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Title: Epigenetic control of atherosclerosis via DNA methylation: a new therapeutic target?

Running title: DNA methylation in atherosclerosis

Armita Mahdavi Gorabi,¹ Peter E. Penson,² Maciej Banach,^{3,4} Morteza Motallebnezhad,^{5,6,7}

Tannaz Jamialahmadi,^{8,9} Amirhossein Sahebkar^{10,11,12*}

¹Research Center for Advanced Technologies in Cardiovascular Medicine, Tehran Heart Center, Tehran University of Medical Sciences, Tehran, Iran

²School of Pharmacy and Biomolecular Sciences, Liverpool John Moores University, Liverpool, UK

³Department of Hypertension, WAM University Hospital in Lodz, Medical University of Lodz, Zeromskiego 113, Lodz, Poland

⁴Polish Mother's Memorial Hospital Research Institute (PMMHRI), Lodz, Poland ⁵Immunology Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

⁶Student Research Committee, Tabriz University of Medical Sciences, Tabriz, Iran

⁷Department of Immunology, Faculty of Medicine, Iran University of Medical Sciences, Tehran, Iran

⁸Halal Research Center of IRI, FDA, Tehran, Iran

⁹Department of Nutrition, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran

¹⁰Biotechnology Research Center, Pharmaceutical Technology Institute, Mashhad University of Medical Sciences, Mashhad, Iran

¹¹Neurogenic Inflammation Research Center, Mashhad University of Medical Sciences, Mashhad, Iran

¹²School of Pharmacy, Mashhad University of Medical Sciences, Mashhad, Iran

*Correspondences: Amirhossein Sahebkar, Biotechnology Research Center, Mashhad University

of Medical Sciences, Mashhad 9177948564, Iran. Tel: +985138002299; Fax: +985138002287; E-

mail: sahebkara@mums.ac.ir; amir_saheb2000@yahoo.com

Abstract

Atherosclerosis is a disease in which lipid-laden plaques are developed inside the vessel walls of arteries. The immune system is activated, resulting in inflammation and oxidative stress. Endothelial cells (ECs) are activated, arterial smooth muscle cells (SMCs) proliferate, macrophages are activated, and foam cells are developed, leading to dysfunctional ECs. Epigenetic regulatory mechanisms, including DNA methylation, histone modifications, and microRNAs are involved in the modulation of genes that play distinct roles in several aspects of cell biology and physiology, hence linking environmental stimuli to gene regulation. Recent research has investigated the involvement of DNA methylation in the etiopathogenesis of atherosclerosis, and several studies have documented the role of this mechanism in various aspects of the disease. Regulation of DNA methylation plays a critical role in the integrity of ECs, SMC proliferation and formation of atherosclerotic lesions. In this review, we seek to clarify the role of DNA methylation in the development of atherosclerosis through different mechanisms.

Keywords: Atherosclerosis; Epigenetic regulation; DNA methylation; Oxidative stress; Inflammation

1. Introduction

Atherosclerosis is considered to be the primary cause of numerous cardiovascular diseases (Hansson et al. , 2006). These disorders have been the predominant cause of mortality in the past decade (Singh and Bedi, 2013). During the development of atherosclerosis, slow and dynamic modifications in the cellular and molecular composition of vessel walls occur, resulting in atherosclerotic plaques (Sipahi and Tuzcu, 2008). Atherosclerosis is a complex disorder that is associated with the accumulation of lipids in the vessel walls, stimulation of the immune system, development of inflammatory responses (with the release of mediators, such as tumor necrosis factor (TNF)- α and interleukin (IL)-1), oxidative stress, development of oxidized low-density lipoproteins (ox-LDLs), activation of ECs, proliferation of arterial SMCs, stimulation of macrophage and promotion of foam cells, and finally endothelial dysfunction (Bennett et al. , 2015, Libby and Ridker, 2006, Sanjadi et al. , 2018, Študentová et al. , 2016, van Diepen et al. , 2013).

Among the epigenetic regulatory mechanisms, DNA methylation is of critical importance (Zhang et al., 2014b). Research investigating abnormalities of DNA methylation in atherosclerotic patients has identified a particular profile of DNA methylation and proposed various pathways and genes in the etiopathogenic mechanism of the disease (Table 1) (Zaina and Lund, 2014). In this article, we summarise the recent findings concerning abnormalities of DNA methylation, and their roles in the pathogenesis and progression of atherosclerosis.

2. Mechanobiology of atherosclerosis

Atherosclerosis is a complex disorder, and several contributing factors have been implicated with its development (Hopkins and Williams, 1981). The strongest risk factors are smoking,

hyperlipidemia, male gender, diabetes, obesity, sedentary lifestyle, and ageing. Hyperlipidemia has been reported to be the critical risk factor, and low lipid levels are associated with a decreased risk for the disease regardless of other risk factors (Poulter, 1999). It is important to note that coronary artery disease (CAD) may also develop in individuals lacking the known risk factors (Wilkins et al., 2012). Genetic variations have partially explained the cumulative risk factors for CAD, and hyperlipidemia is partially explained by genetics (International Consortium for Blood Pressure Genome-Wide Association et al., 2011). Environmental contributing risk factors may be epigenetically involved in the development of atherosclerosis.

According to the 'response to injury' theory, atherosclerosis initially develops at regions with injuries to the endothelium, known as endothelial dysfunction which is characterized by decreased levels of nitric oxide (NO) in the vessel wall and enhanced generation of angiotensin II (ANGII), thromboxane, and endothelin 1 (ET1) (Ross and Glomset, 1976). Reduced generation of NO results in apoptosis of EC (Csiszar et al., 2004) and increased ANGII (Endtmann et al., 2011). In arteries, lipid retention is characterized by slow thickening of the intimal layer resulting from the accumulation of modified LDL in the extracellular region of the sub-intimal layer (Skålén et al., 2002, Ylä-Herttuala et al., 1989). Monocytes are recruited to the affected region and become macrophages, which engulf the excessive LDL and develop to foam cells, resulting in the generation of lesions (Steinberg, 2002). Upon local injury, the SMC of vessel wall lose their function and begin to proliferate, leading to obstruction of the arterial lumen. Modified SMCs secrete mediators and thereby trigger growth of the lesion. Infiltration of immune cells leads to a local inflammatory condition. Atherosclerosis is usually regarded as an inflammatory condition, and enhanced inflammatory cytokines in the blood may be useful in the prediction of complications, such as plaque rupture, fibrosis, thrombosis, and calcification of vessel wall (Glass

and Witztum, 2001). These events are accompanied by elevated levels of extracellular mediators, such as vascular cell adhesion molecule 1 (VCAM1), intercellular adhesion molecule 1 (ICAM1), Monocyte chemoattractant protein 1 (MCP1), vascular endothelial growth factor (VEGF), and IL-8 (Findley et al. , 2008, Hoogeveen et al. , 2005). Among the critical triggering factors of inflammatory responses are monocytes recruited to the vessel wall (Hansson, 2005), modified LDL (Wiesner et al. , 2010), hypomethylated self-DNA (Zhang et al. , 2014a), and Toll-like receptors (TLRs) recognizing self RNA (Fabbri et al. , 2013). Inflammation seems to be present throughout the progression of atherosclerosis, and modifications to the methylation profile of DNA is thought to regulate the continuous state of inflammation.

3. Mechanisms of DNA methylation

The mechanisms underlying the regulation of gene expression through DNA methylation have previously been described (**Figure 1**) (Jones and Takai, 2001, Robertson, 2005). DNA methylation is considered as an epigenetic mechanism in the mammalian cells that is mediated by DNA methyltransferase (DNMT) enzymes, and S-adenosyl-methionine (SAM) acts as the donor of methyl groups. By transfer of the methyl group (CH3) onto the C5 position of cytosines, the 5-methylcytosine (5mC) is generated (Quintero-Ronderos and Montoya-Ortiz, 2012). Currently, five members of DNMT enzymes have been recognized that are categorized into two major groups: maintenance DNMTs (DNMT1, DNMT2) and de novo DNMTs (DNMT3a, DNMT3b, and DNMT3L). While the maintenance DNMTs are involved in the methylation of the cytosine in the hemimethylated DNA during DNA replication, the de novo DNMT enzymes play a specific role in the methylation across the embryonic development (Huang and Fan, 2010). These enzymes have been attributed with reciprocal activities, in which they are involved in adding and removal of

methyl groups. The methylated DNA confers a suppressed transcription state that is mediated through facilitated binding of methyl-CpG-binding domain (MBD) proteins and reduced binding of transcription factors to the methylated DNA sites. In addition, DNA methylation impresses the chromatin structure and causes the generation of co-repressor complexes. On the other hand, the unmethylated DNA confers a euchromatin structure that facilitates the binding of transcription factors to the target sites, leading to gene transcription (Fan and Zhang, 2009).

DNA hypermethylation has been reported to occur as part of the development of numerous human disorders. In the absence of disease, DNA hypomethylation is observed in the CpG islands located in the promoter region of genes, while hypermethylation is seen in the CpG islands found within the non-promoter region of genes. Global hypomethylation of DNA, in which there is a decreased methylation level of DNA within the non-promoter regions, perhaps leads to a structural alteration and chromosome instability, resulting in transcriptional activity in undesirable sites and in normally silent (inactive) regions. That notwithstanding, global hypermethylation of DNA may be accompanied by downregulation of genes which suppress or protect against the development of diseases. As an example, decreased transcription of transposable elements, such as short interspersed nucleotide element (ALU) and long interspersed nucleotide elements-1 (LINE-1) located within the non-promoter sites, has been associated with the regulation of genome integrity by means of enhanced methylation status at their sequences (Zhang and Zeng, 2016). In malignant diseases, a severe hypomethylation is seen in the transposable elements, conferring DNA recombination, mutations, and chromosomal instability, thereby contributing to the development of tumors (Roman-Gomez et al., 2005). DNA hypermethylation is regarded as a critical epigenetic signature seen in the promoter site of tumor suppressor genes in various malignancies (Sharma et al., 2010). Considering the shared risk factors between malignancies and atherosclerotic

cardiovascular disease (CVD) (Koene et al. , 2016), it has been suggested that there might be a dysregulation in the methylome and, therefore transcription, of the cardiovascular-related genes (Table 1).

4. Genome-wide DNA methylation and atherogenesis

Global DNA hypermethylation of cytosines in the CpGs has been identified in both human subjects and animal studies that was attributed to the clinical aspects of atherosclerosis (Rangel-Salazar et al. , 2011, Yoo et al. , 2012). Using genome-wide DNA methylation sequencing, a positive correlation was found between DNA methylation level and the grade of the atherosclerotic lesion in the atherosclerotic human aortas (Morales et al. , 2015). Moreover, using the methylated DNA immunoprecipitation sequencing (meDIP-seq), the differentially methylated regions were observed in the cardiovascular disease-associated genes in the ECs obtained from porcine aortas (Jiang et al. , 2015). Such observations imply that the DNA methylation profiling can divulge the biomarkers of atherosclerosis, proposing a plausible role of DNA methylation in the progression of the disease.

5. DNA methylation abnormalities in atherosclerosis

5.1. Oxidative stress and DNA methylation in atherosclerosis

Oxidative stress is controlled in the body by maintaining a balance between the daily production of reactive oxygen species (ROS) and the systems which remove antioxidants. Under normal physiological conditions, a balance exists between ROS generation and enzymatic and nonenzymatic antioxidant factors, which are involved in reducing or scavenging the ROS (Liu and Timmons, 2016). Dysfunction of the mechanisms which remove antioxidants, or increased generation of ROS can result in a redox imbalance (Kim et al., 2011). It has been demonstrated that prolonged oxidative stress can lead to ageing and a range of disorders, including cancers, inflammation, cardiovascular disorders, and infectious diseases (Berlett and Stadtman, 1997, Finkel and Holbrook, 2000, Kobayashi et al., 2002). Furthermore, investigations have shown that oxidative stress during the development of atherosclerosis can modify the methylation status of DNA (Lim et al., 2008, Nanduri and Prabhakar, 2015, Niu et al., 2015). These observations followed the findings in tumor cells, that oxidative stress is associates with substantial alterations in methylation (Afanas'ev, 2015). Early investigations indicated an association between 8hydroxyguanine (8-OHdG) (a marker of ROS) and inverse alterations of DNA methylation (Weitzman et al., 1994). In addition, it was reported that oxidative damage of guanines through 8-OHdG in the parental DNA strand would allow normal copying of methylation profile through pathways involved in the maintenance of DNA methylation; however oxidative damage of guanines via 8-OHdG in the newly copied DNA strand could suppress the methylation of DNA (Turk et al., 1995). It has been reported that ROS, especially hydrogen peroxide (H_2O_2), can alter the methylation profile of DNA. H₂O₂ was shown to be able to change the DNA methylation through the facilitation of DNA methyltransferase1 (DNMT1) binding to chromatin (O'Hagan et al., 2011). During atherosclerosis, ROS production can alter makers of DNA methylation. Furthermore, increased methylation of the superoxide dismutase 2 (SOD2) gene, leads to its suppression and has been reported to result in SMC proliferation. Treatment with the DNMT inhibitor 5-azacytidine (5-azaC) caused upregulation of SOD2 expression and decreased SMC proliferation. The hypermethylation of SOD2, which causes decreased expression of SOD2, can stimulate the Hypoxia-inducible factor 1-alpha (HIF-1 α); hence the methylation status of this gene is important for the development of atherosclerotic lesions. (Baccarelli and Bollati, 2009). Little is

known about the involvement of oxidative stress in the alteration of DNA methylation during atherosclerotic lesion development. That notwithstanding, due to an evident impact of oxidative stress on the modulation of DNA methylation, and considering the fact that atherosclerosis is associated with chronic oxidative stress, it has been suggested that global modifications of DNA methylation may occur during atherosclerosis.

5.2. DNA methylation during aging and atherosclerosis

Investigations have revealed that the severity of clinical presentations due to atherosclerosis is associated with ageing (Denke and Grundy, 1990, Tanaka et al., 1988). In addition, it has been shown that the expression level of several genes undergoes remarkable alterations as we age, although the mechanisms underlying these changes are not completely understood. However, it is evident that the ageing process is accompanied by an alteration in the extent of DNA methylation (Holliday, 1987). Moreover, in vitro experiments have demonstrated that the continuous passaging of normal fibroblast leads to hypomethylation of DNA (Holliday, 1985, Wilson and Jones, 1983). Additionally, aged tissues have been observed to have a similar DNA hypomethylation pattern (Drinkwater et al., 1989, Mays-Hoopes et al., 1986, Wilson et al., 1987). Among the important genes that have been indicated to undergo a hypomethylation during aging are Estrogen receptor alpha (ERα), BMP/retinoic acid-inducible neural-specific 1 (BRINP1), E-cadherin, insulin-like growth factor-2 (IGF-2), P15, c-Fos, PAX6, c-Myc, versican, myogenic differentiation 1 (MYOD1), HIC ZBTB transcriptional repressor 1 (HIC1) (Issa, 2000). Several genes that had previously been speculated to be methylated solely during tumor developments have recently been observed to be among the age-related methylation alteration genes. Clearly, aging is the primary cause of hypermethylation during malignancies (Issa, 2000). Furthermore, it has been reported that

the methylation of the gene coding for ER α in heart muscle occurs as a result of ageing (Post et al. , 1999). In vitro evaluation of the methylation status of SMCs demonstrated that there was a remarkable divergence in the extent of methylation of the gene coding for ER α in tissues obtained from an infant (19%) and an adult (99%) cadaver (Ying et al. , 2000). Hence, aging-associated alteration in the DNA methylation is not exclusively seen in the tumors, but might be critically involved in other age-related disorders including atherosclerosis. In spite of aging-associated global hypomethylation, gene-specific hypermethylation has also been observed that may result in an increased rate of mutations and DNA instability (Liu et al. , 2003). Thus, it seems that aging is accompanied by alteration in the DNA methylation of several genes that might also be involved in the cardiovascular system, and therefore, etiopathogenesis of atherosclerosis.

5.3. DNA methylation and inflammation in atherosclerosis

Atherosclerosis is an inflammatory condition, in which there is a systemic increased level of cytokines and recruitment of circulating leukocytes, especially monocytes to the endothelium. Monocytes reside in the sub-endothelial layer, and development into macrophages, and alter, foam cells.

A chronic inflammatory condition has been reported to stem partially from modifications in methylation status (**Figure 2**). It has been shown that inflammation might be connected to a hypermethylated status of DNA and that DNA hypermethylation was related to increased mortality in patients with atherosclerosis-related disorders (Stenvinkel et al. , 2007). A global DNA methylation investigation demonstrated a strong association between an altered DNA methylation profile, and inflammation (Karimi et al. , 2006). In addition, altered methylation and expression of cytochrome C oxidase subunit II (Cox-II) has been associated with inflammation in cardiovascular

disease. It has been reported that Cox-II is linked with the progression of atherosclerosis, and its transcription may be triggered through pro-inflammatory mediators, including TNF-α. In addition, a negative correlation was identified between Cox-II protein and mRNA expressions and DNA methylation status. The authors suggested a link between the epigenome and the regulation of the expression of Cox-II (Ianni et al. , 2013). It has been established that downregulation of cyclooxygenase-2 (COX-2) expression in subjects at high-risk and treated with aspirin may confer a protective role against the development of atherothrombosis (Cipollone and Fazia, 2006). However, when the beneficial contribution of COX-2 inhibitors is assessed, it is necessary to consider the multifaceted aspects of the prostanoid biology as well as the important role of the COX-2-derived prostaglandin I2 (PGI2) in the regulation of systemic hemodynamics that may lead to inadequate circulatory volume.

It has been reported that an inflammatory state during atherosclerosis may alter the DNA methylation of NF- κ B coding gene and, therefore, result in altered signaling and the production of further inflammatory mediators that may contribute to the development of atherosclerosis (Ding et al., 2015).

Comparison of genome-wide DNA methylation of 440,292 CpG sites between human monocytes, naïve macrophages, activated macrophages with a pro-inflammatory phenotype or an antiinflammatory state, and monocyte-derived foam cells indicated differences in methylation level between these cells. Moreover, DNA methylation highly different during monocyte-tomacrophage differentiation, that was limited to single CpGs or very short regions, and co-localized with lineage-specific enhancers (Dekkers et al., 2019). These data show that localized modulation of DNA methylation at regulatory regions plays a role in cell differentiation, hence implying the involvement of DNA methylation in pathologic cell differentiation during cardiovascular disorders. Additionally, DNA methylation status of M1/M2 macrophage polarization markers was evaluated in CAD patients, resulting in the identification of differently methylated *STAT1*, *STAT6*, *MHC2*, *IL12b*, *iNOS*, *JAK1*, *JAK2* and *SOCS5* genes (Bakshi et al. , 2019).

5.4. DNA methylation and modulation of SMCs during atherosclerosis

During the development of atherosclerosis, inflammatory mediators cause stimulation of SMCs, and their proliferation leads to plaque development. Moreover, activated SMCs generate numerous extracellular matrix components and produce a fibrous cap on the lesion (Steucke et al., 2015). The high rate of oxidized phospholipids may result in an expansion of vascular SMC (VSMC) (Berliner and Watson, 2005, Krychtiuk et al., 2015, Libby et al., 2011, Sazonova et al., 2015). Given that the VSMCs have critical roles in the development of atherosclerotic plaques, control of VSMC biology could present a promising approach to the management of atherosclerosis. Differentiation and proliferation of VSMCs occur early in the development of atherosclerotic lesions, and the developing plaques are maintained by the fibrous cap to generate a stable mass (Sazonova et al., 2015). Using whole-genome shotgun bisulfite sequencing, Zaina et al. indicated that the atherosclerotic portion of the aorta was hypermethylated in numerous genomic loci. Moreover, high-density DNA methylation microarray led to the recognition of genes involved in the function of endothelial cells as wells SMCs (Zaina et al., 2014). Matrix metalloproteinases (MMPs) have been reported to be associated with VSMC biology and the progression of atherosclerosis. MMP-9 was reported to play a role in the migration of VSMCs to other organs (Choi et al., 2005, Johnson et al., 2005, Luttun et al., 2004). In spite of MMP-9 involvement in the development of primary lesions during atherosclerosis, this enzyme can also play a role in the prevention of the development of end-stage lesions in the process of atherosclerosis (Jeong et al.,

2015, Lacolley et al., 2012). It has been shown that VSMCs take part in the promotion of cholesterol influx, reduced rate of efflux, and development of foam cells during the early stages of development of atherosclerotic lesions. Furthermore, VSMCs undergo a process of senescence and programmed cell death that may be involved in the progression of atherosclerosis.

Reports have indicated that SMCs may develop altered expression of several genes and proteins during atherosclerosis. Profiling of the gene expression signature in the SMCs has demonstrated that several genes are underlying a transcriptional modulation of the epigenetic mechanisms, especially DNA methylation (Alexander and Owens, 2012, Spin et al. , 2012). These genes have been observed to participate in the differentiation and phenotypic alteration of SMCs as well as the migration of these cells, leading to the development of vascular complications. A number of these genes that play a role in the differentiation of SMCs are regulated via DNA methylation. These include SMC-specific SM22 α , platelet-derived growth factor (PDGF), serum response factor (SRF), ER α , and ER β (de Oca et al. , 2010). Animal studies into the proliferation of intimal SMCs indicated a global hypomethylation of DNA (Laukkanen et al. , 1999).

Moreover, investigations conducted on $ApoE^{-/-}$ mice as well as human atherosclerotic lesions demonstrated that there was a hypomethylated state of the genomic DNA (Hiltunen et al., 2002). A low DNMT activity, as well as global hypomethylation of DNA, were reported during proliferation and phenotypic differentiation of SMCs *in vitro* (Chen et al., 2011, Little et al., 2013). Alterations in DNA methylation affect the SMC phenotype via the extracellular matrix, which in turn results in vascular calcification (Couffinhal et al., 1994).

In vitro experiments on human aortic smooth muscle cells and rat aortic rings to evaluate the phenotypic difference of VSMCs triggered by high phosphate indicated that high phosphate level was associated with increased DNMT activity and methylation of the promoter region of SM22α.

This resulted in a gain of the osteoblast transcription factor Cbfa1 by VSMCs. The demethylating compound procaine led to declined DNMT activity and inhibited methylation of the SM22 α , which culminated in an increase in SM22 α transcription and less calcification of VSMCs. Therefore, methylation of SM22 α is critical in VSMC calcification (de Oca et al., 2010), and might offer a pathologic pathway in atherosclerosis or CVD by impairing VSMC normal physiology.

The ten-eleven-translocation (TET) family has been implicated in modulation of the extent of DNA methylation (Tahiliani et al. , 2009). TET proteins, containing TET1–TET3, possess DNA demethylation activity through oxidizing 5- methylcytosine (5mC) and generating 5- Hydroxymethylcytosine (5hmC). In vitro treatment of human VSMCs with 5-azacitidine (5-azaC), which is a DNA methylation inhibitor agent, resulting in the overexpression of MMP1, implying to the regulation of MMP1 in VSMCs through methylation (Azechi et al. , 2014). In addition, 5- azaC was reported to inhibit the expansion and migration of SMCs in the airway by altering the methylation of PDGF as well by enhancing the contractile potential of SMCs (Ning et al. , 2013). These observations established a notion that DNA methylation of SMC might be involved in the progression of atherosclerosis. In general, DNA hypomethylation culminates in the expansion and migration of SMCs, eventuating in the expedition of plaque generation (**Figure 2**).

5.5. DNA methylation and elevated homocysteinemia (eHcy) during atherosclerosis

Homocysteine (Hcy), is a non-classic sulfur-containing amino acid that is produced during the process of methionine metabolism. Hcy plays a physiologic function in DNA metabolism through methylation (Perna and Ingrosso, 2015). Methionine is converted to S-adenosylmethionine (SAM), which is the major donor of the methyl group for DNA methyltransferase enzymes. By losing methyl groups, SAM is converted to S-adenosylhomocysteine (SAH), which is a strong

competitive inhibitor for methyltransferase enzymes. SAH hydrolysis leads to the generation of adenosine and Hcy (Mandaviya et al., 2014). Under normal physiological conditions, Hcy levels in plasma range from 5 μ M to 15 μ M (Ji and Kaplowitz, 2004). It has been reported that elevated levels of Hcy, by modulation of the DNA methylation level, is a predisposing risk factor for the progression of atherosclerosis and plays a role in the proliferation of VSMCs as well as endothelial dysfunction. Dysregulated levels of DNA methylation, alongside with elevated levels of Hcy, has been observed in patients with CVD (Castro et al., 2003). Reduced methylation of Alu and LINE-1 elements were reported upon incubation of VSMCs with high doses of Hcy, which increased DNMT function, increased SAH levels, and reduced SAM levels (Yideng et al., 2007). It was also observed that incubation of VSMCs with different doses of Hcy resulted in the decreased methylation and increased expression of PDGF, hence increasing the expansion of VSMC (Han et al., 2014, Jiang et al., 2007). It has been shown that Hcy might impress the ER, which has been implicated in the pathogenesis of atherosclerosis. It was reported that there was a positive association between *estrogen receptor 1 (ESR1)* gene prompter methylation level and the intensity of plaque lesions in atherosclerosis (Huang et al., 2009). Studies have documented that elevated levels of Hcy may contribute to the onset of atherosclerosis development through mechanisms such as the proliferation of VSMC, stimulation of immune system, and oxidative stress (Bao and Zheng, 2015, Leach et al., 2014). It was also reported that incubation of monocytes with Hcy promoted the methylation of DNA in the promoter region Peroxisome proliferator-activated receptor alpha (PPARA) and PPAR gamma (PPARG) genes, culminating in a decreased levels of genes translation and transcription. Furthermore, there were a decrease and increase in the SAM and SAH levels, respectively. Such observations suggest that PPARA and PPARG gene methylations are stimulated through Hcy that might confer an important tool in the progression of

atherosclerosis, proposing a potential therapeutic target for ameliorating Hcy-triggered atherosclerotic lesions (Yideng et al., 2008). Elevated level of Hcy was shown to result in DNA hypomethylation in the genes implicated in the atherosclerosis pathogenesis like *Cyclin A*, which plays a role in the inhibition of cell cycle progression as well as the endothelial remodeling. Furthermore, elevated concentrations of Hcy resulted in decreased activity of DNMT1; nonetheless, upregulation of DNMT1 reversed the inhibitory effects of Hcy on *Cyclin A* transcription and growth inhibition of EC. As a consequence, hypomethylation of *Cyclin A* through elevated levels of Hcy may be a critical mechanism that is responsible for EC proliferation ultimately leading to progression of atherosclerosis (Jamaluddin et al., 2007).

Circulating concentrations of cholesterol might also be regulated by Hcy. It was reported that clinically important levels of Hcy (100 mM) resulted in increased serum levels of cholesteryl ester, free cholesterol, and total cholesterol. Incubation of 100 mM of Hcy with human monocytes resulted in underexpression of apolipoprotein E (ApoE) (Yi-Deng et al., 2007). Elevated levels of Hcy was shown to regulate DNA hypermethylation of ApoE and, therefore, its expression in atherosclerotic lesions (Wang et al., 2013).

Elevated levels of Hcy may contribute to the abnormal deposition of lipid in the proximal aorta, a manifestation mirroring the progress of atherosclerosis (Chen et al., 2001). Furthermore, Hcy might alter the methylation state of genes involved in cholesterol efflux (Liang et al., 2013). Elevated levels of Hcy may promote a global hypomethylation of DNA and, therefore, modulate the expression profile of genes implicated in atherosclerosis (Hsi et al., 2019, Mandaviya et al., 2014). That notwithstanding, the exact molecular mechanisms underlying the DNA methylation modulation by Hcy and the association between the global DNA methylation and regulation of atherosclerosis-specific genes are not fully understood. Further investigations to resolve these

issues may lead to the identification of novel therapeutic targets to treat atherosclerosis induced by Hcy.

6. Therapeutic potential of controlling DNA methylation in the treatment of atherosclerosis Although relatively little is known concerning the DNA methylation alterations during atherosclerosis, several promising results in the field of pharmacoepigenetics have been obtained from preclinical studies and clinical trials. In a phase III trial, it was reported that apabetalone, which is an inhibitor of bromodomain and extra-terminal proteins (BET, a histone modification reader), resulted in elevated circulating concentrations of HDL, reduced CRP level, and upregulation of ApoAI, that was accompanied by fewer cardiovascular events in the patients (Nicholls et al., 2018). In addition, several agents that target epigenetic regulators are now in preclinical or clinical assessments to treat cancers and may have applications in the treatment of cardiovascular events. It has been reported that Decitabine (5-aza-dC) can stimulate the expression of ESR1 and ESR2 in tumor cells and that it plays a role in maintaining the stability of antiinflammatory phenotype in the ECs obtained from a mouse model of atherosclerosis (Dunn et al. , 2014). Furthermore, decitabine was reported to be involved in the amelioration of atherosclerosis by repressing the recruitment and activation of monocytes and other immune cells (Cao et al., 2014).

7. Conclusions

Although the knowledge on the role of epigenetics in the pathogenesis of atherosclerosis is in infancy, the involvement of alterations of DNA methylation is indisputable. Additionally, other epigenetic regulatory mechanisms, including histone modifications and microRNAs, play critical

roles in the etiology and pathogenesis of atherosclerosis. These epigenetic regulatory mechanisms present a promising approach to the treatment of atherosclerosis. However, to date, there is little evidence relating to the treatment of atherosclerosis by modulation of DNA methylation. Additional research is needed into the environmental stimuli of DNA methylation pattern as well as the molecular mechanisms underlying these methylation alterations implicated in atherogenesis. A more detailed understanding of the mechanisms by which abnormalities of DNA methylation result in atherosclerosis would allow the development of diagnostic biomarkers and efficient therapies. Of note, there is evidence suggesting that statins, as the major class of drugs used for the treatment of atherosclerotic cardiovascular disease, can affect epigenetic events including DNA methylation (Ishikawa et al., 2014, Li et al., 2014, Zhu et al., 2017). This finding may imply that epigenetic effects of statins can explain, at least in part, the putative anti-atherosclerotic as well as the wide range of pleiotropic activities described for these drugs (Allen and Mamotte, 2017, Chruściel et al., 2016, Parizadeh et al., 2011, Sahebkar et al., 2015a, Sahebkar et al., 2015b, Storino Farina et al., 2015). Finally, future investigations may be directed toward epidrugs, such as those that inhibit the DNMT enzymes, in order to treat atherosclerosis.

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Authors' contributions

All authors searched for literature and wrote and edited the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

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Consent for publication

Not applicable.

Competing interests

Dr. Penson owns four shares in Astra Zeneca PLC and has received travel/speaker's fees from Amgen Inc. Dr. Banach has served as speakers bureau: Abbott/Mylan, Abbott Vascular, Actavis, Akcea, Amgen, Biofarm, KRKA, MSD, Sanofi-Aventis and Valeant; consultant to Abbott Vascular, Akcea, Amgen, Daichii Sankyo, Esperion, Lilly, MSD, Resverlogix, Sanofi-Aventis; received grants from Sanofi and Valeant. Other authors have no competing inerests to disclose.

References

Afanas'ev I. Mechanisms of superoxide signaling in epigenetic processes: relation to aging and cancer. Aging and disease. 2015;6:216.

Alexander MR, Owens GK. Epigenetic control of smooth muscle cell differentiation and phenotypic switching in vascular development and disease. Annual review of physiology. 2012;74:13-40.

Allen SC, Mamotte CDS. Pleiotropic and adverse effects of statins-do epigenetics play a role? Journal of Pharmacology and Experimental Therapeutics. 2017;362:319-26.

Azechi T, Sato F, Sudo R, Wachi H. 5-Aza-2-deoxycytidine, a DNA methyltransferase inhibitor, facilitates the inorganic phosphorus-induced mineralization of vascular smooth muscle cells. Journal of atherosclerosis and thrombosis. 2014;21:463-76.

Baccarelli A, Bollati V. Epigenetics and environmental chemicals. Current opinion in pediatrics. 2009;21:243.

Bakshi C, Vijayvergiya R, Dhawan V. Aberrant DNA methylation of M1-macrophage genes in coronary artery disease. Scientific reports. 2019;9:1-22.

Bao Xm, Zheng H. Atorvastatin attenuates homocysteine-induced migration of smooth muscle cells through mevalonate pathway involving reactive oxygen species and p38 MAPK. Clinical and Experimental Pharmacology and Physiology. 2015;42:865-73.

Bennett M, Wang J, Uryga AK, Reinhold J, Figg N, Baker L, et al. Vascular smooth muscle cell senescence promotes atherosclerosis and features of plaque vulnerability. Circulation. 2015:CIRCULATIONAHA. 115.016457.

Berlett BS, Stadtman ER. Protein oxidation in aging, disease, and oxidative stress. Journal of Biological Chemistry. 1997;272:20313-6.

Berliner JA, Watson AD. A role for oxidized phospholipids in atherosclerosis. New England Journal of Medicine. 2005;353:9-11.

Canani RB, Paparo L, Nocerino R, Cosenza L, Pezzella V, Di Costanzo M, et al. Differences in DNA methylation profile of Th1 and Th2 cytokine genes are associated with tolerance acquisition in children with IgE-mediated cow's milk allergy. Clinical epigenetics. 2015;7:38.

Cao Q, Wang X, Jia L, Mondal AK, Diallo A, Hawkins GA, et al. Inhibiting DNA methylation by 5-Aza-2'-deoxycytidine ameliorates atherosclerosis through suppressing macrophage inflammation. Endocrinology. 2014;155:4925-38.

Carless MA, Kulkarni H, Kos MZ, Charlesworth J, Peralta JM, Göring HH, et al. Genetic effects on DNA methylation and its potential relevance for obesity in Mexican Americans. PLoS One. 2013;8:e73950.

Castro R, Rivera I, Struys EA, Jansen EE, Ravasco P, Camilo ME, et al. Increased homocysteine and S-adenosylhomocysteine concentrations and DNA hypomethylation in vascular disease. Clinical chemistry. 2003;49:1292-6.

Chan Y, Fish JE, D'Abreo C, Lin S, Robb GB, Teichert A-M, et al. The cell-specific expression of endothelial nitric oxide synthase: a role for DNA methylation. Journal of Biological Chemistry. 2004.

Chen K-C, Wang Y-S, Hu C-Y, Chang W-C, Liao Y-C, Dai C-Y, et al. OxLDL up-regulates microRNA-29b, leading to epigenetic modifications of MMP-2/MMP-9 genes: a novel mechanism for cardiovascular diseases. The FASEB Journal. 2011;25:1718-28.

Chen Z, Karaplis AC, Ackerman SL, Pogribny IP, Melnyk S, Lussier-Cacan S, et al. Mice deficient in methylenetetrahydrofolate reductase exhibit hyperhomocysteinemia and decreased methylation capacity, with neuropathology and aortic lipid deposition. Human molecular genetics. 2001;10:433-44.

Choi ET, Collins ET, Marine LA, Uberti MG, Uchida H, Leidenfrost JE, et al. Matrix Metalloproteinase-9 Modulation by Resident Arterial Cells Is Responsible for Injury-Induced Accelerated Atherosclerotic Plaque Development in Apolipoprotein E–Deficient Mice. Arteriosclerosis, thrombosis, and vascular biology. 2005;25:1020-5.

Chruściel P, Sahebkar A, Rembek-Wieliczko M, Serban MC, Ursoniu S, Mikhailidis DP, et al. Impact of statin therapy on plasma adiponectin concentrations: A systematic review and metaanalysis of 43 randomized controlled trial arms. Atherosclerosis. 2016;253:194-208.

Cipollone F, Fazia ML. COX-2 and atherosclerosis. Journal of cardiovascular pharmacology. 2006;47:S26-S36.

Couffinhal T, Duplaa C, Moreau C, Lamaziere J, Bonnet J. Regulation of vascular cell adhesion molecule-1 and intercellular adhesion molecule-1 in human vascular smooth muscle cells. Circulation research. 1994;74:225-34.

Csiszar A, Ungvari Z, Koller A, Edwards JG, Kaley G. Proinflammatory phenotype of coronary arteries promotes endothelial apoptosis in aging. Physiol Genomics. 2004;17:21-30.

de Oca AM, Madueño JA, Martinez-Moreno JM, Guerrero F, Muñoz-Castañeda J, Rodriguez-Ortiz ME, et al. High-phosphate-induced calcification is related to SM22α promoter methylation in vascular smooth muscle cells. Journal of Bone and Mineral Research. 2010;25:1996-2005.

Deaton AM, Cook PC, De Sousa D, Phythian-Adams AT, Bird A, MacDonald AS. A unique DNA methylation signature defines a population of IFN- γ /IL-4 double-positive T cells during helminth infection. European journal of immunology. 2014;44:1835-41.

Dekkers KF, Neele AE, Jukema JW, Heijmans BT, de Winther MP. Human monocyte-tomacrophage differentiation involves highly localized gain and loss of DNA methylation at transcription factor binding sites. Epigenetics & chromatin. 2019;12:34.

Denke MA, Grundy SM. Hypercholesterolemia in elderly persons: resolving the treatment dilemma. Annals of Internal Medicine. 1990;112:780-92.

Ding Z, Liu S, Wang X, Deng X, Fan Y, Sun C, et al. Hemodynamic shear stress via ROS modulates PCSK9 expression in human vascular endothelial and smooth muscle cells and along the mouse aorta. Antioxidants & redox signaling. 2015;22:760-71.

Drinkwater RD, Blake TJ, Morley AA, Turner DR. Human lymphocytes aged in vivo have reduced levels of methylation in transcriptionally active and inactive DNA. Mutation Research/DNAging. 1989;219:29-37.

Dunn J, Qiu H, Kim S, Jjingo D, Hoffman R, Kim CW, et al. Flow-dependent epigenetic DNA methylation regulates endothelial gene expression and atherosclerosis. The Journal of clinical investigation. 2014;124:3187-99.

Endtmann C, Ebrahimian T, Czech T, Arfa O, Laufs U, Fritz M, et al. Angiotensin II impairs endothelial progenitor cell number and function in vitro and in vivo: implications for vascular regeneration. Hypertension. 2011;58:394-403.

Fabbri M, Paone A, Calore F, Galli R, Croce CM. A new role for microRNAs, as ligands of Tolllike receptors. RNA biology. 2013;10:169-74.

Fan S, Zhang X. CpG island methylation pattern in different human tissues and its correlation with gene expression. Biochemical and biophysical research communications. 2009;383:421-5.

Findley CM, Mitchell RG, Duscha BD, Annex BH, Kontos CD. Plasma levels of soluble Tie2 and vascular endothelial growth factor distinguish critical limb ischemia from intermittent claudication in patients with peripheral arterial disease. J Am Coll Cardiol. 2008;52:387-93.

Finkel T, Holbrook NJ. Oxidants, oxidative stress and the biology of ageing. Nature. 2000;408:239.

Glass CK, Witztum JL. Atherosclerosis. the road ahead. Cell. 2001;104:503-16.

Han XB, Zhang HP, Cao CJ, Wang YH, Tian J, Yang XL, et al. Aberrant DNA methylation of the PDGF gene in homocysteine-mediated VSMC proliferation and its underlying mechanism. Molecular medicine reports. 2014;10:947-54.

Hansson GK. Inflammation, atherosclerosis, and coronary artery disease. N Engl J Med. 2005;352:1685-95.

Hansson GK, Robertson A-KL, Söderberg-Nauclér C. Inflammation and atherosclerosis. Annu Rev Pathol Mech Dis. 2006;1:297-329.

Hara S, Takano T, Fujikawa T, Yamada M, Wakai T, Kono T, et al. Forced expression of DNA methyltransferases during oocyte growth accelerates the establishment of methylation imprints but not functional genomic imprinting. Human molecular genetics. 2014;23:3853-64.

Hastings NE, Simmers MB, McDonald OG, Wamhoff BR, Blackman BR. Atherosclerosis-prone hemodynamics differentially regulates endothelial and smooth muscle cell phenotypes and promotes pro-inflammatory priming. American Journal of Physiology-Cell Physiology. 2007;293:C1824-C33.

Hiltunen MO, Turunen MP, Häkkinen TP, Rutanen J, Hedman M, Mäkinen K, et al. DNA hypomethylation and methyltransferase expression in atherosclerotic lesions. Vascular medicine. 2002;7:5-11.

Holliday R. The significance of DNA methylation in cellular aging. Molecular biology of aging: Springer; 1985. p. 269-83.

Holliday R. The inheritance of epigenetic defects. Science. 1987;238:163-70.

Hoogeveen RC, Morrison A, Boerwinkle E, Miles JS, Rhodes CE, Sharrett AR, et al. Plasma MCP-1 level and risk for peripheral arterial disease and incident coronary heart disease: Atherosclerosis Risk in Communities study. Atherosclerosis. 2005;183:301-7.

Hopkins PN, Williams RR. A survey of 246 suggested coronary risk factors. Atherosclerosis. 1981;40:1-52.

Hsi E, Wang Y-S, Huang C-W, Yu M-L, Juo S-HH, Liang C-L. Genome-wide DNA hypermethylation and homocysteine increase a risk for myopia. International journal of ophthalmology. 2019;12:38.

Huang K, Fan G. DNA methylation in cell differentiation and reprogramming: an emerging systematic view. Regenerative medicine. 2010;5:531-44.

Huang Y-S, Zhi Y-F, Wang S-R. Hypermethylation of estrogen receptor- α gene in atheromatosis patients and its correlation with homocysteine. Pathophysiology. 2009;16:259-65.

Ianni M, Porcellini E, Carbone I, Potenzoni M, Pieri A, Pastizzaro C, et al. Genetic factors regulating inflammation and DNA methylation associated with prostate cancer. Prostate cancer and prostatic diseases. 2013;16:56.

International Consortium for Blood Pressure Genome-Wide Association S, Ehret GB, Munroe PB, Rice KM, Bochud M, Johnson AD, et al. Genetic variants in novel pathways influence blood pressure and cardiovascular disease risk. Nature. 2011;478:103-9.

Ishikawa S, Hayashi H, Kinoshita K, Abe M, Kuroki H, Tokunaga R, et al. Statins inhibit tumor progression via an enhancer of zeste homolog 2-mediated epigenetic alteration in colorectal cancer. International Journal of Cancer. 2014;135:2528-36.

Issa J. CpG-island methylation in aging and cancer. Current topics in microbiology and immunology. 2000;249:101-18.

Jamaluddin MS, Yang X, Wang H. Hyperhomocysteinemia, DNA methylation and vascular disease. Clinical Chemical Laboratory Medicine. 2007;45:1660-6.

Jeong J-W, Kim JW, Ku SK, Kim SG, Kim KY, Kim G-Y, et al. Essential oils purified from Schisandrae semen inhibits tumor necrosis factor- α -induced matrix metalloproteinase-9 activation and migration of human aortic smooth muscle cells. BMC complementary and alternative medicine. 2015;15:7.

Ji C, Kaplowitz N. Hyperhomocysteinemia, endoplasmic reticulum stress, and alcoholic liver injury. World journal of gastroenterology: WJG. 2004;10:1699.

Jiang Y-Z, Manduchi E, Stoeckert CJ, Davies PF. Arterial endothelial methylome: differential DNA methylation in athero-susceptible disturbed flow regions in vivo. BMC genomics. 2015;16:506.

Jiang Y, Sun T, Xiong J, Cao J, Li G, Wang S. Hyperhomocysteinemia-mediated DNA hypomethylation and its potential epigenetic role in rats. Acta biochimica et biophysica Sinica. 2007;39:657-67.

Johnson JL, George SJ, Newby AC, Jackson CL. Divergent effects of matrix metalloproteinases 3, 7, 9, and 12 on atherosclerotic plaque stability in mouse brachiocephalic arteries. Proceedings of the National Academy of Sciences. 2005;102:15575-80.

Jones PA, Takai D. The role of DNA methylation in mammalian epigenetics. Science. 2001;293:1068-70.

Joyce BT, Gao T, Liu L, Zheng Y, Liu S, Zhang W, et al. Longitudinal study of DNA methylation of inflammatory genes and cancer risk. Cancer Epidemiology and Prevention Biomarkers. 2015:cebp. 0198.2015.

Karimi M, Johansson S, Stach D, Corcoran M, Grandér D, Schalling M, et al. LUMA (LUminometric Methylation Assay)—a high throughput method to the analysis of genomic DNA methylation. Experimental cell research. 2006;312:1989-95.

Kennedy A, Schmidt EM, Cribbs AP, Penn H, Amjadi P, Syed K, et al. A novel upstream enhancer of FOXP3, sensitive to methylation-induced silencing, exhibits dysregulated methylation in rheumatoid arthritis Treg cells. European journal of immunology. 2014;44:2968-78.

Kim D, Kubzansky LD, Baccarelli A, Sparrow D, Spiro A, Tarantini L, et al. Psychological factors and DNA methylation of genes related to immune/inflammatory system markers: the VA Normative Aging Study. BMJ open. 2016;6:e009790.

Kim GH, Ryan JJ, Marsboom G, Archer SL. Epigenetic mechanisms of pulmonary hypertension. Pulmonary circulation. 2011;1:347-56.

Kobayashi S, Inoue N, Azumi H, Seno T, Hirata K-i, Kawashima S, et al. Expressional changes of the vascular antioxidant system in atherosclerotic coronary arteries. Journal of atherosclerosis and thrombosis. 2002;9:184-90.

Koene RJ, Prizment AE, Blaes A, Konety SH. Shared Risk Factors in Cardiovascular Disease and Cancer. Circulation. 2016;133:1104-14.

Krychtiuk KA, Kastl SP, Hofbauer SL, Wonnerth A, Goliasch G, Ozsvar-Kozma M, et al. Monocyte subset distribution in patients with stable atherosclerosis and elevated levels of lipoprotein (a). Journal of clinical lipidology. 2015;9:533-41.

Lacolley P, Regnault V, Nicoletti A, Li Z, Michel J-B. The vascular smooth muscle cell in arterial pathology: a cell that can take on multiple roles. Cardiovascular research. 2012;95:194-204.

Laukkanen MO, Mannermaa S, Hiltunen MO, Aittomäki S, Airenne K, Jänne J, et al. Local hypomethylation in atherosclerosis found in rabbit ec-sod gene. Arteriosclerosis, thrombosis, and vascular biology. 1999;19:2171-8.

Leach NV, Dronca E, Vesa SC, Sampelean DP, Craciun EC, Lupsor M, et al. Serum homocysteine levels, oxidative stress and cardiovascular risk in non-alcoholic steatohepatitis. European Journal of internal medicine. 2014;25:762-7.

Li L, Hou JJ, Qiu RR, Jia SB, Cong GZ, Sun N. Influence of atorvastatin in Bcl-2 methylation in cultured human umbilical endothelial cells treated with homocysteine and its mechanism of antiarteriosclerosis. Journal of Jilin University Medicine Edition. 2014;40:1002-6.

Liang Y, Yang X, Ma L, Cai X, Wang L, Yang C, et al. Homocysteine-mediated cholesterol efflux via ABCA1 and ACAT1 DNA methylation in THP-1 monocyte-derived foam cells. Acta Biochim Biophys Sin. 2013;45:220-8.

Libby P, Ridker PM. Inflammation and atherothrombosis: from population biology and bench research to clinical practice. Journal of the American College of Cardiology. 2006;48:A33-A46.

Libby P, Ridker PM, Hansson GK. Progress and challenges in translating the biology of atherosclerosis. Nature. 2011;473:317.

Lim S-O, Gu J-M, Kim MS, Kim H-S, Park YN, Park CK, et al. Epigenetic changes induced by reactive oxygen species in hepatocellular carcinoma: methylation of the E-cadherin promoter. Gastroenterology. 2008;135:2128-40. e8.

Little PJ, Rostam MA, Piva TJ, Getachew R, Kamato D, Guidone D, et al. Suramin inhibits PDGFstimulated receptor phosphorylation, proteoglycan synthesis and glycosaminoglycan hyperelongation in human vascular smooth muscle cells. Journal of Pharmacy and Pharmacology. 2013;65:1055-63.

Liu C, Xu D, Sjöberg J, Forsell P, Björkholm M, Claesson H-E. Transcriptional regulation of 15lipoxygenase expression by promoter methylation. Experimental cell research. 2004;297:61-7.

Liu L, Wylie RC, Andrews LG, Tollefsbol TO. Aging, cancer and nutrition: the DNA methylation connection. Mech Ageing Dev. 2003;124:989-98.

Liu M, Timmons BW. The effect of acute exercise on neutrophil reactive oxygen species production and inflammatory markers in healthy prepubertal and adult males. Pediatric exercise science. 2016;28:55-63.

Luttun A, Lutgens E, Manderveld A, Maris K, Collen D, Carmeliet P, et al. Loss of matrix metalloproteinase-9 or matrix metalloproteinase-12 protects apolipoprotein E-deficient mice against atherosclerotic media destruction but differentially affects plaque growth. Circulation. 2004;109:1408-14.

Ma S, Zhang H, Sun W, Gong H, Wang Y, Ma C, et al. Hyperhomocysteinemia induces cardiac injury by up-regulation of p53-dependent Noxa and Bax expression through the p53 DNA methylation in ApoE^{-/-} mice. Acta Biochim Biophys Sin. 2013;45:391-400.

Mandaviya PR, Stolk L, Heil SG. Homocysteine and DNA methylation: a review of animal and human literature. Molecular genetics and metabolism. 2014;113:243-52.

Mays-Hoopes L, Chao W, Butcher HC, Huang RCC. Decreased methylation of the major mouse long interspersed repeated DNA during aging and in myeloma cells. Developmental genetics. 1986;7:65-73.

Morales V, del Pilar M, Zaina S, Heyn H, Carmona FJ, Varol N, et al. The DNA methylation drift of the atherosclerotic aorta increases with lesion progression. BMC Medical Genomics, 2015, vol 8, num 7. 2015.

Nanduri J, Prabhakar NR. Epigenetic regulation of carotid body oxygen sensing: clinical implications. Arterial Chemoreceptors in Physiology and Pathophysiology: Springer; 2015. p. 1-8.

Nicholls SJ, Ray KK, Johansson JO, Gordon A, Sweeney M, Halliday C, et al. Selective BET protein inhibition with apabetalone and cardiovascular events: a pooled analysis of trials in patients with coronary artery disease. American Journal of Cardiovascular Drugs. 2018;18:109-15.

Ning Y, Huang H, Dong Y, Sun Q, Zhang W, Xu W, et al. 5-Aza-2'-deoxycytidine inhibited PDGF-induced rat airway smooth muscle cell phenotypic switching. Archives of toxicology. 2013;87:871-81.

Niu Y, DesMarais TL, Tong Z, Yao Y, Costa M. Oxidative stress alters global histone modification and DNA methylation. Free Radical Biology and Medicine. 2015;82:22-8.

O'Hagan HM, Wang W, Sen S, Shields CD, Lee SS, Zhang YW, et al. Oxidative damage targets complexes containing DNA methyltransferases, SIRT1, and polycomb members to promoter CpG Islands. Cancer cell. 2011;20:606-19.

Parizadeh SMR, Azarpazhooh MR, Moohebati M, Nematy M, Ghayour-Mobarhan M, Tavallaie S, et al. Simvastatin therapy reduces prooxidant-antioxidant balance: Results of a placebo-controlled cross-over trial. Lipids. 2011;46:333-40.

Park J, Jang KL. Hepatitis C virus represses E-cadherin expression via DNA methylation to induce epithelial to mesenchymal transition in human hepatocytes. Biochemical and biophysical research communications. 2014;446:561-7.

Perna AF, Ingrosso D. Atherosclerosis determinants in renal disease: how much is homocysteine involved? Nephrology Dialysis Transplantation. 2015;31:860-3.

Post WS, Goldschmidt-Clermont PJ, Wilhide CC, Heldman AW, Sussman MS, Ouyang P, et al. Methylation of the estrogen receptor gene is associated with aging and atherosclerosis in the cardiovascular system. Cardiovascular research. 1999;43:985-91.

Poulter N. Coronary heart disease is a multifactorial disease. American Journal of Hypertension. 1999;12:928-58.

Quintero-Ronderos P, Montoya-Ortiz G. Epigenetics and autoimmune diseases. Autoimmune diseases. 2012;2012.

Rangel-Salazar R, Wickström-Lindholm M, Aguilar-Salinas CA, Alvarado-Caudillo Y, Døssing KB, Esteller M, et al. Human native lipoprotein-induced de novo DNA methylation is associated with repression of inflammatory genes in THP-1 macrophages. BMC genomics. 2011;12:582.

Rasmussen MA, Holst B, Tümer Z, Johnsen MG, Zhou S, Stummann TC, et al. Transient p53 suppression increases reprogramming of human fibroblasts without affecting apoptosis and DNA damage. Stem cell reports. 2014;3:404-13.

Robertson KD. DNA methylation and human disease. Nat Rev Genet. 2005;6:597-610.

Roman-Gomez J, Jimenez-Velasco A, Agirre X, Cervantes F, Sanchez J, Garate L, et al. Promoter hypomethylation of the LINE-1 retrotransposable elements activates sense/antisense transcription and marks the progression of chronic myeloid leukemia. Oncogene. 2005;24:7213.

Ross R, Glomset JA. The pathogenesis of atherosclerosis. New England journal of medicine. 1976;295:369-77.

Sahebkar A, Kotani K, Serban C, Ursoniu S, Mikhailidis DP, Jones SR, et al. Statin therapy reduces plasma endothelin-1 concentrations: A meta-analysis of 15 randomized controlled trials. Atherosclerosis. 2015a;241:433-42.

Sahebkar A, Serban C, Mikhailidis DP, Undas A, Lip GYH, Muntner P, et al. Association between statin use and plasma d-dimer levels: A systematic review and meta-analysis of randomised controlled trials. Thrombosis and Haemostasis. 2015b;114:546-57.

Sanjadi M, Rezvanie Sichanie Z, Totonchi H, Karami J, Rezaei R, Aslani S. Atherosclerosis and autoimmunity: a growing relationship. International journal of rheumatic diseases. 2018;21:908-21.

Sazonova OV, Isenberg BC, Herrmann J, Lee KL, Purwada A, Valentine AD, et al. Extracellular matrix presentation modulates vascular smooth muscle cell mechanotransduction. Matrix Biology. 2015;41:36-43.

Sharma S, Kelly TK, Jones PA. Epigenetics in cancer. Carcinogenesis. 2010;31:27-36.

Singh M, Bedi US. Is atherosclerosis regression a realistic goal of statin therapy and what does that mean? Current atherosclerosis reports. 2013;15:294.

Sipahi I, Tuzcu EM. Candidate Mechanisms for Regression of Coronary Atherosclerosis with High-Dose Statins. American journal of cardiovascular drugs. 2008;8:365-71.

Skålén K, Gustafsson M, Rydberg EK, Hultén LM, Wiklund O, Innerarity TL, et al. Subendothelial retention of atherogenic lipoproteins in early atherosclerosis. Nature. 2002;417:750.

Spin JM, Maegdefessel L, Tsao PS. Vascular smooth muscle cell phenotypic plasticity: focus on chromatin remodelling. Cardiovascular research. 2012;95:147-55.

Steinberg D. Atherogenesis in perspective: hypercholesterolemia and inflammation as partners in crime. Nature medicine. 2002;8:1211.

Stenvinkel P, Karimi M, Johansson S, Axelsson J, Suliman M, Lindholm B, et al. Impact of inflammation on epigenetic DNA methylation–a novel risk factor for cardiovascular disease? Journal of internal medicine. 2007;261:488-99.

Steucke KE, Tracy PV, Hald ES, Hall JL, Alford PW. Vascular smooth muscle cell functional contractility depends on extracellular mechanical properties. Journal of biomechanics. 2015;48:3044-51.

Storino Farina M, Rojano Rada J, Molina Garrido A, Martínez X, Pulgar A, Paniagua R, et al. Statins and atherosclerosis: the role of epigenetics. Medwave. 2015;15:e6324.

Študentová H, Indráková J, Petrová P, Kamínek M, Kalábová H, Šrámek V, et al. Risk factors of atherosclerosis during systemic therapy targeting vascular endothelial growth factor. Oncology letters. 2016;11:939-44.

Tahiliani M, Koh KP, Shen Y, Pastor WA, Bandukwala H, Brudno Y, et al. Conversion of 5methylcytosine to 5-hydroxymethylcytosine in mammalian DNA by MLL partner TET1. Science. 2009;324:930-5.

Tanaka K, Masuda J, Imamura T, Sueishi K, Nakashima T, Sakurai I, et al. A nation-wide study of atherosclerosis in infants, children and young adults in Japan. Atherosclerosis. 1988;72:143-56.

Turk PW, Laayoun A, Smith SS, Weitzman SA. DNA adduct 8-hydroxyl-2'-deoxyguanosine (8-hydroxyguanine) affects function of human DNA methyltransferase. Carcinogenesis. 1995;16:1253-5.

van Diepen JA, Berbée JF, Havekes LM, Rensen PC. Interactions between inflammation and lipid metabolism: relevance for efficacy of anti-inflammatory drugs in the treatment of atherosclerosis. Atherosclerosis. 2013;228:306-15.

Wang L, Jia X-J, Jiang H-J, Du Y, Yang F, Si S-Y, et al. MicroRNAs 185, 96, and 223 repress selective high-density lipoprotein cholesterol uptake through posttranscriptional inhibition. Molecular and cellular biology. 2013;33:1956-64.

Wang T, Chen M, Liu L, Cheng H, Yan Y-E, Feng Y-H, et al. Nicotine induced CpG methylation of Pax6 binding motif in StAR promoter reduces the gene expression and cortisol production. Toxicology and applied pharmacology. 2011;257:328-37.

Weitzman SA, Turk PW, Milkowski DH, Kozlowski K. Free radical adducts induce alterations in DNA cytosine methylation. Proceedings of the National Academy of Sciences. 1994;91:1261-4.

Wierda RJ, Kuipers HF, van Eggermond MC, Benard A, van Leeuwen JC, Carluccio S, et al. Epigenetic control of CCR5 transcript levels in immune cells and modulation by small molecules inhibitors. Journal of cellular and molecular medicine. 2012;16:1866-77.

Wiesner P, Choi SH, Almazan F, Benner C, Huang W, Diehl CJ, et al. Low doses of lipopolysaccharide and minimally oxidized low-density lipoprotein cooperatively activate macrophages via nuclear factor kappa B and activator protein-1: possible mechanism for acceleration of atherosclerosis by subclinical endotoxemia. Circulation research. 2010;107:56-65. Wilkins JT, Ning H, Berry J, Zhao L, Dyer AR, Lloyd-Jones DM. Lifetime risk and years lived free of total cardiovascular disease. JAMA. 2012;308:1795-801.

Wilson VL, Jones PA. DNA methylation decreases in aging but not in immortal cells. Science. 1983;220:1055-7.

Wilson VL, Smith R, Ma S, Cutler R. Genomic 5-methyldeoxycytidine decreases with age. Journal of Biological Chemistry. 1987;262:9948-51.

Yang K, He YS, Wang XQ, Lu L, Chen QJ, Liu J, et al. MiR-146a inhibits oxidized low-density lipoprotein-induced lipid accumulation and inflammatory response via targeting toll-like receptor 4. FEBS letters. 2011;585:854-60.

Yi-Deng J, Tao S, Hui-Ping Z, Jian-Tuan X, Jun C, Gui-Zhong L, et al. Folate and ApoE DNA methylation induced by homocysteine in human monocytes. DNA and Cell Biology. 2007;26:737-44.

Yideng J, Jianzhong Z, Ying H, Juan S, Jinge Z, Shenglan W, et al. Homocysteine-mediated expression of SAHH, DNMTs, MBD2, and DNA hypomethylation potential pathogenic mechanism in VSMCs. DNA and cell biology. 2007;26:603-11.

Yideng J, Zhihong L, Jiantuan X, Jun C, Guizhong L, Shuren W. Homocysteine-mediated PPAR α , γ DNA methylation and its potential pathogenic mechanism in monocytes. DNA and cell biology. 2008;27:143-50.

Ying AK, Hassanain HH, Roos CM, Smiraglia DJ, Issa J-PJ, Michler RE, et al. Methylation of the estrogen receptor- α gene promoter is selectively increased in proliferating human aortic smooth muscle cells. Cardiovascular research. 2000;46:172-9.

Ylä-Herttuala S, Palinski W, Rosenfeld M, Parthasarathy S, Carew T, Butler S, et al. Evidence for the presence of oxidatively modified low density lipoprotein in atherosclerotic lesions of rabbit and man. The Journal of clinical investigation. 1989;84:1086-95.

Yoo T, Yoon YS, Ryu SH, Ahn JY, Kim S, Ha TY, et al. Hypermethylation of repetitive DNA elements in livers of mice fed an atherogenic diet. Nutrition. 2012;28:127-30.

Yu X-H, Fu Y-C, Zhang D-W, Yin K, Tang C-K. Foam cells in atherosclerosis. Clinica chimica acta. 2013;424:245-52.

Zaina S, Heyn H, Carmona FJ, Varol N, Sayols S, Condom E, et al. DNA methylation map of human atherosclerosis. Circulation: Cardiovascular Genetics. 2014;7:692-700.

Zaina S, Lund G. Cardiovascular epigenome-wide association studies: is epigenetics falling short? Current opinion in lipidology. 2014;25:474-5.

Zhang J-Z, Liu Z, Liu J, Ren J-X, Sun T-S. Mitochondrial DNA induces inflammation and increases TLR9/NF-κB expression in lung tissue. Int J Mol Med. 2014a;33:817-24.

Zhang X, Fu R, Yu J, Wu X. DNA demethylation: where genetics meets epigenetics. Current pharmaceutical design. 2014b;20:1625-31.

Zhang Y, Zeng C. Role of DNA methylation in cardiovascular diseases. Clinical and Experimental Hypertension. 2016;38:261-7.

Zheng J, Cheng J, Zhang Q, Xiao X. Novel insights into DNA methylation and its critical implications in diabetic vascular complications. Bioscience reports. 2017;37.

Zhu B, Gong Y, Yan G, Wang D, Wang Q, Qiao Y, et al. Atorvastatin treatment modulates p16 promoter methylation to regulate p16 expression. FEBS Journal. 2017;284:1868-81.

Gene	Involvement in	Ref
Insulin like growth factor 2 (IGF-II)	Cell differentiation and expansion	(Hara et al. , 2014)
Paired box 6 (PAX6)	Cell differentiation and expansion	(Wang et al., 2011)
Interferon-γ (IFN-γ)	Inflammatory response	(Canani et al. , 2015)
Intercellular adhesion molecule 1(ICAM-I)	Inflammatory response	(Joyce et al. , 2015)
Interleukin 4 (IL-4)	Inflammatory reaction	(Deaton et al., 2014)

 Table 1. Atherosclerosis-specific genes modulated via DNA methylation during the disease.

Tumor protein p53 (P53) Apoptosis	Apoptosis	(Rasmussen
		et al.,
		2014)
R call lymphome 2 (RCL 2)	Apoptosis	(Ma et al.,
B-cell lymphoma 2 (BCL-2)		2013)
District derived encycle forten recenter sinks (DDCE a)	SMCs multiferation	(Ning et al.,
r latelet derived growth factor receptor alpha (r DOI-a)	Swics promeration	2013)
Estrogen receptor alpha/beta (ERα/β)	Atherosclerotic tissues remodeling	(Baccarelli
		and Bollati,
		2009)
Myogenic differentiation 1 (MYOD1)	Atherosclerotic tissues remodeling	(Carless et
Atherosci	Atteroscierotic tissues remodeling	al., 2013)
Nitric oxide synthese 3 (eNOS)	Endothelial cell remodeling	(Chan et al.
		, 2004)
Fos proto-oncogene (c-Fos)	Shear stress	(Hastings et
	Shour Stress	al., 2007)
Cadherin 1 (E-cadherin)	Extracellular matrix Extracellular matrix Extracellular matrix	(Park and
		Jang, 2014)
TIMP metallopeptidase inhibitor 3 (TIMP-3)		(Yang et al.
		, 2011)
Matrix metallopeptidase (MMP)-2, MMP-7, MMP-9 Extracellular matrix		(Choi et al.,
		2005,
		Johnson et
		al., 2005,
		Luttun et
		al., 2004)
C-C motif chemokine receptor 5 (CCR5)	Inflammation	(w) erua et
Forkhead box P3 (Foxp3)	Inflammation	(Kannady
		(Keinleuy et al
		2014
		(Kim et al
Nitric oxide synthase 2 (iNOS)	Inflammation, macrophage activation	2016)
	Plaque development	(Lin et al
15-Lipoxygenase (15-LO)		2004)
Fatty acid desaturase 2 (Fads2)	Plaque development	(Hastings et
		al., 2007)
Superoxide dismutase 3 (SOD)	Plaque development	(Laukkanen
		et al., 1999)

Figure legends

Figure 1. Molecular mechanism of DNA methylation. Reproduced from (Zheng et al., 2017)

Figure 2. DNA methylation regulation of pathogenic pathways during atherosclerosis.