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Applying Adverse Outcome Pathways (AOPs) to support Integrated Approaches to Testing and Assessment (IATA)

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Abstract

Chemical regulation is challenged by the large number of chemicals requiring assessment for potential human health and environmental impacts. Current approaches are too resource intensive in terms of time, money and animal use to evaluate all chemicals under development or already on the market. The need for timely and robust decision making demands that regulatory toxicity testing becomes more cost-effective and efficient. One way to realize this goal is by being more strategic in directing testing resources; focusing on chemicals of highest concern, limiting testing to the most probable hazards, or targeting the most vulnerable species. Hypothesis driven Integrated Approaches to Testing and Assessment (IATA) have been proposed as practical solutions to such strategic testing. In parallel, the development of the Adverse Outcome Pathway (AOP) framework, which provides information on the causal links between a molecular initiating event (MIE), intermediate key events (KEs) and an adverse outcome (AO) of regulatory concern, offers the biological context to facilitate development of IATA for regulatory decision making. This manuscript summarizes discussions at the Workshop entitled “Advancing AOPs for Integrated Toxicology and Regulatory Applications” with particular focus on the role AOPs play in informing the development of IATA for different regulatory purposes.

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Adverse Outcome Pathway (AOP); Integrated Approaches to Testing and Assessment (IATA); (Q)SAR; Read-across; High Throughput screening (HTS); High Content Screening (HCS); Cross Species extrapolation

Highlights

- AOPs provide a mechanistic basis for IATA development
- The elements of an AOP-informed IATA and how to integrate data are described
- A conceptual framework based on the AOP concept is proposed
- Examples are presented to illustrate the framework for different regulatory uses

1. Introduction

AOPs to support IATA in regulatory decision-making

Within the last decade, the global economy has witnessed a significant shift in the regulatory environment coupled with the volume and diversity of industrial chemicals being manufactured. Despite some regional differences, regulatory management in general comprises hazard identification/characterization, an exposure assessment and a risk assessment as its main steps. In some cases, the identification of hazards is prior to market approval and certain hazards e.g. carcinogenicity (C), mutagenicity (M), or reproductive (R) effects (CMRs) may lead to restrictions on use irrespective of any subsequent risk assessment.

The hazard identification step is driven by a desire to identify all the hazards of potential concern and assign the appropriate hazard classification (i.e. classification & labelling requirements) regardless of the relevance of these hazards as a consequence of exposure. The hazard characterization step is often associated with extensive *in vivo* toxicity testing using standardized guidelines or protocols. The time, cost and animal use to generate such hazard data are significant and difficult to achieve in practice given the large number of chemicals that need to be evaluated. Furthermore, the classical *in vivo* testing approach is based on apical endpoints, which typically provide minimal information on the mode or mechanism of action. This limits the development and application of new *in vitro* assays, read-across approaches or inter-species extrapolation, all of which could facilitate an initial hazard assessment. In addition, the societal demand to use (vertebrate) animal tests only as a last resort for obtaining hazard information coupled with the increasing number of different regulatory programs worldwide calls for a re-consideration of traditional assessment strategies and incorporation of

26 alternative approaches. At the same time, substantial advances have been made in the
27 use of high throughput (HT) and high content (HC) screening assays to quantify and
28 characterize molecular and cellular responses to chemicals (Kavlock et al., 2007; Judson
29 et al., 2014; Kleinstreuer et al., 2014). A shift towards more mechanistically-based
30 alternative approaches represents a promising opportunity for assessing hazards of
31 regulatory concern. To that end the Adverse Outcome Pathway (AOP) framework
32 provides the biological context and supporting weight of evidence (WoE) to facilitate the
33 interpretation of such alternative data. An AOP represents the existing knowledge
34 concerning the causal linkages between the molecular initiating event (MIE) and the
35 cascade of intermediate or key events (KEs) at the subcellular, cellular, tissue and organ
36 level that lead to a specific adverse outcome (AO) at the individual or population level
37 (Ankley et al., 2010; OECD, 2013). This conceptual framework enables information and
38 data from different chemicals, different levels of biological organization, and different
39 taxonomic domains relevant for one AOP to be assembled. Well-developed AOPs may
40 therefore be expected to help guide identification of experimental testing (e.g. *in vivo*, *in*
41 *vitro*, *in chemico*) and non-testing (*in silico*) approaches to support regulatory decision
42 making. There is now a need for an objective framework to interpret the results from
43 novel test methods and their prediction models in order to facilitate their application in
44 regulatory decision making. Such a framework will conceivably consist of three main
45 elements: the AOP itself, non-animal (alternative) test methods and *in silico* approaches
46 targeting key components of the AOP, and their associated prediction models for a
47 particular regulatory context. The synthesis and integration of these elements form the
48 basis for developing Integrated Approaches to Testing and Assessment (IATA) that may
49 be used in regulatory applications.

50 This manuscript summarises discussions from the Workshop entitled “Advancing AOPs
51 for Integrated Toxicology and Regulatory Applications” held in Somma Lombardo, Italy
52 on the 2-7th March, 2014 (<https://aopkb.org/saop/workshops/somma.html>).
53 Specifically it captures the discussions and insights derived within the workgroup that
54 discussed the role that AOPs can play in informing the development of IATA for
55 regulatory purposes. The next section defines IATA and related terms. Following that,
56 the main elements or components that make up IATA are described including
57 considerations, (*e.g.* scientific confidence), that are associated with these different
58 elements and their integration. Once the components have been defined, the overall
59 applicability and limitations of IATA for different regulatory purposes are considered.
60 These concepts are then illustrated by way of three examples that are supported by
61 specific AOPs at different levels of development. A final summary considers how the
62 proposed conceptual framework may impact different regulatory applications.

63

64 ***2. IATA and related concepts***

65 Integrated Approaches to Testing and Assessment (IATA) are structured approaches
66 that integrate and weigh different types of data for the purposes of performing hazard
67 identification (i.e. the potential to cause a hazard), hazard characterization (e.g. the toxic
68 potency) and/or safety assessment (i.e. the potential/toxicity potency related to
69 exposure) of a chemical or group of chemicals. For the purposes of this paper, IATA will
70 be generally referred to in a singular form to represent a specific case rather than a
71 collective approach. An IATA should be viewed as an iterative process that includes
72 efficiency analyses to determine whether more data, and what type of data, are required
73 to make effective regulatory decisions while reducing reliance on animal testing. An

74 IATA is not a novel concept per se, indeed it has been discussed at a special OECD
75 workshop on IATA in 2007 (OECD, 2008) and described by the US EPA as part of a FIFRA
76 Scientific Advisory Panel document in 2011 (US EPA, 2011).

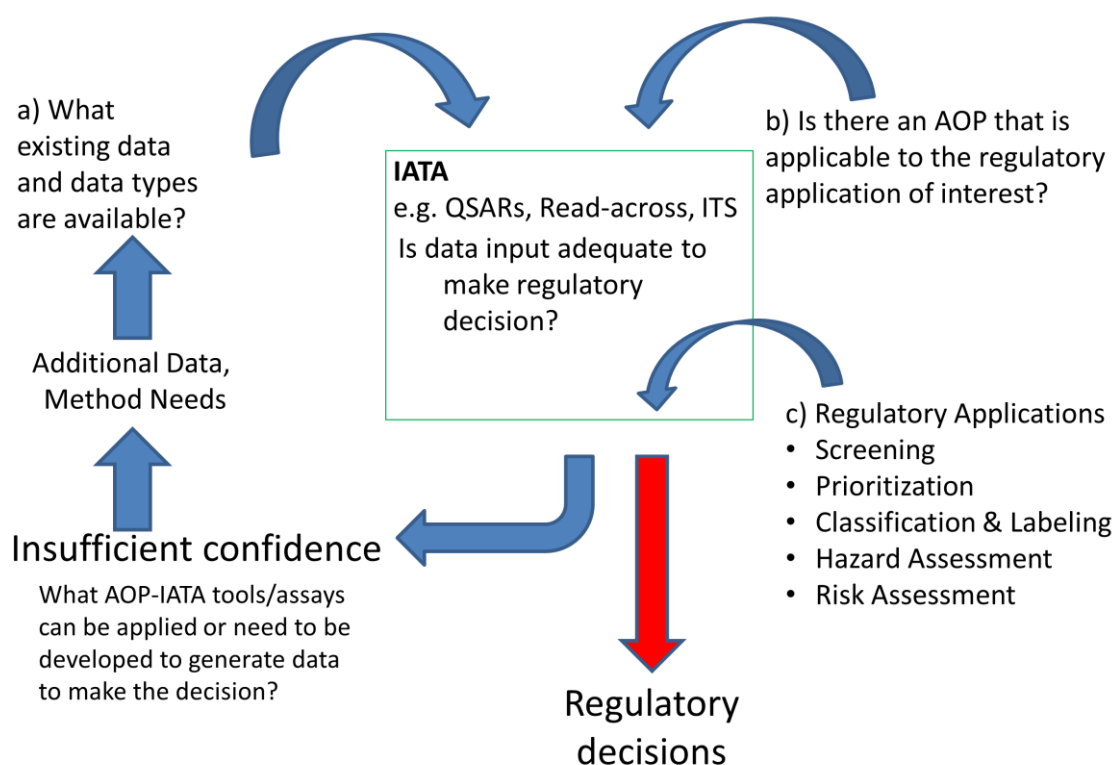
77 An IATA initially gathers and weighs relevant existing information to derive an initial
78 conclusion. If the existing information is insufficient to address the regulatory or safety
79 decision under consideration, it guides the generation of new data using a hypothesis-
80 driven approach with the goal of addressing the residual uncertainty preventing a
81 regulatory decision. The benefit of an IATA lies in the potential breadth of information
82 that can be used in the assessment, as it may exploit both non-testing (*in silico*) and
83 experimental (*in vivo*, *in vitro* and *in chemico*) approaches. The IATA is considered a
84 generic approach and may encompass testing strategies such as integrated testing
85 strategies (ITS), sequential testing strategies (STS), as well as weight of evidence (WoE)
86 considerations (OECD, 2014). Both ITS, i.e. the fixed and structured integration and
87 weighing of relevant information to support the final decision (Ahlers et al., 2008;
88 Hartung et al., 2013), and STS, i.e. the fixed stepwise approach involving interim decision
89 steps to reach a decision, represent structured and formal processes to derive a
90 conclusion (OECD, 2014). In contrast, WoE considerations, i.e. the structured,
91 systematic, independent and transparent review of existing and available data without
92 use of experimental or computational efforts, aim to perform a reliable and relevant
93 compilation of knowledge intended for a certain regulatory purpose (Balls et al., 2006;
94 OECD, 2014). Whilst an IATA provides a structure for data integration and a means for
95 targeting testing for particular uses, it is not necessarily framed by any mechanistic
96 rationale. There is a growing support for using AOPs to provide such a mechanistic basis
97 (OECD, 2013). Thus, AOP-informed IATA development may drive the development of *in*
98 *silico*, *in vitro*, or *in chemico* approaches that are anchored in well-developed knowledge

99 captured within an AOP. Exposure considerations and the use of exposure assessment
100 tools may also form an integral part of an IATA.

101 AOPs are expected to provide insight into the biological relevance, reliability, and
102 uncertainties associated with the results from *in silico*, *in chemico* and *in vitro*
103 approaches for regulatory use. AOPs also have substantial merit in traditional
104 assessment strategies. For instance, they can assist manufacturers and regulators to
105 identify whether a potential hazard can be expected that justifies subsequent detailed
106 testing. Furthermore, in the environmental hazard and risk assessment, they show great
107 promise in the species-to-species extrapolation critical for protection of endangered
108 species (Perkins et al., 2013). AOPs could also help to design ITS, which ideally cover the
109 relevant key events of an AOP. AOPs are intended to provide a transparent evaluation of
110 available evidence and relevant data, scientific confidence is envisioned to be evaluated
111 through approaches akin to the “Bradford Hill Considerations” developed originally in
112 epidemiology (Hill, 1965; OECD, 2013). Briefly, by examining: (1) biological
113 concordance, (2) essentiality of Key Events, (3) concordance of empirical observation
114 (encompasses dose response and temporal concordance and beyond), (4) consistency
115 (among different biological contexts) and (5) analogy (consistency across chemicals), a
116 clear statement regarding the supporting evidence for the AOP can be developed (Meek
117 et al., 2014a, b). Depending on the outcomes for these considerations, a given AOP may
118 differ in its level of scientific rigor and confidence, which in turn will drive its practical
119 suitability in addressing different regulatory applications (Perkins et al., *submitted*;
120 Patlewicz et al., *submitted*).

121 The practical implementation of an AOP-informed IATA for a given chemical or group of
122 chemicals considers problem formulation based on the risk management scope and

123 goals, the selection and evaluation of suitable AOPs to inform the IATA and existing
 124 information that is available for the chemical(s) of interest. All these considerations will
 125 influence the makeup of an IATA in terms of the different types of testing (e.g. *in*
 126 *chemico*, *in vitro* and *in vivo*), non-testing (e.g. *in silico*), or data integration approaches
 127 (e.g. ITS, STS, WoE or other IATA strategies) that can be exploited (Figure 1). Figure 1
 128 outlines a proposed framework to guide how existing information (e.g. hazard and
 129 exposure information) needs to be evaluated and what new data, if any, needs to be
 130 generated, so that the IATA can lead to a regulatory decision.



131

132

133 *Figure 1. Conceptual framework for an AOP-informed IATA to support regulatory*
 134 *decisions. The framework is driven by the problem formulation, which involves a*
 135 *consideration of the risk management scope, the data requirements and the level of*
 136 *acceptable uncertainty associated with the decision being made. The regulatory*

137 *application will also provide an indication of the level of AOP confidence, ideally needed.*
138 *The framework, which comprises different elements (testing and non-testing approaches,*
139 *etc.), will evaluate the existing information that is available for the chemical(s) of interest*
140 *(a), the type of information that might be required as defined by the AOP itself (b), and*
141 *other relevant information that is pertinent in making a regulatory decision (c). If the*
142 *outcome generated based on the framework is of sufficient confidence for the regulatory*
143 *purpose of interest, no further action is warranted. If the outcome derived from the*
144 *framework is of insufficient confidence, then additional data might need to be generated*
145 *through new testing and assessment. The new information derived will then be passed back*
146 *into the framework for re-evaluation. Indeed a decision outcome could result in more*
147 *thorough regulatory follow up or implementation of measures to reduce use and/or*
148 *exposure. Any new information generated will also be used to augment the corresponding*
149 *AOP.*

150

151 **3. Elements for developing AOP-informed IATA**

152 Non-testing and testing approaches as well as data integration strategies form the
153 elements or building blocks that are necessary to derive IATA. These elements are
154 described in more detail in terms of the applicability and limitations in the following
155 sections.

156 *3.1 Non-testing approaches*

157 Non-testing or *in silico* approaches serve two functions within an IATA, they either
158 provide a way to organize existing information or they are used to make predictions of
159 molecular initiating events (MIE) or other key events (KEs) as defined in an associated
160 AOP. The breadth of non-testing approaches is extensive. They range from the search
161 and retrieval of existing data, to the identification of structural fragments to indicate
162 activity and assist grouping (e.g. structure-activity relationships (SARs), read-across), to
163 quantitative models (e.g. quantitative structure-activity relationships (QSARs)). Some of

164 these SARs or QSARs may be housed in software tools known as expert systems for ease
 165 of use. A summary of non-testing approaches that may be useful in the development of
 166 IATA are described in more detail in Cronin and Madden, (2010).

167 Within IATA, non-testing approaches will most likely be exploited to characterize the
 168 MIE within AOPs either qualitatively or quantitatively (Table 1). A number of different
 169 types of MIEs, and thus AOPs, may be identified for a given IATA in order to generate
 170 sufficient information for the decision to be made.

171
 172 Table 1. Examples of MIEs within AOPs that may be relevant to IATA endpoints derived
 173 from non-testing approaches.

MIE	Effect	Examples of In Silico Tool(s)
Unspecific – no definable single molecular site of action	General accumulation in cellular membranes leading to e.g. narcosis, basal cytotoxicity etc.	Classification schemes e.g. Verhaar implemented in the OECD QSAR Toolbox or Toxtree. QSARs based on hydrophobicity.
Non-specific covalent binding (and formation of radicals)	Irreversible binding to cellular protein and /or DNA which may lead to a variety of effects; e.g. fibrosis	OECD QSAR Toolbox profilers for protein and DNA binding. Quantum chemical calculations.
Redox cycling leading to disruption of specific pathways	Mitochondrial toxicity	Structural alerts e.g. Nelms et al., 2014
Receptor mediated effects on signalling pathways	A wide variety of acute and non-lethal effects e.g. estrogen receptor binding	3-D molecular modelling. Toxicophores / alerts e.g. EPA ER Binding Expert System as encoded in the OECD QSAR Toolbox, DART system (Wu et al., 2014)

Physical effects	Skin corrosion	Structural alerts, physicochemical properties e.g. pH
Unknown or very poorly defined MIE	Idiosyncratic drug toxicity	Structural alerts

174

175 3.1.1 Confidence Factors for In Silico Models in IATA

176 Assuring scientific confidence in the validity of *in silico* models and their outcomes are
 177 key considerations for their application. For (Q)SARs, the OECD has developed
 178 validation principles which provide a framework for assuring the scientific validity
 179 (relevance and reliability) of a (Q)SAR model (OECD, 2004; 2007). The (Q)SAR model
 180 reliability is a relative concept, depending on the context in which the model is applied;
 181 meeting each and all of the OECD principles is not necessarily warranted. It is worth
 182 noting that the OECD principles only focus on the scientific validity of a given (Q)SAR
 183 model, and not on the prediction it generates. The adequacy of a (Q)SAR result for a
 184 given compound also needs to be considered before use. In the context of the European
 185 Regulation for registration, evaluation, authorisation and restriction of chemicals
 186 (REACH) (EC, 2006; ECHA, 2008) the following specific conditions are considered when
 187 evaluating *in silico* models:

- 188 1. the estimate should be generated by a valid (relevant and reliable) model;
- 189 2. the model should be applicable to the chemical of interest with the necessary
 190 level of reliability;
- 191 3. the model endpoint should be relevant for the regulatory purpose.

192

193 Whilst these were outlined specifically for REACH, the conditions could be conceivably
 194 adapted to address other regulatory purposes.

195 For chemical categorization (OECD, 2014) (e.g. read-across), no such principles have
196 been formalized. Systematic frameworks to aid in the evaluation of read-across and
197 identify associated uncertainties are in development by European Chemicals Agency
198 (ECHA) (known as the Read-Across Assessment Framework (RAAF)) as well as by
199 Industry (Blackburn and Stuard, 2014). To date these frameworks do not specifically
200 consider the role of AOPs or how alternative data characterizing MIEs or other KEs may
201 be conceivably used to address uncertainties. Work underway within the SEURAT
202 program (ChemWatch, 2014 – see: <http://chemicalwatch.com/19594/seurat-1-homes->
203 [in-on-test-chemicals-for-read-across](http://chemicalwatch.com/19594/seurat-1-homes-in-on-test-chemicals-for-read-across)) and independently by DECO-2, a Cefic-LRI AIMT-4
204 project (Patlewicz et al., *in preparation*) are both aiming to investigate the feasibility of
205 enhancing read-across by using the AOP concept.

206 It is noteworthy to mention that there will be clear instances when *in silico* approaches
207 will not provide meaningful information in the context of an IATA, for example if there is
208 no direct linkage to the MIE because the MIE is unknown or ill-defined. Predictions from
209 *in silico* approaches will also be inappropriate, when the target substance is outside of
210 the applicability domain of the model.

211

212 *3.2 Testing approaches*

213 There are many testing approaches that can form key elements within an IATA – from *in*
214 *chemico*, *in vitro* to *in vivo* experimental efforts. Testing elements such as
215 toxicogenomics, high content/high-throughput screening (HC/HT) in particular will play
216 a crucial role in shifting IATA away from a reliance on *in vivo* information addressing one
217 or multiple adverse outcomes.

218

219 3.2.1 *In chemico tests*

220 Biological effects of chemicals can be provoked by an initial covalent modification of a
221 biological macromolecule. The covalent modification of DNA leading to mutagenesis or
222 the reaction with immunoproteins resulting in immunosuppression represent
223 prominent examples (Cronin et al., 2009). *In chemico* tests are experimental
224 measurements that address these covalent modifications without involving biological
225 organisms (reviewed in Schwöbel et al., 2011). These assays are usually used to identify,
226 and in some cases estimate, the intrinsic reactivity of substances to a specific biological
227 target and in that respect are best suited to target the MIE within an AOP. Most *in*
228 *chemico* tests relevant to toxicity prediction have investigated the reaction of an
229 electrophilic molecule (normally assumed to be the toxicant) with a model nucleophile
230 (representing a surrogate for the target biological macromolecule) (e.g. Roberts et al.,
231 2008; Aptula and Roberts, 2006; Schultz et al; 2005; Thaens et al., 2012). Also included
232 in this type of data could be the assessment of oxidizing behavior and the role of other
233 reactive species (nucleophiles, reactive oxygen species, radicals) principally amenable to
234 *in chemico* testing (Cronin et al., 2009).

235

236 3.2.2 *In vitro and alternative test systems*

237 Cellular *in vitro* systems, lower vertebrate embryos and invertebrates are proposed and
238 used as alternative test system to indicate toxic potential to various organisms. Relevant
239 information on the toxic potential of a chemical can be obtained via e.g. comparison of
240 the toxicity to baseline toxicity as an indicator of a non-narcotic or specific mode of

241 action (Escher and Schwarzenbach, 2002). By including appropriate endpoints it is
242 possible to target MIEs or KEs relevant for an AOP-informed assessment. Extrapolations
243 from alternative test systems, however, have to consider that the toxicokinetic
244 properties may greatly differ and result in deviating effect concentrations between e.g. *in*
245 *vitro* and *in vivo* tests. Fish/amphibian embryos or invertebrates – despite their
246 evolutionary distance to e.g. mammals or other vertebrate classes – may provide in
247 some cases a higher predictive capacity than *in vitro* systems given that they
248 represent/accommodate the complexity of a whole organism (Perkins et al., 2013).

249

250 3.2.3 High throughput screening assays

251 High throughput screening assays (HT) comprise *in chemico* and certain *in vitro* test
252 methods such as receptor binding or receptor transactivation assays (Romanov et al.,
253 2008), cellular reporter assays (Romanov et al., 2008; Kleinstreuer et al., 2014), assays
254 using invertebrate (e.g. *C. elegans*, *Drosophila*, algae, crustaceans, see Perkins et al.,
255 2013) or fish embryos (Truong et al., 2014). Toxicogenomic (transcriptomics,
256 proteomics, and metabolomics), utilizing non-biased screening approaches may play a
257 more important role in the future within IATA, since they allow more detailed insights
258 into mechanisms of action and can be applied to survey the breadth of
259 molecular/cellular effects relevant for a wide variety of AOPs (Garcia-Reyero et al.,
260 2014a,b).

261 Assays targeted towards MIEs, can be very specific for a distinct target (e.g., receptor,
262 enzyme) that leads to an AO. This specificity will also provide the chemical structure and
263 bioactivity data needed to foster development of *in silico* models (as described in section

264 3.1). Assays that target downstream KEs such as more generic stress responses
 265 (Simmons et al., 2009) may not have this specificity, but may provide an approach to
 266 integrate multiple MIEs (Miller et al., 2009). More importantly, analysis of downstream
 267 KEs provide the opportunity to predict an AO even in cases where the precise MIE is not
 268 known or is not fully understood. Table 2 presents several examples of alternative
 269 experimental testing approaches that may be relevant to predict AOs within an IATA.

270

271 Table 2. Summary and examples of the different types of experimental testing

272 approaches in AOP-informed IATAs.

Approach	Usage	AOP target	Example(s) of HT/HC compatible assays	Adverse outcome
<i>In chemico</i>	Indicate reactivity or covalent interaction with a biomolecule	MIE	GSH (Schultz et al., 2005);dNTP adduct formation (Zhao et al. 2002)	Unspecific (excess toxicity), genotoxicity/mutagenicity, immunosuppression, skin sensitization
<i>In vitro</i> (cellular)	Confirm toxicity pathway Confirm the (absence of) need for higher-tier testing Can be HT/HC compatible	MIE, KE	Cell lines, transactivation and reporter cell assays, subcellular assays, e.g. HTS assays for endocrine disruption (Cox et al., 2014, Murk et al., 2013)	Through targeting specific toxicity pathways, a wide range of AOs can be targeted (Bhattacharya et al. 2011), e.g. for endocrine disruption sexual development, reproductive disorders. Many different endpoints are possible through targeting specific toxicity pathways.
Invertebrates	Replace (vertebrate)	MIE, KE	<i>C. elegans</i> (Leung et al.	e.g. acute toxicity, developmental toxicity, neurotoxicity,

	animal tests		2008),	genotoxicity,
Fish or amphibian embryos	Replace (adult vertebrate) animal tests	MIE, KE	<i>D. rerio</i> embryos (Truong et al., 2014)	Acute and chronic fish toxicity, hepatotoxicity neurotoxicity, teratogenicity, endocrine disruption (reviewed in Scholz et al., 2013a,b),

273 **GSH:** Reduced glutathion, dNTP: Deoxyribonucleotide triphosphate, HT/HC: High-throughput/High-
274 content.

275 3.2.4 Confidence factors and limitations for testing approaches in IATA

276 The use of alternative testing approaches provide higher confidence when they are
277 scientifically and technically valid for use. Validation of alternative assays in particular
278 HT/HC assays has been subject of several publications such as Judson *et al.* (2013),
279 Hartung *et al.* (2013) and Patlewicz *et al.* (2013). In the latter scientific confidence was
280 discussed in the context of the existing validation frameworks for (Q)SARs and
281 biomarkers (Institute of Medicine, 2010). In Cox *et al.* (2014), a scientific confidence
282 framework was proposed comprising three inter-related elements to facilitate the
283 systematic, transparent and objective evaluation and documentation of HT/HC assays
284 and their associated prediction models. The elements comprise analytical validation,
285 qualification and utilization. Analytical validation would entail an assessment of the
286 biological basis and analytical performance of the assays. This would involve a
287 consideration of what events within the AOP the assay(s) were mapped to – whether
288 they target the MIE or other downstream KEs. The applicability domain of the assay in
289 terms of the chemical coverage and the typical performance statistics – sensitivity,
290 specificity, accuracy, would be considered as well. The qualification step would involve
291 an assessment of the associated prediction models derived from such assays and

292 utilization would consider the intended regulatory application based on the previous 2
293 steps.

294 Even when assays have been scientifically and technically validated, they may exhibit
295 certain limitations. Most assays do not consider the impact of potential metabolic
296 transformation, which can lead to reduced sensitivity (in case of *in vivo* metabolic
297 activation) or to a high number of false positives (in case of *in vivo* inactivation) or false
298 negatives (in case of *in vivo* bioactivation). Furthermore, certain compounds are difficult
299 or impossible to test using *in vitro* systems, for example due to their poor solubility in
300 the culture medium, aggregation potential, volatility, or partitioning behavior (tendency
301 to adsorb onto plastic). In such cases *in silico* methods could provide a more appropriate
302 approach (Zaldivar et al, 2010, 2011).

303

304 3.3 Data-integration strategies

305 Whilst there has been a tendency to define one “definitive” test for hazard assessment in
306 the past, increasingly the need for more than one piece of evidence for hazard
307 assessment has become evident. This need is fundamental in both the AOP concept and
308 the AOP-informed IATA. Therefore, data integration strategies are needed to integrate *in*
309 *silico*, *in chemico*, *in vitro*, *in vivo*, and available epidemiological or clinical data which

310 1. Allow for the combination of low-cost (sensitive) screening assays with more
311 sophisticated (specific) confirmatory assays.

312 2. Consider the incomplete coverage of one assay in the chemical universe
313 (applicability domain), severity classes or modes of action.

314 3. Compensate for the insufficient reliability of a single test.

315 4. Combine kinetic and exposure information, with (quantitative) *in vitro* to *in*
316 *vivo* extrapolation.

317 Testing and non-testing outcomes can be manually integrated together to derive an
318 outcome for specific regulatory purposes. This is relatively straightforward for a simple
319 linear AOP with a limited number of KEs, such as skin sensitization (OECD, 2012a,b). As
320 more AOPs are developed, and KEs are identified that cut across different AOPs into
321 networks of interlinked AOPs, the complexity of data integration supporting an IATA will
322 increase. Manual integration of a myriad of KEs may not be feasible to do. Moreover,
323 some of the assay outcomes or prediction models derived may require interpretation, a
324 translation step to convert the raw test outcome into a form that addresses the
325 information need for the regulatory purpose under consideration (see Weinberg, 1972
326 for detailed discussion). Note this interpretation step is not specific to IATA, but as the
327 complexity of IATA increases, more formalized systematic and transparent translation
328 approaches will be required. Integration of many information sources can be addressed
329 in different ways from:

- 330 1. Battery approaches, i.e. all results are collected and then interpreted
- 331 2. Sequential / tiered approach, i.e. in a given sequence results are
332 collected stopping when sufficient information is available through to
- 333 3. Result-driven further testing, e.g. determination of next most valuable
334 test or branching of test strategies depending on previous test results
335 (prioritization).

336 Integration of results derived from these information sources in turn occurs on different
337 levels, from the raw data level to the summary (categorical) level where certain
338 information is lost. Examples of data integration approaches include:

- 339 1. Boolean AND / OR / NOT combinations of categorized results (e.g. overall call
340 is denoted as positive if any of the test outcomes are positive)
- 341 2. Scoring approaches (e.g. various tests contributing to an overall score)
- 342 3. Decision trees (typically sequential with branching)
- 343 4. Deterministic, i.e. a point of departure for assessments is derived (e.g. lowest
344 active concentration) possibly combined with assessment factors to derive a
345 threshold value
- 346 5. Probabilistic, i.e. probabilities are assigned as a function of different
347 information leading typically to distributions of probabilities / uncertainties
- 348 6. Prediction based on machine learnings (e.g. PCA, random forest, multiple
349 regression) applied to a training set of compounds

350 IATA does extend beyond hazard information and will often also include kinetics and
351 exposure data, which in turn augments the complexity of the data integration
352 approaches applied. At this stage, no general guidance can be proposed, although it is
353 envisaged that a learning-by-doing is necessary and the advantages (and possible
354 disadvantages) of formally integrated data will emerge and can be resolved.

355

356 **4. Applicability of AOP-informed IATA for regulatory purposes**

357 Any non-standard approach needs to be fit for purpose whether it will be used for
358 prioritization, hazard identification, classification & labelling and/or risk assessment.
359 This is true for IATA as a whole, as well as the respective IATA elements themselves; the

360 latter of which have already been discussed in the previous sections. Specific criteria to
361 define fitness for specific regulatory applications have not been defined but guiding
362 principles are being proposed. Becker et al. (2014) outlined a scientific confidence
363 framework first proposed for HT/HC screening assays (Patlewicz et al., 2013) and their
364 prediction models (Cox et al., 2014) but adapted it to help in the evaluation of AOPs for
365 different purposes including IATA. Specific guidance for the assessment of IATA is not
366 currently available but recent initiatives taken up by the OECD Task Force for Hazard
367 Assessment (TFHA) are aiming to develop general principles for the evaluation and
368 documentation of IATA using skin sensitization as an initial case study (Worth and
369 Patlewicz, *submitted*). The initial principles proposed are framed by a clear identification
370 of the regulatory requirement as well as the applicability domain of the IATA itself:

- 371 a) define the endpoint of regulatory concern being assessed;
- 372 b) define the purpose/application for which the IATA is proposed;
- 373 c) describe the rationale, including mechanistic basis (e.g. AOP), according to which
374 the IATA is constructed;
- 375 d) describe the individual information sources constituting the IATA;
- 376 e) characterize the predictive performance and applicability domain of the IATA, or
377 IATA subcomponent(s) that can be expressed as a prediction model(s).

378

379 **5. Examples of AOP-informed IATAs in regulatory decision-making**

380 There are many potential regulatory applications for IATA. In this section, we highlight
381 three case study examples, which target different regulatory scenarios and hence are
382 characterized by differing levels of scientific confidence.

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5.1. Identification of chemicals disrupting estrogen, androgen, and thyroid hormone pathways

Endocrine disruption, particularly disruption of estrogen, androgen and thyroid pathways, is considered as an endpoint of high regulatory concern, given the potential adverse impact on human and environmental health, particularly sexual differentiation, reproduction and population development. AOPs linked to endocrine disruption of these three hormonal pathways represent examples where links between the MIE and KEs and the final AO have been reasonably established (Ankley et al., 2005; Miller et al., 2009; Volz et al., 2011). The OECD has already provided a conceptual framework describing the assays that would be available to target the different MIE and KE for endocrine disruption (OECD 2012c). Recent suggestions for developing scientific criteria for identification of an endocrine disrupting chemical (EDC) also conform to the principle of providing evidence of causality between mechanistic information (e.g. KEs) and AOs for endocrine disruption (Munn and Goumenou, 2013). Principally there is no single AOP for endocrine disruption. Depending on the targeted hormonal pathway or whether it is applied in the environmental or human health context, multiple AOPs could be defined. However, they share great commonalities at the different levels of biological complexity and are therefore described here.

Table 3. Examples of MIEs and KEs relevant for different levels of the AOPs for endocrine disruption. Given the large number of assays available for the different MIEs, KEs, and AOs, only selected examples are presented. For further assays descriptions refer to OECD (2012c).

AOP level (MIE	Description	Level	of	Test/Non	test	method
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and KE not in sequential order)	(examples)	biological organization	examples
MIE1	Hormone receptor binding and activation	Molecular level	Receptor-ligand binding assays (Tollefsen and Nilsen, 2008); Transactivation assays (Legler et al., 1999); QSARs for hormone receptor binding (Lo Piparo and Worth, 2010; Novic and Vracko, 2010)
MIE2	Interference with hormone synthesis	Molecular level	Steroidogenesis in vitro (OECD TG 456); In vitro assays for induction and inhibition of enzymes for TH metabolism (Murk et al., 2013); Zebrafish embryo assay for disruption of thyroid gland function (Raldua, 2009)
KE 1	Cell proliferation	Cellular	MCF7 cell proliferation assay (Körner et al., 1998)
KE 2	Increased vitellogenin	Cellular	<i>In vitro</i> fish hepatocyte vitellogenin production

	production		(Tollefsen et al., 2008)
KE3	Proliferation of uterus Metamorphosis	Organ	Uterotrophic assay (OECD TG 440) Amphibian metamorphosis assay (OECD TG 231)
KE4	Vitellogenin induction, 2 nd sex characteristics, fecundity, gonad development	Organ Organism	Fish Reproductive Screening Assay (OECD TG 229)
AO	Reproduction	Population	Reproductive toxicity studies Fish full life cycle assays (TG 415, 443)

406

407 In the US, the Endocrine Disruption Screening Program (EDSP) was established
408 in an effort to identify substances with the potential to interact with components of the
409 endocrine system. The Program comprises two Tiers; Tier 1 consists of a battery of *in*
410 *vitro* and *in vivo* assays that are intended to determine the potential of a chemical to
411 interact with the estrogen (E), androgen (A), or thyroid (T) hormone pathways whereas
412 Tier 2 comprises multigenerational reproductive and developmental toxicity tests in
413 several species to determine whether a substance can cause adverse effects resulting
414 from effects on the E, A, or T pathways. In Tier 2, the tests to be run are selected by a

415 WoE evaluation of Tier 1 results. The Tier 1 battery itself is expensive, time consuming
416 and does not lend itself to the assessment of large numbers of chemicals (each Tier 1
417 costs of the order of 1 million US dollars). Furthermore, it still relies to a large extent on
418 *in vivo* assays. Hence, more cost-efficient processes relying on *in silico* (QSAR and Expert
419 Systems) and HT screening data for prioritizing large numbers of chemicals for hazard
420 assessment purposes are being developed (Figure S1, supplementary information
421 outlines the use of the framework for prioritizing substances for their potential E, A and
422 T effects). When coupled with exposure predictions (from ADME and exposure models),
423 such a combination of non-testing and resource-efficient testing approaches could
424 provide sufficient confidence in prioritization decisions for subsequent testing
425 requirements. The EDSP represents an opportunity where relevant HTS assays can be
426 mapped to associated AOPs that are already well understood (e.g. Schmieder et al., 2003;
427 Crofton and Zoeller et al., 2005) and where confidence in the HTS predictive power for
428 higher levels of the AOP are well established.

429 While the goal of the US EDSP and application of corresponding HTS assays is clearly one
430 of prioritization and directing of testing, there is also scope to apply a tiered approach
431 for defined testing schemes such as required by European Union regulation. Substances
432 with endocrine disrupting capacity are conditionally exempted from exposure criteria,
433 i.e. higher tier assays for these compounds are required also at lower production
434 volumes. Similarly, Tier 1 *in vivo* assays to analyze the endocrine disruption potential are
435 required in environmental hazard assessment for the regulation of pesticides, biocides
436 and pharmaceuticals (Scholz et al., 2013a, b). It is however, not yet clear how the
437 endocrine disrupting potency will be identified but HT assays may provide a cost-
438 effective and reliable approach.

439 HT screening assays for determination of interference with hormone production,
440 hormone receptor binding and activation are currently available for a number of
441 hormone pathways. Of these, assays to target interference with the estrogen pathway
442 seems to be best developed with HTS methods for steroidogenesis and aromatase
443 inhibition (Villeneuve et al., 2007; Vinggaard et al., 2000), estrogen receptor (ER)
444 binding and activation (Legler et al., 1999; Tollefsen and Nilsen; 2008; Tollefsen et al.,
445 2008) and *in silico* (QSARs and docking models) for interaction with the ER (Schmieder
446 et al., 2003; Mombelli, 2012). A similar suite of bioassays exists for androgen signaling
447 pathways, although the role of androgen agonists or antagonists in endocrine disruption
448 is not as well developed. Nevertheless, assays such as a transcriptional activation assay
449 for the detection of the androgenic and anti-androgenic activity of chemicals have been
450 developed to support the assessment of disruption of the androgen axis (Rostkowski et
451 al., 2011). HT assays for detecting thyroid receptor agonists and antagonists also exist
452 (Murk et al., 2013), however, the majority of thyroid disruptors act via a variety of MIEs
453 that alter cellular TH signaling pathways via modulation of the TH levels. Thus, for
454 thyroid disrupting compounds the most relevant KE with respect to AO is the reduction
455 of thyroid hormone synthesis and homeostasis (Capen, 1997; Crofton, 2008).
456 Appropriate thyroid hormone-relevant assays are missing for many of the targets, and
457 development of appropriate assays that cover relevant MIEs and KEs are strongly
458 needed (Murk et al., 2013). As an interim approach, TR transcription assays such as
459 ToxCast and Tox21-TR assays can be applied. Assays of fish embryos targeting reduced
460 T4-levels (Thienpoint et al., 2011; Opitz et al., 2012) can be employed to identify
461 goitrogens. Despite the remaining high uncertainty for thyroid hormone disruption, a
462 significant reduction of higher tier testing could be achieved by including exposure
463 modelling into the screening approach. Wambaugh et al (2013) have developed a high-

464 throughput exposure model that uses data on production and use of chemicals, in
465 combination with a Bayesian statistical approach to describe the degree of uncertainty,
466 to provide exposure estimates for thousands of chemicals. Combining this with hazard
467 data allows for a rapid estimate of margins of exposure and prioritization of further
468 testing using both exposure and hazard data. Whilst the IATA framework has been
469 illustrated for prioritization per se, it could be refined for other purposes such as
470 classification and labelling, or hazard assessment both of which would be pertinent for
471 registration of chemicals in Europe.

472

473 **5.2 Skin sensitization**

474 Skin sensitization has been well studied over many decades. The chemical and biological
475 pathway driving the induction and elicitation of allergic contact dermatitis is relatively
476 well understood (see Lepoittevin J-P et al, 1997; Smith Pease CK, 2003, Adler et al, 2011)
477 and this knowledge has helped shape the development of alternative non-animal test
478 methods. Most recently the knowledge has been structured and documented in an AOP
479 construct and published by the OECD (OECD, 2012a, b). The OECD documentation for
480 this AOP summarizes the scientific evidence and assesses the overall WoE supporting
481 the AOP. There is strong evidence for the qualitative sequence of events from the MIE to
482 AO. Indeed empirical evidence from various elements of the AOP has value in assessing
483 the *potential* of a chemical to be a skin sensitizer but, with few exceptions, it is
484 insufficient to predict the relative *potency* of a chemical. As such, animal methods, in
485 particular the Local Lymph Node Assay (LLNA) are at present still needed to provide a
486 quantitative measure of relative sensitizing potency, which is critical for risk assessment
487 applications.

488 In order for the AOP for skin sensitization to be applied in practice, available test/non
 489 test approaches that characterize each of the KEs need to be mapped to the AOP. This
 490 mapping provides a perspective of what practical testing/non testing strategies could be
 491 derived as IATA. For skin sensitization, there has been considerable progress in
 492 developing specific test methods that target MIEs and many of the KEs relevant for the
 493 AO (see Table 4 for examples of appropriate assays).

494

495 Table 4. A summary of *in silico* and experimental testing approaches targeting MIEs and
 496 KEs of skin sensitization.

AOP level	Description	Level of biological organization	Test/Non test method
Dermal exposure	Dermal metabolism, epidermal disposition	Chemical structure & properties	(Q)SARs
MIE	Covalent binding between electrophile and skin protein	Molecular level	DRPA (Gerberick et al., 2004; 2007), GSH depletion assay (Schultz et al., 2005), QSARs/read-across
KE 1	Activation of inflammatory cytokines	Cellular response	KeratinoSens™ (Emter et al., 2010, 2013) read-across

KE 2	Maturation and mobilization of dendritic cells	Cellular response	MUSST (Python et al., 2007), h-CLAT (Sakaguchi et al., 2007), read-across
KE 3	T-cell proliferation	Organ response	LLNA (OECD Test Guideline (TG) 429), QMM, read-across
Adverse Outcome (AO)	Allergic contact dermatitis	Organism response	GPMT (OECD TG 406); HRIPT

497 **DRPA:** Direct Peptide Reactivity Assay, **GSH:** Reduced glutathione, **MUSST:** MYELOID U937 *SKIN*
498 *SENSITIZATION TEST*, **h-CLAT:** human Cell Line Activation Test, **LLNA:** Mouse Local lymph Node
499 Assay, **QMM:** quantitative mechanistic model, **GPMT:** Guinea Pig Maximization Test, **HRIPT:**
500 Human Repeat Insult Patch Test

501

502 A specific framework for the assessment of skin sensitization potential was adapted
503 from Figure 1 (shown in Figure S2 of the supplementary information). In applying the
504 framework, two outcomes can be envisaged – either the evaluation of the model/assay
505 outcomes will result in a consistent profile enabling an assessment of skin sensitization
506 hazard to be made (i.e. the substance is (not) a skin sensitizer with high confidence) or
507 the outcomes are insufficient to conclude with any great certainty that the substance is
508 (not) as skin sensitizer. The latter could be due to inadequacies in the model/assay
509 domains of applicability either on the basis of the underlying training sets or due to
510 technical limitations in the assays themselves (volatility, solubility, metabolic
511 competence). These insufficiencies however inform the development or refinement of

512 new test assays or refinement/extension of the *in silico* models. Any new information
513 then generated can be passed back to refine and improve the original AOP for
514 sensitization. A more detailed example for this IATA for skin sensitization has been
515 discussed in a separate manuscript (see Patlewicz et al, 2014).

516

517 **5.3 AChE inhibition leading to lethality**

518 Organophosphate and carbamate insecticides, which are widely used for agricultural
519 and residential purposes, have frequently been reported to cause toxicity to organisms
520 ranging from invertebrates to vertebrates and mammals (McHenry et al., 1997; Fulton
521 and Key, 2001). The toxicity of these compounds is mainly due to the selective inhibition
522 of acetylcholinesterase (AChE), leading to accumulation of acetylcholine (ACh) in the
523 synaptic cleft, subsequent overstimulation, and the disruption of nerve impulses
524 ultimately leading to ataxia, central respiratory paralysis, seizures, coma and death
525 (Costa, 2006, Bradbury et al. 2008). The well-developed knowledge on how these
526 chemicals cause lethality has led to the development of an AOP for acetylcholinesterase
527 inhibition leading to acute mortality (Russom et al., 2014). This AOP is characterized by
528 a clear mechanistic understanding of the MIE, KEs and AOs (Table 5) for a number of
529 species (Russom et al., 2014). The available information on relevant chemical structures,
530 the overall weight of evidence and the broad taxonomic applicability domain of this AOP
531 are of particular value to inform and provide input to IATAs, particularly for cross-
532 species extrapolations.

533

534 Table 5. A summary of *in silico* and experimental testing approaches relevant for
 535 different levels of the AOP – Acetylcholine esterase (AChE) inhibition leading to lethality
 536 (Russom, et al., 2014). References represent examples only. See Russom, et al. (2014)
 537 for a more extensive review of the literature supporting this AOP.

AOP level	Description	Level of biological organization	Test/Non test method
MIE	Inhibition of AChE activity. Inhibition caused by non-reversible or reversible inhibition.	Molecular level	QSARs/read-across Inhibition of AChE activity (<i>in vitro</i>) (Garcia-Reyero et al., 2014b; Holth and Tollefsen, 2012)
KE1	Accumulation of acetylcholine (ACh) in the synaptic cleft	Cellular level	No direct test-method available; biological plausibility well established; many studies linking MIE with downstream KEs & AOs across a variety of species (Bianco et al., 2013); Brain ACh levels can serve as a surrogate biomarker for associated KEs (Kobayash et al., 1985);

KE2	Excitatory responses in muscle and brain	Organ level	Electrophysiology in isolated neurons (Oyama, et al., 1989); Contractile response in muscle (Kobayash et al., 1994); Altered response in brain (biological plausibility well established)
AO	Neurotoxic symptomology (increased respiration, bradycardia, seizures) leading to death	Organism	Respiratory/cardiovascular responses (McKim, et al., 1987); Altered photomotor or locomotor response (Kokel et al. 2010, Irons et al. 2010, Garcia-Reyero et al 2014b);
AO	Population decline	Population	Inferred based on measured effects on mortality (Barata et al., 2004) and feeding behavior (Hunt, et al., 1991)

538

539 Since Acetylcholine esterase (AChE) inhibition is a well-established AOP, it can support a
540 variety of regulatory uses. The WoE supporting this AOP is strong (Russom, et al., 2014),
541 and there is extensive toxicity data for a number of chemicals in a variety of species that

542 is consistent with mechanistic knowledge assembled in the AOP
543 (<http://www.epa.gov/ecotox/>). Information from *in vitro* results could potentially be
544 used under certain circumstances, but the use of *in vitro* AChE inhibition alone may not
545 be sufficient (Knudsen et al., 2011) possibly due to lack of these assays accounting for
546 bioactivation of certain chemicals such as Diazinon by metabolism (Aylward et al., 2011)
547 or mitigation of effects by metabolic degradation such as observed for malathion (de
548 Bruijn and Hermens, 1993). *In silico* approaches might be sufficient for some uses
549 (Fukuto et al., 1990; El Yazal et al., 2001; Wong et al., 2012), but should be used with
550 caution particularly in cases where metabolic activation is required (de Bruijn and
551 Hermens, 1993). Extensive *in vivo* data exist with reasonable concordance seen between
552 sequence similarity among AChE enzymes and *in vivo* activity across non-vertebrate
553 species (Russom, 2014). For animals, including humans, determination of AChE
554 inhibition in both the central and peripheral nervous systems are considered crucial for
555 a thorough evaluation of potential hazard
556 (<http://www.epa.gov/pesticides/trac/science/cholin.pdf>). However, blood
557 cholinesterase inhibition is accepted as a surrogate parameter in humans, when data for
558 AChE inhibition in peripheral and central nervous system are not available.
559 Recommendations on surrogate parameters in wildlife have currently not been
560 developed sufficiently to support a WoE approach to identify potential hazard.

561

562 To illustrate how this AOP could be used in IATA, consider the classification of a
563 pesticide known to act via AChE inhibition as a potential application. A particular
564 concern in this case is the biological impact on non-target organisms (see Figure S3 in
565 supplementary information). If this is a crop use that is expected to result in minimal

566 exposure through either application or ingestion, the species of concern might be
567 restricted to non-target organisms that would be exposed during the application or via
568 interactions with the treated crops and possibly aquatic organisms from run off
569 following application. Demonstration of low level of exposure in combination with low
570 sensitivity for AchE in vertebrates, would be expected to limit potential hazards to non-
571 target invertebrates. If toxicity data from the target species (e.g. insects for use of
572 insecticides) exist, hazard assessment could be facilitated by sequence alignments to
573 predict cross-species susceptibility to non-target species where exposure is considered
574 relevant (Lalone et al., 2013; Russom et al., 2014). Documentation of potential risk
575 scenarios (e.g. small margin of safety between exposure and potential effects) based on
576 the non-testing approaches proposed herein, may lead to a decision to generate
577 additional testing data using *in vivo* studies with the appropriate species or relevant
578 surrogate species in cases where testing is not feasible (e.g. endangered species, lack of
579 appropriate laboratory strains etc.).

580 This hypothetical case study illustrates how a well-defined AOP could be used for certain
581 regulatory purposes independent of chemical specific information at the intermediate
582 key events. The weight of evidence incorporates over 50 years of research including
583 basic biochemistry as well as toxicology. Given the strong support and conservation of
584 the AOP across taxa, a wealth of toxicological data at the organism level can be leveraged
585 for the decision at hand. This allows the use of *in silico* predictions for cross-species
586 extrapolations in combination with use of data from experimentally tractable species to
587 limit the need for additional studies to characterize intermediate events of well-
588 developed AOPs. If this were not the case, other approaches such as *in vitro* screening
589 and *in vivo* measurement of intermediate KE (Figure S3 in supplementary information)
590 would likely be required to safeguard against adversely impacting non-target species.

591

592 **6. Implications for Integrated Toxicology and Regulatory Applications**

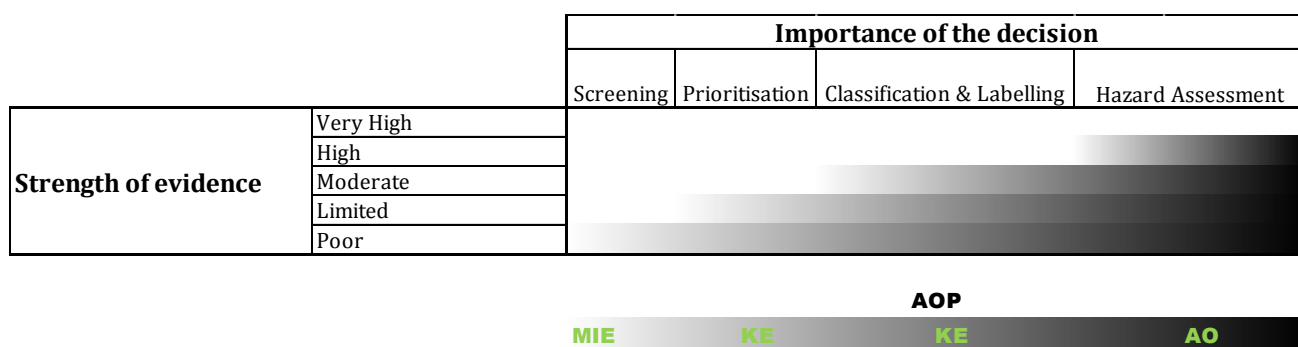
593 Development and application of AOP-informed IATA represents a new way to evaluate
594 and generate information to meet different regulatory purposes. A conceptual
595 framework for applying IATA has been proposed that considers existing information
596 (from a hazard and exposure perspective) in the context of an AOP to make an informed
597 decision based on the regulatory context. Frameworks to characterize the scientific
598 confidence of an AOP that are required to meet different regulatory needs are in
599 development (Becker et al, 2014; Patlewicz et al., *submitted*; Perkins et al., *submitted*).
600 These will shape the structure of the IATA and its elements in terms of the test methods
601 and non-testing approaches. Establishing scientific confidence is critical for both the
602 elements making up the IATA as well as the IATA as a whole. Three case studies have
603 been described in detail to illustrate how the conceptual framework proposed in Section
604 1 can be adapted to meet different regulatory purposes (e.g. prioritization, hazard
605 assessment, classification and labelling and other applications such as cross-species
606 extrapolations).

607 The EDC example shows how a battery of AOPs and associated HT assays can be used in
608 a prioritization scenario. This addresses the first 4 principles for IATA development and
609 application as outlined in Section 4. The skin sensitization example, which is aimed at
610 addressing hazard assessment, arguably addresses all 5 principles. The AchE inhibition
611 example illustrates how an established AOP can be used for classification and labelling in
612 certain regulatory contexts despite a lack of properly developed testing and non-testing
613 methods spanning the full AOP continuum. In the regulatory context considered for that
614 example, the need for explicit tests of intermediate KEs is avoided by the wealth of data

615 available. A well-developed AOP and by demonstration of phylogenetically-conserved
616 MIEs across taxa enable identification of susceptible species being particularly relevant
617 or tractable to cost-efficient *in vivo* testing (e.g. invertebrates). Clearly the degree to
618 which these principles need to be characterized can and will differ based on the level of
619 uncertainty that can be tolerated for the regulatory purpose under consideration.
620 Scientific confidence of the AOP and its associated IATA will be strongest where there is
621 a close link between the MIE and KEs to the AO.

622 There is a desire to exploit *in silico* and HTS testing tools to populate an IATA. One
623 starting point for such AOP-informed IATAs could be to apply *in silico* methods or HT
624 approaches for providing information about the MIE to determine what data if any
625 would need to be generated for different KEs or the AO for a given chemical. The
626 stronger the evidence coming from non-testing or alternative testing approaches, the
627 less additional information would, in theory, need to be generated for a given decision.
628 Thus, a moderate level of confidence might be sufficient for a prioritization purpose, but
629 in order to make a decision related to hazard and risk assessment, assays or a
630 combination of assays closely linked to the MIE and with high predictivity of the AO may
631 be required (Figure 2). Additional information on one or more KEs along the pathway
632 generated from *in vitro*, *in chemico* or HT/HC assays would serve to provide increasing
633 confidence for a given decision.

634



635

636 *Figure 2. Relationship between strength of evidence (reliability, relevance and concordance*
637 *etc.) for the IATA supporting the AO of regulatory concern and the importance of the*
638 *regulatory decision to be made. The figure shows the possible combination of AOP based*
639 *information and available data, and how the use of one could be strengthened by the other.*
640 *The color of the cells represent the amount of additional information from other sources*
641 *needed for a decision (the darker the color, the more additional information is needed to*
642 *reach a decision with confidence).*

643

644 Of course implicit in such a confidence determination, is the WoE evaluation of the AOP
645 itself. The OECD AOP guidance entails completing a template, using evolved and tailored
646 Bradford Hill (BH) considerations, in which each KEs and KE relationships (KERs) in an
647 AOP are evaluated and are scored as high, moderate and low (OECD, 2013). The
648 outcomes of these WoE determinations for the KEs and KERs of an AOP help in making a
649 determination of whether different decisions are feasible based on the outcomes of MIEs
650 or other KEs and the extent to which they are predictive of the AO.

651

652 The case studies presented could in theory be applied in practice now, although the
653 number of well-developed AOPs is currently limiting the practical applicability for larger
654 scale regulatory deployment. Furthermore, consideration needs to be given to the

655 analytical validation of testing and non-testing approaches in order to better
656 characterize their applicability domain i.e. the types of chemicals that can be reliably
657 assessed. A detailed description of AOPs of regulatory relevance and the establishment
658 of qualitative and quantitative links between MIEs, KEs and AOs will additionally help
659 foster application for different regulatory decisions. While qualitative links are already
660 established for a number of the AOPs so far developed and supported by visualization
661 and description tools such as the AOP Knowledge Base (<https://aopkb.org>), appropriate
662 quantitative approaches for confidence evaluation by WoE assessments of KERs are
663 currently being critically assessed (Becker et al., *in preparation*). Recent initiatives to
664 provide quantitative assessment of the role of MIE and KE proximity to the AO for the
665 confidence of predictions to regulatory-relevant endpoints will likely also assist in
666 developing pragmatic tools for IATA development. Additional improvements of IATAs by
667 including toxicokinetics and reverse dosimetry into extrapolations to regulatory-
668 relevant endpoints would further increase the applicability of IATAs for practical use.

669

670 Although not necessarily applicable to the case studies highlighted here, many of the
671 AOPs in development have been data-rich and based on historical *in vivo* data. Thus the
672 body of evidence to justify the essentiality of KEs and the linkages has facilitated
673 different use scenarios including risk assessment where the KEs proximal to the AO are
674 better defined. Going forward, the challenges foreseen will be to identify the data gaps
675 and assay needs, to integrate different AOPs together to provide a more holistic
676 assessment of likely effects. The latter is a major issue as an AOP by its nature assumes
677 that adversity can be described by a relevant assembly of MIEs and KEs, whereas the

678 question remains of how many AOPs need to be integrated into IATA to assure that there
679 is no important hazard or adversity overlooked.

680

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710

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