardtke, T. (2010a) Landscape-moderated biodiversity
383 effects of agri-environmental management: a meta-analysis. *Proceedings of the Royal*
384 *Society B: Biological Sciences*, **278**, 1894-1902.

385 Batáry, P., Matthiesen, T., & Tschardtke, T. (2010b) Landscape-moderated importance of
386 hedges in conserving farmland bird diversity of organic vs. conventional croplands and
387 grasslands. *Biological Conservation*, **143**, 2020-2027.

388 Benton, T.G., Vickery, J.A., & Wilson, J.D. (2003) Farmland biodiversity: is habitat
389 heterogeneity the key?. *Trends in ecology & evolution*, **18**, 182-188.

390 Biesmeijer, J.C., Roberts, S.P., Reemer, M., Ohlemüller, R., Edwards, M., Peeters, T.,
391 Schaffers, A.P., Potts, S.G., Kleukers, R., Thomas, C.D., Settele, J., & Kunin, W.E.
392 (2006) Parallel declines in pollinators and insect-pollinated plants in Britain and the
393 Netherlands. *Science*, **313**, 351-354.

394 Boyle, P., Hayes, M., Gormally, M., Sullivan, C., & Moran, J. (2015) Development of a nature
395 value index for pastoral farmland—A rapid farm-level assessment. *Ecological*
396 *indicators*, **56**, 31-40.

- 397 Brothers, D.J. (1999) Phylogeny and evolution of wasps, ants and bees (Hymenoptera,
398 Chrysoidea, Vespoidea and Apoidea). *Zoologica Scripta*, **28**, 233-250.
- 399 Campbell, J.W., & Hanula, J.L. (2007) Efficiency of Malaise traps and colored pan traps for
400 collecting flower visiting insects from three forested ecosystems. *Journal of Insect*
401 *Conservation*, **11**, 399-408.
- 402 Carey, J.G., Brien, S., Williams, C.D., & Gormally, M.J. (2017) Indicators of Diptera diversity
403 in wet grassland habitats are influenced by environmental variability, scale of
404 observation, and habitat type. *Ecological Indicators*, **82**, 495-504.
- 405 Carvell, C. (2002) Habitat use and conservation of bumblebees (*Bombus* spp.) under different
406 grassland management regimes. *Biological conservation*, **103**, 33-49.
- 407 Clemente, M.A., Lange, D., Del-Claro, K., Prezoto, F., Campos, N.R., & Barbosa, B.C. (2012)
408 Flower-visiting social wasps and plants interaction: Network pattern and environmental
409 complexity. *Psyche: A Journal of Entomology*, 2012.
- 410 Cranmer, L., McCollin, D., & Ollerton, J. (2012) Landscape structure influences pollinator
411 movements and directly affects plant reproductive success. *Oikos*, **121**, 562-568.
- 412 Dainese, M., Luna, D.I., Sitzia, T., & Marini, L. (2015) Testing scale-dependent effects of
413 seminatural habitats on farmland biodiversity. *Ecological Applications*, **25**, 1681-1690.
- 414 Defra (2007) *Hedgerow Survey Handbook*. A standard procedure for local surveys in the UK,
415 London.
- 416 Devigne, C., & De Biseau, J.C. (2014) Urban ecology: comparison of the effectiveness of five
417 traps commonly used to study the biodiversity of flying insects. *Biodiversity*, **5**, 165-174.
- 418 Dvořák, L., & Roberts, S.P. (2006) Key to the paper and social wasps of Central Europe
419 (Hymenoptera: Vespidae). *Acta Entomologica Musei Nationalis Pragae*, **46**, 221-244.
- 420 Else, G.R., Bolton, B., & Broad, G.R. (2016) Checklist of British and Irish Hymenoptera-
421 aculeates (Apoidea, Chrysoidea and Vespoidea). *Biodiversity data journal*, (4).

- 422 Fahrig, L. (2003) Effects of habitat fragmentation on biodiversity. *Annual review of ecology,*
423 *evolution, and systematics*, **34**, 487-515.
- 424 Falk, S., & Lewington, R. (2017) *Field Guide to the Bees of Great Britain and Ireland.*
425 Bloomsbury Wildlife Guides.
- 426 Garibaldi, L.A., Steffan-Dewenter, I., Kremen, C., Morales, J.M., Bommarco, R.,
427 Cunningham, S.A., Carvalheiro, L.G., Chacoff, N.P., Dudenhöffer, J.H., Greenleaf, S.S.,
428 Holzschuh, A., Isaacs, R., Krewenka, K., Mandelik, Y., Mayfield, M.M., Morandin,
429 L.A., Potts, S.G., Ricketts, T.H., Szentgyörgyi, H., Viana, B.F., Westphal, C., Winfree,
430 R., & Klein, A.M. (2011) Stability of pollination services decreases with isolation from
431 natural areas despite honey bee visits. *Ecology Letters*, **14**, 1062-1072.
- 432 Garratt, M.P., Senapathi, D., Coston, D.J., Mortimer, S.R., & Potts, S.G. (2017) The benefits
433 of hedgerows for pollinators and natural enemies depends on hedge quality and landscape
434 context. *Agriculture, Ecosystems & Environment*, **247**, 363-370.
- 435 Gittings, T., O'Halloran, J., Kelly, T., & Giller, P.S. (2006) The contribution of open spaces to
436 the maintenance of hoverfly (Diptera, Syrphidae) biodiversity in Irish plantation
437 forests. *Forest ecology and management*, **237**, 290-300.
- 438 Goulson, D., Nicholls, E., Botías, C., & Rotheray, E.L. (2015) Bee declines driven by
439 combined stress from parasites, pesticides, and lack of flowers. *Science*, **347**, 1255957.
- 440 Graham, L., Gaulton, R., Gerard, F., & Staley, J. T. (2018) The influence of hedgerow
441 structural condition on wildlife habitat provision in farmed landscapes. *Biological*
442 *conservation*, **220**, 122-131.
- 443 Grundel, R., Frohnapple, K.J., Jean, R.P., & Pavlovic, N.B. (2011) Effectiveness of bowl
444 trapping and netting for inventory of a bee community. *Environmental Entomology*, **40**,
445 374-380.

- 446 Haddad, N.M., Haarstad, J., & Tilman, D. (2000) The effects of long-term nitrogen loading on
447 grassland insect communities. *Oecologia*, **124**, 73-84.
- 448 Haines-Young, R., & Potschin, M. (2010) The links between biodiversity, ecosystem services
449 and human well-being. *Ecosystem Ecology: a new synthesis*, **1**, 110-139.
- 450 Hanna, C., Foote, D., & Kremen, C. (2012) Short- and long-term control of *Vespula*
451 *pennsylvanica* in Hawaii by fipronil baiting. *Pest management science*, **68**, 1026-1033.
- 452 Hannon, L.E., & Sisk, T.D. (2009) Hedgerows in an agri-natural landscape: potential habitat
453 value for native bees. *Biological conservation*, **142**, 2140-2154
- 454 Happe, A.K., Riesch, F., Rösch, V., Gallé, R., Tschardt, T., & Batáry, P. (2018) Small-scale
455 agricultural landscapes and organic management support wild bee communities of cereal
456 field boundaries. *Agriculture, Ecosystems & Environment*, **254**, 92-98.
- 457 Harris, R.J. (1991) Diet of the wasps *Vespula vulgaris* and *V. germanica* in honeydew beech
458 forest of the South Island, New Zealand. *New Zealand journal of zoology*, **18**, 159-169.
- 459 Heithaus, E.R. (1979) Community structure of neotropical flower visiting bees and wasps:
460 diversity and phenology. *Ecology*, **60**, 190-202.
- 461 Hermes, M.G., & Köhler, A. (2006) The flower-visiting social wasps (Hymenoptera, Vespidae,
462 Polistinae) in two areas of Rio Grande do Sul State, southern Brazil. *Revista Brasileira*
463 *de Entomologia*, **50**, 268-274.
- 464 Herzon, I., & Helenius, J. (2008) Agricultural drainage ditches, their biological importance and
465 functioning. *Biological conservation*, **141**, 1171-1183.
- 466 Hill, D.S. (1987) *Agricultural insect pests of temperate regions and their control*. CUP
467 Archive.
- 468 Hyvönen, T., Ketoja, E., Salonen, J., Jalli, H., & Tiainen, J. (2003) Weed species diversity and
469 community composition in organic and conventional cropping of spring
470 cereals. *Agriculture, Ecosystems & Environment*, **97**, 131-149.

471 Kells, A.R., & Goulson, D. (2003) Preferred nesting sites of bumblebee queens (Hymenoptera:
472 Apidae) in agroecosystems in the UK. *Biological conservation*, **109**, 165-174.

473 Kennedy, C.M., Lonsdorf, E., Neel, M.C., Williams, N.M., Ricketts, T.H., Winfree, R.,
474 Bommarco, R., Brittain, C., Burley, A.L., Cariveau, D., Carvalheiro, L.G., Chacoff, N.P.,
475 Cunningham, S.A., Danforth, B.N., Dudenhöffer, J., Elle, E., Gaines, H.R., Garibaldi,
476 L.A., Gratton, C., Holzschuh, A., Isaacs, R., Javorek, S.K., Jha, S., Klein, A.M.,
477 Krewenka, K., Mandelik, Y., Mayfield, M.M., Morandin, L., Neame, L.A., Otieno, M.,
478 Park, M., Potts, S.G., Rundlöf, M., Saez, A., Steffan-Dewenter, I., Taki, H., Viana, B.F.,
479 Westphal, C., Wilson, J.K., Greenleaf, S.S., & Kremen, C. (2013) A global quantitative
480 synthesis of local and landscape effects on wild bee pollinators in
481 agroecosystems. *Ecology Letters*, **16**, 584-599.

482 Kleijn, D., Kohler, F., Báldi, A., Batáry, P., Concepción, E.D., Clough, Y., Díaz, M., Gabriel,
483 D., Holzschuh, A., Knop, E., Kovács, A. Marshall, E.J.P., Tschardtke, T., & Verhulst, J.
484 (2009) On the relationship between farmland biodiversity and land-use intensity in
485 Europe. *Proceedings of the Royal Society B: Biological Sciences*, **276**, 903-909.

486 Kleijn, D., Rundlöf, M., Scheper, J., Smith, H.G., & Tschardtke, T. (2011) Does conservation
487 on farmland contribute to halting the biodiversity decline?. *Trends in ecology &*
488 *evolution*, **26**, 474-481.

489 Kleijn, D., & Van Langevelde, F. (2006) Interacting effects of landscape context and habitat
490 quality on flower visiting insects in agricultural landscapes. *Basic and Applied*
491 *Ecology*, **7**, 201-214.

492 Klein, A.M., Vaissiere, B.E., Cane, J.H., Steffan-Dewenter, I., Cunningham, S.A., Kremen, C.,
493 & Tschardtke, T. (2007) Importance of pollinators in changing landscapes for world
494 crops. *Proceedings of the royal society B: biological sciences*, **274**, 303-313.

495 Klimek, S., Hofmann, M., & Isselstein, J. (2007) Plant species richness and composition in
496 managed grasslands: the relative importance of field management and environmental
497 factors. *Biological conservation*, **134**, 559-570.

498 Kremen, C., Williams, N.M., Bugg, R.L., Fay, J.P., & Thorp, R.W. (2004) The area
499 requirements of an ecosystem service: crop pollination by native bee communities in
500 California. *Ecology letters*, **7**, 1109-1119.

501 Kruess, A., & Tschardtke, T. (2002) Grazing intensity and the diversity of grasshoppers,
502 butterflies, and trap-nesting bees and wasps. *Conservation Biology*, **16**, 1570-1580.

503 LaSalle, J., & Gauld, I.D. (1993) Hymenoptera: their biodiversity, and their impact on the
504 diversity of other organisms. *Hymenoptera and biodiversity*. 1-26.

505 McMahon, B.J., Anderson, A., Carnus, T., Helden, A.J., Kelly-Quinn, M., Maki, A., Sheridan,
506 H., & Purvis, G. (2012) Different bioindicators measured at different spatial scales vary
507 in their response to agricultural intensity. *Ecological Indicators*, **18**, 676-683.

508 Melo, G.A., & Gonçalves, R.B. (2005) Higher-level bee classifications (Hymenoptera,
509 Apoidea, Apidae sensu lato). *Revista Brasileira de Zoologia*, **22**, 153-159.

510 Morandin, L.A., & Kremen, C. (2013) Hedgerow restoration promotes pollinator populations
511 and exports native bees to adjacent fields. *Ecological Applications*, **23**, 829-839.

512 Nielsen, A., Steffan-Dewenter, I., Westphal, C., Messinger, O., Potts, S.G., Roberts, S.P.,
513 Settele, J., Szentgyörgyi, H., Vaissière, B.E., Vaitis, M., Woyciechowski, M., Bazos, I.,
514 Biesmeijer, J.C., Bommarco, R., Kunin, W.E., Tscheulin, T., Lamborn, E., & Petanidou,
515 T. (2011) Assessing bee species richness in two Mediterranean communities: importance
516 of habitat type and sampling techniques. *Ecological Research*, **26**, 969-983.

517 Öckinger, E., & Smith, H.G. (2007) Semi-natural grasslands as population sources for
518 pollinating insects in agricultural landscapes. *Journal of applied ecology*, **44**, 50-59.

- 519 Ozanne, C.M. (2005) Sampling methods for forest understory vegetation. *Insect sampling in*
520 *forest ecosystems*, 58-76.
- 521 Pereira, E.J.G., Picanço, M.C., Bacci, L., Crespo, A.L.B., & Guedes, R.N.C. (2007a) Seasonal
522 mortality factors of the coffee leafminer, *Leucoptera coffeella*. *Bulletin of entomological*
523 *research*, **97**, 421-432.
- 524 Pereira, E.J.G., Picanço, M.C., Bacci, L., Della Lucia, T.M.C., Silva, É.M., & Fernandes, F.L.
525 (2007b) Natural mortality factors of *Leucoptera coffeella* (Lepidoptera: Lyonetiidae) on
526 *Coffea arabica*. *Biocontrol science and technology*, **17**, 441-455.
- 527 Picanço, M.C., Bacci, L., Queiroz, R.B., Silva, G.A., Miranda, M.M.M., Leite, G.L.D., &
528 Suinaga, F.A. (2011) Social wasp predators of *Tuta absoluta*. *Sociobiology*, **58**, 621-633.
- 529 Plantureux, S., Peeters, A., & McCracken, D. (2005) Biodiversity in intensive grasslands:
530 Effect of management, improvement and challenges. *Agronomy research*, **3**, 153-164.
- 531 Pollard, K.A., & Holland, J.M. (2006) Arthropods within the woody element of hedgerows and
532 their distribution pattern. *Agricultural and Forest Entomology*, **8**, 203-211
- 533 Potter-Craven, J., Kirkpatrick, J.B., McQuillan, P.B., & Bell, P. (2018) The effects of
534 introduced vespine wasps (*Vespula germanica* and *V. vulgaris*) on threatened native
535 butterfly (*Oreixenica ptunarra*) populations in Tasmania. *Journal of insect*
536 *conservation*, **22**, 521-532.
- 537 Potts, S.G., Biesmeijer, J.C., Kremen, C., Neumann, P., Schweiger, O., & Kunin, W.E. (2010)
538 Global pollinator declines: trends, impacts and drivers. *Trends in ecology &*
539 *evolution*, **25**, 345-353
- 540 Prys-Jones, O.E., & Corbet, S.A. (1991) *Naturalists' Handbooks 6: Bumblebees*. Slough:
541 Richmond Publishing Co. Ltd.

542 Pywell, R.F., Warman, E.A., Carvell, C., Sparks, T.H., Dicks, L.V., Bennett, D., Wright, A.,
543 Critchley, C.N.R., & Sherwood, A. (2005) Providing foraging resources for bumblebees
544 in intensively farmed landscapes. *Biological Conservation*, **121**, 479-494.

545 Richards, O.W. (1980) Scolioida, Vespoidea and Sphecoidea. Hymenoptera,
546 Aculeata. *Handbooks for the identification of British insects*, **6**(3 (b)).

547 Richter, M.R. (2000) Social wasp (Hymenoptera: Vespidae) foraging behavior. *Annual review*
548 *of entomology*, **45**, 121-150

549 Robinson, R.A., & Sutherland, W.J. (2002) Post-war changes in arable farming and
550 biodiversity in Great Britain. *Journal of applied Ecology*, **39**, 157-176.

551 Rollin, O., Bretagnolle, V., Decourtye, A., Aptel, J., Michel, N., Vaissière, B.E., & Henry, M.
552 (2013) Differences of floral resource use between honey bees and wild bees in an
553 intensive farming system. *Agriculture, Ecosystems & Environment*, **179**, 78-86.

554 Sánchez-Bayo, F., & Wyckhuys, K.A. (2019) Worldwide decline of the entomofauna: A
555 review of its drivers. *Biological Conservation*, **232**, 8-27.

556 Saville, N.M., Dramstad, W.E., Fry, G.L., & Corbet, S.A. (1997) Bumblebee movement in a
557 fragmented agricultural landscape. *Agriculture, Ecosystems & Environment*, **61**, 145-
558 154.

559 Spears, L.R., & Ramirez, R.A. (2015) Learning to love leftovers: using by-catch to expand our
560 knowledge in entomology. *American Entomologist*, **61**, 168-173.

561 Steffan-Dewenter, I., Münzenberg, U., Bürger, C., Thies, C., & Tschardtke, T. (2002) Scale-
562 dependent effects of landscape context on three pollinator guilds. *Ecology*, **83**, 1421-
563 1432.

564 Steffan-Dewenter, I., & Leschke, K. (2003) Effects of habitat management on vegetation and
565 above-ground nesting bees and wasps of orchard meadows in Central
566 Europe. *Biodiversity & Conservation*, **12**, 1953-1968.

- 567 Stoate, C., Boatman, N.D., Borralho, R.J., Carvalho, C.R., De Snoo, G.R., & Eden, P. (2001)
568 Ecological impacts of arable intensification in Europe. *Journal of environmental*
569 *management*, **63**, 337-365.
- 570 Sumner, S., Law, G., & Cini, A. (2018) Why we love bees and hate wasps. *Ecological*
571 *entomology*, **43**, 836-845
- 572 Svensson, B., Lagerlöf, J., & Svensson, B.G. (2000) Habitat preferences of nest-seeking
573 bumble bees (Hymenoptera: Apidae) in an agricultural landscape. *Agriculture,*
574 *Ecosystems & Environment*, **77**, 247-255.
- 575 Townes, H. (1972) A light-weight Malaise trap. *Entomological news*, **83**, 239-247.
- 576 Tschamtker, T., Gathmann, A., & Steffan-Dewenter, I. (1998) Bioindication using trap-nesting
577 bees and wasps and their natural enemies: community structure and interactions. *Journal*
578 *of applied ecology*, **35**, 708-719.
- 579 Tschamtker, T., Klein, A.M., Kruess, A., Steffan-Dewenter, I., & Thies, C. (2005) Landscape
580 perspectives on agricultural intensification and biodiversity–ecosystem service
581 management. *Ecology letters*, **8**, 857-874.
- 582 Vanbergen, A. J., & Insect Pollinators Initiative. (2013) Threats to an ecosystem service:
583 pressures on pollinators. *Frontiers in Ecology and the Environment*, **11**, 251-259.
- 584 Vickery, J.A., Tallwin, J.R., Feber, R.E., Asteraki, E.J., Atkinson, P.W., Fuller, R.J., &
585 Brown, V.K. (2001) The management of lowland neutral grasslands in Britain: effects of
586 agricultural practices on birds and their food resources. *Journal of Applied Ecology*, **38**,
587 647-664.
- 588 Wenseleers, T., Badcock, N.S., Erven, K., Tofilski, A., Nascimento, F.S., Hart, A.G., Burke,
589 T.A., Archer, M.E., & Ratnieks, F.L.W. (2005) A test of worker policing theory in an
590 advanced eusocial wasp, *Vespula rufa*. *Evolution*, **59**, 1306-1314.

591 Westphal, C., Bommarco, R., Carré, G., Lamborn, E., Morison, N., Petanidou, T., Potts, S.G.,
592 Roberts, S.P.M., Szentgyörgyi, H., Tscheulin, T., Vaissière, B.E., Woyciechowski, M.,
593 Biesmeijer, J.C., Kunin, W.E., Settele, J., & Steffan-Dewenter, I. (2008) Measuring bee
594 diversity in different European habitats and biogeographical regions. *Ecological*
595 *Monographs*, **78**, 653-671.

596 Williams, P., Whitfield, M., Biggs, J., Bray, S., Fox, G., Nicolet, P., & Sear, D. (2004)
597 Comparative biodiversity of rivers, streams, ditches and ponds in an agricultural
598 landscape in Southern England. *Biological conservation*, **115**, 329-341.

599 Winfree, R., Griswold, T., & Kremen, C. (2007) Effect of human disturbance on bee
600 communities in a forested ecosystem. *Conservation biology*, **21**, 213-223.

601 Wolton, R.J., Bentley, H., Chandler, P.J., Drake, C.M., Kramer, J., Plant, A.R., & Stubbs, A.E.
602 (2014) The diversity of Diptera associated with a British hedge. *Dipterists Digest*, **21**, 1-
603 36.

604 Yeo, P. F., & Corbet, S. A. (1983) *Solitary wasps*. Cambridge University Press.

605 Zhang, W., Ricketts, T.H., Kremen, C., Carney, K., & Swinton, S.M. (2007) Ecosystem
606 services and dis-services to agriculture. *Ecological economics*, **64**, 253-260.

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616 **Tables and Figures**

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619 **Table 1.** Total numbers of aculeate Hymenoptera (Aculeate) captured on each sampling period
620 (2018) across all selected farms in Co. Sligo (Ireland).

621

| Aculeate | 24 May 7 Jun | 7 Jun 21 Jun | 21 Jun 5 Jul | 5 Jul 19 Jul | 19 Jul 2 Aug | 2 Aug 16 Aug | 16 Aug 30 Aug | 30 Aug 13 Sep | Abundance |
|------------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|------------------|------------------|-----------|
| <i>Apoidea</i> | | | | | | | | | |
| Apidae | 239 | 162 | 87 | 155 | 189 | 68 | 39 | 15 | 954 |
| Crabronidae | 1 | 2 | 13 | 2 | 13 | 3 | 2 | - | 36 |
| <i>Vespoidea</i> | | | | | | | | | |
| Eumeninae | 3 | - | 12 | 1 | - | - | - | - | 16 |
| Vespinae | 43 | 17 | 38 | 48 | 54 | 49 | 37 | 42 | 328 |
| Total | 286 | 181 | 150 | 206 | 256 | 120 | 78 | 57 | 1334 |

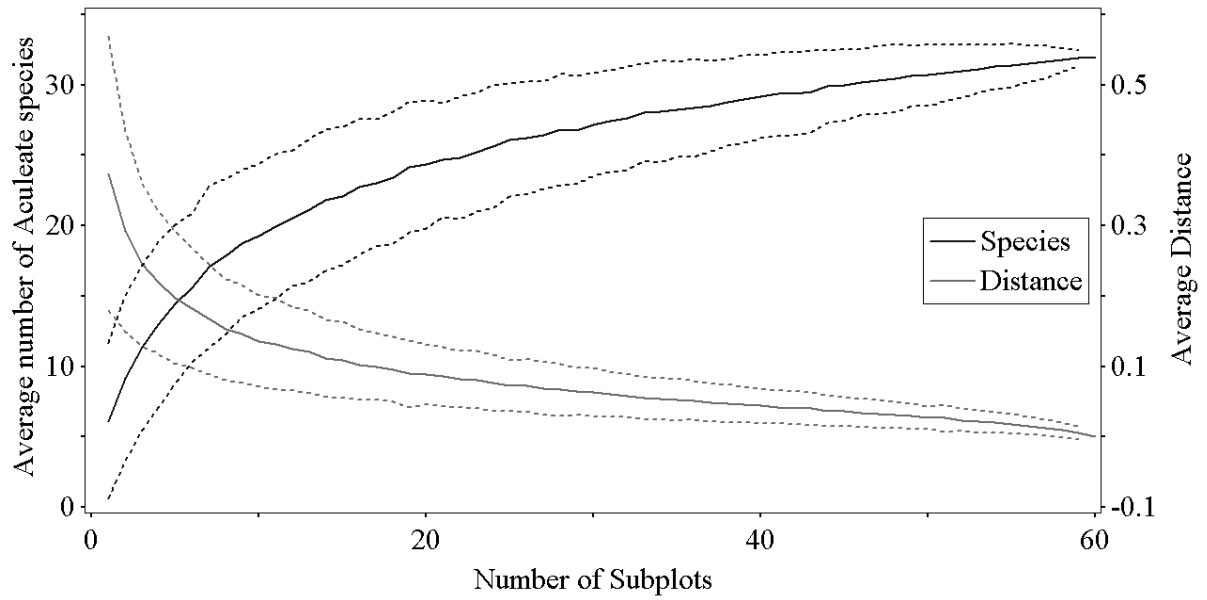
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625 **Table 2.** F-value (*F*) and level of significance (*P*) for aculeate Hymenoptera (Aculeate) species
626 richness and abundance at farms in Co. Sligo (Ireland) in 2018 with regard to farming intensity
627 (Intensity), farmland linear habitat (Habitat) and trap side (Trap Side). Numbers in bold
628 indicate significance ($P < 0.05$).

629

| | Intensity | | Habitat | | Trap Side | |
|------------------|-----------|--------------|----------|--------------|-----------|------------------|
| | <i>F</i> | <i>P</i> | <i>F</i> | <i>P</i> | <i>F</i> | <i>P</i> |
| <i>Aculeate</i> | | | | | | |
| Species Richness | 5.43 | 0.007 | 3.49 | 0.037 | 6.97 | 0.011 |
| Abundance | 1.94 | 0.165 | 2.49 | 0.103 | 15.84 | <0.001 |

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633 **Fig. 1.** Species-area curves for aculeate Hymenoptera (Aculeate) collected from Malaise traps
 634 across all selected farms in Co. Sligo (Ireland) in 2018. Dotted lines represent \pm SDs. First-
 635 Order Jackknife estimate of total species richness was 38.88.

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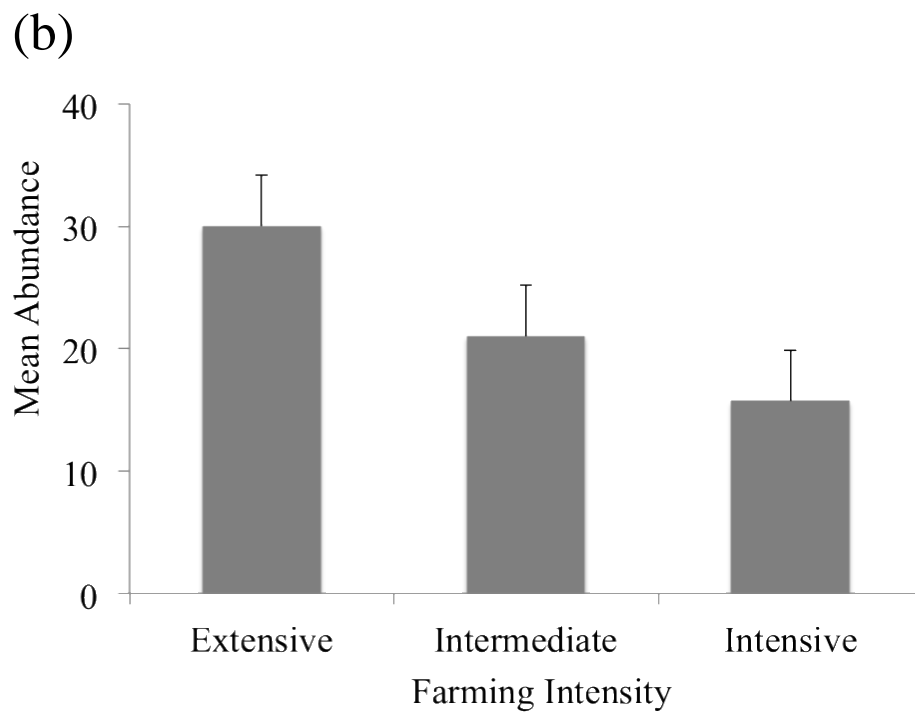
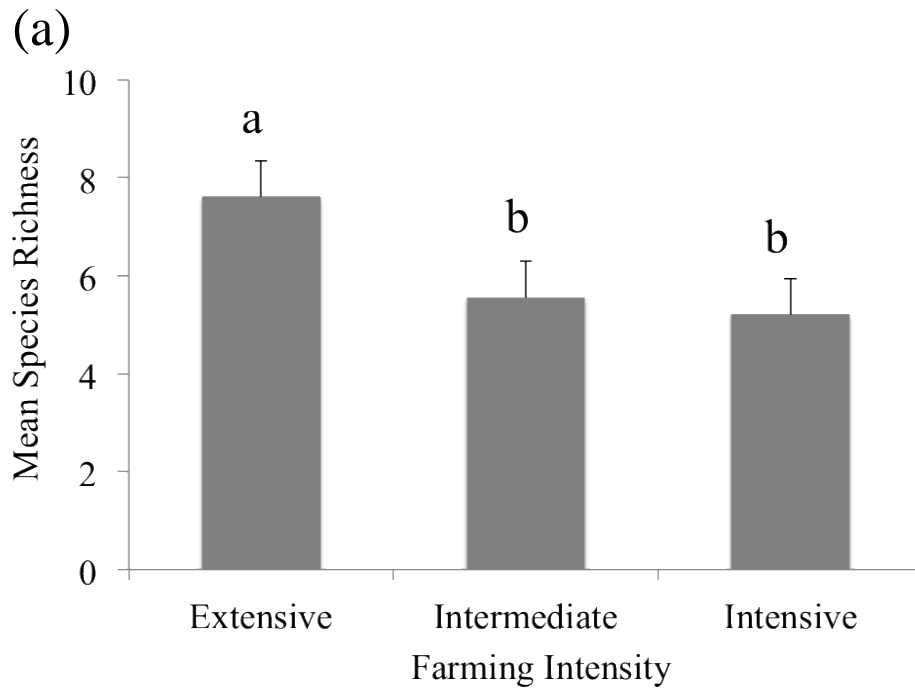
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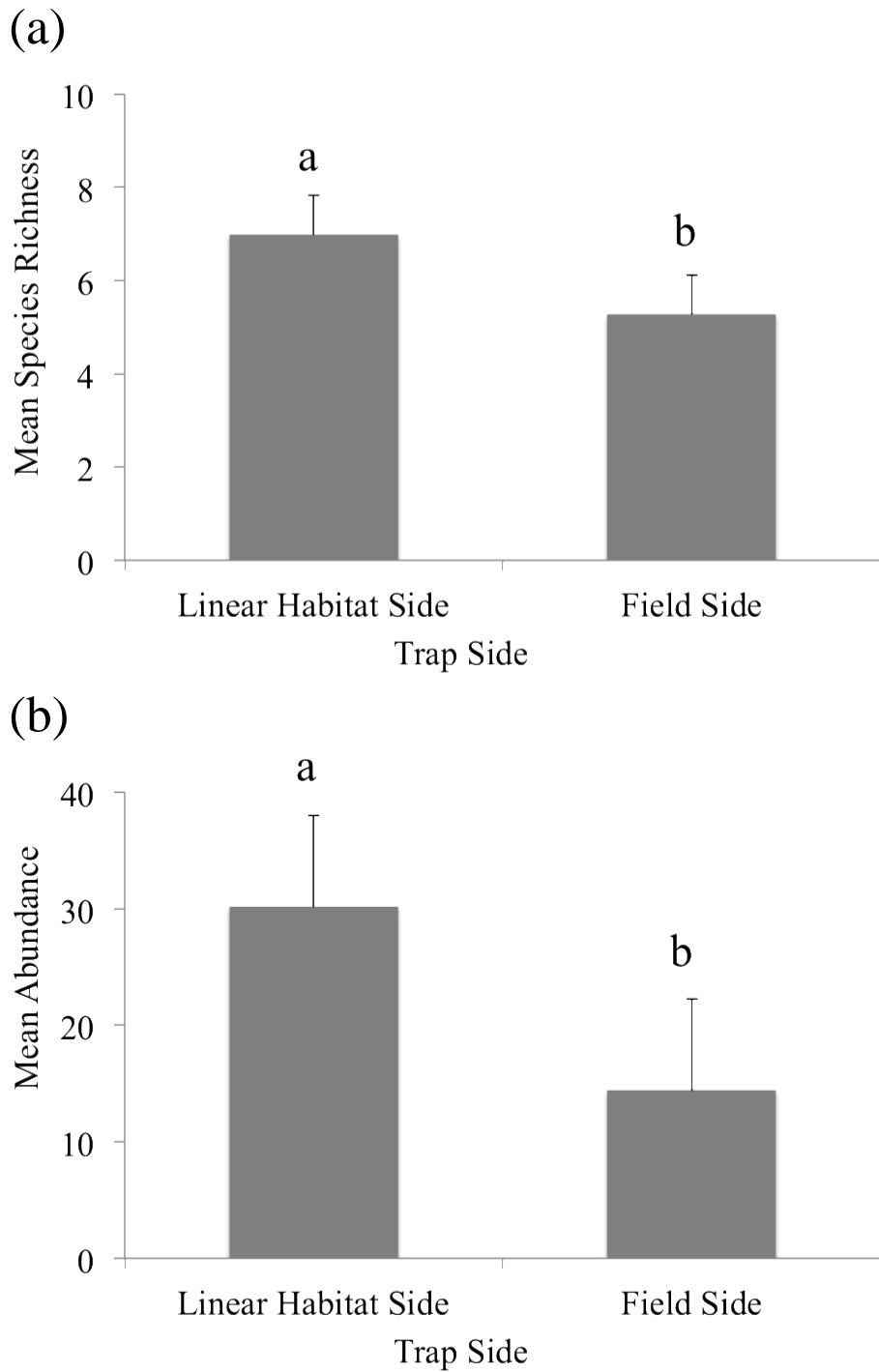
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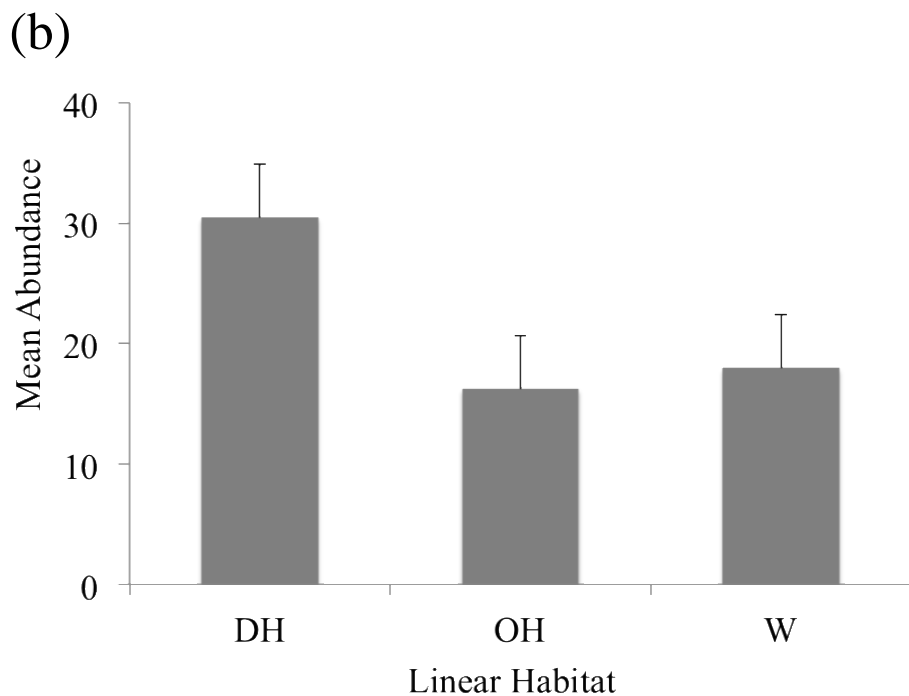
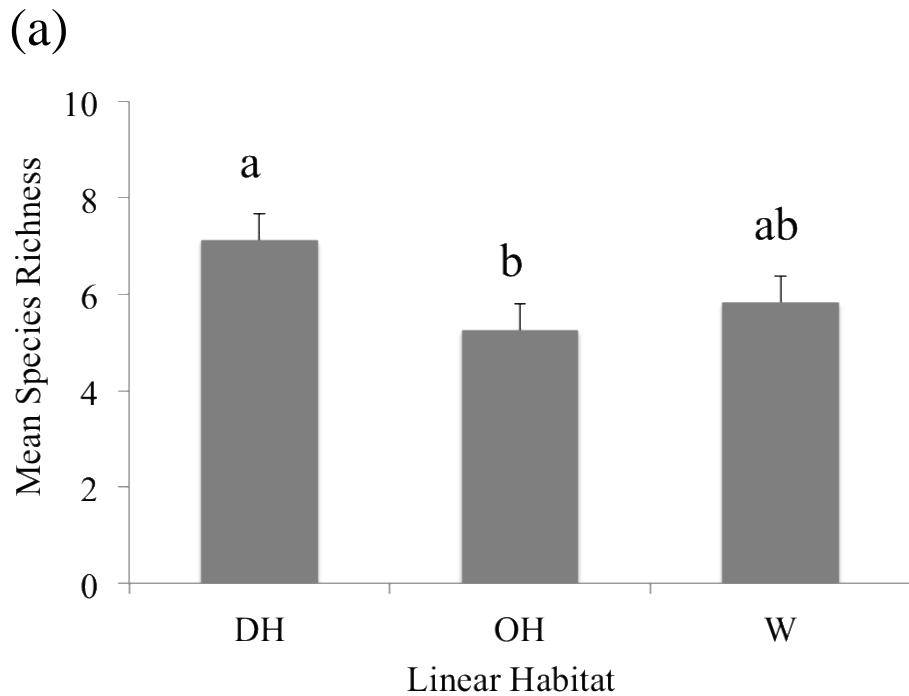
650 **Fig. 2.** Mean (a) species richness and (b) abundance of aculeate Hymenoptera across each
651 farming intensity in Co. Sligo (Ireland) in 2018. Error bars represent SE. Different letters over
652 the bars indicate significant differences between categories ($P < 0.05$).

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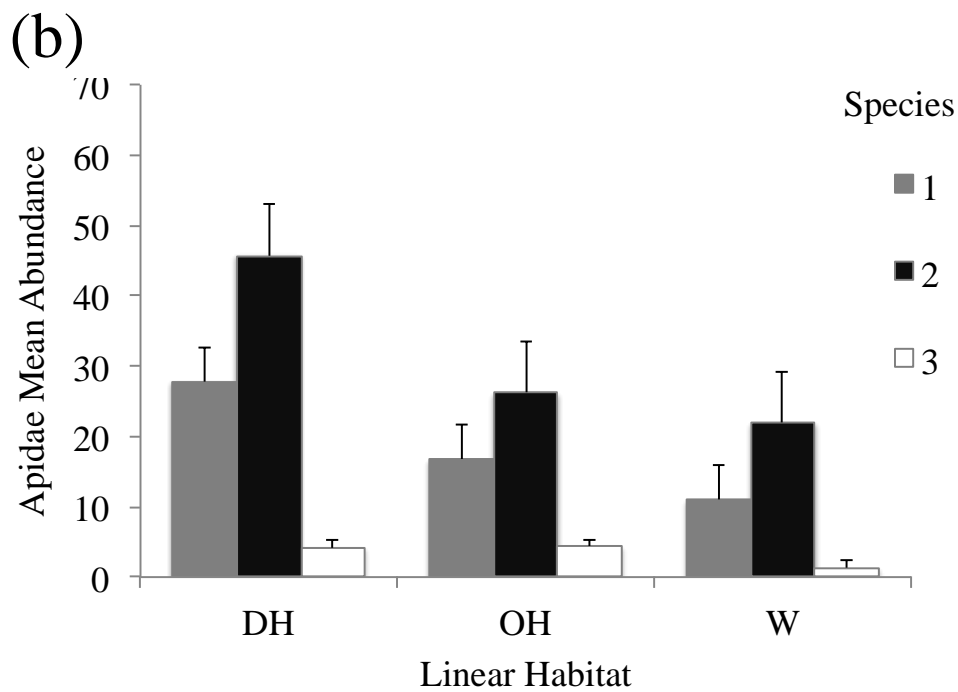
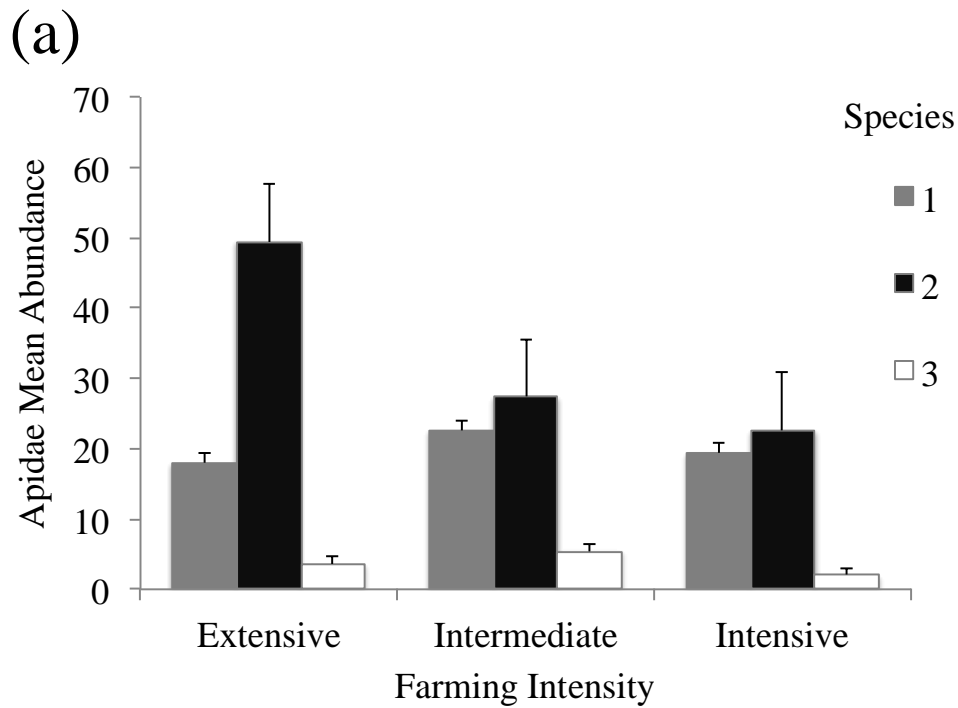
655 **Fig. 3.** Mean (a) species richness and (b) abundance of aculeate Hymenoptera with reference
 656 to trap side in Co. Sligo (Ireland) in 2018: side of the trap facing the farmland linear habitat
 657 (Linear Habitat Side) and side of the trap facing open field (Field Side). Error bars represent
 658 SE. Different letters over the bars indicate significant differences between categories ($P <$
 659 0.05).



660

661 **Fig. 4.** Mean (a) species richness and (b) abundance of aculeate Hymenoptera across each
 662 farmland linear habitat type in Co. Sligo (Ireland) in 2018: DH (dense hedgerow); OH (open
 663 hedgerow); and W (watercourse). Error bars represent SE. Different letters over the bars
 664 indicate significant differences between categories ($P < 0.05$).

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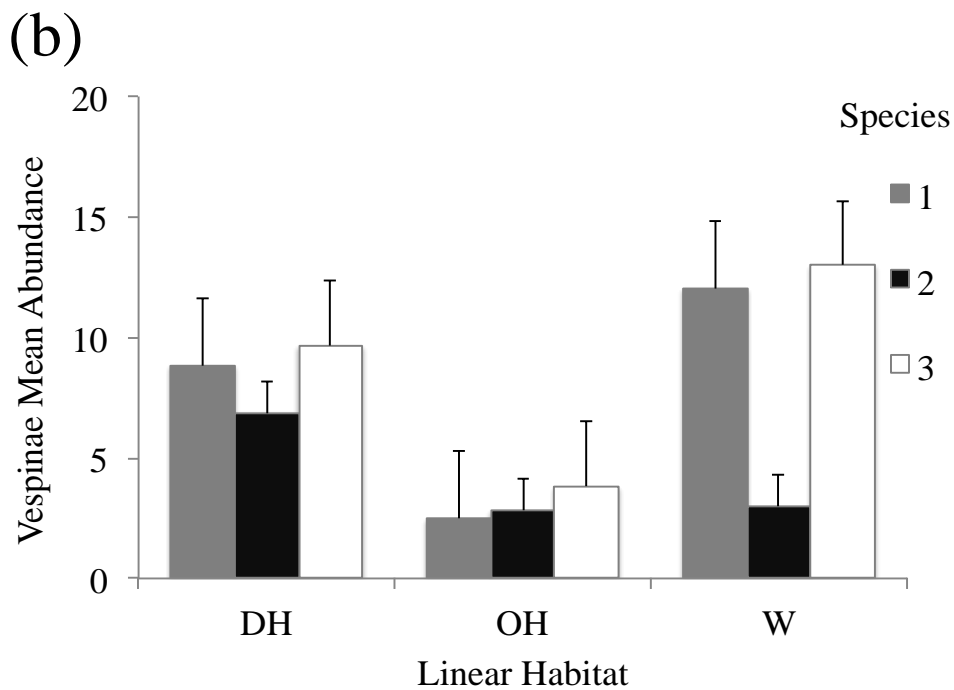
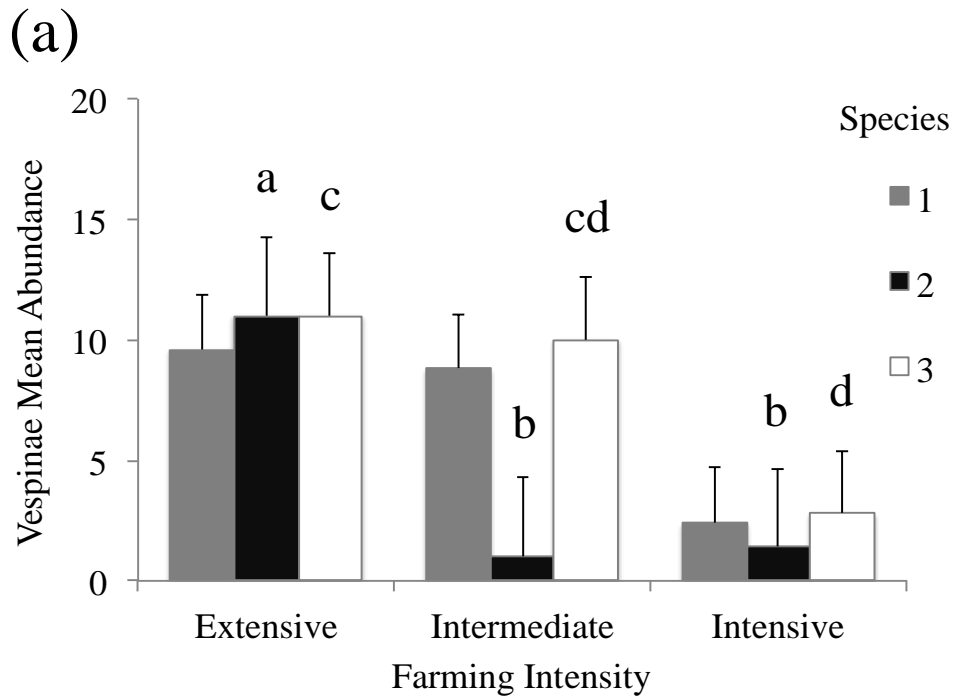
667 **Fig. 5.** Mean abundance of the three most abundant Apidae species across: (a) farming intensity

668 and (b) farmland linear habitat at farms in Co. Sligo (Ireland) in 2018: DH (dense hedgerow);

669 OH (open hedgerow); and W (watercourse). 1 (*Bombus lucorum*); 2 (*Bombus pascuorum*); 3

670 (*Bombus pratorum*). Error bars represent SE.

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673 **Fig. 6.** Mean abundance of the three most abundant Vespinae species across: (a) farming
 674 intensity and (b) farmland linear habitat at farms in Co. Sligo (Ireland) in 2018: DH (dense
 675 hedgerow); OH (open hedgerow); W (watercourse). 1 (*Vespula germanica*); 2 (*Vespula rufa*);
 676 3 (*Vespula vulgaris*). Error bars represent SE. Different letters over the bars indicate significant
 677 differences when they occur within each species ($P < 0.05$).

678 **Supporting Information**

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680 **Appendix S1.** Site classification based on farming intensity across all selected farms in Co.

681 Sligo, Ireland: extensive, intermediate and intensive; and farmland linear habitat type: (DH)

682 dense hedgerow, (OH) open hedgerow, and W (watercourse).

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| Sites | Farmland Linear Habitat | HNV ¹ |
|---------------------|-------------------------|------------------|
| <i>Extensive</i> | | |
| 1 | DH | 6.9 |
| 2 | OH | 6.9 |
| 3 | OH | 7.5 |
| 4 | DH | 7.5 |
| 5 | W | 8.2 |
| <i>Intermediate</i> | | |
| 6 | DH | 3.8 |
| 7 | OH | 3.8 |
| 8 | DH | 4.1 |
| 9 | W | 3.9 |
| 10 | OH | 4.6 |
| <i>Intensive</i> | | |
| 11 | DH | 3.4 |
| 12 | DH | 3.4 |
| 13 | W | 3.4 |
| 14 | OH | 3.3 |
| 15 | OH | 3.3 |

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686 ¹ HNV (High Nature Value) indices were obtained from the maps of each farm and calculated through the web
 687 page <http://www.high-nature-value-farmland.ie/is-your-farm-hnv/>. The score is based on stocking rates (LU/ha),
 688 area of improved grasslands, area owned and farmed, and the visual observations of the size of the farm fields and
 689 field boundaries

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699 **Appendix S2.** List of aculeate Hymenoptera captured during this investigation in 2018 at
 700 selected farms in Co. Sligo (Ireland) separated into each (sub-) family.

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| Apidae - Species List | Overall abundance | Percentage of total abundance |
|---------------------------------------|-------------------|-------------------------------|
| <i>Andrena fucata</i> Smith | 7 | 0.7 |
| <i>Andrena haemorrhoa</i> Fabricius | 2 | 0.2 |
| <i>Andrena scotica</i> Perkins | 14 | 1.5 |
| <i>Bombus hortorum</i> L. | 22 | 2.3 |
| <i>Bombus jonellus</i> Kirby | 34 | 3.6 |
| <i>Bombus lapidaries</i> L. | 5 | 0.5 |
| <i>Bombus lucorum</i> L. | 300 | 31.5 |
| <i>Bombus muscorum</i> L. | 1 | 0.1 |
| <i>Bombus pascuorum</i> Scopoli | 497 | 52.1 |
| <i>Bombus pratorum</i> L. | 55 | 5.8 |
| <i>Bombus sylvestris</i> Lepeletier | 8 | 0.8 |
| <i>Lasioglossum albipes</i> Fabricius | 2 | 0.2 |
| <i>Megachile versicolor</i> Smith | 1 | 0.1 |
| <i>Nomada marshamella</i> Kirby | 2 | 0.2 |
| <i>Nomada ruficornis</i> L. | 1 | 0.1 |
| <i>Sphecodes ephippius</i> L. | 2 | 0.2 |
| <i>Sphecodes monilicornis</i> Kirby | 1 | 0.1 |

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| Crabronidae - Species List | Overall abundance | Percentage of total abundance |
|---|-------------------|-------------------------------|
| <i>Crossocerus dimidiatus</i> Fabricius | 1 | 2.8 |
| <i>Crossocerus megacephalus</i> Rossi | 10 | 27.8 |
| <i>Ectemnius continuus</i> Fabricius | 3 | 8.3 |
| <i>Ectemnius lapidaries</i> Panzer | 8 | 22.2 |
| <i>Mellinus arvensis</i> L. | 13 | 36.1 |
| <i>Pemphredon lugubris</i> Fabricius | 1 | 2.8 |

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| Eumeninae - Species List | Overall abundance | Percentage of total abundance |
|---|-------------------|-------------------------------|
| <i>Symmorphus bifasciatus</i> L. | 12 | 75.0 |
| <i>Ancistrocerus nigricornis</i> Curtis | 1 | 6.2 |
| <i>Ancistrocerus oviventris</i> Wesmael | 3 | 18.8 |

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| Vespinae - Species List | Overall abundance | Percentage of total abundance |
|---|-------------------|-------------------------------|
| <i>Dolichovespula norwegica</i> Fabricius | 24 | 7.4 |
| <i>Dolichovespula sylvestris</i> Scopoli | 7 | 2.1 |
| <i>Vespula austriaca</i> Panzer | 6 | 1.8 |
| <i>Vespula germanica</i> Fabricius | 104 | 31.7 |
| <i>Vespula rufa</i> L. | 67 | 20.4 |
| <i>Vespula vulgaris</i> L. | 120 | 36.6 |

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737 **Appendix S3.** Level of significance (P) from Mann-Whitney U test for the three most abundant
738 Apidae species (*Bombus lucorum*, *Bombus pascuorum*, *Bombus pratorum*) and Vespinae
739 species (*Vespula germanica*, *Vespula rufa*, *Vespula vulgaris*) at farms in Co. Sligo (Ireland) in
740 2018 with regard to farming intensity: extensive, intermediate and intensive; and farmland
741 linear habitat types: DH (dense hedgerow), OH (open hedgerow) and W (watercourse).
742 Numbers in bold indicate significance ($P < 0.05$).

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| | Intensity | | | Habitat | | |
|--------------------------|------------------|--------------|------------------|------------------|----|-------|
| | Comparison types | | P | Comparison types | | P |
| Apidae | | | | | | |
| <i>Bombus lucorum</i> | Extensive | Intermediate | 0.414 | DH | OH | 0.220 |
| | Extensive | Intensive | 0.805 | DH | W | 0.101 |
| | Intermediate | Intensive | 0.300 | OH | W | 0.454 |
| <i>Bombus pascuorum</i> | Extensive | Intermediate | 0.439 | DH | OH | 0.068 |
| | Extensive | Intensive | 0.170 | DH | W | 0.066 |
| | Intermediate | Intensive | 0.327 | OH | W | 0.625 |
| <i>Bombus pratorum</i> | Extensive | Intermediate | 0.400 | DH | OH | 0.833 |
| | Extensive | Intensive | 0.248 | DH | W | 0.156 |
| | Intermediate | Intensive | 0.075 | OH | W | 0.121 |
| Vespinae | | | | | | |
| <i>Vespula germanica</i> | Extensive | Intermediate | 0.931 | DH | OH | 0.119 |
| | Extensive | Intensive | 0.119 | DH | W | 0.957 |
| | Intermediate | Intensive | 0.116 | OH | W | 0.399 |
| <i>Vespula rufa</i> | Extensive | Intermediate | <0.001 | DH | OH | 0.351 |
| | Extensive | Intensive | <0.001 | DH | W | 0.486 |
| | Intermediate | Intensive | 0.710 | OH | W | 0.124 |
| <i>Vespula vulgaris</i> | Extensive | Intermediate | 0.446 | DH | OH | 0.074 |
| | Extensive | Intensive | 0.005 | DH | W | 0.918 |
| | Intermediate | Intensive | 0.056 | OH | W | 0.183 |

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