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**Synthesis, Ni(II) Schiff base complexation and structural analysis
of fluorinated analogs of the ligand (*S*)-2-[N-(N'-
benzylpropyl)amino]benzophenone (BPB)**

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Abstract

Herein we report the first X-ray crystal structure of the well-known (*S*)-L-ala-Ni-BPB complex (**1**) and compare this with the X-ray crystal structures obtained for two novel fluorinated (*S*)-L-ala-Ni-BPB complexes (**8** and **12**) that contain either an *S*- or *R*-fluorine atom on the proline ring of the BPB ligand. The preparation of complexes **8** and **12** has been enabled by the synthesis of two new fluorinated BPB ligands (**7** and **11**). In this work we looked to observe structural effects that the introduction of a single fluorine atom has on the known complex **1**. Arising from this, we highlight a novel fluorine – nickel interaction that may provide additional stabilisation to complex **8**.

Keywords: *fluoro-proline, fluorinated ligands, gauche effect, nickel complex, X-ray crystallography*

1. Introduction

The introduction of a fluorine atom in to a molecule can have a remarkable effect on its physical and chemical properties, metabolic stability and its biological activity. This has been widely exploited within medicinal chemistry and is discussed in detail in a number of recent review articles [1]. Fluorine substitution can also be used to fine-tune the stereo-electronic properties of a molecule and in doing so allow conformational control [2,3]. For example β -fluoro imines have been shown by *X*-ray crystallographic analysis and theoretical DFT calculations, to have a torsion angle (ϕ) of approximately 60° about the NCCF bonds (Figure 1), therefore, preferentially adopting a *gauche* conformation [4]. This is explained by firstly, the avoidance of an electronically destabilising *anti*-arrangement around the NCCF bond (Figure 1), in which the electron-withdrawing fluorine atom lies directly opposite to the electron deficient imine or ammonium group. Secondly, the favoured conformation puts a hydrogen atom *anti*- to the iminium or ammonium group, thus allowing a stabilising electron-donation from $\sigma_{\text{C-H}}$ to $\sigma^*_{\text{C-N}^+}$ by hyperconjugation (Figure 1). This has been utilised as a tool in medicinal chemistry, where introduction of a single fluorine atom with the correct stereochemistry selectively stabilises a compound in a biologically active conformation *e.g.* 3-fluoro-N-methyl-D-aspartic acid, whereas, the opposite enantiomer stabilises an inactive conformer [5]. Furthermore, 4-fluoroprolines have been incorporated in to collagen triple helices with the observation that this modification confers hyperstability within these structures due to the above stereoelectronic effects [6]. A similar approach using fluoropyrrolidine derivatives (Figure 1) has also been pioneered recently by the Gilmour group as a method to control molecular conformation in acyclic systems, and thus, probe catalyst mechanism, reactivity and selectivity in asymmetric organocatalysis [7].

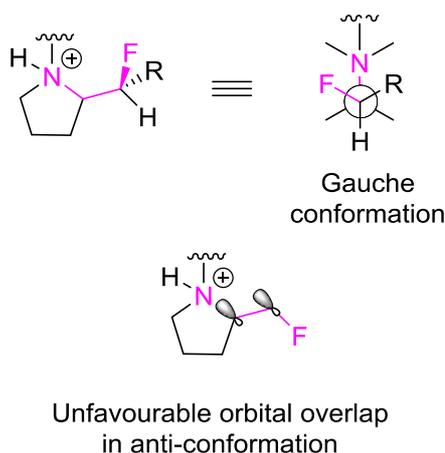
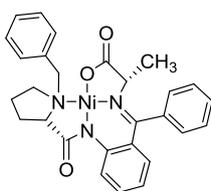


Figure 1 – Fluorine-ammonium *gauche* effect in 5-membered nitrogen-containing heterocycles [6].

Nickel (II) Schiff base complexes of (*S*)-2-[N-(N'-benzylpropyl)amino]benzophenone (BPB) and amino acids (*e.g.* **1**) have been widely used as nucleophilic glycine equivalents in the asymmetric synthesis of C α -substituted amino acids[8]. Soloshonok *et al* have also investigated the modification of the chelating ligand, replacing the proline ring in **1** to prepare novel NH-Ni ligands and observing the resulting diastereomeric ratio of the Ni-complexes and the consequences in alkylation and Michael addition reactions [9, 10]. Incorporation of fluorine at the 3-position of protonated pyrrolidines has been shown to stabilise particular conformations through β -fluoro-ammonium interactions [11]. In light of this we hypothesised that incorporation of a single fluorine atom on the proline ring within the BPB ligand could offer a novel route to modulate the conformation and hence the reactivity of a BPB-nickel (II) type complex. As such, we sought to probe whether the introduction of a fluorine atom in the correct configuration at the carbon beta to the proline nitrogen atom in **1** could invoke conformational effects, such as a fluorine-ammonium ion *gauche* effect, leading to complexes with an altered chelate ring configuration around the nickel atom.



1

Here we report the synthesis and the first ever X-ray crystal structure of the well-known (*S*)-L-Ala-Ni-BPB Schiff base complex (**1**). This structure has been compared with the X-ray crystal structures obtained for two novel fluorinated (*S*)-L-Ala-Ni-BPB complexes (**8** and **12**) that

contain either an *S*- or *R*-fluorine atom on the proline ring of the BPB ligand. The preparation of complexes **8** and **12** has been enabled by the synthesis of two new fluorinated BPB ligands (**7** and **11**). Furthermore, we highlight a novel Ni-F interaction and use DFT calculations to support a role for this in stabilizing the complex configuration.

2. Results and discussion

We initially prepared the reported (*S*)-BPB ligand [12] and corresponding (*S*)-L-Ala-Ni-BPB complex (**1**) [13] according to the published methods and obtained crystals of **1** by slow evaporation from acetone and methylethylketone. This was to provide a comparison of the overall structure of **1** with the fluorinated derivatives in X-ray diffraction studies (Figure 2). In spite of being synthesized and used many times by researchers in the field of peptide chemistry [14], the structure of complex (**1**) has not yet been reported in the literature (CCDC ver. 2013). Herein, we report the first X-ray structure of (*S*)-L-Ala-Ni-complex (**1**) (in form of MEK solvate).

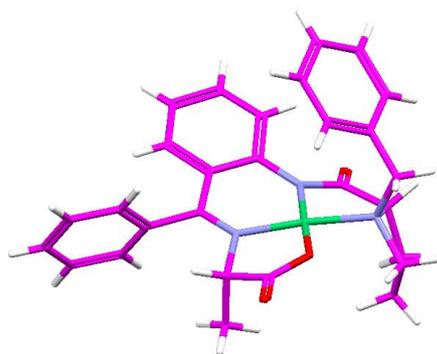
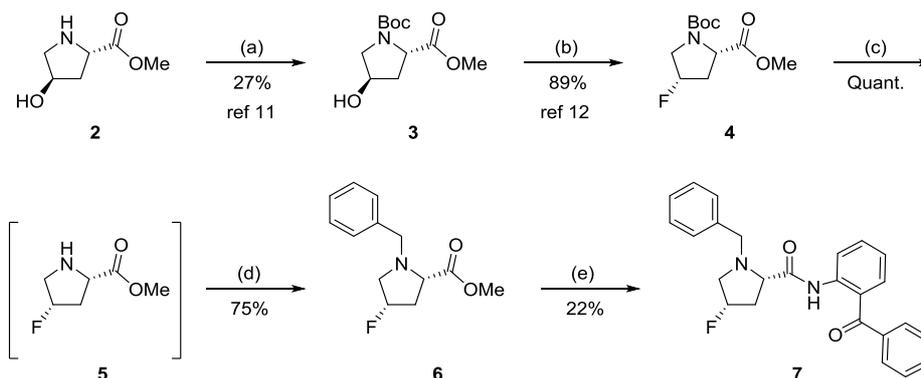


Figure 2. First reported X-ray crystal structure of (*S*)-L-Ala-Ni-BPB complex **1**. The MEK solvent molecule and one of the component of disordered benzyl group are omitted for clarity. All crystal structure drawings were produced with Olex2 [15]. All structural images produced in Hermes [16].

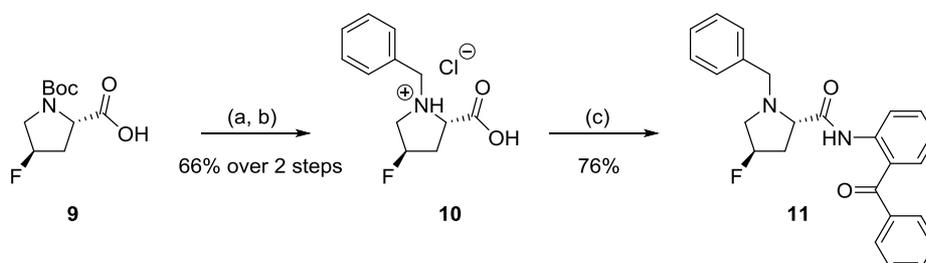
We attempted to prepare the analogous *R*- and *S*- fluorinated nickel (II) complexes of the L-Schiff base of (*S*)-2-[*N*-(*N'*-benzylprolyl)amino]benzophenone (BPB) and L-alanine (**7** and **12**) starting from commercially available proline derivatives (Scheme 1 and Scheme 2, respectively). *Trans*-hydroxyproline methyl ester (*trans*-HypOMe, **2**) was protected as the reported [17] *N*-(*tert*-butylcarbamate) (**3**) prior to a 2-step fluorination procedure employing tetrabutylammonium difluorotriphenylsilicate (TBAT) followed by treatment with perfluoro-

1-butanefluoride (PBSF), with inversion of stereochemistry at the 4-position [18]. Removal of the Boc protecting group preceded *N*-benzylation using standard conditions. Conversion of the methyl ester (**6**) in to the required amide (**7**) was achieved directly through an oxidative process involving potassium *tert*-butoxide and exposure to atmospheric oxygen [19].



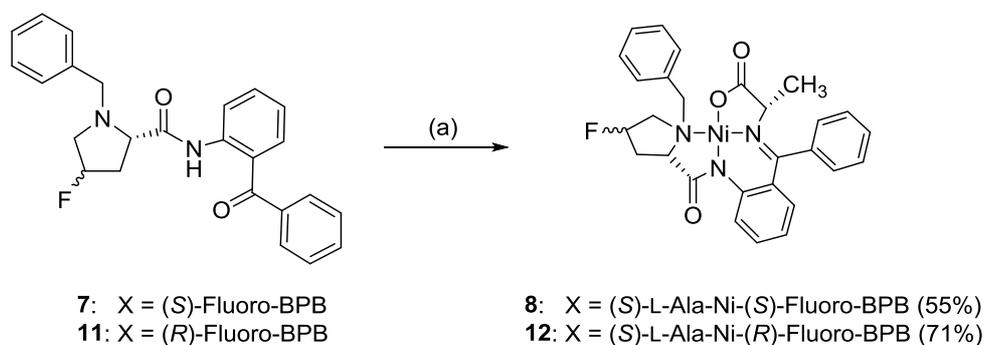
Scheme 1. Reagents and conditions: a) Boc_2O , NEt_3 , CH_2Cl_2 , rt, 18 h; b) i) TBAT, DIPEA, CH_2Cl_2 , ii) PBSF, rt, 72 h; c) TFA, CH_2Cl_2 , rt, 2 h (used in next step without purification); d) benzyl bromide, K_2CO_3 , DMF, rt, 20 h; e) 3-aminobenzophenone, $t\text{-BuOK}$, THF, open to air, rt, 70 min.

To prepare the alternative (*R*)-fluorinated ligand (**Scheme 2**), commercially sourced *N*-Boc-*trans*-4-fluoro-L-proline (**9**) was deprotected by treatment with trifluoroacetic acid and used directly in an *N*-benzylation reaction without purification (**Scheme 2**) [20]. On this occasion, the free acid of **10** was coupled with 2-aminobenzophenone to afford amide **11** by initial conversion of the acid in to a mixed anhydride intermediate [21].



Scheme 2. Reagents and conditions: a) TFA, CH_2Cl_2 , rt, 2 h (used in next step without purification); b) benzyl chloride, KOH, $i\text{-PrOH}$, 50°C , 6 h; c) MsCl, 2-aminobenzophenone, Et_3N , CH_2Cl_2 .

With the fluorinated BPB ligands (**7** and **11**) in hand, we subsequently attempted to prepare the corresponding Ni-complexes (**Scheme 3**). Following the standard protocol for complexation with Ni(II), chiral ligand **7** was treated with nickel (II) nitrate hexahydrate and L-alanine under basic conditions with gentle heating. This reaction was performed in parallel with the analogous reaction to prepare the reported des-fluoro complex **1**. After 2 h formation of **1** was complete and afforded a deep red solution. By comparison, more than 72 h were required for the (*S*)-fluorinated derivative (**8**) to reach completion. *R*-fluoro-BPB ligand **11**, however, formed the desired Ni(II) complex (**12**) rapidly, within 20 min according to TLC, however, heating was continued for 24 h to ensure complete conversion. Crystals of complexes **8** and **12** were obtained as dark red needles by slow evaporation from diethyl ether and the structures were determined by X-ray analysis.



Scheme 3. Reagents and conditions: a) L-alanine, Ni(NO₃)₂·6H₂O, MeOH, KOH, 60 °C, 24-72 h.

It was proposed that, with the correct (*S*)-stereochemistry at the fluorine position, we would be able to stabilise the existing conformation by replacement of a hydrogen atom already orientated in the *gauche* conformation with respect to the nearby nickel-coordinating nitrogen atom (Figure 2).

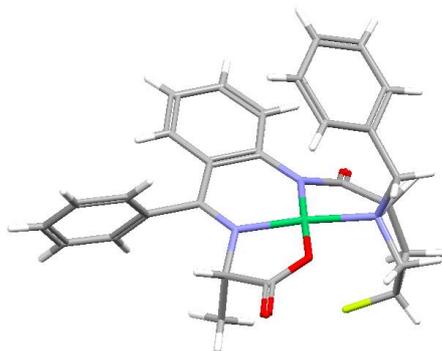


Figure 3. X-ray crystal structure of L-Ala-Ni-(S)-FBPB complex **8**.

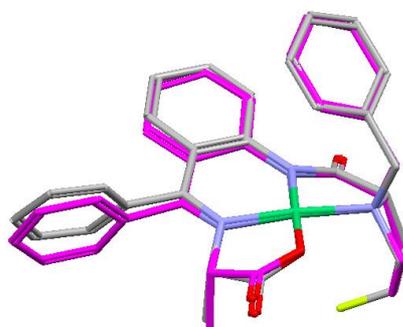


Figure 4. Least-squares superposition of fluorinated complex **8** (grey) with non-fluorinated complex **1** (pink).

The fluorinated and non-fluorinated complexes were aligned using the central nickel and its coordinated heteroatoms in molecular viewer Hermes [16] to an RMSD of less than 0.01 Å (Figure 3). The structure **8** contained two independent molecules with virtually identical configuration of the coordination polyhedron of metal atom. Overlay of **1** and **8** (Figure 4) showed that replacement of the corresponding *gauche* hydrogen with fluorine does not perturb the favoured conformation with respect to the benzyl group. We observed that the fluorine atom of **8** lies in the anticipated *gauche* conformation (Figure 5), with a measured NCCF bond torsion angle of 76° (slightly greater than 60° due to ring strain in the pyrrolidine) that is consistent with the corresponding non-fluorinated complex (**1**).

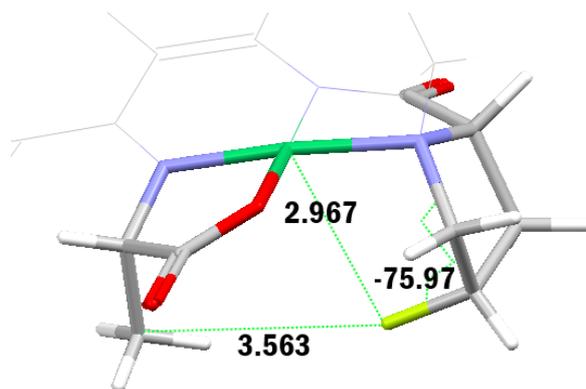


Figure 5. Expansion of a section of complex **8** showing the measured NCCF bond torsion angle of 76° and also displaying a putative C-H \cdots F bond [**Error! Bookmark not defined.**].

Replacement of the hydrogen atom with fluorine introduces two further interesting intramolecular features in **8**. Firstly, a putative intramolecular C-H \cdots F bond may be observed (Figure 5). The existence of such interactions is controversial and many authors have concluded that any interaction may be only providing van der Waals stabilisation [22]. However, others have reported that despite being very weak non-covalent interactions, C-H \cdots F interactions have been observed in organic structures, notably in some fluorobenzenes [23], and that these may be enhanced when the proton of the C-H bond is more acidic [24]. In complex **8** the fluorine atom is found to be within hydrogen bonding distance (2.64 \AA) of the proximal methyl hydrogens (Figure 5). Although these are likely to be weak interactions, this could confer additional stability to the molecule. Secondly, as observed in the crystal structure, fluorine is positioned directly below the central nickel ion at a distance of 2.97 \AA , which is smaller than the sum of their van der Waals radii (3.10 \AA). As no alteration in the crystal structure of **8** is observed relative to the non-fluorinated compound **1** (Figure 4) *e.g.* due to repulsion between the F and Ni atoms, this may suggest that a non-bonding interaction arises, which may stabilise the complex.

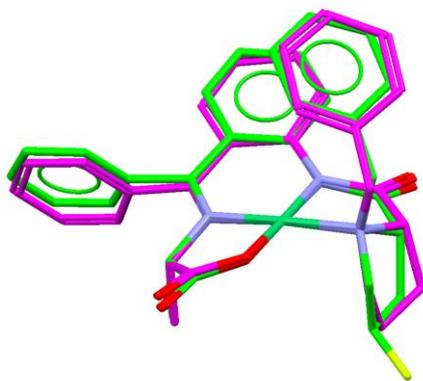


Figure 6. Least squares superposition of complex **1** (pink) and complex **12** (green).

Complex **12** was aligned to complex **1** as described above, to an RMSD of 0.05 Å. The greatest difference in the structures, as seen in Figure 6, is in the fluorinated proline ring. The position of the fluorine in complex **12** abolishes the potential nickel interaction observed in complex **8**. Furthermore, the proline ring in complex **12** is twisted, where it is in the envelope conformation in complex **8** (Figure 7). A comparison of complex **12** with the CSD (version 5.35) using Mogul shows the torsions within this ring to be well represented, with the nearest comparable structure only 0.89° RMSD difference over ring torsions, and with 40% of the returned structures within 10°. Finally, the NCCF torsion angle of complex **12** differs greatly from that of complex **8** (Table 1, Figure 8), which is an expected change resulting from introduction of equatorial and axial substituents.

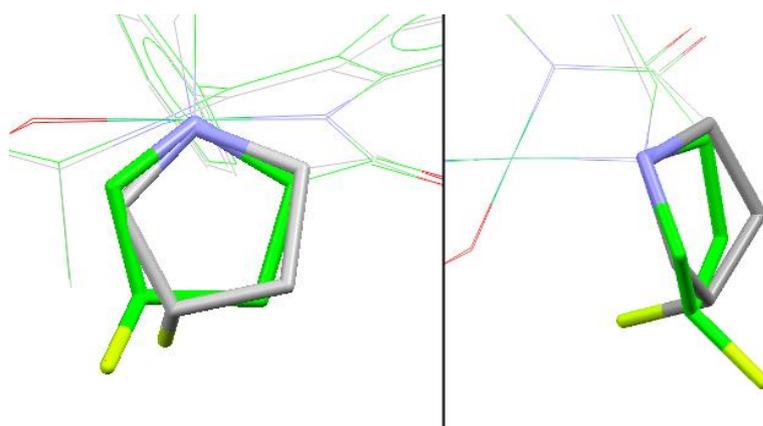


Figure 7. The proline ring of complex **12** (green) is twisted, whereas the ring of complex **8** (grey) adopts an envelope conformation.

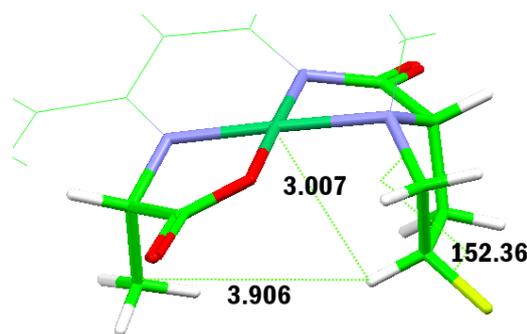


Figure 8. The NCCF torsion angle of complex **12** is 152.4°.

Table 1: Torsion angles for ΦNCCH and ΦNCCF . *For complex **1**, NCCH_2 angle is given as NCCF for comparison.

Ni Complex	$\Phi\text{NCCH} / ^\circ$	$\Phi\text{NCCF} / ^\circ$
1	-73.31	161.70*
8	162.98	-75.97
12	-86.23	152.36

To investigate the differences between the complexes in more detail DFT, calculations were carried out using the crystal structures of complexes **8** and **12**. By comparing the calculated energies of the R-fluorine and S-fluorine complexes it would be possible to estimate the strength of the interaction between S-fluorine and nickel. DFT calculations were performed as implemented in the Gaussian03 package. The B3LYP functional was used in conjunction with the 6-311 + +G** basis set. The difference in calculated energies between the two complexes (**8** and **12**) was determined to be 5.99 kJ/mol (in favour of the S-fluorine complex **8**). This energy difference supports the notion of a stabilizing non-bonding interaction between the S-fluorine and the nickel in complex **8** as observed in the X-ray structure.

3. Conclusions

Here we report the first X-ray crystal structure of the known nickel (II) Schiff base complex (**1**). This structure has been compared with the X-ray crystal structures obtained for two novel fluorinated nickel (II) Schiff base complexes that contain either an S- and R-fluorine atom beta to the proline nitrogen on the BPB ligand (**8** and **12** respectively). We discovered that the introduction of an S-fluorine atom (complex **8**) has no significant effect upon the overall structural geometry of the complex. However, whilst we accept that these may be artifacts of

the structures being solved in the crystal form, we speculate that inclusion of a fluorine may enhance the conformational rigidity of complex **8** through a combination of the formation of a C-H ... F bond and a novel Ni ... F non-bonding interaction, as well as through a fluorine-ammonium *gauche* interaction. DFT calculations also supported this hypothesis with a clear difference in the calculated energies of 5.99 kJ/mol between the S-fluorinated complex **8** and the R-fluorinated complex **12** (no Ni... F interaction observed in the X-ray structure). Introduction of the *R*-fluorine atom substituent (complex **12**) also afforded no significant overall changes to the structure although as expected some minor conformational changes in the proline ring of complex **12** were observed. Further investigations to probe any differences between complex **1** and the fluorinated complexes **8** and **12**, with regard to reactivity and stability, are currently underway.

Acknowledgements

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4. Experimental

4.1 Materials

L-4-trans-hydroxyproline methyl ester hydrochloride (**2**) (Novabiochem) and *N*-Boc-*cis*-4-fluoro-L-proline methyl ester (**9**) (Sigma Aldrich) were purchased from commercial suppliers and used directly as indicated in the appropriate experimental procedures. All other chemicals were purchased from Sigma Aldrich. Complex **1** [**Error! Bookmark not defined.**] and intermediates **3** [17] and **4** [18] were prepared according to literature procedures and the obtained analytical data was in accordance with the published characterizations. The preparation of *N*-benzyl-(*S*)-4-fluoro-L-proline methyl ester (**6**) followed a previously undescribed route (details below) but analysis indicated that this was identical to the reported substance [19].

4.2 Characterization

NMR spectra were collected using Bruker Avance 400 MHz or Varian VNMRs 700 MHz spectrometers. Multiplicities are reported as: s = singlet, d = doublet, m = multiplet, t = triplet or combinations thereof *e.g.* dd = doublet of doublets. Chemical shifts are reported in ppm and are referenced to residual solvent peaks; CHCl₃ (¹H 7.26 ppm, ¹³C 77.2 ppm), H₂O (¹H 4.79 ppm). *J* couplings are measured in Hertz (Hz). Reactions were monitored by TLC using Merck precoated silica gel plates. Column chromatography was performed using silica gel with the solvent system indicated. All reported yields refer to the isolated yield and the product purity was estimated to be >95% by ¹H NMR. IR spectra were recorded on a Perkin Elmer Spectrum RX1 fitted with an ATR attachment. IR absorptions reported are in cm⁻¹. Mass spectra were collected on a Waters TQD mass spectrometer and accurate mass spectra were collected on a Waters LCT Premier XE mass spectrometer. Optical rotations were measured using a Jasco P-1020 polarimeter with samples in the solvents, temperatures and concentrations as indicated.

4.2.1 *N*-benzyl-(*S*)-4-fluoro-*L*-proline methyl ester (6) [19]. A solution of **4** (0.50 g, 2.00 mmol) in CH₂Cl₂ (25 mL) and trifluoroacetic acid (2.5 mL) was stirred at room temperature for 2 h. Upon completion of the reaction, volatiles were removed under reduced pressure, affording a colourless solid (0.29 g, quant.). Trifluoroacetate salt **5** was used directly in the subsequent step without further purification. Potassium carbonate (1.47 g, 10.6 mmol) was added to a rapidly stirred suspension of **5** (0.55 g, 2.12 mmol) in anhydrous DMF (50 mL) and gradually reached homogeneity after 1 h at room temperature. Benzyl bromide was added in one portion and the reaction mixture was stirred for a further 48 h at room temperature until TLC indicated the complete consumption of starting materials. The crude mixture was poured over crushed ice and stirred until thawing was complete and the emulsion was extracted with Et₂O (100 mL), which was then washed with brine (sat.) (3 × 30 mL). Purification by column chromatography (silica; EtOAc – hexane 0% to 50%) afforded the desired product (0.39 g, 75%). All analyses were consistent with the reported preparation [25].

Pale yellow oil, ¹H NMR (400 MHz, CDCl₃): δ 7.35 – 7.22 (m, 5H, Ph), 5.08 (dm, *J* = 54.7 Hz, 1H, CHF), 4.02 (d, *J* = 13.1 Hz, 1H, benzyl CH), 3.69 (s, 3H, OMe), 3.58 (d, *J* = 13.1 Hz, 1H, benzyl CH), 3.33 – 3.20 (m, 2H, CH₂), 2.63 – 2.42 (m, 2H, CH₂), 2.27 (m, 1H, CH); ¹³C NMR (101 MHz, CDCl₃): δ 172.2, 136.4, 128.1, 127.3, 126.3, 91.0 (d, *J*_{C-F} = 173.2 Hz), 62.7, 58.2 (d, *J*_{C-F} = 22.1 Hz), 56.8, 51.0, 36.2 (d, *J*_{C-F} = 22.1 Hz); ¹⁹F NMR (376 MHz, CDCl₃): δ -168.99 (m, 1F); LCMS (ESI+) 238.09 [M+H]⁺.

4.2.2 (S)-2-[N-(N'-benzyl-(S)-4-fluoro-L-prolyl)amino]benzophenone (7). The methyl ester **6** (0.26 g, 1.1 mmol) was added to a solution of ^tBuOK (0.25 g, 2.19 mmol) in moist THF (15 mL), which was pre-stirred open to the air for 1 minute. Addition of 2-aminobenzophenone (0.22 g, 1.1 mmol). After stirring at room temperature for 1 h TLC indicated that the reaction was incomplete. Additional ^tBuOK (123 mg, 1.1 mmol) was added and again stirred for 1 h. The crude mixture was concentrated *in vacuo* and partitioned between CH₂Cl₂ (20 mL) and water (20 mL). The organic extract was dried (MgSO₄) and concentrated to an oil. The title compound was isolated after column chromatography (silica; EtOAc – hexane 0% to 30%) (97 mg, 22%) as a pale yellow oil.

$[\alpha]_D^{23} - 41.1$ ($c = 1.0$, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 11.61 (s, 1H, NH), 8.55 (d, $J = 8.3$ Hz, 1H, Ar), 7.83 – 7.75 (m, 2H, Ar), 7.62 – 7.56 (m, 1H, Ar), 7.56 – 7.45 (m, 4H, Ar), 7.45 – 7.38 (m, 2H, Ar), 7.22 – 7.15 (m, 3H, Ar), 7.09 (t, $J = 7.6$ Hz, 1H, Ar), 5.10 (dt, $J = 52.9, 4.1$ Hz, 1H, CHF), 4.00 (d, $J = 13.0$ Hz, 1H, benzyl CH), 3.63 (d, $J = 13.0$ Hz, 1H, benzyl CH), 3.55 – 3.39 (m, 2H, CH), 2.70 – 2.43 (m, 2H, CH₂), 2.31 (m, 1H, CFCH), 1.26 (s, 1H, CH); ¹³C NMR (101 MHz, CDCl₃): δ 199.1, 172.2, 139.7, 139.0, 137.6, 133.5, 132.7, 128.3, 127.5, 127.3, 126.2, 125.2, 121.7, 120.9, 92.2 (d, $J_{C-F} = 179.5$ Hz), 62.4, 58.4 (d, $J_{C-F} = 22.0$ Hz), 57.0, 51.1, 36.6 (d, $J_{C-F} = 22.1$ Hz); ¹⁹F NMR (376 MHz, CDCl₃): δ -172.11 (m, 1F); IR (neat) ν 3320, 2945, 1675, 1655, 1287, 1115, 703 cm⁻¹; LCMS (ESI+) $m/z = 403.19$ [M+H]⁺. HRMS (ESI) calcd for C₂₅H₂₄FN₂O₂ [M+H]⁺: $m/z = 403.1812$, found 403.1822.

4.2.3 L-Ala-Ni-(S)-fluoro-BPB complex (8). To a solution of **7** (27 mg, 0.08 mmol) and Ni(NO₃)₂·6H₂O (44 mg, 0.15 mmol) in anhydrous methanol (2 mL) was added a solution of KOH (34 mg, 0.60 mmol) and L-alanine (34 mg, 0.38 mmol) in methanol (1 mL). This mixture was heated at 60 °C with stirring for 72 h – TLC indicated the reaction was complete and the deep red solution concentrated *in vacuo* before partitioning between water (30 mL) and CHCl₃ (20 mL). Trituration of the concentrated organic extract under Et₂O afforded a dark red solid that was isolated by filtration under suction and washed with more Et₂O. Purification of the crude material by preparative TLC (silica; acetone – CHCl₃ 20%) afforded a red/orange solid (22 mg, 55%).

mp 288 – 290 °C (dec.); $[\alpha]_D^{23} + 255$ ($c = 2.0$, CHCl_3); ^1H NMR (700 MHz, CDCl_3): δ 8.09 (d, $J = 7.9$ Hz, 3H, Ar), 7.54 – 7.47 (m, 2H, Ar), 7.47 – 7.42 (m, 1H, Ar), 7.38 (t, $J = 7.7$ Hz, 2H, Ar), 7.25 – 7.21 (m, 2H, Ar), 7.14 (ddd, $J = 8.7, 6.9, 1.8$ Hz, 1H, Ar), 6.97 (dm, $J = 7.7$ Hz, 1H, Ar), 6.67 (m, 1H, Ar), 6.62 (dd, $J = 7.7, 1.8$ Hz, 1H, Ar), 5.49 (dm, $J = 52.7$ Hz, 1H, CHF), 4.59 (d, $J = 12.7$ Hz, 1H, benzyl CH), 3.99 (m, 1H, CH), 3.90 (m, 1H, CH), 3.65 (dd, $J = 11.6, 4.7$ Hz, 1H, CH), 3.58 (d, $J = 12.7$ Hz, 1H, benzyl CH), 2.91 (m, 1H, CH), 2.76 (m, 1H, CH), 2.25 (ddd, $J = 36.4, 11.6, 3.5$ Hz, 1H, CH), 1.60 (m, 3H, CH_3); ^{13}C NMR (101 MHz, CDCl_3): δ 179.9, 170.1, 141.9, 133.4, 133.1, 132.6, 131.9, 131.7, 129.6, 129.2, 129.0, 128.8, 127.5, 127.2, 126.7, 123.8, 120.8, 91.4 (d, $J = 179.0$ Hz), 68.6, 66.9, 62.9, 61.5 (d, $J = 20.4$ Hz), 38.4 (d, $J = 22.4$ Hz), 20.3; ^{19}F NMR (376 MHz, CDCl_3): δ -167.18 (m, 1F); LCMS (ESI+) $m/z = 530.12$ $[\text{M}+\text{H}]^+$; HRMS (ESI) calcd for $\text{C}_{28}\text{H}_{27}\text{FN}_3\text{O}_3\text{Ni}$ $[\text{M}+\text{H}]^+$: $m/z = 530.1382$, found 530.1390.

4.2.4 N-benzyl-(R)-4-fluoro-L-proline hydrochloride salt (10). A solution of *N*-Boc-(*R*)-4-fluoro-L-proline (0.5 g, 2.18 mmol) in CH_2Cl_2 (25 mL) and trifluoroacetic acid (2.5 mL) was stirred at room temperature for 2 h. Upon completion of the reaction, volatiles were removed under reduced pressure, affording a colourless solid (0.52 g, quant.). The trifluoroacetate salt was used directly in the subsequent step without further purification. A solution of the salt in isopropanol (5 mL) was added to a fully dissolved solution of potassium hydroxide (0.61 g, 10.9 mmol) in isopropanol (25 mL), followed by dropwise addition of benzyl chloride (350 μL , 3.1 mmol) with stirring at 50 °C for 6 h. The pH was adjusted to around 5 by addition of concentrated hydrochloric acid and extracted with CHCl_3 (20 mL). Solids were removed by filtration and washed with CHCl_3 (2×10 mL). The organic extracts were combined and concentrated *in vacuo* to a pale oil, which was triturated under cold acetone (20 mL) and the resulting analytically pure white solid was isolated by filtration and washed with more cold acetone (10 mL). Additional product was isolated from the acetone washings and was combined with the initial crop (0.37 g, 66%).

mp 213 - 215 °C; $[\alpha]_D^{27} + 18.9$ ($c = 0.95$, H_2O); ^1H NMR (400 MHz, D_2O): δ 7.49 – 7.34 (m, 5H, Ph), 5.41 (dt, $J = 51.7, 3.9$ Hz, 1H, CHF), 4.54 – 4.38 (m, 3H, CH and benzyl CH_2), 3.98 – 3.63 (m, 2H, CH_2), 2.78 (m, 1H, CH), 2.25 (dm, $J = 39.3$ Hz, 1H, CHF); ^{13}C NMR (101 MHz, D_2O): δ 170.7, 130.7, 130.3, 129.3, 91.8 (d, $J_{\text{C-F}} = 171.7$ Hz), 66.5, 60.6, 59.8 (d, $J_{\text{C-F}} =$

21.7 Hz), 36.3 (d, $J_{C-F} = 21.7$ Hz); ^{19}F NMR (376 MHz, D_2O): δ -173.53 (m, 1F); IR (neat) ν 3037, 2944, 1644, 1159 cm^{-1} ; LCMS (ESI+) 224.28 $[\text{M}+\text{H}]^+$.

4.2.5 (S)-2-[N-(N'-benzyl-(R)-4-fluoro-L-prolyl)amino]benzophenone (11).

Methanesulfonyl chloride (105 μL , 1.36 mmol) was added slowly to a solution containing *N*-methylimidazole (216 μL , 2.72 mmol) and proline acid **10** (0.28 g, 1.24 mmol) in CH_2Cl_2 , cooled to 0°C . After stirring for 10 min, the flask was removed from the ice bath, and 2-aminobenzophenone (0.22 g, 1.12 mmol) was added and the reaction mixture was stirred at 50°C for 20 h. The reaction was quenched by addition of sat. NH_4Cl aq. (10 mL), the organic phase was extracted CH_2Cl_2 (2×20 mL) and volatiles were removed under reduced pressure. The crude mixture was purified by column chromatography (silica; 1:9 ethyl acetate-hexane) and isolated as a pale brown oil (0.38 g, 76%).

$[\alpha]_D^{24} -14.1$ ($c = 1$, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 11.59 (s, 1H, NH), 8.59 (dd, $J = 8.4, 1.1$ Hz, 1H, Ar), 7.81 – 7.74 (m, 2H, Ar), 7.65 – 7.59 (m, 1H, Ar), 7.58 – 7.47 (m, 4H, Ar), 7.46 – 7.41 (m, 2H, Ar), 7.19 – 7.13 (m, 3H, Ar), 7.10 (ddd, $J = 8.2, 7.3, 1.1$ Hz, 1H, Ar), 5.35 (dm, $J = 52.0$, Hz, 1H, CHF), 4.03 (d, $J = 13.0$ Hz, 1H, benzyl CH), 3.78 – 3.65 (m, 2H, CH and benzyl CH), 3.48 (ddd, $J = 34.8, 13.1, 4.2$ Hz, 1H, CH), 2.90 (ddt, $J = 28.5, 13.0, 1.8$ Hz, 1H, CH), 2.72 – 2.55 (m, 1H, CH), 2.22 – 2.01 (m, 1H, CH); ^{13}C NMR (101 MHz, CDCl_3): δ 198.3, 172.9, 139.2, 138.5, 137.9, 133.6, 132.8, 132.6, 130.1, 129.0, 128.4, 128.3, 127.3, 124.9, 122.4, 121.6, 93.6 (d, $J_{C-F} = 180.1$ Hz), 67.8, 60.6, 59.3 (d, $J_{C-F} = 22.0$ Hz), 38.7 (d, $J_{C-F} = 22.1$ Hz); ^{19}F NMR (376 MHz, CDCl_3): δ -171.49 (m, 1F); IR (neat) ν 3296, 1703, 1521, 1264, 1169, 733 cm^{-1} ; LCMS (ESI+) $m/z = 402.98$ $[\text{M}+\text{H}]^+$; HRMS (ESI) calcd for $\text{C}_{25}\text{H}_{24}\text{FN}_2\text{O}_2$ $[\text{M}+\text{H}]^+$: $m/z = 403.1810$, found 403.1822;

4.2.6 L-Ala-Ni-(S)-fluoro-BPB complex (12).

To a solution of **11** (0.10 g, 0.27 mmol) and $\text{Ni}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$ (0.16 g, 0.55 mmol) in anhydrous methanol (3 mL) was added a solution of KOH (0.12 mg, 2.20 mmol) and L-alanine (0.12 g, 1.37 mmol) in methanol (2 mL). This mixture was heated at 60°C with stirring for 24 h – TLC indicated the reaction was complete and the deep red solution concentrated *in vacuo* before partitioning between water (30 mL) and CHCl_3 (20 mL). Trituration of the concentrated organic extract under Et_2O afforded a dark red solid that was isolated by filtration under suction and washed with more Et_2O . The washings

were combined and evaporated and triturated again to obtain further material, which was analytically pure without further purification (0.10 mg, 71%).

mp 296 – 298 °C; $[\alpha]_D^{24} + 241$ ($c = 1.0$, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 8.28 – 8.22 (m, 2H, Ar), 8.03 (dd, $J = 8.7, 1.1$ Hz, 1H, Ar), 7.55 – 7.47 (m, 2H, Ar), 7.47 – 7.41 (m, 1H, Ar), 7.38 (t, $J = 7.7$ Hz, 2H, Ar), 7.30 – 7.17 (m, 2H, Ar), 7.12 (ddd, $J = 8.7, 6.8, 1.9$ Hz, 1H, Ar), 6.96 – 6.90 (m, 1H, Ar), 6.70 – 6.59 (m, 2H, Ar), 5.68 (dm, $J = 55.1$ Hz, 1H, CHF), 4.42 (d, $J = 12.4$ Hz, 1H, benzyl CH), 4.25 (ddd, $J = 32.8, 14.4, 4.3$ Hz, 1H, CH), 3.95 – 3.81 (m, 2H, CH), 3.65 (d, $J = 12.4$ Hz, 1H, benzyl CH), 3.32 (dddd, $J = 35.7, 15.1, 10.9, 4.5$ Hz, 1H, CH), 2.94 (m, 2H, CH), 1.54 (d, $J = 7.0$ Hz, 3H, CH_3); ^{13}C NMR (101 MHz, CDCl_3): δ 180.4, 178.9, 170.8, 142.0, 133.5, 133.4, 133.3, 132.4, 131.8, 129.9, 129.4, 129.1, 129.0, 127.6, 127.3, 126.4, 123.8, 121.1, 91.4 (d, $J_{\text{C-F}} = 178.6$ Hz), 70.1, 66.5, 65.5, 64.0 (d, $J_{\text{C-F}} = 21.7$ Hz), 39.4 (d, $J_{\text{C-F}} = 21.6$ Hz), 21.8; ^{19}F NMR (376 MHz, CDCl_3) δ -173.86 (m, 1F); LCMS (ESI+) $m/z = 530.06$ $[\text{M}+\text{H}]^+$; HRMS (ESI) calcd for $\text{C}_{28}\text{H}_{27}\text{FN}_3\text{O}_3\text{Ni}$ $[\text{M}+\text{H}]^+$: $m/z = 530.1385$, found 530.1390.

X-ray crystal structure determination and structural analysis

The X-ray single crystal data have been collected on a Bruker SMART CCD 6000 (compound **8**, sealed fine-focus tube, graphite monochromator, $\lambda\text{MoK}\alpha$, $\lambda = 0.71073$ Å) and D8Venture (compounds **1** and **12**, μS -microsource, focusing mirrors, Photon 100 CMOS detector, $\lambda\text{CuK}\alpha$, $\lambda = 1.54178$ Å and $\lambda\text{MoK}\alpha$, $\lambda = 0.71073$ Å, respectively,) diffractometers equipped with the Cryostream (Oxford Cryosystems) open-flow nitrogen cryostats at the temperature 120.0(2) K. All structures were solved by direct method and refined by full-matrix least squares on F2 for all data using Olex2 [15] and SHELXTL [26] software. All non-disordered non-hydrogen atoms were refined anisotropically, hydrogen atoms in structure **8** were refined isotropically. The atoms of disordered groups in structure **1** were refined with fixed SOF values in isotropic approximation. The hydrogen atoms in these groups and in the structures **1** and **12** were placed in the calculated positions and refined in riding mode.

Crystal data for **1**: $\text{C}_{28}\text{H}_{27}\text{N}_3\text{NiO}_3 \cdot \text{C}_4\text{H}_8\text{O}$ ($M = 584.34$), orthorhombic, space group $\text{P2}_1\text{2}_1\text{2}_1$, $a = 9.0963(2)$, $b = 12.1090(3)$, $c = 26.0025(6)$ Å, $V = 2864.10(12)$ Å³, $Z = 4$, $\mu(\text{CuK}\alpha) = 1.312$ mm⁻¹, $D_{\text{calc}} = 1.355$ g/mm³, 21312 reflections measured, 5425 unique ($R_{\text{int}} = 0.0260$) were used

in all calculations. The final R_1 was 0.0470 (5218 $>2\sigma(I)$) and wR_2 was 0.1297 (all data), $S = 1.051$.

Crystal data for **8**: $C_{28}H_{26}N_3O_3FNi$ ($M = 530.23$): orthorhombic, space group $P2_12_12_1$, $a = 9.4749(5)$, $b = 11.3449(7)$, $c = 44.685(3)$ Å, $V = 4803.2(5)$ Å³, $Z = 8$, $\mu(\text{MoK}\alpha) = 0.852$ mm⁻¹, $D_{\text{calc}} = 1.466$ g/mm³, 78710 reflections measured, 12765 unique ($R_{\text{int}} = 0.0738$) were used in all calculations. The final R_1 was 0.0364 (10213 $>2\sigma(I)$) and wR_2 was 0.0766 (all data), $S = 0.978$.

Crystal data for **12**: $C_{28}H_{26}N_3O_3FNi$ ($M = 530.23$): orthorhombic, space group $P2_12_12_1$, $a = 9.6711(8)$, $b = 10.0617(8)$, $c = 24.6671(19)$ Å, $V = 2400.3(3)$ Å³, $Z = 4$, $\mu(\text{MoK}\alpha) = 0.853$ mm⁻¹, $D_{\text{calc}} = 1.467$ g/cm³, 33745 reflections measured, 6370 unique ($R_{\text{int}} = 0.0834$) which were used in all calculations. The final R_1 was 0.0584 (5748 $>2\sigma(I)$) and wR_2 was 0.1369 (all data), $S = 1.077$.

CCDC [xxx,xxx](#) and [xxx](#) contain the supplementary crystallographic data for this paper. These data can be viewed free of charge via <http://www.ccdc.cam.ac.uk/cont/retrieving.html> or from the CCDC, 12 Union Road, Cambridge, CB2 1EZ; fax: +44 1223 336033. E-mail: deposit@ccdc.cam.ac.uk

References

1. For reviews see:

- a) J. Wang, M. Sanchez-Rosello, J.L. Aceña, C. del Pozo, A.E. Sorochinsky, S. Fustero, V.A. Soloshonok, H. Liu. Fluorine in pharmaceutical industry: Fluorine-containing drugs introduced to the market in the last decade (2001–2011), *Chem. Rev.* 114, (2013) 2432-2506.
 - b) D. O'Hagan, Fluorine in Health Care: Organofluorine Containing Blockbuster Drugs, *J. Fluor. Chem.* 131 (2010) 1071–1081.
 - c) S. Purser, P.R. Moore, S. Swallow, V. Gouverneur, Fluorine in Medicinal Chemistry, *Chem. Soc. Rev.*, 37 (2008) 320–330.
 - d) I. Ojima, Exploration of Fluorine Chemistry at the Multidisciplinary Interface of Chemistry and Biology, *J. Org. Chem.* 78 (2013) 6358–6383.
 - e) R. Filler, R. Saha, Fluorine in Medicinal Chemistry: A Century of Progress and a 60-Year Retrospective of Selected Highlights, *Future Med. Chem.* 1 (2009) 777-791.
2. D.Y. Buissonneaud, T. van Mourik, D. O'Hagan, A DFT study on the origin of the fluorine gauche effect in substituted fluoroethanes, *Tetrahedron* 66 (2010) 2196–2202.
 3. D. Cahard, V. Bizet, The influence of fluorine in asymmetric catalysis, *Chem. Soc. Rev.* 43 (2014) 135-147.
 4. C. Sparr, E. Salamanova, W.B. Schweizer, H.M. Senn, R. Gilmour, Theoretical and X-ray Crystallographic Evidence of a Fluorine-Imine Gauche Effect: An Addendum to Dunathan's Stereoelectronic Hypothesis, *Chem. Eur. J.* 17 (2011) 8850 – 8857.
 5. P.W. Chia, M.R. Livesey, A.M.Z. Slawin, T. van Mourik, D.J.A. Wyllie, D. O'Hagan, 3-Fluoro-N-methyl-d-aspartic acid (3F-NMDA) Stereoisomers as Conformational Probes for Exploring Agonist Binding at NMDA Receptors, *Chem. Eur. J.* 18 (2012) 8813 – 8819.
 - 6 J.A. Hodges, R.T. Raines, Stereoelectronic and Steric Effects in the Collagen Triple Helix: Toward a Code for Strand Association, *J. Am. Chem. Soc.* 127 (2005) 15923–15932.

7. For examples see:

- a) C. Sparr, W.B. Schweizer, H.M. Senn, R. Gilmour, The Fluorine-Iminium Ion Gauche Effect: Proof of Principle and Application to Asymmetric Organocatalysis, *Angew. Chem. Int. Ed.* 48 (2009) 3065 – 3068.
- b) C. Sparr, E.-M. Tanzer, J. Bachmann R. Gilmour, A Synthesis of (S)-2-(Fluorodiphenylmethyl)-Pyrrolidine: A Novel Organocatalyst for the Stereoselective Epoxidation of α,β -Unsaturated Aldehydes, *Synthesis*, 8 (2010) 1394 - 1397.
- c) C. Sparr, R. Gilmour, Fluoro-Organocatalysts: Conformer Equivalents as a Tool for Mechanistic Studies, *Angew. Chem.* 122 (2010) 6670 –6673.
- d) C. Sparr, L.E. Zimmer, R. Gilmour. Exploiting Fluorine Conformational Effects in Organocatalyst Design: The Fluorine-Iminium Ion Gauche Effect. (2011) in *Asymmetric Syntheses. More Methods and Applications* (Eds. S. Bräse and M. Christmann), Wiley-VCH Verlag GmbH & Co, Weinheim, 117-124.
- e) E.-M. Tanzer, R. Gilmour, Fluorinated Cinchona Alkaloids in Phase Transfer Catalysis: Controlling Internal Rotation by a Fluorine-Ammonium Ion Gauche Effect ($\text{\textcircled{O}}\text{NCCF}$), *Chem. Eur. J.* 18 (2012) 2006.
- f) E.-M. Tanzer, L.E. Zimmer, W.B. Schweizer R. Gilmour, Fluorinated Organocatalysts for the Enantioselective Epoxidation of Enals: Molecular Pre-organisation by the Fluorine-Iminium Ion Gauche Effect, *Chem. Eur. J.* 18 (2012) 11334.
- g) Y.P. Rey, R. Gilmour, R. Modulating NHC catalysis with fluorine, *Beilstein J. Org. Chem.* 9 (2013) 2812-2820.
- h) I.G. Molnar, E.-M. Tanzer, C. Daniliuc, R. Gilmour, Enantioselective Aziridination of Cyclic Enals Facilitated by the Fluorine-Iminium Ion Gauche Effect, *Chem. Eur. J.* 20 (2014) 794-800.
- i) Y.P. Rey, L.E. Zimmer, C. Sparr, E.-M. Tanzer, W.B. Schweizer, H.M. Senn, S. Lakhdar R. Gilmour, Molecular Design Exploiting a Fluorine Gauche Effect as a Stereoelectronic Trigger, *Eur. J. Org. Chem.* (2014) 1202-1211.

8. For examples see:

- a) A.E. Sorochinsky, J.L. Aceña, H. Moriwaki, T. Sato, V.A. Soloshonok, Asymmetric synthesis of alpha-amino acids via homologation of Ni(II) complexes of glycine Schiff bases; Part 1: Alkyl halide alkylations, *Amino Acids* 45 (2013) 691-718
- b) A.E. Sorochinsky, J.L. Aceña, H. Moriwaki, T. Sato, V.A. Soloshonok, Asymmetric synthesis of alpha-amino acids via homologation of Ni(II) complexes of glycine Schiff bases. Part 2: Aldol, Mannich addition reactions, deracemization and (S) to (R) interconversion of alpha-amino acids, *Amino Acids* 45 (2013) 1017-1033
- c) J.L. Aceña, A.E. Sorochinsky, V. Soloshonok, Asymmetric synthesis of α -amino acids via homologation of Ni(II) complexes of glycine Schiff bases. Part 3: Michael addition reactions and miscellaneous transformations, *Amino Acids* 46 (2014) 2047-2073.
- d) J.L. Aceña, A.E. Sorochinsky, H. Moriwaki, T. Sato, V.A. Soloshonok, Synthesis of fluorine-containing α -amino acids in enantiomerically pure form via homologation of Ni (II) complexes of glycine and alanine Schiff bases, *J. Fluor. Chem.* 155 (2013) 21-38.
- e) Y.N. Belokon, Y.E. Zeltzer, V.I. Bakhmutov, M.B. Saporovskaya, M.G. Ryzhov, A.I. Yanovsky, Y.T. Struchkov, V.M. Belikov, Asymmetric Synthesis of Threonine and Partial Resolution and Retroracemization of α -Amino Acids via Copper(II) Complexes of Their Schiff Bases with (S)-2-N-N(N'-Benzylpropyl)aminobenzaldehyde and (S)-2-N-(N'-benzylpropyl)aminoacetophenone. Crystal and Molecular Structure of a Copper(II) Complex of Glycine Schiff Base with (S)-2-N-N(N'-Benzylpropyl)aminoacetophenone, *J. Am. Chem. Soc.* 105 (1983) 2010-2017.
- f) Y.N. Belokon, A.G. Bulychev, S.V. Vitt, Y.T. Struchkov, A.S. Batsanov, T.V. Timofeeva, V.A. Tsyryapkin, M.C. Ryzhov, L.A. Lysova, V.I. Bakhmutov, V.M. Belikov, General Method of Diastereo- and Enantioselective Synthesis of β -Hydroxy- α -amino Acids by Condensation of Aldehydes and Ketones with Glycine, *J. Am. Chem. Soc.* 107 (1985) 4252-4259.
9. H. Moriwaki, D. Resch, H. Li, I. Ojima, R. Takeda, J.L. Aceña, V.A. Soloshonok, Synthesis and stereochemical assignments of diastereomeric Ni (II) complexes of glycine Schiff base with (R)-2-(N-{2-[N-alkyl-N-(1-phenylethyl) amino] acetyl} amino) benzophenone; a case of configurationally stable stereogenic nitrogen, *Beilstein J. Org. Chem.* 10 (2014) 442-448.

10. M. Bergagnini, K. Fukushi, J. Han, N. Shibata, C. Roussel, T.K. Ellis, J.L. Aceña, V.A. Soloshonok, NH-type of chiral Ni (ii) complexes of glycine Schiff base: design, structural evaluation, reactivity and synthetic applications, *Org. Biomol. Chem.* **12** (2014) 1278-1291.
11. N.H. Campbell, D.L. Smith, A.P. Reszka, S. Neidle, D. O'Hagan, Fluorine in medicinal chemistry: β -fluorination of peripheral pyrrolidines attached to acridine ligands affects their interactions with G-quadruplex DNA, *Org. Biomol. Chem.* **9** (2011) 1328–1331.
12. H. Ueki, T.K. Ellis, C.H. Martin, T.U. Boettiger, S.B. Bolene, V.A. Soloshonok, Improved synthesis of proline-derived Ni (II) complexes of glycine: versatile chiral equivalents of nucleophilic glycine for general asymmetric synthesis of α -amino acids, *J. Org. Chem.* **68** (2003) 7104-7107.
13. Y.N. Belokon, V.I. Tararov, V.I. Maleev, T.F. Savel'eva, M.G. Ryzhov, Improved procedures for the synthesis of (S)-2-[N-(N'-benzylpropyl)amino]benzophenone (BPB) and Ni(II) complexes of Schiff's bases derived from BPB and amino acids, *Tetrahedron: Asymmetry* **9** (1998) 23 4249-4252.
14. **For examples see:**
 - a) Y.W. Kim, T.N. Grossmann, G.L. Verdine, Synthesis of all-hydrocarbon stapled α -helical peptides by ring-closing olefin metathesis, *Nat. Protoc.* **6** (2011) 761-771.
 - b) G.H. Bird, W.C. Crannell, L.D. Walensky, Chemical Synthesis of Hydrocarbon - Stapled Peptides for Protein Interaction Research and Therapeutic Targeting, *Curr. Protoc. Chem. Biol.* (2011) 99-117.
 - c) W. Qiu, V.A. Soloshonok, C. Cai, X. Tang, V.J. Hruby, Convenient, Large-Scale Asymmetric Synthesis of Enantiomerically Pure trans-Cinnamylglycine and- α -Alanine, *Tetrahedron* **56** (2000) 2577-2582.
 - d) Y.N. Belokon, N.I. Chernoglazova, C.A. Kochetkov, N.S. Garbalinskaya V.M. Belikov, Preparation of optically pure α -methyl- α -amino acids via alkylation of the nickel(II) Schiff base of (R,S)-alanine with (S)-2-N-(N-benzylpropyl)aminobenzaldehyde, *J. Chem. Soc., Chem. Commun.* **3** (1985) 171-172.

- 15 O. V. Dolomanov, L. J. Bourhis, R. J. Gildea, J. A. K. Howard and H. Puschmann, OLEX2: a complete structure solution, refinement and analysis program *J. Appl. Cryst.* **42** (2009) 339-341.
16. Hermes - Accessed on web (08/04/2014) at <http://www.ccdc.cam.ac.uk/Solutions/GoldSuite/pages/GoldSuite.aspx>
17. Q. Jing, H. Li, L.J. Roman, P. Martásek, T.L. Poulos, R.B. Silverman, Accessible Chiral Linker to Enhance Potency and Selectivity of Neuronal Nitric Oxide Synthase Inhibitors, *ACS Med. Chem. Lett.* **5** (2014) 56–60.
18. W. Zhuang, X. Zhao, G. Zhao, L. Guo, Y. Lian, J. Zhou, D. Fang, Synthesis and biological evaluation of 4-fluoroproline and 4-fluoropyrrolidine-2-acetic acid derivatives as new GABA uptake inhibitors, *Bioorg. Med. Chem.* **17** (2009) 6540–6546.
19. B. R. Kim, H. G. Lee, S. B. Kang, G. H. Sung, J. J. Kim, J. K. Park & Y. J. Yoon, tert-Butoxide-Assisted Amidation of Esters under Green Conditions. *Synthesis*, **44** (2012) 42-50.
- 20 Y.Q. Tang, J.M. Lu, X.R. Wang, & L.X. Shao, Palladium (II)–N-heterocyclic carbene complexes derived from proline: synthesis and characterization, *Tetrahedron*, **66** (2010) 7970-7974.
21. H. Ueki, T.K. Ellis, C.H. Martin, T.U. Boettiger, S.B. Bolene, & V.A.J. Soloshonok, Improved synthesis of proline-derived Ni (II) complexes of glycine: versatile chiral equivalents of nucleophilic glycine for general asymmetric synthesis of α -amino acids. *J. Org. Chem.* **68** (2003) 7104-7107.
22. J.A.K. Howard, V.J. Hoy, D. O'Hagan, G.T. Smith, How good is fluorine as a hydrogen bond acceptor? *Tetrahedron* **52** (1996) 12613.
23. V.R. Thalladi, H.-C. Weiss, D. Bläser, R. Boese, A. Nangia, G.R. Desiraju, C-H \cdots F Interactions in the Crystal Structures of Some Fluorobenzenes, *J. Am. Chem. Soc.* **120** (1998) 8702-8710.
24. H.-C. Weiss, R. Boese, H.L. Smith, M.M. Haley, CH \cdots Pi versus CH \cdots Halogen Interactions: The Crystal Structures of The 4-Halogenoethynylbenzenes, *Chem. Commun.* **24** (1997) 2403-2404.

25. A. Cochi, D. Gomez Pardo, J. Cossy, Access to Optically Active 3-Azido-and 3-Aminopiperidine Derivatives by Enantioselective Ring Expansion of Prolinols. *Org. Lett.* 13 (2011) 4442-4445.
26. G.M. Sheldrick, A short history of SHELX. *Acta Cryst.* (2008), A64, 112-122