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Interactions between labile carbon, temperature and land use regulate carbon dioxide and methane production in tropical peat

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KEYWORDS: Tropical peat; Carbon dioxide; Methane; Root exudates; Land use change

Abstract

1 Tropical peatlands are a significant carbon store and contribute to global carbon dioxide (CO₂) and methane (CH₄) 2 emissions. Tropical peatlands are threatened by both land use and climate change, including $3 - 4^{\circ}$ C predicted 3 warming by 2100 and the alteration of regional precipitation patterns. Plant communities in tropical peatlands can 4 regulate greenhouse gas (GHG) fluxes through labile carbon inputs, but the extent to which these inputs regulate 5 the temperature response of CO₂ and CH₄ production in tropical peat remains unclear. We conducted an anoxic 6 incubation experiment using three peat types derived from contrasting botanical origins to assess how carbon 7 addition affects the temperature response (Q10) of CO2 and CH4 production. Peats from a forested peatland in 8 Panama and Malaysia, and a converted oil palm and pineapple intercropping system in Malaysia differed 9 significantly in terms of redox potential, total carbon and carbon: nitrogen ratios. The production of CO₂ and CH₄ 10 varied significantly among peat types and increased with increasing temperature. Carbon addition further 11 increased gas fluxes, but did not influence the Q₁₀ for CO₂ or CH₄ production. These findings demonstrate that 12 CO₂ and CH₄ production in tropical peats are sensitive to warming, that potential emissions vary among contrasting peat types, but that the effect of root inputs in altering Q_{10} appears to be limited. 13

14 Introduction

Global atmospheric concentrations of carbon dioxide (CO₂) and methane (CH₄) are increasing, driven by anthropogenic activities and accelerating climate change (IPCC, 2013). Tropical peatlands represent an important component of the carbon cycle, being an important source of both CO₂ and CH₄ and a major store of carbon. Indeed, tropical peatlands account for 11% of total peatland area but 15-19% of peat carbon stocks worldwide, equivalent to approximately 104.7 Gt C, with significant deposits reported throughout the tropics in Central and South America, Central Africa, and Southeast Asia (Dargie et al., 2017; Page et al., 2011).

21

Greenhouse gas (GHG) emissions from tropical peatlands are strongly regulated by peat temperature, with increased temperature associated with greater fluxes *in situ* (Jauhiainen et al., 2014). This is particularly important in the context of predicted climate change for tropical peatlands globally. Current estimates of air temperature changes in the Neotropics and Southeast Asia are for $3 - 4^{\circ}$ C warming by 2100 (IPCC, 2013). Previous *ex situ* studies have demonstrated that the temperature response of tropical peats to warming is not linear (Sjögersten et al., 2018), meaning that relatively small increases in temperature have the potential to dramatically increase emissions.

29

30 In addition to changing regional climate patterns, many tropical peatlands are under significant threat from 31 changes in land use. In Southeast Asia, the drainage, deforestation and expansion of plantation agriculture has the 32 potential to significantly alter the balance of emissions from peatland sites (Hergoualc'h and Verchot, 2014). 33 Conversion to plantation agriculture has previously been shown to significantly alter organic matter properties 34 (Cooper et al., 2019; Tonks et al., 2017). In turn, this may affect the response of these ecosystems to environmental change, including to elevated temperatures and altered patterns of precipitation. Differences in organic matter 35 36 properties between contrasting vegetation types have also been reported in Neotropical systems (Girkin et al., 37 2019; Hoyos-Santillan et al., 2015; Upton et al., 2018).

38

Root exudates represent an important plant carbon input which, depending on their composition, can contribute significantly to net greenhouse gas emissions in tropical peats even at relatively low addition rates (Girkin et al., 2018a, 2018b). Root respiration, which includes microbial use of exudates, can be the dominant driver of CO₂ fluxes *in situ* in tropical peatlands (Girkin et al., 2018c; Melling et al., 2013). Emissions of both CO₂ and CH₄ are also driven by the decomposition of peat organic matter, with the extent of production determined by substrate

quality (Bridgham and Richardson, 1992; Hoyos-Santillan et al., 2015). Under low oxygen to anoxic conditions that can predominate in peatlands, methanogenic archaea produce CH₄ through the reduction of acetate (acetoclastic methanogenesis), the reduction of CO₂ and H₂ (hydrogenotrophic methanogenesis), or the cleavage of methylated organic compounds (Holmes et al., 2015; Kolton et al., 2019; Le Mer and Roger, 2001). These pathways are strongly influenced by prevalent environmental conditions *in situ*, including substrate and nutrient availability, microbial community structure, water table depth, and temperature (Couwenberg et al., 2009; Gorham, 1991; Le Mer and Roger, 2001).

51

52 The temperature sensitivity of soil respiration under aerobic conditions increases with the recalcitrance of organic 53 matter due to the higher activation energies required (Fierer et al., 2005; Lloyd and Taylor, 1994). Tropical (and 54 some temperate) peats have been shown to have a lower carbohydrate content and a higher proportion of aromatic compounds compared to boreal peatlands (Hodgkins et al., 2018), therefore requiring higher energy yield terminal 55 56 electron acceptors to drive decomposition, which are likely less available in the low oxygen to anaerobic conditions of peatlands (Keiluweit et al., 2016). The degradation of root exuded labile carbon compounds, which 57 can include a range of organic acids, sugars and amino acids, are likely to therefore have a lower sensitivity than 58 59 other organic matter components, the oxidation of which may be coupled to lower yielding terminal electron 60 acceptors (Davidson and Janssens, 2006; Keiluweit et al., 2016). Previous ex situ studies of the response of tropical 61 peat have not accounted for the influence of continuous low input rates of labile carbon (Sjögersten et al., 2018), 62 for example low concentrations of glucose, a common component of plant root exudate profiles (Smith, 1976).

63

64 Developing an understanding of temperature sensitivity of peat and soils is important in accurately assessing future changes in the global carbon cycle in response to rises in temperature. Strong responses of greenhouse gas 65 66 production to increased temperature, and greater temperature sensitivity with increasing carbon substrate 67 recalcitrance are predicted by kinetic theory (Davidson and Jannssens, 2006) and supported by some experimental 68 data (Conant et al., 2008). Decomposition of recalcitrant peatland carbon (particularly highly aromatic tropical 69 peat) may also be subject to thermodynamic limitation due to a shortage of terminal electron acceptors (Beer and 70 Blodau, 2007). Previous studies in mineral soils have reported an increase in the temperature sensitivity of aerobic 71 respiration on the addition of labile carbon (Gershenson et al., 2009; Nianpeng et al., 2013; Song et al., 2010), 72 most likely because microbial respiration in these soils was limited by carbon lability. High latitude peatlands 73 have previously been demonstrated to have high temperature sensitivity for CH₄ production (Bergman et al., 1998). Increased temperatures have also been shown to result in substantial changes in microbial community structure, possibly resulting in changes in methanogenic pathways (Lupascu et al., 2012). However, it is unclear how this applies in tropical peatlands with high organic matter content (> 60%), but a high aromatic content, and how responses vary between dominant vegetation types with different litter inputs (Upton et al., 2018, Cooper et al., 2019), and under waterlogged and low oxygen conditions (Wright et al., 2013).

79

In this study, we assessed the response of CO_2 and CH_4 production of three tropical peats to elevated temperatures and carbon addition, in the form of glucose. Based on the strong role of organic matter quality in regulating greenhouse gas emissions we predicted that i) basal CO_2 and CH_4 production would differ among peat types, and ii) temperature sensitivity would differ among peat types. As labile carbon can be rapidly depleted during decomposition, and peat is predominantly composed of recalcitrant biomolecules, we also hypothesised that iii) glucose addition would increase Q_{10} for all peat types due to waterlogged, low oxygen conditions, and strong substrate limitation resulting in thermodynamic limitation.

87

88 Methods

89 Study sites

This study was conducted using peat samples collected from sites in Panama and Malaysia between May 2016 90 91 and July 2017. Panamanian peat samples were collected in May 2016 from the 80 km² ombrotrophic peatland at 92 Changuinola, part of the San San Pond Sak freshwater and marine wetland located in Bocas del Toro province (Figure 1a). The site was located approximately 600 m from the coast (09° 18' 13.00"N, 82° 21' 13.80"W) in a 93 94 mixed forest stand. The central peat dome is approximately 8 m deep and was initiated approximately 4000–5000 years ago (Phillips et al., 1997). The site features seven distinct plant phasic communities beginning with a 95 96 Rhizophora mangle mangrove swamp on the coastal margins, which is succeeded by palm swamp dominated by 97 Raphia taedigera, a mixed forest stand, a monodominant Campnosperma panamensis forest stand, and a Myrica-98 Cyrilla bog-plain (Phillips et al., 1997). This vegetation gradient is matched by a pronounced decrease in nutrient 99 availability, particularly phosphorus (Cheesman et al., 2012; Sjögersten et al., 2011). The microbial community 100 throughout the peatland is dominated by Acidobacteria, with precise community composition clustered by 101 phosphorus availability (Troxler et al., 2012). Sampling was conducted in the mixed forest stand dominated by C. 102 panamensis, R. taedigera, and Symphonia globulifera.

104 The Malaysian forested peatland site is located in Terengganu state, in northeastern Peninsular Malaysia. The site 105 is approximately 8.42 km² and, and is 11.3 km from the coast, located in Kampung Mat Jintan (5°25'16.2"N, 106 102°55'46.2"E) in the boundary between Kula Nerus and Setiu districts (Figure 1b). The forest vegetation comprised trees that were up to 40 m tall and with a diameter-at-breast-height (DBH) of 40 - 50 cm. Common 107 108 species included Antisoptera sp., Shorea sp., Calophyllum sclerophyllum, Calophylum sp., Blumeondendron 109 tokbrai, Durio carinatus, Gonostylus bancanus, Elateriospermum tapos, and Syzgium sp. Both Macaranga 110 pruinosa and M. gigantean were present on the forest edges. Pandanus helicopus and Nepenthes ampullaria were 111 common understory species. Peat depth was approximately 2 m.

112

The Malaysian oil palm and pineapple intercropping site is located in Selangor State, approximately 14 km from the west coast of Peninsular Malaysia (3°25'20.6"N, 101°19'56.6"E). The site is surrounded by recently planted (2014) 2nd generation oil palm mono-cropping and is drained by two drainage ditches along the borders at opposite sides of the site (Dhandapani et al., 2019b). During sampling the site was not flooded, although the peat was moist, with high gravimetric moisture content.

118

119 At each site, three $10 \times 10 \times 10$ cm bulk peat samples were collected using a hand trowel, excluding any recent 120 surface litter. Samples were shipped to the University of Nottingham and kept at 4°C for one month prior to 121 analysis.

122

123 Experimental design

124 Peat samples (20 g) from each site, were placed in stainless steel chambers (0.6 dm³) with polypropylene lids and 125 a silicon ring seal. Two holes were drilled in the lids and fitted with three-way stopcock valves silicon-sealed in 126 place. The gas-tightness of chambers was tested by closing all valves and submersing in water, and through 127 flushing the chamber with N₂ and observing for changes in headspace CO₂ and CH₄ concentrations over time prior 128 to adding peat. Each peat sample was mixed with 20 ml of deionised water to mimic in situ flooded conditions. 129 Chambers were flushed with N₂ to displace accumulated headspace gases for two minutes and were placed in 130 incubators set at 25, 30 and 35°C for acclimation of microbial communities for seven weeks following sample storage and preparation. temperatures were selected to represent broad scale warming of peatlands in the Central 131 132 Americas and in Southeast Asia (IPCC, 2013). Chambers were subsequently opened to displace accumulated headspace gases, flushed again with N_2 for two minutes and sealed. Overall conditions most closely match the flooded oxic conditions outlined in Sjogersten et al. (2018), namely, water saturated with low oxygen, but not entirely anoxic, mostly closely resembling peat *in situ* conditions in the 0 – 10 cm horizon (Girkin, 2018; Hoyos-Santillan et al., 2016).

137

Glucose solutions were prepared for an input rate equivalent to 0.1 mg C g^{-1} soil per day. This addition rate represents a relatively low daily addition rate for plant carbon inputs (Grayston and Campbell, 1996). Solutions were adjusted to a pH of 5.5 using HCl sterilised, and stored at 4°C prior to addition. Oxygen was not removed from exudate solutions prior to addition to better mimic the combined inputs of oxygen and labile carbon at the root-peat interface (Hoyos-Santillan et al., 2016).

143

During headspace sampling, chambers were connect to a Los Gatos ultraportable greenhouse gas analyser (San Jose, California), sampling at 0.5 Hz. Fluxes were subsequently measured over 90 seconds, with measurement occurring immediately prior to glucose addition, and at 6, 24, 72 and 120 hours following the first addition. Glucose (or deionised water) was added to each chamber at the rate of 1 ml per day. Gas concentrations were adjusted for incubation temperature ($25 - 35^{\circ}$ C), as well tube and optical bench volume, according to the ideal gas law. The rate of potential gas production, expressed as µg CO₂ g⁻¹ hr⁻¹ or µg CH₄ g⁻¹ hr⁻¹, was calculated assuming a linear accumulation rate of gases in the headspace (Hogg et al., 1992).

- 151
- 152 Q_{10} , describing the temperature sensitivity of CO_2 and CH_4 production was calculated as:
- 153 $Q_{10} = 10^{(m \times 10)}$
- 154 Where m is the regression slope of a $\log_{10} \text{CO}_2$ or CH₄ flux versus temperature plot.
- 155

156 Peat characterisation

Sub-samples from each site were used to characterise peat physiochemical properties. Gravimetric water content was determined by analysis of the mass of water lost from 10 g fresh peat oven dried at 105°C for 24 hours. Organic matter content was determined as the mass lost after ignition for 7 hours at 550°C. Total carbon (C) and total nitrogen (N) content were determined from 0.2 g of dry, homogenised peat combusted using a total element analyser (Flash EA 1112, CE Instruments, Wigan, UK). pH and redox potential were measured using a Hanna 209 meter coupled with pH and redox probes following one week acclimation but prior to beginning the
experiment. After measurement, chambers were flushed with N₂ and resealed.

164

165 Statistical analysis

166 Differences in CO₂ and CH₄ production and Q_{10} s were tested using the restricted maximum likelihood method 167 (REML), including site, temperature and presence/absence of glucose as fixed effects and sample replicate as a 168 random effect. CO₂ and CH₄ fluxes were log-transformed to meet test assumptions. Significance was assessed at 169 p < 0.05. All statistical analyses were conducted using Genstat v17.

170

171 **Results**

172 Peat biochemical properties

Peats were acidic (pH < 5), with high gravimetric moisture and organic matter contents. These properties were not significantly different among peat types (p > 0.05). There was, however, a significant difference in redox potential among sites (p = 0.003), with substantially lower redox potential in the Malaysian primary forest peats compared to the Panamanian forest or intercropping site (Table 1). Total carbon was also significantly different among sites (p = 0.021), and was greatest in pineapple intercropping sites. Total nitrogen did not differ significantly among peat types (p = 0.134). C:N was significantly different among peat types (p = 0.003), and was lowest in Panamanian forest peat but broadly comparable between Malaysian peats.

180

181 Basal peat CO₂ and CH₄ production

Mean basal CO₂ production varied between $69.0 - 77.7 \ \mu g \ CO_2 \ g^{-1} \ hr^{-1}$ (Figure 2) but did not differ significantly among peat types (p = 0.151). CH₄ production, however, varied significantly among peat types (p = 0.02). The magnitude of CH₄ production compared to CO₂ was considerably lower, ranging from 0.06 – 0.08 $\mu g \ CH_4 \ g^{-1} \ hr^{-1}$. Greatest CO₂ and CH₄ production occurred in Malaysian primary forest peats, with the most variable production

186 occurring in peats from the intercropping sites.

187

188 Temperature and labile carbon sensitivity

189 Temperature strongly influenced CO_2 fluxes, with increased production relative to basal rates for all sites at both

190 30° C and 35° C (p < 0.001). Glucose addition significantly increased CO₂ production (p < 0.001), with a significant

(p < 0.001) interaction between glucose addition and peat type, with a 178% increase in mean fluxes from
 Malaysian primary forest peats, but only 34% for the Panamanian mixed forest peat.

193

194 CH₄ production increased significant with temperature (p < 0.001), with mean production increasing at both 30 195 and 35°C (Figure 3b,d,f). Glucose addition increased mean CH₄ fluxes by 11% (p = 0.034). There was also a 196 significant interaction between glucose addition and peat type (p < 0.001). In Malaysian forest and Panamanian 197 forest peats, there were 5 and 7% declines in mean CH₄ fluxes respectively on glucose addition, but there was a 198 56% increase in mean fluxes for the intercropping site.

199

There were no significant differences in temperature sensitivity (Q_{10}) of CO₂ or CH₄ production in the presence or absence of glucose (p > 0.05, Table 2), between sites (p > 0.05), or in the interaction between treatment and sites (p > 0.05).

203

204 Discussion

205 Warming promoted CO_2 and CH_4 production in all peat types, in common with previous studies in both temperate 206 and boreal (Dunfield et al., 1993, Inglett et al., 2012), and tropical peats (Sjögersten et al., 2018), as well as drained 207 lowland tropical soils from Peru (Nottingham et al., 2015). Previously, it has been proposed that heterotrophic 208 microbial communities in tropical peat respond weakly to warming primarily due to adaptation to high peat 209 temperatures, and because optimal temperature for respiration and methanogenesis is approximately 25°C (Kolton et al., 2019; Menichetti et al., 2015; Sjögersten et al., 2018). Q₁₀ for both CO₂ (1.07 - 2.25) and CH₄ (1.10 - 2.39) 210 211 production were relatively low in our study, particularly when compared to 6.1 for anaerobic CH₄ production 212 reported for Panamanian peats (Sjögersten et al., 2018), although they are closer to the 2.2 - 3.7 reported from studies of CH₄ production in Central Kalimantan, Indonesia (Brady, 1997; Hirano et al., 2009; Jauhiainen et al., 213 214 2014), and 1.8 under flooded oxic conditions from Panamanian peats (Sjögersten et al., 2018). Previous studies have reported a higher temperature sensitivity of CH₄ production than CO₂ production, although we found no 215 216 supporting evidence for this (Table 2) (Sjögersten et al., 2018, and references therein). These results have clear implications in assessing the impact of future environmental change on gaseous carbon emissions from tropical 217 peatlands: warming peat will result in increased CO₂ and CH₄ emissions from flooded, low oxygen peats. 218

220 Although there was no difference in basal CO_2 production (from unamended peats at 25°C), or in temperature 221 sensitivity among peat types, both CO₂ and CH₄ fluxes varied significantly among sites across the three 222 temperatures. While organic matter content was similar among peat types, there were significant differences in total carbon and C:N, likely reflecting contrasting aboveground vegetation and management practices. Previous 223 224 work in Malaysia (Tonks et al., 2017, Cooper et al., 2019) and Panama (Hoyos-Santillan et al., 2015, Upton et al., 225 2018, Girkin et al., 2019) has demonstrated substantial differences in organic matter properties between contrasting plant functional types and land uses, driven by differences in plant litter inputs and decomposition 226 227 rates. While various studies have assessed the impacts of land use change on organic matter properties (Cooper et 228 al., 2019; Könönen et al., 2018; Tonks et al., 2017), there is limited data available regarding the role of 229 intercropping systems on organic matter properties, meaning differences in responses cannot be readily ascribed to contrasts in organic matter properties alone. The low CH4 fluxes from the Malaysian intercropping system may 230 reflect significant depletion of labile carbon (Cooper et al., 2019), but could also be driven by a poorly adapted 231 232 methanogenic community, or a highly active methanotrophic community under partially aerobic conditions 233 (Andersen et al., 2013), as well as a redox potential c. 300 mV.

234

235 Changes in CO_2 and CH_4 production in response to glucose addition varied between peat types. At 35°C, CH_4 236 fluxes were greater from unamended Panamanian and Malaysian forest peats compared to carbon amended peats, 237 implying that labile carbon availability was not a limiting factor for production at this temperature. Similarly, CO₂ production in peat from the Panamanian primary forest was comparable at 30 - 35°C even with glucose 238 239 amendment implying an additional limiting factor other than temperature. Rates of CO₂ and CH₄ production in 240 tropical peatlands have previously been reported as lower at low fertility (Sjögersten et al., 2011), with litter decomposition partially constrained by nutrient availability (Hoyos Santillan et al., 2018). The higher response of 241 242 the Malaysian intercropping peat to glucose addition may reflect the effects of management practices, specifically 243 fertiliser addition. This may substantially alter the temperature sensitivity of the system by alleviating inorganic 244 nitrogen (NH₄⁺/NO₂⁻/NO₃⁻) limitation (Liu et al., 2016; Wang et al., 2010), although in this study only total peat 245 nitrogen was assessed and this did not differ significantly between peat types. The lack of significant difference in temperature sensitivity of CO_2 and CH_4 production with glucose addition may be because despite an increase 246 247 in carbon lability, unamended peats still had sufficient available carbon for respiration due to high organic matter content (Dai et al., 2017). Alternatively, the system is thermodynamically limited due to a shortage of high energy 248 terminal electron acceptors necessary for the decomposition of recalcitrant aromatic carbon which is likely under 249

the lox oxygen conditions (Hodgkins et al., 2018; Kolton et al., 2019), and with the relatively low concentration
glucose additions being rapidly consumed (Girkin et al., 2018a).

252

Fully understanding the impact of increased temperature on fluxes in situ is more complex due to the additional 253 254 regulatory roles of microtopography (Jauhiainen et al., 2005), water table changes (Wright et al., 2013), plant inputs of oxygen (Hoyos-Santillan et al., 2016), and small scale heterogeneity of peat organic matter properties 255 256 (Girkin et al., 2019). In temperate peatlands, vegetation cover has been shown to also have a significant impact 257 on the temperature sensitivity of GHG production (Leroy et al., 2017). Increasing atmospheric CO₂ has been found 258 to increase rates of root exudation in wetland ecosystems (Sánchez-Carrillo et al., 2018) and increases in 259 temperature have also been reported to enhance rates of root exudation in some tree species (Uselman et al., 2000), and alter the composition of exudate profiles (Badri and Vivanco, 2009; Vančura, 1967), both known to be critical 260 261 regulators of GHG emissions and peat properties (Girkin et al., 2018a & 2018b). As a consequence, the true 262 response of *in situ* net emissions of GHGs will comprise components driven by both the temperature sensitivity of the peat itself, and any changes in root inputs. 263

264

265 Taken together, our results demonstrate that increased temperatures will substantially increase CO₂ and CH₄ 266 production in tropical peats, regardless of current land cover. As a consequence, predicted warming for Central 267 America and Southeast Asia will be associated with substantial increases in gaseous carbon emissions (IPCC, 2013). However, the extent of this increase is likely to be lower than the response of higher latitude peatlands for 268 which higher Q_{108} have been reported, including 2.4 – 5.8 for a *Sphagnum* peatland (Lupascu et al., 2012) and 2.5 269 270 -35 (Bergman et al., 1998). Differences in fluxes between peat types are likely driven by contrasts in organic 271 matter properties (Cooper et al., 2019) and/or nutrient availability (Hoyos Santillan et al., 2018; Sjögersten et al., 272 2011) and microbial community structure and function. Processes that alter organic matter properties, for example 273 the conversion of pristine forest to oil palm, or the use of intercropping species, will further affect fluxes. In 274 addition, our findings demonstrate that the temperature sensitivity of CO₂ and CH₄ production is not affected by 275 labile carbon addition. This is important because plants can input significant quantities of labile carbon which are 276 not accounted for in the majority of studies of GHG temperature sensitivity.

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287 **References**

- Andersen, R., Chapman, S.J., Artz, R.R.E., 2013. Microbial communities in natural and disturbed peatlands: A
 review. Soil Biology & Biochemistry 57, 979–994. doi:10.1016/j.soilbio.2012.10.003
- Badri, D. V, Vivanco, J.M., 2009. Regulation and function of root exudates. Plant Cell and Environment 32, 666–
 681. doi:DOI 10.1111/j.1365-3040.2009.01926.x
- Beer, J., Blodau, C., 2007. Transport and thermodynamics constrain belowground carbon turnover in a northern
 peatland>. Geochimica et Cosmochimica Acta. doi:10.1016/j.gca.2007.03.010
- 294 Bergman, I., Svensson, B.H., Nilsson, M., 1998. Regulation of methane production in a swedish acid mire by pH,
- temperature and substrate. Soil Biology and Biochemistry. doi:10.1016/S0038-0717(97)00181-8
- Brady, M.A., 1997. Organic matter dynamics of coastal peat deposits in Sumatra, Indonesia. Department of
 Forestry. University of British Columbia.
- Bridgham, S.D., Richardson, C.J., 1992. Mechanisms controlling soil respiration (CO2 and CH4)in southern
 peatlands. Soil Biology & Biochemistry 24, 1089–1099.
- Cheesman, A.W., Turner, B.L., Reddy, K.R., 2012. Soil Phosphorus Forms along a Strong Nutrient Gradient in a
 Tropical Ombrotrophic Wetland. Soil Science Society of America Journal 76, 1496–1506.
 doi:10.2136/sssaj2011.0365
- Conant, R.T., Drijber, R.A., Haddix, M.L., Parton, W.J., Paul, E.A., Plante, A.F., Six, J., Steinweg, M.J., 2008.
 Sensitivity of organic matter decomposition to warming varies with its quality. Global Change Biology.
- 305 doi:10.1111/j.1365-2486.2008.01541.x
- Cooper, H. V., Vane, C.H., Evers, S., Aplin, P., Girkin, N.T., Sjögersten, S., 2019. From peat swamp forest to oil
 palm plantations: The stability of tropical peatland carbon. Geoderma 342, 109–117.

- Couwenberg, J., Dommain, R., Joosten, H., 2009. Greenhouse gas fluxes from tropical peatlands in south-east
 Asia. Global Change Biology 16, 1715–1732. doi:10.1111/j.1365-2486.2009.02016.x
- Dai, S.S., Li, L.J., Ye, R., Zhu-Barker, X., Horwath, W.R., 2017. The temperature sensitivity of organic carbon
 mineralization is affected by exogenous carbon inputs and soil organic carbon content. European Journal of

312 Soil Biology. doi:10.1016/j.ejsobi.2017.06.010

- Dargie, G.C., Lewis, S.L., Lawson, I.T., Mitchard, E.T.A., Page, S.E., Bocko, Y.E., Ifo, S.A., 2017. Age, extent
 and carbon storage of the central Congo Basin peatland complex. Nature. doi:10.1038/nature21048
- Davidson, E.A., Janssens, I.A., 2006. Temperature sensitivity of soil carbon decomposition and feedbacks to
 climate change. Nature. doi:10.1038/nature04514
- Dhandapani, S., Ritz, K., Evers, S., Sjögersten, S., 2019a. Environmental impacts as affected by different oil palm
 cropping systems in tropical peatlands. Agriculture, Ecosystems and Environment.
 doi:10.1016/j.agee.2019.02.012
- Dhandapani, S., Ritz, K., Evers, S., Yule, C.M., Sjögersten, S., 2019b. Are secondary forests second-rate?
 Comparing peatland greenhouse gas emissions, chemical and microbial community properties between
 primary and secondary forests in Peninsular Malaysia. Science of the Total Environment.
 doi:10.1016/j.scitotenv.2018.11.046
- Fierer, N., Craine, J.M., Mclauchlan, K., Schimel, J.P., 2005. Litter quality and the temperature sensitivity of
 decomposition. Ecology. doi:10.1890/04-1254
- Gershenson, A., Bader, N.E., Cheng, W., 2009. Effects of substrate availability on the temperature sensitivity of
 soil organic matter decomposition. Global Change Biology. doi:10.1111/j.1365-2486.2008.01827.x
- 328 Girkin, N.T., 2018. Tropical forest greenhouse gas emissions: root regulation of soil processes and fluxes.
 329 University of Nottingham.
- Girkin, N.T., Turner, B.L., Ostle, N., Craigon, J., Sjögersten, S., 2018a. Root exudate analogues accelerate CO2
 and CH4 production in tropical peat. Soil Biology and Biochemistry 117, 48–55.
 doi:10.1016/j.soilbio.2017.11.008
- Girkin, N.T., Turner, B.L., Ostle, N., Sjögersten, S., 2018b. Composition and concentration of root exudate
 analogues regulate greenhouse gas fluxes from tropical peat. Soil Biology and Biochemistry 127, 280–285.
 doi:S0038071718303444
- Girkin, N.T., Turner, B.L., Ostle, N., Sjögersten, S., 2018c. Root-derived CO 2 flux from a tropical peatland.
 Wetlands Ecology and Management 26, 985–991. doi:10.1007/s11273-018-9617-8

- 338 Girkin, N.T., Vane, C.H., Cooper, H. V., Moss-Hayes, V., Craigon, J., Turner, B.L., Ostle, N., Sjögersten, S.,
- 2019. Spatial variability of organic matter properties determines methane fluxes in a tropical forested
 peatland. Biogeochemistry 142, 231–245.
- Gorham, E., 1991. Northern Peatlands Role in the Carbon-Cycle and Probable Responses to Climatic Warming.
 Ecological Applications 1, 182–195. doi:Doi 10.2307/1941811
- Grayston, S.J., Campbell, C.D., 1996. Functional biodiversity of microbial communities in the rhizospheres of
 hybrid larch (Larix eurolepis) and Sitka spruce (Picea sitchensis). Tree Physiology 16, 1031–1038.
- Hergoualc'h, K., Verchot, L. V, 2014. Greenhouse gas emission factors for land use and land-use change in
 Southeast Asian peatlands. Mitigation and Adaptation Strategies for Global Change 19, 789–807. doi:DOI
 10.1007/s11027-013-9511-x
- Hirano, T., Jauhiainen, J., Inoue, T., Takahashi, H., 2009. Controls on the Carbon Balance of Tropical Peatlands.
 Ecosystems 12, 873–887. doi:DOI 10.1007/s10021-008-9209-1
- 350 Hodgkins, S.B., Richardson, C.J., Dommain, R., Wang, H., Glaser, P.H., Verbeke, B., Winkler, B.R., Cobb, A.R.,
- 351 Rich, V.I., Missilmani, M., Flanagan, N., Ho, M., Hoyt, A.M., Harvey, C.F., Vining, S.R., Hough, M.A.,
- 352 Moore, T.R., Richard, P.J.H., De La Cruz, F.B., Toufaily, J., Hamdan, R., Cooper, W.T., Chanton, J.P.,
- 2018. Tropical peatland carbon storage linked to global latitudinal trends in peat recalcitrance. Nature
 Communications. doi:10.1038/s41467-018-06050-2
- Hogg, E.H., Lieffers, V.J., Wein, R.W., 1992. Potential Carbon Losses from Peat Profiles Effects of
 Temperature, Drought Cycles, and Fire. Ecological Applications 2, 298–306. doi:Doi 10.2307/1941863
- Holmes, M.E., Chanton, J.P., Tfaily, M.M., Ogram, A., 2015. CO2 and CH4 isotope compositions and production
 pathways in a tropical peatland. Global Biogeochemical Cycles 29, 1–18. doi:10.1002/2014GB004951
- 359 Hoyos-Santillan, J., Craigon, J., Lomax, B.H., Lopez, O.R., Turner, B.L., Sjögersten, S., 2016. Root oxygen loss
- from Raphia taedigera palms mediates greenhouse gas emissions in lowland neotropical peatlands. Plant
 and Soil 404, 47–60. doi:10.1007/s11104-016-2824-2
- Hoyos-Santillan, J., Lomax, B.H., Large, D., Turner, B.L., Boom, A., Lopez, O.R., Sjögersten, S., 2015. Getting
 to the root of the problem: litter decomposition and peat formation in lowland Neotropical peatlands.
 Biogeochemistry 126, 115–129. doi:10.1007/s10533-015-0147-7
- 365 Hoyos Santillan, J., Lomax, B.H., Turner, B.L., Sjögersten, S., 2018. Nutrient limitation or home field advantage:
- 366 Does microbial community adaptation overcome nutrient limitation of litter decomposition in a tropical
 367 peatland? Journal of Ecology 00, 1–12.

- 368 IPCC, 2013. Climate Change 2013: The Physical Science Basis, Climate Change 2013: The Physical Science
 369 Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on
- 370 Climate Change. doi:10.1017/CBO9781107415324
- 371 Jauhiainen, J., Kerojoki, O., Silvennoinen, H., Limin, S., Vasander, H., 2014. Heterotrophic respiration in drained
- tropical peat is greatly affected by temperature-a passive ecosystem cooling experiment. Environmental
 Research Letters 9. doi:Artn 105013Doi 10.1088/1748-9326/9/10/105013
- Jauhiainen, J., Takahashi, H., Heikkinen, J.E.P., Martikainen, P.J., Vasander, H., 2005. Carbon fluxes from a
 tropical peat swamp forest floor. Global Change Biology. doi:10.1111/j.1365-2486.2005.001031.x
- Keiluweit, M., Nico, P.S., Kleber, M., Fendorf, S., 2016. Are oxygen limitations under recognized regulators of
 organic carbon turnover in upland soils? Biogeochemistry. doi:10.1007/s10533-015-0180-6
- Kolton, M., Marks, A., Wilson, R.M., Chanton, J.P., Kostka, J.E., 2019. Impact of warming on greenhouse gas
 production and microbial diversity in anoxic peat from a Sphagnum-dominated bog (Grand Rapids,
 Minnesota, United States). Frontiers in Microbiology. doi:10.3389/fmicb.2019.00870
- Könönen, M., Jauhiainen, J., Straková, P., Heinonsalo, J., Laiho, R., Kusin, K., Limin, S., Vasander, H., 2018.
 Deforested and drained tropical peatland sites show poorer peat substrate quality and lower microbial
 biomass and activity than unmanaged swamp forest. Soil Biology and Biochemistry.
 doi:10.1016/j.soilbio.2018.04.028
- Le Mer, J., Roger, P., 2001. Production, oxidation, emission and consumption of methane by soils: A review.
 European Journal of Soil Biology 37, 25–50.
- Leroy, F., Gogo, S., Guimbaud, C., Bernard-Jannin, L., Hu, Z., Laggoun-Défarge, F., 2017. Vegetation
 composition controls temperature sensitivity of CO2 and CH4 emissions and DOC concentration in
 peatlands. Soil Biology and Biochemistry. doi:10.1016/j.soilbio.2017.01.005
- Liu, Q., Wang, R., Li, R., Hu, Y., Guo, S., 2016. Temperature sensitivity of soil respiration to nitrogen
 fertilization: Varying effects between growing and non-growing seasons. PLoS ONE.
 doi:10.1371/journal.pone.0168599
- Lloyd, J., Taylor, J., 1994. On the temperature dependence of soil respiration. Functional Ecology.
 doi:10.2307/2389824
- Lupascu, M., Wadham, J.L., Hornibrook, E.R.C., Pancost, R.D., 2012. Temperature Sensitivity of Methane
 Production in the Permafrost Active Layer at Stordalen, Sweden: a Comparison with Non-permafrost
 Northern Wetlands. Arctic Antarctic and Alpine Research 44, 469–482. doi:10.1657/1938-4246-44.4.469

- Melling, L., Tan, C.S.Y., Goh, K.J., Hatano, R., 2013. Soil Microbial and Root Respirations from Three
 Ecosystems in Tropical Peatland of Sarawak, Malaysia. Journal of Oil Palm Research 25, 44–57.
- Menichetti, L., Reyes Ortigoza, A.L., García, N., Giagnoni, L., Nannipieri, P., Renella, G., 2015. Thermal
 sensitivity of enzyme activity in tropical soils assessed by the Q10and equilibrium model. Biology and
 Fertility of Soils. doi:10.1007/s00374-014-0976-x
- 403 Nianpeng, H., Ruomeng, W., Yang, G., Jingzhong, D., Xuefa, W., Guirui, Y., 2013. Changes in the temperature
- sensitivity of SOM decomposition with grassland succession: Implications for soil C sequestration. Ecology
 and Evolution. doi:10.1002/ece3.881
- 406 Page, S.E., Rieley, J.O., Banks, C.J., 2011. Global and regional importance of the tropical peatland carbon pool.
 407 Global Change Biology. doi:10.1111/j.1365-2486.2010.02279.x
- Phillips, S., Rouse, G.E., Bustin, R.M., 1997. Vegetation zones and diagnostic pollen profiles of a coastal peat
 swamp, Bocas del Toro, Panama. Palaeogeography, Palaeoclimatology, Palaeoecology 128, 301–338.
- 410 Sánchez-Carrillo, S., Álvarez-Cobelas, M., Angeler, D.G., Serrano-Grijalva, L., Sánchez-Andrés, R., Cirujano,
- S., Schmid, T., 2018. Elevated Atmospheric CO2 Increases Root Exudation of Carbon in Wetlands: Results
 from the First Free-Air CO2 Enrichment Facility (FACE) in a Marshland. Ecosystems. doi:10.1007/s10021017-0189-x
- Sjögersten, S., Aplin, P., Gauci, V., Peacock, M., Siegenthaler, A., Turner, B.L., 2018. Temperature response of
 ex-situ greenhouse gas emissions from tropical peatlands: Interactions between forest type and peat moisture
 conditions. Geoderma. doi:10.1016/j.geoderma.2018.02.029
- Sjögersten, S., Cheesman, A.W., Lopez, O., Turner L., B., 2011. Biogeochemical processes along a nutrient
 gradient in a tropical ombrotrophic peatland. Biogeochemistry 104, 147–163. doi:DOI 10.1007/s10533010-9493-7
- Smith, W.H., 1976. Character and Significance of Forest Tree Root Exudates. Ecology 57, 324–331. doi:Doi
 10.2307/1934820
- Song, M., Jiang, J., Cao, G., Xu, X., 2010. Effects of temperature, glucose and inorganic nitrogen inputs on carbon
 mineralization in a Tibetan alpine meadow soil. European Journal of Soil Biology.
 doi:10.1016/j.ejsobi.2010.09.003
- 425 Tonks, A.J., Aplin, P., Beriro, D.J., Cooper, H., Evers, S., Vane, C.H., Sjogersten, S., 2017. Impacts of conversion
- 426 of tropical peat swamp forest to oil palm plantation on peat organic chemistry, physical properties and
- 427 carbon stocks. Geoderma 289, 36–45. doi:10.1016/j.geoderma.2016.11.018

- Troxler, T.G., Ikenaga, M., Scinto, L., Boyer, J.N., Condit, R., Perez, R., Gann, G.D., Childers, D.L., 2012.
 Patterns of Soil Bacteria and Canopy Community Structure Related to Tropical Peatland Development.
 Wetlands 32, 769–782. doi:10.1007/s13157-012-0310-z
- 431 Upton, A., Vane, C.H., Girkin, N., Turner, B.L., Sjögersten, S., 2018. Does litter input determine carbon storage
 432 and peat organic chemistry in tropical peatlands? Geoderma 326, 76–87.
- 433 doi:10.1016/j.geoderma.2018.03.030
- 434 Uselman, S.M., Qualls, R.G., Thomas, R.B., 2000. Effects of increased atmospheric CO₂, temperature, and soil
- N availability on root exudation of dissolved organic carbon by a N-fixing tree (*Robinia pseudoacacia L.*).
 Plant and Soil. doi:Doi 10.1023/A:1004705416108
- Vančura, V., 1967. Root exudates of plants III. Effect of temperature and "cold shock" on the exudation of
 various compounds from seeds and seedlings of maize and cucumber. Plant and Soil.
 doi:10.1007/BF01376325
- Wang, X., Li, X., Hu, Y., Lv, J., Sun, J., Li, Z., Wu, Z., 2010. Effect of temperature and moisture on soil organic
 carbon mineralization of predominantly permafrost peatland in the Great Hing'an Mountains, Northeastern
 China. Journal of Environmental Sciences. doi:10.1016/S1001-0742(09)60217-5
- Wright, E.L., Black, C.R., Turner, B.L., Sjögersten, S., 2013. Environmental controls of temporal and spatial
 variability in CO2 and CH4 fluxes in a neotropical peatland. Global Change Biology.
 doi:10.1111/gcb.12330





450 Figure 1: Study site locations in (a) Panama: Changuinola, Bocas del Toro province. (b) Malaysia: Kampung Raja
451 Musa, North Selangor and Kampung Mat Jintan, Terenganu,

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453 Table 1: Peat properties for Panamanian and Malaysian forested peatlands, and converted Malaysian oil palm with

454 pineapple intercropping. Means \pm one SE (n = 3). (^{a.} Phillips et al., 1997; ^{b.} Isla Colon, STRI Environmental

455 Monitoring; ^{c.} (Wright et al., 2013); ^{d.} (Dhandapani et al., 2019b); ^{e.} (Dhandapani et al., 2019a); ^{f.} (Global

	Panamanian	Malaysian	Malaysian	p-value
	forest	forest	intercropping	
Peat depth (m)	5 ^a	2 ^d	3 ^e	-
pH	4.0 ± 0.01	4.4 ± 0.4	4.2 ± 0.1	0.457
Redox potential (mV)	303.7 ± 6.9	185.3 ± 21.2	294.3 ± 3.5	0.001
Moisture content (%)	77.00 ± 9.5	90.4 ± 0.2	77.2 ± 1.2	0.225
Organic matter content (%)	93.8 ± 1.4	78.9 ± 11.8	86.3 ± 2.5	0.378
<i>C</i> (%)	43.8 ± 3.9	51.3 ± 0.6	59.4 ± 2.8	0.021
N (%)	2.6 ± 0.4	1.9 ± 0.1	2.3 ± 0.05	0.134
C:N	17 ± 1.0	26.9 ± 1.3	25 ± 1.7	0.003
Peat temperature (°C)	23.9 ± 0.1	27.3 ± 0.1	27.7 ± 0.5	-
Air temperature (°C)	26.3	26.8	26.6	-
Mean annual rainfall (mm)	3206 ^b	1000 ^d	1359- 2480 ^f	-
Water table range (cm)	$-20 - 20^{\circ}$	$5 - 10^{d}$	-15050^{f}	-

456 Environment Centre, 2014).





460 Figure 2: *Ex situ* basal CO₂ and CH₄ fluxes at 25°C and in the absence of glucose. Means \pm one SE (n = 3).





Figure 3: Mean CO_2 and CH_4 fluxes for (a-b) Panamanian forest, (c-d) Malaysian forest, (e-f) Malaysian pineapple intercropping. Means \pm one SE (n = 3).

464 Table 2: Q₁₀ for Panamanian and Malaysian forests, and Malaysian pineapple intercropping sites in the presence

		CO ₂		CH ₄	
Site	Glucose	Q10	R ²	Q10	\mathbb{R}^2
Panamanian forest	+	1.3 ± 0.4	0.67	1.1 ± 0.04	0.48
	-	1.5 ± 0.2	0.78	1.5 ± 0.3	0.74
Malaysian forest	+	1.4 ± 0.1	0.31	1.2 ± 0.1	0.50
	-	1.3 ± 0.01	0.80	1.5 ± 0.3	0.57
Malaysian intercropping	+	1.2 ± 0.04	0.83	1.3 ± 0.2	0.04
	-	1.3 ± 0.3	0.81	1.3 ± 0.2	0.68

465 and absence of glucose. Mean \pm one SE (n = 3).