



## LJMU Research Online

**Bosch, J, Elvira, S, Sausor, C, Bielby, J, González-Fernández, I, Alonso, R and Bermejo-Bermejo, V**

**Increased tropospheric ozone levels enhance pathogen infection levels of amphibians**

<http://researchonline.ljmu.ac.uk/id/eprint/14013/>

### Article

**Citation** (please note it is advisable to refer to the publisher's version if you intend to cite from this work)

**Bosch, J, Elvira, S, Sausor, C, Bielby, J, González-Fernández, I, Alonso, R and Bermejo-Bermejo, V (2020) Increased tropospheric ozone levels enhance pathogen infection levels of amphibians. Science of the Total Environment. ISSN 0048-9697**

LJMU has developed [LJMU Research Online](http://researchonline.ljmu.ac.uk/) for users to access the research output of the University more effectively. Copyright © and Moral Rights for the papers on this site are retained by the individual authors and/or other copyright owners. Users may download and/or print one copy of any article(s) in LJMU Research Online to facilitate their private study or for non-commercial research. You may not engage in further distribution of the material or use it for any profit-making activities or any commercial gain.

The version presented here may differ from the published version or from the version of the record. Please see the repository URL above for details on accessing the published version and note that access may require a subscription.

For more information please contact [researchonline@ljmu.ac.uk](mailto:researchonline@ljmu.ac.uk)

<http://researchonline.ljmu.ac.uk/>

1 **Increased tropospheric ozone levels enhance pathogen**  
2 **infection levels of amphibians**

3 Jaime Bosch<sup>a,b,c\*</sup>, Susana Elvira<sup>d</sup>, Cristina Sausor<sup>b</sup>, Jon Bielby<sup>e</sup>, Ignacio González-  
4 Fernández<sup>d</sup>, Rocío Alonso<sup>d</sup>, Victoria Bermejo-Bermejo<sup>d</sup>

5  
6  
7 <sup>a</sup>Research Unit of Biodiversity - CSIC/UO/PA, Universidad de Oviedo, Edificio de  
8 Investigación, 5<sup>a</sup> planta, 3600 Mieres, Spain

9 <sup>b</sup>Museo Nacional de Ciencias Naturales CSIC, José Gutiérrez Abascal 2, 28006 Madrid,  
10 Spain

11 <sup>c</sup>Centro de Investigación, Seguimiento y Evaluación, Parque Nacional de la Sierra de  
12 Guadarrama, 28740 Rascafría, Spain

13 <sup>d</sup>CIEMAT, Ecotoxicology of Air Pollution. Environmental Dept., Avda. Complutense 40,  
14 28040 Madrid, Spain

15 <sup>e</sup>Liverpool John Moores University, School of Natural Sciences and Psychology, James  
16 Parsons Building, Byrom Street, Liverpool, L3 3AF, United Kingdom

17  
18 \*Corresponding author. Email: [bosch@mncn.csic.es](mailto:bosch@mncn.csic.es)

19  
20 Keywords: Air pollution, chytridiomycosis, mountain areas, global change, amphibian  
21 declines

24 **ABSTRACT**

25 As a result of anthropogenic activities, changes to the chemistry of Earth's  
26 atmosphere pose a threat to ecosystem health and biodiversity. One such change is  
27 the increase in tropospheric ozone (O<sub>3</sub>), which is particularly severe in the  
28 Mediterranean basin area, where the levels of this pollutant are chronically high  
29 during spring and summer time. Within this region, Mediterranean mountain  
30 ecosystems are hot spots for biodiversity which may be especially vulnerable to  
31 changes in O<sub>3</sub> levels. Declines in montane amphibian populations have been  
32 recorded worldwide, including the Mediterranean basin. A significant driver of  
33 these declines is the emerging infection disease, chytridiomycosis, caused by the  
34 aquatic fungus *Batrachochytrium dendrobatidis* (*Bd*). Chytridiomycosis has  
35 negatively affected populations of several amphibian species in the Spanish Central  
36 Range, including in the Sierra Guadarrama, and interactions with other biotic and  
37 abiotic factors are an important part of these declines. However, there is little  
38 evidence or knowledge of whether tropospheric O<sub>3</sub> levels may be another factor in  
39 the outbreaks of this disease. To test the hypothesis that O<sub>3</sub> levels are another  
40 interactive driver of *Bd* infection dynamics, two different approaches were  
41 followed: 1) an experimental study in open top chambers was used to quantify the  
42 aspects of how *Bd* infection progressed throughout the metamorphic process  
43 under four different O<sub>3</sub> levels; and 2) a field epidemiological study was used to  
44 analyse the relationship between the *Bd* infection load in the Sierra de Guadarrama  
45 and tropospheric O<sub>3</sub> levels during a 9 year period. Our results suggest that high O<sub>3</sub>  
46 levels significantly delayed the rate of development of tadpoles and increased *Bd*  
47 infection, providing empirical evidence of two new separate ways that may explain  
48 population declines of montane amphibians.

49 **2. INTRODUCTION**

50 Air pollution is causing rapid changes to the chemistry of Earth's  
51 atmosphere, posing a major threat to our environment. Tropospheric ozone (O<sub>3</sub>) is  
52 a major air pollutant, widely affecting rural and forested areas of the Northern  
53 hemisphere, causing harmful impacts on agricultural production, natural  
54 ecosystems and loss of the services they provided (Sutton et al., 2011; CLRTAP  
55 2017). Background O<sub>3</sub> levels have been increasing since the 19<sup>th</sup> century due to the  
56 industrial revolution and the increased anthropogenic production of industrial and  
57 urban emissions (Young et al., 2013; Nopmongcol et al., 2016; Ainsworth et al.,  
58 2020). Ozone precursors (mainly NO<sub>x</sub>, CO and non-methane volatile organic  
59 compounds-NMVOCs) react photochemically to form O<sub>3</sub> and can be transported  
60 long distances in the atmosphere, enhancing O<sub>3</sub> background levels in rural and  
61 natural areas. This local and regional transport is accompanied by long-range and  
62 intercontinental transport, causing high O<sub>3</sub> concentration in regions located far  
63 from sources of pollutant emissions (Cristofanelle et al., 2009, Chen et al., 2017).  
64 Moreover, the link between the O<sub>3</sub> problem with the climate change phenomena is  
65 widely accepted, considering the future meteorological factors like solar radiation  
66 or air temperatures, that enhance atmospheric photochemistry, will also play an  
67 important role on the future O<sub>3</sub> levels (Colette et al., 2013; Lefhon et al., 2018).

68 High O<sub>3</sub> levels have significant physiological effects on humans and other  
69 mammals (Lippmann 1989, U.S. EPA 2013, WHO 2013; Fleming et al., 2018). Ozone  
70 exposure induces an oxidative stress at the respiratory tract that affects  
71 pulmonary function, bronchial airway reactivity or lung permeability, and depletes  
72 the antioxidant defences (Schelegle et al., 2009; Tighe et al., 2015; Brand et al.,  
73 2016). Epidemiological studies evaluating chronic long-term effects suggest that

74 daily exposures to O<sub>3</sub> increase mortality rates and respiratory morbidity of the  
75 European human populations (Jarret et al., 2009; Díaz et al., 2018). An important  
76 effect of O<sub>3</sub> on mammalian physiology is related to the alteration of the immune  
77 system, and in particular of the function of alveolar macrophages, like decreased  
78 phagocytosis of particulate immune complexes, enhanced production of prostaglandin  
79 E<sub>2</sub> or increased superoxide production (Hollingsworth et al., 2007; Tighe et al., 2015).  
80 These functional changes in macrophages are associated with impaired antimicrobial  
81 host defense; i.e. the pollutant can enhance pulmonary infections in mice caused by  
82 streptococci, which are able to proliferate and more fully express virulence factors  
83 after an exposure to the pollutant (Canning et al., 1991; Gilmour et al., 1993). This  
84 immunosuppression, expressed as increased sensitivity to bacterial infections, has  
85 been considered as a general O<sub>3</sub>-response of the mammals in the review of Lacroix  
86 et al. (1998).

87         Our knowledge of O<sub>3</sub> impacts on the health of non-mammalian taxa is  
88 scarce, but some studies focussing on amphibians do exist. Toads may exhibit a  
89 reduction in lung ventilation and a decline in oxygen consumption after O<sub>3</sub>  
90 exposure, which is linked to stressful physiological effects (Mautz et al., 2004;  
91 Dohm et al., 2001, 2008). Exposure to O<sub>3</sub> can also alter the water balance and  
92 thermal preferences in anuran amphibians (Dohm et al., 2001, 2005). O<sub>3</sub> effects on  
93 the immune defence system of amphibians are consistent with those found in  
94 mammalian species. In marine toads, O<sub>3</sub> can reduce the capacity of the alveolar  
95 macrophages to phagocytize foreign particles and microorganisms (Dohne et al.,  
96 2005). These results suggest a possible role of oxidant air pollutants, such as O<sub>3</sub>, in  
97 regional declines of amphibian populations, especially considering their potential

98 interactions with pathogen infections, which are considered to be a major driver of  
99 amphibian declines.

100         Global amphibian populations are threatened by an emerging infectious  
101 disease, chytridiomycosis, caused by the aquatic generalist fungus  
102 *Batrachochytrium dendrobatidis* (hereafter *Bd*) (Fisher et al., 2009). The impacts of  
103 this pathogenic fungus are taxonomically, spatially and temporally heterogeneous,  
104 depending on different biotic and abiotic factors, some related to climate change  
105 parameters (Bosch et al., 2018, Garner et al., 2011). One consistency is that  
106 montane species of amphibians have been particularly heavily affected by the  
107 disease. *Bd* infects and reproduces in the amphibian skin feeding on keratin, a  
108 structural protein found in the mouthparts of larval amphibians and throughout  
109 the body of post-metamorphic individuals. *Bd* can cause amphibian mortality by  
110 interfering with the multiple physiological processes undertaken by the amphibian  
111 skin, such as electrolyte exchange and respiration (Voyles et al., 2009). Immune  
112 responses to *Bd* exposure may be also centre on the amphibian skin, with  
113 symbiotic bacteria living on the epidermis, either directly inhibiting pathogen  
114 growth, or indirectly by priming the amphibian immune system to resist infection.  
115 Given our knowledge of how O<sub>3</sub> can reduce individual respiratory activity and  
116 suppress immune systems, it seems biologically plausible that tropospheric levels  
117 of O<sub>3</sub> could interact with and potentially increase the impacts of pathogens such as  
118 *Bd*.

119         Studies on O<sub>3</sub>-fungal-pathogen interactions have largely focussed on plants,  
120 with the findings being heavily context-dependent. On one hand, O<sub>3</sub> may act as a  
121 fungicide and mitigate impacts of pathogens on the host plant (Dohmen et al.,  
122 1987), whereas in some host-pathogen systems the pollutant enhances the fungi

123 infection (Tiedemann et al., 1991). The direction of the response is therefore likely  
124 to depend on the complex interactions between host and pathogen and their  
125 relative O<sub>3</sub>-sensitivity. To date, we are aware of no studies investigating the  
126 relationship between O<sub>3</sub> and fungal pathogens on wild fauna.

127         The climatic characteristics of the Mediterranean basin favour the  
128 photochemical reactions among O<sub>3</sub> precursors and the formation of the pollutant  
129 (Millán et al., 1997, Cristofanelli and Bonasoni, 2009). These conditions, such as  
130 high solar radiation and temperature, and prevailing stable atmospheric  
131 conditions result in some of the highest surface O<sub>3</sub> concentrations in Europe (EEA  
132 2011). In the Iberian Peninsula, O<sub>3</sub> levels chronically exceed the current thresholds  
133 established for plant ecosystems protection (Ribas and Peñuelas, 2006; Adame and  
134 Sole, 2013) and frequently exceeds the thresholds for human health (MITECO,  
135 2018). Experimental assays have already demonstrated that these O<sub>3</sub> levels are  
136 high enough to reduce crop yield and quality (González-Fernández et al., 2014,  
137 2016). On natural vegetation, including forest (Alonso et al., 2013; Marzoulli et al.,  
138 2018) and herbaceous species (Sanz et al., 2011; Calvete-Sogo et al., 2014), O<sub>3</sub>  
139 concentrations affect parameters related to growth and reproductive fitness that  
140 may lead to changes in the structure and diversity of communities (Calvete-Sogo et  
141 al., 2016).

142         Mediterranean mountains, which are hot spots for biodiversity (Myers et al.,  
143 2000) and frequently belong to protected areas like National Parks or Nature 2000  
144 Network, are currently suffering extensively from elevated levels of tropospheric  
145 ozone (Saavedra et al., 2012; Adame and Sole, 2013; Elvira et al., 2016). In the  
146 Sierra de Guadarrama mountains, the O<sub>3</sub> levels recorded during the 2005-2011  
147 period indicated that concentrations of this pollutant exceed the thresholds for

148 human health, and can be more than three-fold above the standard values for plant  
149 protection, according to the Air Quality Directive EU/50/2008 (Elvira et al., 2016).  
150 Thus, a tropospheric O<sub>3</sub> increase should be considered as a stress factor for the  
151 health of these ecosystems and their constituent parts. Although there are no  
152 standard values for fauna protection, the O<sub>3</sub> seasonal and daily pattern at the  
153 highest altitudes, with high background values maintained during the night (Elvira  
154 et al., 2016) might increase the potential negative effect for nocturnal fauna like  
155 amphibians, which are already experiencing population declines in this region.

156 Chytridiomycosis in the Spanish Central Range negatively affects the  
157 population-level dynamics of several amphibian species. The variability of its  
158 effects has been associated with water temperature variability (Fernandez-  
159 Beaskoetxea et al., 2015) and UV-B exposure (Ortíz-Santaliestra et al., 2011; Hite et  
160 al., 2016). However, the relationship between the presence of *Bd* and abiotic  
161 factors is not always clear, and the relationship between environmental variables  
162 and the prevalence of the infections is weak (Walker et al., 2010). A recent study  
163 based on long-term monitoring in the area indicates that the threat posed by  
164 chytridiomycosis is ongoing after two decades, and even highlighted a positive  
165 effect of climate warming on populations of three out of the nine species present  
166 (Bosch et al., 2018). However, to date, there are no studies that incorporate air  
167 quality parameters and their interactions with *Bd*, despite the possibility that they  
168 may influence *Bd* infection dynamics.

169 In order to study the potential role of the high O<sub>3</sub> levels at Sierra de  
170 Guadarrama area in disease-related amphibian declines two different approaches  
171 were followed: an experimental assay where *Bd* infected common midwife toad,  
172 *Alytes obstretricans*, tadpoles were exposed to different O<sub>3</sub> levels in an Open-Top-



173 Chambers (OTCs) facility; and a field study relating the significance of the O<sub>3</sub> factor  
174 on the prevalence of *Bd* infection in metamorphs of spiny common toads (*Bufo*  
175 *spinosus*) at Sierra de Guadarrama during a 9 year period. The main hypothesis is  
176 that the high oxidative capacity of the pollutant would affect amphibian survival  
177 rates by reducing their capacity to combat *Bd* infection, even though the influence  
178 of ozone on amphibian survival rates could be moderated by other environment  
179 variables which we account for in our analyses.

180

### 181 **3. MATERIALS AND METHODS**

#### 182 3.1. Open-top-chamber experimental study

##### 183 3.1.1. Experimental design and ozone treatments

184 The experiment was performed in the CIEMAT Open-Top-Chamber  
185 experimental facility (hereafter OTC) located in the Spanish central plateau at  
186 Santa Olalla municipality (450 m.a.s.l.; 40°3'N, 4°26'W) at the public research farm  
187 La Higuera (MNCN-CSIC). This location is a rural area far from local sources of  
188 air pollution and 80 km away from Madrid City. Chronic O<sub>3</sub> levels are the only air  
189 quality problem in the area. OTC facilities were developed in the 1980s to study  
190 the effects of O<sub>3</sub> on vegetation and crops (Heck et al., 1982) and they are commonly  
191 used to establish exposure and dose-response functions and threshold values for  
192 plant protection under international forums like the Air Convention of the United  
193 Nations (CLRTAP 2017).

194 Twelve National Crop Loss Assessment Network (NCLAN)-type chambers  
195 (Heck et al., 1982) with a 3-m diameter, allowed an experimental random block  
196 design with four O<sub>3</sub> treatments, each replicated three times (three OTCs per O<sub>3</sub>  
197 treatment). An additional three chamberless ambient plots (AA) were included to

198 control for chamber effect. Ozone treatments were: charcoal filtered air (FA)  
199 mirroring the natural preindustrial background levels, non-filtered air (NFA)  
200 reproducing ambient levels of the farm and non-filtered air supplemented with 20  
201 and 40 nL L<sup>-1</sup> of O<sub>3</sub> (NFA+ and NFA++ respectively) over an 8-hour period (07:00  
202 to 15:00 GTM). Maximum hourly values at NFA++ during the exposure period  
203 ranged between 90-110 nL L<sup>-1</sup> to achieve the sporadically maximum levels  
204 observed on the 10-year study of the O<sub>3</sub> levels at Sierra de Guadarrama Mountains  
205 (Elvira et al., 2016).

206         Within each OTC, O<sub>3</sub> for the NFA+ and NFA++ treatments was supplied by  
207 means of an O<sub>3</sub>-generator (Model 16, A2Z Ozone Systems Inc., USA) system fed  
208 with pure oxygen. The concentration of O<sub>3</sub> (ML® 9810B, Teledyne, USA), sulphur  
209 dioxide (SO<sub>2</sub>; ML®9850B UV, Teledyne, USA), and nitrogen oxides (NO<sub>2</sub> and NO;  
210 ML®9841, Teledyne, USA) inside each OTC and AA plot were monitored  
211 continuously using an automated time-sharing system which sampled each AA plot  
212 and OTC for 10 min, thus sampled all the field each 2.5 h. The air temperature and  
213 relative humidity within each OTC and AA plot was monitored with a  
214 meteorological sensor (HOBO® Pro v2, Onset, USA) and the water temperature of  
215 the tadpole containers was also monitored (TMC6-HD HOBO®, Onset, USA). A more  
216 detailed description of the facility can be consulted from Calvete et al., (2014).

### 217 3.1.2. Animal collection and maintenance

218         *Alytes obstetricans* tadpoles at Gosner stage 36 (no, or rudimentary, hind  
219 limbs present; Gosner 1960) were captured in April 2016 from Toro, a mid-  
220 altitude site (Zamora, Central Spain, 740 m a.s.l.; 41°22'N, 5°26'W), where the  
221 prevalence of *Bd* infection in larval stages is known to approach 100% during  
222 colder months (Fernández-Beaskoetxea et al., 2015). The oral disc of a subset of 20

223 animals was swabbed to quantify *Bd* infection levels (see methods below), and  
224 yielded a 100% prevalence.

225 Individual tadpoles were placed at ground level within a container filled  
226 with 2 L of clean spring water, and between six and eight containers were placed in  
227 each treatment replicate. Thus, a total of 20-24 individual tadpoles were exposed  
228 to each O<sub>3</sub> treatment. Tadpoles were fed *ad libitum* with ground fish food, and  
229 water was changed twice a week. Once the forelimbs of a tadpole had emerged the  
230 water in the container was reduced by 70% and a piece of plastic mesh was placed  
231 inside to provide terrestrial habitat. Individuals were monitored until the  
232 completion of metamorphosis.

### 233 3.1.3. Survival and rate of development

234 Mortality was recorded every day. Rate of development was measured once  
235 a week as the proportion of individuals within each O<sub>3</sub> treatments to have reached  
236 the phenological stage of Gosner stage 44 and 46 by day 27 from the start of the O<sub>3</sub>  
237 exposure. Gosner stage 44 (hereafter, forelimbs stage) is reached when forelimbs  
238 emerge, mouthparts are restructured for terrestrial foraging (teeth present, mouth  
239 fully formed), but the tail stub is still present. Gosner stage 46 is reached when  
240 metamorphosis just complete (hereafter, toadlet stage).

### 241 3.1.4. *Bd* infection

242 *Bd* samples were collected when individuals reached forelimbs stage by  
243 swabbing both feet and the belly with a sterile cotton swab (MW 100–100, Medical  
244 Wire & Equipment) and at toadlet stage by removing a small portion of tissue of  
245 the regressing tail and storing it in 70% ethanol (following Geiger et al. (2013)  
246 who found accumulation of *Bd* on this body area of *A. obstetricans* undergoing  
247 metamorphosis).

248 DNA extractions from swab and tissue samples were performed using  
249 PrepMan Ultra (Applied biosystems) and the amount of *Bd* DNA present in each  
250 sample was measured through a CFX96™ Real-Time PCR Detection System (BIO-  
251 RAD) with a *Bd*-specific Taqman Assay (Boyle et al., 2004). Each 96-well assay  
252 plate included two negative controls and four different standards per duplicate  
253 containing DNA from 100, 10, 1 and 0.1 *Bd* genome equivalents (GE). Each sample  
254 was performed in duplicate and considered *Bd*-positive when the results of the  
255 two replicates were consistent and > 0.1 zoospore genome equivalents. If not, the  
256 sample was re-run a third time and considered positive only if another positive  
257 result occurred.

#### 258 3.1.5. Statistical analyses

259 Difference in the proportion of individuals to reach forelimbs stage at day  
260 27 between the O<sub>3</sub>-filtered air treatment (FA) and the rest of non-filtered air O<sub>3</sub>  
261 treatments was compared with a Fisher's exact test. Differences in *Bd* infection  
262 load across O<sub>3</sub> treatments were analysed using a general lineal model analysis on  
263 log-transformed infection load data from both swabs (forelimbs stage) and tissue  
264 samples (toadlet stage), considering experimental block as a random factor.  
265 Normal probability plots and scatter plots of residuals were used to determine  
266 whether assumptions regarding the distribution of residuals were validated.  
267 Levenne's test was applied to check variance homoscedasticity. Outliers were tested  
268 considering the studentized residuals procedure, but only 5 % of the dataset was  
269 rejected. When significant differences among treatments were detected (p<0.05),  
270 those treatments differing significantly from one another were identified using  
271 Tukey Honestly Significant Difference test (HSD). Differences among O<sub>3</sub> treatments  
272 were also tested with *a priori* planned comparisons considering linear and

273 quadratic responses. Differences in the proportion of survival toadlets among  
274 treatments at the end of the experiment were analysed with a Fisher's exact test.

275 Ozone exposure indexes to relate O<sub>3</sub> levels and effects on wild fauna have  
276 not been defined up to now, although for human health or plant damage a  
277 complete methodology for risk assessment has been developed in the last decade  
278 within the United Nations Air Convention (CLRTAP 2017) and World Health  
279 Organization (WHO 2013). Thus, for the present study, different O<sub>3</sub> exposure  
280 indexes weekly calculated were tested: 24h-mean for the 7-days (24M) before  
281 reaching the forelimbs stage and the toadlet stage, 7-days total accumulated hourly  
282 mean values (AOT00), and accumulated hourly mean values above 20, 30 and 40  
283 nL L<sup>-1</sup> thresholds (AOT20, AOT30, AOT40) for the same 7-days period. The later  
284 indexes are calculated as the sum of the differences between hourly concentrations  
285 greater than each threshold and the threshold over the considered period (CLRTAP,  
286 2017). Due to the nocturnal activity of toads, accumulated indexes included the  
287 whole day period (contrasting with the indexes considered for plants which only  
288 considered the daily hours). However, for comparison between the O<sub>3</sub> levels  
289 during the experiment and previous field data registered at the Sierra de  
290 Guadarrama (Elvira et al., 2016), accumulated AOT40 values for diurnal hours  
291 thorough the whole experiment (48 days) were also calculated.

292 Statistical analyses were carried out using Statistica v.11 (StatSoft Inc.,  
293 USA).

### 294 3.2. Long term field epidemiological study

295 We screened 175 toadlets of spiny common toad (*Bufo spinosus*) for *Bd*  
296 infection that were found dead at Laguna de Pájaros (Peñalara Massif, Sierra de  
297 Guadarrama National Park, Spain) from 2004 to 2012 and preserved in 70%

298 ethanol. All specimens were collected over a two-weeks period every year and had  
299 finished their metamorphosis and, therefore were at the stage at which they were  
300 reliant on atmospheric air for respiration. Toe clips of 17-20 individuals per year  
301 were used for DNA extractions and qPCR analyses were performed as described  
302 above.

303 Data from the CIEMAT monitoring station located less than two kilometres  
304 away from Laguna de Pájaros at Cotos (1850 m a.s.l., 40°49' 31"N, 3°57'40" W,  
305 Sierra de Guadarrama National Park, Spain) were used to record daily O<sub>3</sub>  
306 concentrations (Elvira et al., 2016). Ozone values were registered considering  
307 standardized conditions, following the procedure of the air quality networks. The  
308 O<sub>3</sub> exposure indexes considered where 24 h mean, and the AOT40 index  
309 accumulated for the previous week and for the two weeks preceding the date of  
310 death for each of the 175 individuals. We chose a time span of one week to  
311 calculate O<sub>3</sub> concentrations because in our experimental setup it took a median of 7  
312 days to pass from forelimbs stage to toadlet stage, the breakpoint when usually  
313 internal gills are lost. However, we also calculated the average mean air  
314 temperatures of the three days, instead a week, preceding the date of death of each  
315 metamorphic individual because Fernández-Beaskoetxea et al. (2015) found short-  
316 term impacts of temperature on *Bd* load. After *Bd* infection load were log-  
317 transformed, we fitted different general linear models using JMP 14 (SAS Inc.) to  
318 detect differences in infection intensity among years of collection and across  
319 variation on air temperatures, O<sub>3</sub> raw concentrations and AOT40 index calculated.  
320 Finally, we ordered all possible models that included at least two explanatory  
321 variables according to the corrected AICc. We considered the best explanatory

322 models to be the model with the lowest AICc score, as well as any other models  
323 that differed from the top model by  $< 2$  AICc.

324

## 325 **4. RESULTS**

### 326 4.1. OTC experimental study

327 Accumulated AOT40 indexes though the whole experiment (48 days) were  
328 0, 421, 4.802 and 10.062 nL L<sup>-1</sup> h for FA, NFA, NFA+ and NFA++ respectively.

329 Considering the 24 h-mean index, the value for the different O<sub>3</sub> treatments were  
330 13, 26, 33 and 41 nL L<sup>-1</sup> for FA, NFA, NFA+ and NFA++ respectively.

#### 331 4.1.1. Survival and rate of development

332 At the end of the experiment, individual survival at toadlet stage was in the  
333 range of 83-100% and no significant differences among treatments were found  
334 ( $p=0.2378$ ). Animals from the FA treatment showed the lowest survival,  
335 meanwhile maximum survival was for the AA treatment.

336 From the start of the O<sub>3</sub> exposure, a range of 27 days was necessary for all  
337 the individuals to achieve the forelimb stage. Table 1 shows timetable of  
338 phenological events during the experiment. As expected, water temperature was  
339 lower in the chamberless plots (AA): during May OTC averaged temperature was  
340 1.4 °C higher than AA plots (18.7°C vs 16.9°C), this difference increased till 1.8°C  
341 during June (22.5°C vs 20.7°C). It took up to 48 days from initial exposure until all  
342 individuals reached forelimbs stage: using the mean values of time until  
343 metamorphosis across O<sub>3</sub> treatments, after 21 days of O<sub>3</sub> exposure 6% of the  
344 tadpoles reached forelimbs stage; a maximum 35% of the experimental population  
345 reached this stage between 27 and 34 days of exposure and a cumulative total of  
346 100% reached forelimbs stage after 48 days of O<sub>3</sub> exposure. O<sub>3</sub> treatment affected

347 this phenological pattern. The pollutant tended to delay tadpole phenology:  
348 tadpoles grown under O<sub>3</sub>-filtered air arrived earlier at the forelimbs stage  
349 compared with the other three O<sub>3</sub>-treatments (p=0.0496). Consistently with the  
350 observed pattern of water temperatures, tadpoles grown in the AA plots were the  
351 most delayed (Figure 1).

352 Ozone affected the phenological pattern of the metamorphosis to reach the  
353 toadlet stage (toadlet stage; Figure 1). Individuals grown under clean air (FA)  
354 reached the toadlet stage earlier. At day 41, when maximum peak of the toadlet  
355 stage was observed, 81% of the individuals that completed their metamorphosis  
356 were grown under clean atmospheres (FA), while in the other treatments this  
357 percentage was 47% (p=0.0212). Considering the cumulate values (Figure 1), all  
358 the individuals in the FA plots completed the toadlet stage at this date, but only  
359 55% of the individuals in the AA plots reached this stage.

#### 360 4.1.2. *Bd* infection

361 The *Bd* loads measured prior to the beginning of the experiment were the  
362 maximum values found throughout this work, and the interindividual variation  
363 was low (mean ± SE of log transformed genomic equivalents of zoospores + 1: 4.81  
364 ± 0.12). When tadpoles reached forelimbs stage, *Bd* infection was much lower  
365 (Figure 2). At this stage, although differences between treatments were not  
366 significant, the response of the infection to the pollutant showed minimum values  
367 in the FA treatment and maximum in the NFA treatment.

368 At the toadlet stage, ozone effect on the infection intensity was significant  
369 when mean values of the *Bd* infection for each O<sub>3</sub> treatment was considered: toads  
370 developed under NFA++ treatment presented the highest levels of *Bd* infection  
371 compared with the other three treatments (F<sub>3,37</sub>=3.66, p=0.0210; Figure 2).



372 When the pattern of the *Bd* infection at the toadlet stage was analyzed  
373 (Figure 3), it could be noticed the different pattern of the NFA++ treatment: time  
374 range of *Bd* is wider and kept high values at day 34.

375 Different O<sub>3</sub> indexes were tested to express quantitatively the significant O<sub>3</sub>  
376 effect observed at the toadlet stage (Table 2). The AOT30 accumulated index  
377 presented the best correlation with *Bd* infection, considering both quadratic  
378 (R<sup>2</sup>=0.89) or linear (R<sup>2</sup>=0.50) relationship, compared with other accumulated  
379 indexes based on lower thresholds (AOT20, AOT00) or indexes based in mean  
380 values ( 24h-mean), showing the importance of O<sub>3</sub> values over the O<sub>3</sub>-  
381 preindustrial background on *Bd* spread. However, for quantifying the quadratic  
382 relationship the behaviour of all the indexes tested were similar (R<sup>2</sup> values in the  
383 range 0.71-0.89).

#### 384 4.2. Long term field epidemiological study

385 The O<sub>3</sub> raw concentrations and the AOT40 index of the week before, as well  
386 as the averaged air temperature for the three days preceding the date of death,  
387 were related to *Bd* infection loads of animals found dead, as shown by the increase  
388 in AICc values and the decrease in model weights in Table 3. All these three  
389 variables were included into the top three models that did not differ noticeable  
390 among them, while the year of collection was not (Figure 4). *Bd* infection load has a  
391 negative relationship with averaged air temperature for the three days preceding  
392 the date of death and with O<sub>3</sub> raw concentrations of the week before, while the  
393 AOT40 index for the week before has a positive relationship.

394

395 **5. DISCUSSION**

396 Our results suggest that O<sub>3</sub> concentration can have a significant effect on  
397 both the rate of development in larval amphibians, and on the progression of  
398 parasite infection within those hosts. Increased levels of O<sub>3</sub> delayed the rate of  
399 development, and when they did finally metamorphose, those larvae exposed to  
400 the highest concentration of O<sub>3</sub> had significantly higher levels of *Bd* infection than  
401 other treatments. Individually and combined, these results provide empirical  
402 evidence of two novel separate mechanisms that may help to explain the high  
403 incidence of decline in populations of montane amphibians. Ozone levels at Sierra  
404 de Guadarrama show significant interannual variability, with dry years having the  
405 highest values (Elvira et al., 2016). The range of the diurnal 3-month AOT40 (May –  
406 July) index in the mountains ranged between 6,100 and 30,300 nL L<sup>-1</sup> h; meaning  
407 around 3,050 and 15,150 nL L<sup>-1</sup> h for 1.5-month period. These values are also in  
408 the range of the 48 days-AOT40 index calculated for the present experiment  
409 considering diurnal hours and O<sub>3</sub> treatments (NFA+ and NFA++): 4.802- 10.062 nL  
410 L<sup>-1</sup> h; even some years the values recorded in the mountain exceed those of the  
411 experiment. On the base of this, current O<sub>3</sub> levels at the mountains might be  
412 enough to produce the observed effects here.

413 In the experiment, individuals grown under filtered air developed more  
414 rapidly than those exposed to increased levels of O<sub>3</sub>. Individuals in the filtered air  
415 were the quickest to develop front limbs (forelimbs stage) and this higher rate was  
416 maintained and still evident in the rate at which treatments reached the stage of  
417 tail absorption (toadlet stage). O<sub>3</sub> is known to have significant negative effects on  
418 respiration in a range of taxa, including amphibians. These effects may manifest  
419 themselves in pulmonary function, lung permeability and lung ventilation rate

420 (Mautz et al., 2004, Dohm et al., 2001). Metamorphosis is a period in which the  
421 methods and mechanisms of respiration of developing amphibians alter radically,  
422 via a suite of behavioural, physiological and morphological changes (Duellman and  
423 Trueb, 1994). While amphibians are known to increase the rate of development to  
424 escape stressful, threatening situations (e.g. the presence of predators, pollutants  
425 or parasites), our data suggest that they may not be able to do so when exposed to  
426 increased levels of O<sub>3</sub>. Alternatively, the observed delay in metamorphosis could be  
427 a life-history strategy to maintain the gills for a longer time; at intermediate  
428 Gosner stages individuals are respiring via both larval (gills) and metamorphosed  
429 (cutaneous and lungs) mechanisms. If cutaneous and lung cells affected by  
430 increased O<sub>3</sub> levels at these early stage, it may be a viable strategy to delay  
431 metamorphosis for an extended period. Finally, perhaps reduced oxygen uptake  
432 caused by the exposure to high levels of O<sub>3</sub>, place a rate-limiting step on amphibian  
433 metabolic pathways, thereby dictating the maximum speed on how quickly  
434 development can occur.

435         Our results suggest that exposure to a high level of O<sub>3</sub> can also have  
436 significant effects on the progression of *Bd* infection; those individuals in the  
437 NFA++ treatment had significantly higher infection levels compared to other  
438 treatments. O<sub>3</sub>-driven changes to host-parasite dynamics are likely to be complex  
439 and multi-factorial, given the multiple points at which these changes could be  
440 caused. The reduced developmental rate of exposed individuals would result in  
441 more contact with other infected individuals and with waterborne *Bd* zoospores,  
442 thereby increasing the chances of parasite proliferation and infection progression.  
443 Further, there are multiple ways in which amphibian immune functioning could be  
444 affected by increased O<sub>3</sub> exposure. For example, we are increasingly getting a

445 better understanding of how the host ecology, genetics, and ontogeny and  
446 environment – all of which could be affected by exposure to O<sub>3</sub> over different  
447 timescales - shape amphibian skin microbiome (Bates et al., 2018; Jani and Briggs,  
448 2018; Griffiths et al., 2018), an important part of amphibian immune function in  
449 response to parasites (Bates et al., 2018; Campbell et al., 2019). Further, given the  
450 reduced rate of development of O<sub>3</sub>-exposed animals, the development and function  
451 of the immune system may also be compromised.

452         The complexity of how O<sub>3</sub> could affect multiple elements of amphibian  
453 development, infection, and, ultimately, their population status, is highlighted by  
454 the non-linear way in which *Bd* infection changes with O<sub>3</sub> exposure level. Our data  
455 suggest that *Bd* infection increase in a non-linear way with the level of O<sub>3</sub> exposure.  
456 Both were better explained by a quadratic function, rather than a linear one, with  
457 lower and higher O<sub>3</sub> concentrations being associated with higher infection levels. It  
458 might be possible that, in accordance with results found for the O<sub>3</sub>-plant fungal  
459 pathogen interactions, the direction of the response depends on their relative O<sub>3</sub>  
460 sensitivity. Low O<sub>3</sub> levels would allow the spread of the infection; medium levels of  
461 the pollutant, without being toxic for the host, could deplete the fungi infection due  
462 to its fungicide capacity; meanwhile high ozone levels would be toxic for the host,  
463 weakening the host and favouring fungal infection. This may explain why O<sub>3</sub> shows  
464 a quadratic relationship with the infection loads of toadlets found dead in the field.  
465 While a moderate increment of O<sub>3</sub> values can contribute to reduce *Bd* infection, a  
466 strong increment produces the opposite effect. However, given the data in hand, it  
467 is not possible to identify a mechanism by which this quadratic relationship  
468 between O<sub>3</sub> and *Bd* infection happens, and it would be interesting to use a wider  
469 range of concentrations to identify more precisely how infection changes as a

470 function of O<sub>3</sub> exposure. Amphibian declines were first observed and recorded in  
471 the 1990s, since then multiple drivers of change have been identified and proposed  
472 as either 'the' cause of decline or, more realistically in most cases, one of an  
473 interactive whole causing amphibian populations to reduce in size and viability. In  
474 particular, montane amphibian species and populations have been consistently  
475 observed to decline, and a range of factors have been linked to this reduction.  
476 Climate change, disease, low genetic variability, and habitat modification have all  
477 been proposed as interacting threats to amphibian populations.

#### 478 5.1. Conclusions

479 Our field and experimental research suggests that the level of O<sub>3</sub> is another,  
480 perhaps underreported threat to amphibian populations, either alone or in  
481 combination with other factors. Understanding better how O<sub>3</sub> levels affect  
482 amphibian biology and interact with intrinsic (e.g. development, immune function,  
483 breeding biology, genetic diversity) and extrinsic factors (parasite infection,  
484 habitat degradation, climate) could provide a more holistic understanding of how  
485 amphibian individuals and populations respond to global change and how we may  
486 better mitigate these changes with a view to conserving amphibian populations.

487

#### 488 **6. ACKNOWLEDGEMENTS**

489 C. Serrano helps in the field and in the laboratory. We thank J. A. Vielva and  
490 all people working at Sierra de Guadarrama National Park for continuous support.  
491 We also thank La Higuera-MNCN-CSIC Research Farm support, specially the  
492 valuable participation of José María Gómez Camacho caring the OTC experiment  
493 development. Experiments were conducted in accordance with guidelines and  
494 recommendations outlined by the Consejería de Medio Ambiente de la Comunidad

495 de Madrid. The Consejería de Medio Ambiente of Castilla y León provided permits  
496 for animal collection. Funding was provided by Comunidad Autónoma de Madrid  
497 and the projects AGRISOST (P2018/BAA-4330), EDEN-MED (CGL2017-84687-C2-  
498 1-R) and the Agreement between CIEMAT and MITECO for the definition of critical  
499 levels and loads of atmospheric pollutants.

500

## 501 **7. REFERENCES**

- 502 Adame, J.A., Solé, J.G., 2013. Surface ozone variations at a rural area in the northeast of the Iberian  
503 Peninsula. *Atmos. Pollut. Res.* 4, 130-141.
- 504 Ainsworth, E.A., Lemonnier, P., Wedow, J. M., 2020. The influence of rising tropospheric carbon  
505 dioxide and ozone on plant productivity. *Plant Biol.* 22, 5-11.
- 506 Alonso, R., Elvira, S., González-Fernández, I., Calvete, H., García-Gómez, H., Bermejo, V., 2014.  
507 Drought stress does not protect *Quercus ilex* L. from ozone effects: results from a  
508 comparative study of two subspecies differing in ozone sensitivity. *Plant Biol.* 16, 375-384.
- 509 Bates, K.A., Clare, F.C., O'Hanlon, S., Bosch, J., Brookes, L., Hopkins, K., McLaughlin, E., Daniel, O.,  
510 Garner, T.W.J., Fisher, M.C., Harrison, X.A., 2018. Amphibian chytridiomycosis outbreak  
511 dynamics are linked with host skin bacterial community structure. *Nat. Commun.* 9, 693.
- 512 Bates, K.A., Shelton, J.M.G., Mercier, V.L., Hopkins, K.P., Harrison, X.A., Petrovan, S.O., Fisher, M.C.,  
513 2019. Captivity and infection by the fungal pathogen *Batrachochytrium salamandrivorans*  
514 perturb the amphibian skin microbiome. *Front. Microbiol.* 10, 1834.
- 515 Bergmann, E., Bender, J., Weigel, H.J., 2017. Impact of tropospheric ozone on terrestrial biodiversity:  
516 A literature analysis to identify ozone sensitive taxa. *J. Appl. Bot. Food Qual.* 90, 83-105.
- 517 Bosch, J., Fernández-Beaskoetxea, S., Garner, T.W.J., Carrascal, L.M., 2018. Long-term monitoring of  
518 an amphibian community after a climate change and infectious disease-driven species  
519 extirpation. *Global Change Biol.* 24, 2622-2632.
- 520 Boyle, D.G., Boyle, D.B., Olsen, V., Morgan, J.A.T., Hyatt, A.D., 2004. Rapid quantitative detection of  
521 chytridiomycosis (*Batrachochytrium dendrobatidis*) in amphibian samples using real-time  
522 Taqman PCR assay. *Dis. Aquat. Organ.* 60, 141-148.

523 Calvete-Sogo, H., Elvira, S., Sanz, J., González-Fernández, I., García-Gómez, H., Sánchez-Marín, L.,  
524 Alonso, R., Bermejo-Bermejo, V., 2014. Current ozone levels threaten gross primary  
525 production and yield of Mediterranean annual pastures and nitrogen modulates the  
526 response. *Atmos. Environ.* 95, 197-206.

527 Calvete-Sogo, H., González-Fernández, I., Sanz, J., Elvira, S., Alonso, R., García-Gómez, H., Ibáñez-Ruiz,  
528 M.A., Bermejo-Bermejo, V., 2016. Heterogeneous responses to ozone and nitrogen alter the  
529 species composition of Mediterranean annual pastures. *Oecologia* 181, 1055-1067.

530 Campbell, L.J., Garner, T.W.J., Hopkins, K., Griffiths, A.G., Harrison, X.A., 2019. Outbreaks of an  
531 emerging viral disease covary with differences in the composition of the skin microbiome  
532 of a wild United Kingdom amphibian. *Front. Microbiol.* 10, 1245.

533 Canning, B.J., Hmieleski, R.R., Spannhake, E.W., Jakab, G.J., 1991. Ozone reduces murine alveolar and  
534 peritoneal macrophage phagocytosis: the role of prostanoids. *Am. J. Physiol-Lung Cell. Mol.*  
535 *Physiol.* 261, 277-282.

536 Chen, X., Liu, Y., Lai, A., Han, S., Fan, Q., Wang, X., Lin, Z., Huang, F., Fan, S., 2018. Factors dominating  
537 3-dimensional ozone distribution during high tropospheric ozone period. *Environ. Pollut.*  
538 232, 55-64.

539 CLRTAP, 2017. Mapping critical levels for vegetation. In: Manual on methodologies and criteria for  
540 modelling and mapping critical loads and levels and air pollution effects, risks and trends.  
541 Umweltbundesamt, Berlin.

542 Colette, A., Bessagnet, B., Vautard, R., Szopa, S., Rao, S., Schucht, S., Klimont, Z., Menut, L., Clain, G.,  
543 Meleux, F., Curci, G., Rouïl, L., 2013. European atmosphere in 2050, a regional air quality  
544 and climate perspective under CMIP5 scenarios. *Atmos. Chem. Phys.* 13, 7451-7471.

545 Cristofanelli, P., Bonasoni, P., 2009. Background ozone in the southern Europe and Mediterranean  
546 area: influence of the transport processes. *Environ. Pollut.* 157, 1399-1406.

547 Diaz, J., Ortiz, C., Falcon, I., Salvador, C., Linares, C., 2018. Short-term effect of tropospheric ozone on  
548 daily mortality in Spain. *Atmos. Environ.* 187, 107-116.

549 Dohm, M.R., Mautz, W.J., Looby, P.G., Gellert, K.S., Andrade, J.A., 2001. Effects of ozone on evaporative  
550 water loss and thermoregulatory behavior of marine toads (*Bufo marinus*). *Environ. Res.*  
551 86, 274-286.

552 Dohm, M. R., Mautz, W.J., Andrade, J. A., Gellert, K.S., Salas-Ferguson, L. J., Nicolaisen, N., Fujie, N.,  
553 2005. Effects of ozone exposure on nonspecific phagocytic capacity of pulmonary  
554 macrophages from an amphibian, *Bufo marinus*. Environ. Toxicol. Chem. 24, 205-210.

555 Dohmen, G., 1987. Secondary effects of air pollution: Ozone decreases brown rust disease potential  
556 in wheat. Environ. Pollut. 43, 189-194.

557 Duellman, W.E., Trueb, L., 1994. Biology of Amphibians. Johns Hopkins University Press, Baltimore.

558 EEA, 2011. Air Quality in Europe - 2011 Report Technical Report No. 12/2011. European  
559 Environment Agency, Copenhagen.

560 Elvira, S., González-Fernández, I., Alonso, R., Sanz, J., Bermejo-Bermejo, V., 2016. Ozone levels in the  
561 Spanish Sierra de Guadarrama mountain range are above the thresholds for plant  
562 protection: analysis at 2262, 1850, and 995 m a.s.l. Environ. Monit. Assess. 188, 593.

563 Fernández-Beaskoetxea, S., Carrascal, L.M., Fernández-Loras, A., Fisher, M.C., Bosch, J., 2015. Short  
564 term minimum water temperatures determine levels of infection by the amphibian chytrid  
565 fungus in *Alytes obstetricans* tadpoles. PLoS ONE 10, e0120237.

566 Fisher, M.C., Garner, T.W.J., Walker, S.F., 2009. Global emergence of *Batrachochytrium dendrobatidis*  
567 and amphibian chytridiomycosis in space, time, and host. Ann. Rev. Microbiol. 63, 291-310.

568 Fleming, Z.L., Doherty, R.M., von Schneidemesser, E., Malley, C.S., Cooper, O.R., Pinto, J.P., Colette, A.,  
569 Xu, X., Simpson, D., Schultz, M.G., Lefohn, A.S., Hamad, S., Moolla, R., Solberg, S., Feng, Z.,  
570 2018. Tropospheric ozone assessment report: present-day ozone distribution and trends  
571 relevant to human health. Elem. Sci. Anth. 6, 12.

572 Garner, T.W.J., Rowcliffe, J.M., Fisher, M.C., 2011. Climate change, chytridiomycosis or condition: an  
573 experimental test of amphibian survival. Global Change Biol. 17, 667-675.

574 Geiger, C.C., Schmidt, B.R., Origi, F.C., 2013. Accumulation of the pathogenic fungus  
575 *Batrachochytrium dendrobatidis* on the regressing tail of midwife toads *Alytes obstetricans*  
576 undergoing metamorphosis. Amphibia-Reptilia 34, 255-258.

577 Gilmour, M.I., Park, P., Selgrade, M.K., 1993. Ozone-enhanced pulmonary infection with  
578 *Streptococcus zooepidemicus* in mice. Am. Rev. Respir. Dis. 147, 753-760.

579 González-Fernández, I., Calvo, E., Gerosa, G., Bermejo-Bermejo, V., Marzuoli, R., Calatayud, V., Alonso,  
580 R., 2014. Setting ozone critical levels for protecting horticultural Mediterranean crops: case  
581 study of tomato. Environ. Pollut. 185, 178-187.



582 González-Fernández, I., Elvira, S., Calatayud, V., Calvo, E., Aparicio, P., Sánchez, M., Alonso, R.,  
583 Bermejo-Bermejo, V., 2016. Ozone effects on the physiology and marketable biomass of  
584 leafy vegetables under Mediterranean conditions: Spinach (*Spinacia oleracea* L.) and Swiss  
585 chard (*Beta vulgaris* L. var. *cycla*). *Agr. Ecosyst. Environ.* 35, 215-228.

586 Gosner, K., 1960. A simplified table for staging anuran embryos and larvae with notes on  
587 identification. *Herpetologica* 16, 183-190.

588 Griffiths, S.M., Harrison, X.A., Weldon, C., Wood, M.D., Pretorius, A., Hopkins, K., Fox, G., Preziosi, R.F.,  
589 Antwis, R.E., 2018. Genetic variability and ontogeny predict microbiome structure in a  
590 disease-challenged montane amphibian. *ISME J.* 12, 2506-2517.

591 Heck, W.W., Taylor, O.C., Adams, R., Bingham, G., Miller, H., Preston, E., Weinstein, L., 1982.  
592 Assessment of crop loss from ozone. *J. Air Pollut. Control Assoc.* 32, 353-361.

593 Hite, J.L., Bosch, J., Fernández-Beaskoetxea, S., Medina, D., Hall, S.R., 2016. Joint effects of habitat,  
594 zooplankton, host stage structure, and diversity on amphibian chytrid. *P. Roy. Soc. B*, 283,  
595 20160832.

596 Hollingsworth, J.W., Kleeberger, S.R., Foster, W.M., 2007. Ozone and pulmonary innate immunity.  
597 *Proc. Am. Thorac. Soc.* 4, 240-246.

598 Hu, L., Jacob, D.J., Liu, X., Zhang, Y., Zhang, L., Kim, P.S., Sulprizio, M.P., Yantosca, R.M., 2017. Global  
599 budget of tropospheric ozone: evaluating recent model advances with satellite (OMI),  
600 aircraft (IAGOS), and ozone sonde observations. *Atmos. Environ.* 167, 323-334.

601 Jani, A.J., Briggs, C.J., 2028. Host and aquatic environment shape the amphibian skin microbiome but  
602 effects on downstream resistance to the pathogen *Batrachochytrium dendrobatidis* are  
603 variable. *Front. Microbiol.* 9, 487.

604 Lacroix, G., Lambre, C., 1998. Ozone and the immune system. *Rev. Mal Respir.* 15, 699-711.

605 Lefohn, AS, Malley, CS, Smith, L, Wells, B, Hazucha, M, Simon, H, Naik, V, Mills, G, Schultz, MG,  
606 Paoletti, E, De Marco, A, Xu, X, Zhang, L, Wang, T, Neufeld, HS, Musselman, RC, Tarasick, D,  
607 Brauer, M, Feng, Z, Tang, H, Kobayashi, K, Sicard, P, Solberg, S and Gerosa, G., 2018.  
608 Tropospheric ozone assessment report: global ozone metrics for climate change, human  
609 health, and crop/ecosystem research. *Elem. Sci. Anth.* 6, 28.

610 Lippmann, M., 1989. Health effects of ozone a critical review. *Japca* 39, 672-695.

611 Marzuoli, R., Bussotti, F., Calatayud, V., Calvo, E., Alonso, R., Bermejo, V., Pollastrini, M., Monga, R.,  
612 Gerosa, G., 2018. Dose-response relationships for ozone effect on the growth of deciduous  
613 broadleaf oaks in mediterranean environment. *Atmos. Environ.* 190, 331-341.

614 Mautz, W.J., Dohm, M.R., 2004. Respiratory and behavioral effects of ozone on a lizard and a frog.  
615 *Comp. Biochem. Phys A* 139, 371-377.

616 Millán, M., Salvador, R., Mantilla, E., Kallos G., 1997. Photo-oxidant dynamics in the western  
617 Mediterranean in summer: results from European research projects. *J. Geophys. Res.* 102,  
618 8811-8823.

619 Myers, N., Mittermeier, R.A., Mittermeier, C.G., Da Fonseca, G.A., Kent, J., 2000. Biodiversity hotspots  
620 for conservation priorities. *Nature* 403, 853.

621 Nopmongcol, U., Jung, J., Kumar, N., Yarwood, G., 2016. Changes in US background ozone due to  
622 global anthropogenic emissions from 1970 to 2020. *Atmos. Environ.* 140, 446-455.

623 Ortíz-Santaliestra, M.E., Fisher, M.C., Fernández-Beaskoetxea, S., Fernández-Benéitez, M.J., Bosch J.,  
624 2011. Ambient ultraviolet B radiation decreases the prevalence of infection by  
625 *Batrachochytrium dendrobatidis* in two amphibian species. *Conserv. Biol.* 25, 975-982.

626 Ribas, A., Peñuelas, J., 2006. Surface ozone mixing ratio increase with altitude in a transect in the  
627 Catalan Pyrenees. *Atmos. Environ.* 40, 7308-7315.

628 Saavedra, S., Rodríguez, A., Taboada, J.J., Souto, J.A., Casares, J.J., 2012. Synoptic patterns and air  
629 mass transport during ozone episodes in northwestern Iberia. *Sci. Total Environ.* 441, 97-  
630 110.

631 Sanz, J., Bermejo-Bermejo, V., Muntifering, R.B., Gonzalez-Fernandez, I., Gimeno, B.S., Elvira, S.,  
632 Alonso, R., 2011. Plant phenology, growth and nutritive quality of *Briza maxima*: responses  
633 induced by enhanced ozone atmospheric levels and nitrogen enrichment. *Environ. Pollut.*  
634 159, 423-430.

635 Sutton, M.A., Howard, C.M., Erisman, J.W., Billen, G., Bleeker, A., Grennfelt, P., Grinsven, H.V.,  
636 Grizzetti, B., 2011. *The European Nitrogen Assessment*. Cambridge University Press.

637 Tiedemann, Av., Weigel, H.J., Jäger, H.J., 1991. Effects of open-top chamber fumigations with ozone  
638 on three fungal leaf diseases of wheat and the mycoflora of the phyllosphere. *Environ.*  
639 *Pollut.* 72, 205-224.

640 Tighe, R.M., Wheeler, J., Hollingsworth, J.W., 2015. Air pollution and immune function. In: Air  
641 Pollution and Health Effects. Springer, London.

642 U.S. EPA, 2013. Integrated Science Assessment (ISA) of ozone and related photochemical oxidants  
643 (Final Report, Feb 2013). U.S. Environmental Protection Agency, Washington, DC,  
644 EPA/600/R-10/076F.

645 Villanueva, F., Tapia, A., Notario, A., Albaladejo, J., Martínez, E., 2014. Ambient levels and temporal  
646 trends of VOCs, including carbonyl compounds, and ozone at Cabañeros National Park  
647 border, Spain. *Atmos. Environ.* 85, 256–265.

648 Voyles, J., Young, S., Berger, L., Campbell, C., Voyles, W.F., Dinudom, A., Cook, D., Webb, R., Alford,  
649 R.A., Skerratt, L.F., Speare, R., 2009. Pathogenesis of chytridiomycosis, a cause of  
650 catastrophic amphibian declines. *Science* 326, 582–585.

651 Walker, S.F., Bosch, J., Gomez, V., Garner, T.W.J., Cunningham, A.A., Schmeller, D.S., Ninyerola, M.,  
652 Henk, D., Ginestet, C., Arthur, C.P., Fisher, M.F., 2010. Factors driving pathogenicity versus  
653 prevalence of amphibian panzootic chytridiomycosis in Iberia. *Ecol. Lett.*, 13, 372-382.

654 WHO, 2013. Review of evidence on health aspects of air pollution- REVIHAAP Project, Copenhagen:  
655 WHO Regional Office for Europe.

656 Young, P.J., Archibald, A.T., Bowman, K.W., Lamarque, J., Naik, V., Stevenson, D.S., Tilmes, S.,  
657 Voulgarakis, A., Wild, O., Bergmann, D., Cameron-Smith, P., Cionni, I., Collins, W.J., Dalsoren,  
658 S.B., Doherty, R.M., Eyring, V., Faluvegi, G., Horowitz, L.W., Josse, B., Lee, Y.H., Mackenzie,  
659 I.A., Nagashima, T., Plummer, D.A., Righi, M., Rumbold, S.T., Skeie, R.B., Shindell, D.T., Strode,  
660 S.A., Sudo, K., Szopa, S., Zeng, G., 2013. Pre-industrial to end 21st century projections of  
661 tropospheric ozone from the Atmospheric Chemistry and Climate Model Intercomparison  
662 Project (ACCMIP). *Atmos. Chem. Phys.* 13, 2063–2090.

663 **Table 1.** Timetable of the experimental events and proportion of individuals at the  
 664 two considered development stages (the cumulative proportion of individual at  
 665 that stage is given in brackets).

666

667	event	date	days after	forelimbs	toadlet
668			start exposure	stage	stage
669				(Gosner 44)	(Gosner 46)
670					
671	Start O <sub>3</sub> exposure	28 April	1	0	0
672	Sampling <i>Bd</i>	19 May	21	0.06 (0.06)	0
673	Sampling <i>Bd</i>	25 May	27	0.25 (0.31)	0.04 (0.04)
674	Sampling <i>Bd</i>	1 June	34	0.34 (0.65)	0.33 (0.37)
675	Sampling <i>Bd</i>	8 June	41	0.25 (0.90)	0.42 (0.79)
676	Last sampling <i>Bd</i>	15 June	48	0.10 (1.00)	0.21 (1.00)

677

678 **Table 2.** Quadratic and linear relationships between *Bd* infection (dependent  
679 variable in log transformed genomic equivalents of zoospores) at the toadlet stage  
680 and different O<sub>3</sub> exposure indexes. AOT30 index is calculated as the sum of the  
681 differences between hourly concentrations greater than 30 nL L<sup>-1</sup> and 30 nL L<sup>-1</sup>  
682 over the weekly period (nL L<sup>-1</sup> h). AOT00 index is the sum of accumulated hourly  
683 values over the weekly period (nL L<sup>-1</sup> h). 24h mean is the O<sub>3</sub> daily average for the  
684 week (nL L<sup>-1</sup>).

685

686	O <sub>3</sub> index		R <sup>2</sup>
687		quadratic	
688	AOT30	$y = 5E-07 x^2 - 0.001 x + 1.081$	0.89
689	AOT00	$y = 2E-07 x^2 - 0.002 x + 3.912$	0.71
690	24h mean	$y = 0.006 x^2 - 0.271 x + 3.790$	0.72
691		linear	
692	AOT30	$y = 4E-04 x + 0.635$	0.50
693	AOT00	$y = 2E-04 x + 0.037$	0.33
694	24h mean	$y = 0.040 x + 0.026$	0.35

695 **Table 3.** Candidate general linear models to determine the best predictors of *Bd*  
696 infection loads of 175 toadlets of spiny common toads found dead between 2004  
697 and 2012 at Sierra de Guadarrama National Park by the year of collection, the  
698 averaged values of air temperature (Temp), O<sub>3</sub> raw values and the AOT40 index  
699 recorded in the area for the three days (temperature) and the week (ozone values)  
700 preceding the date of death of each individual. AOT40 index is calculated as the  
701 sum of the differences between hourly concentrations greater than 40 nL L<sup>-1</sup> and  
702 40 nL L<sup>-1</sup> over the weekly period (nL L<sup>-1</sup> h).

703

704	Rank	model	AICc	R <sup>2</sup>	ΔAICc	k	weight
705	1	O <sub>3</sub> + Temp	385	0.26	0.0	5	0.4
706	2	AOT40 + Temp	385	0.26	0.1	5	0.4
707	3	O <sub>3</sub> + AOT40	386	0.23	1.6	5	0.2
708	4	O <sub>3</sub> + AOT40 + Temp	389	0.25	4.2	6	0
709	5	year + Temp	390	0.23	5.6	12	0
710	6	year + AOT40	391	0.22	6.4	12	0
711	7	year + O <sub>3</sub>	391	0.23	6.8	12	0
712	8	year + AOT40 + Temp	395	0.23	10.8	13	0
713	9	year + O <sub>3</sub> + Temp	396	0.23	11.8	13	0
714	10	year + O <sub>3</sub> + AOT40 + Temp	401	0.23	16.0	14	0

715

716 Figure 1. Proportion of individuals (accumulated values across all replicates)  
717 reaching the Gosner stage 44 (all four limbs developed; A) and the toadlet stage (B)  
718 per O<sub>3</sub> treatment according to the number of days from the beginning of the  
719 experiment. FA, Filtered Air; NF, Non Filtered Air; NFA+, Non Filtered Air +20 nL L<sup>-1</sup>  
720 of O<sub>3</sub>; NFA++, Non Filtered Air +40 nL L<sup>-1</sup> of O<sub>3</sub>; AA, Ambient plots.

721

722 Figure 2. The effect of ozone on *Bd* infection (mean ± SE of log transformed  
723 genomic equivalents of zoospores) at the Gosner stage 44 (all four limbs  
724 developed; grey bars) and the toadlet stage (black bars) per O<sub>3</sub> treatment. FA,  
725 Filtered Air; NFA, Non Filtered Air; NFA+, Non Filtered Air +20 nL L<sup>-1</sup> of O<sub>3</sub>; NFA++,  
726 Non Filtered Air +40 nL L<sup>-1</sup> of O<sub>3</sub>; AA, Ambient plots. Different letters indicate  
727 statistically significant differences (p<0.05).

728

729 Figure 3. Mean *Bd* infection ± SE of log transformed genomic equivalents of  
730 zoospores for individuals in the toadlet stage at the different sampling dates and  
731 O<sub>3</sub> treatments. FA, Filtered Air; NF, Non-Filtered Air; NFA+, Non-Filtered Air +20  
732 nL L<sup>-1</sup> of O<sub>3</sub>; NFA++, Non-Filtered Air +40 nL L<sup>-1</sup> of O<sub>3</sub>; AA, Ambient plots.

733

734 Figure 4. Predicted values (marginal model plots) of *Bd* infection loads of toadlets  
735 of spiny common toads found dead between 2004 and 2012 at Sierra de  
736 Guadarrama National Park by the three top models relating infection and the  
737 averaged values of air temperature, O<sub>3</sub> and AOT40 recorded in the area for the  
738 three days (temperature) and the week (ozone values) preceding the date of death.

739