

# Climate forcing of an emerging fungal pathogen across a multihost community

**Frances C. Clare<sup>1,3\*</sup>, Julia B. Halder<sup>3</sup>, Olivia Daniel<sup>2</sup>, Jon Bielby<sup>1</sup>,  
Mikhail A. Semenov<sup>4</sup>, Thibaut Jombart<sup>3</sup>, Adeline Loyau<sup>5,6,7</sup>, Dirk  
S. Schmeller<sup>5,6</sup>, Andrew A. Cunningham<sup>1</sup>, Marcus Rowcliffe<sup>1</sup>,  
Trenton W. J. Garner<sup>1</sup>, Jaime Bosch<sup>8</sup>, Matthew C. Fisher<sup>3\*</sup>**

1. Institute of Zoology, Zoological Society of London, Regent's Park, London, NW1 4RY, UK

2. Department of Life Sciences, Silwood Park Campus, Imperial College London SL5 9PU, UK

3. Department of Infectious Disease Epidemiology, Imperial College London W2 1PG, UK

4. Computational and Systems Biology, Rothamsted Research, West Common, Harpenden, Hertfordshire,  
AL5 2JQ

5. Université de Toulouse; UPS, INPT; EcoLab (Laboratoire Ecologie Fonctionnelle et Environnement); 118  
route de Narbonne, 31062 Toulouse, France

6. Helmholtz Centre for Environmental Research- UFZ, Department of Conservation Biology, Permoserstr.  
15, 04318 Leipzig, Germany

7. Helmholtz Centre for Environmental Research - UFZ, Department of System Ecotoxicology, Permoserstr.  
15, 04318 Leipzig, Germany

8. Departamento de Biodiversidad y Biología Evolutiva, Museo Nacional de Ciencias Naturales, CSIC, Jose  
Gutierrez Abascal, 2 28006 Madrid, Spain

## Corresponding authors:

\*Frances C. Clare, Institute of Zoology, Zoological Society of London, Regent's Park, London, NW1 4RY,  
UK; +44 2074496626; [frances.clare@ioz.ac.uk](mailto:frances.clare@ioz.ac.uk)

\*Matthew C. Fisher, Dpt. Infectious Disease Epidemiology, Norfolk Place, London W2 1PG; +44 2073727733  
[matthew.fisher@imperial.ac.uk](mailto:matthew.fisher@imperial.ac.uk)

**Keywords:** climate change, chytridiomycosis, multi-host communities, epidemiology, mountain  
ecosystems, host x pathogen x environment interactions

---

## Summary

Changes in the timings of seasonality as a result of anthropogenic climate change are predicted to occur over the coming decades. While this is expected to have widespread impacts on the dynamics of infectious disease through environmental forcing, empirical data is lacking. Here, we investigated whether seasonality affected susceptibility to infection by the emerging fungal pathogen *Batrachochytrium dendrobatidis* (*Bd*) across a montane community of amphibians that are suffering declines and extirpations as a consequence of this infection. We found a robust temporal association between the timing of the spring thaw and *Bd* infection in two host species, where we show that an early onset of spring forced high prevalences of infection. A third highly susceptible species (the midwife toad *Alytes obstetricans*) maintained a high prevalence of infection independent of seasonality. Our data show that perennially overwintering midwife toad larvae act as a year-round reservoir of infection with seasonality determining the extent to which infection spills over into sympatric species. We used future temperature projections based on global climate models to demonstrate that the timing of spring thaw in this region will advance markedly by the 2050's, indicating that climate change will further force the severity of infection. Our findings of the effect of seasonality on multi-host infection dynamics show that the community-level impact of fungal infectious disease on biodiversity will need to be re-evaluated in the face of climate change.

## Introduction

Climate change is likely to influence infectious disease dynamics, with many pathogens, especially those with complex life cycles or those infecting ectothermic hosts, predicted to increase in severity or range as the earth continues to warm (1-3). *Batrachochytrium dendrobatidis* (*Bd*), one of two fungi known to cause amphibian chytridiomycosis, is associated with the decline and extinction of amphibians worldwide (4, 5). In common with other infections, it is widely assumed that climate change has facilitated epizootics of chytridiomycosis, thus allowing *Bd* to establish in naïve ecosystems (6-8).

A link between climatic variables, host phenology (the timing of recurring natural phenomena) and the population level impact of *Bd* has been investigated by both *in situ* (6, 7), and *ex situ* (8-10) studies. However, the hypothesis that climate change dictates *Bd* infection dynamics in nature has not been proven. This is because the key *in situ* studies to date have confounded two variables, pathogen introduction and environmental variation (6, 7), and the required longitudinal epidemiological studies in established diseased ecosystems have not yet been undertaken (11, 12). Further, *Bd* is a generalist pathogen (13), exhibiting broad variation in its ability to infect and cause disease across species (14). Within multi-species amphibian assemblages, different host species exhibit a range of responses to pathogen exposure. These responses include resistance to infection (diluters of infection), infection tolerance (reservoirs of infection), and variation in susceptibility to lethal disease (15, 16). Yet, studies to date have focussed on the most readily infected species assuming homogeneous host response over time, and data that addresses the medium to long-term temporal impact of *Bd* across all members of a host community in concert with local climatic data, is absent.

Climate variation is pronounced in montane systems, and it is in these environments that epizootics of chytridiomycosis predominantly occur (17-20) presenting an opportunity to untangle the complex relationship between disease dynamics and climate. One such environment, the Pyrenean mountain range, contains many lakes housing multi-species amphibian assemblages within an expanding epizootic of *Bd* infection (20). Across the core outbreak region of the Western Pyrenees, the midwife toad (*Alytes obstetricans*) experiences annual mass mortality due to chytridiomycosis. Two other anuran species also commonly breed in this area, the common toad (*Bufo spinosus*, previously known as *Bufo bufo* (21)) and the common frog (*Rana temporaria*).

Across Europe, the prevalence of *Bd* within affected common toads is usually low (14), with the majority of individuals tolerating infection (22). In comparison, the common frog is considered to be resistant to *Bd* infection (14, 23), and until now there has been no evidence of disease in this species. As a result of the differences that these three species exhibit in their response to exposure to *Bd*, they constitute an ideal study-system for exploring the temporal trends in pathogen infection at a community level. To understand the inter-relationship between infection, community and climate, we investigated seasonality in this montane ecosystem and studied how this affects infection dynamics at the present, and when extrapolated into the future using downscaled outputs of global climate change models.

## Results and Discussion

We monitored amphibians at a key remote infected site, Lac Arlet (altitude 1,986 m absl), in the French Pyrenees over seven years. Across this period we found a robust temporal link between disease dynamics (Fig. 1a & b) and the timing of spring across this community of amphibians (Fig. 1c & d). We uncovered a significant effect of the onset of spring on the prevalence of *Bd* infection in both *B. spinosus* (slope = -0.1000;  $z = -4.715$ , d.f. = 149,  $p < 0.001$ ; adjusted  $R^2 = 0.19$ ), and *R. temporaria* (slope = -0.039,  $z = -4.356$ , d.f. = 207,  $p < 0.001$ ; adjusted  $R^2 = 0.08$ ), with early spring onset resulting in a higher prevalence of *Bd* infection (Fig. 1d). The highly susceptible species, *A. obstetricans*, maintained a high prevalence of infection throughout the study independent of the onset of spring for both overwintering tadpoles and metamorphs ( $p = 0.09$ ). The timing of the end of season showed little variation between years (Fig. 1c; Table 1), with the corollary that the length of the amphibian activity period was increased in years with an early spring onset.

The fact that species previously resistant to infection and disease alters in different environmental / climatic contexts illustrates the importance of understanding community-level dynamics when considering the impacts of infections. Our data show that a species thought to be highly resistant to infection and disease can suffer mortality (Fig. 2a); prior to the current study, *R. temporaria* has rarely been found to be infected with *Bd* (14) and has never been observed to suffer chytridiomycosis despite widespread surveillance across Europe. However, we detected widespread infection in this species, along with mortality due to chytridiomycosis, when seasonal onset was early. We also found concurrent temporal changes in the prevalence of *Bd* infection in the more susceptible species, *B. spinosus*, which

became locally extinct in 2013 (Fig. 1a & b). We believe that the abrupt decline of *B. spinosus* was disease-driven, due to the high prevalence of infection and widespread mortality detected in metamorphs in the years leading to its disappearance. The number of *A. obstetricans* OW larvae has also declined steadily at Lac Arlet during the seven-year course of our study alongside high recorded mortality (Fig 2b; Table 2), showing that mortality due to chytridiomycosis is leading to synchronous multi-species declines across this site.

Our data suggest that the susceptible and highly parasitised *A. obstetricans* are acting as the key biological reservoir of *Bd* in this system, and that seasonality is determining the extent to which infection spills over into sympatric species. At the altitude of Lac Arlet, *A. obstetricans* tadpoles over-winter for a number of years before completing development. The tadpole stage of the majority of amphibians rarely suffer mortality or visible effects of *Bd* infection (24, 25) and these life-history traits result in over-wintering tadpoles playing a leading a role in the maintenance of *Bd* across seasons, as has been proposed previously for other systems (19, 26). The yearly changes in infection prevalence that we detected in hosts that are less susceptible to infection could imply that the density of one generally heavily infected species, *A. obstetricans*, drives infection levels within this system. We detected the highest abundance of *A. obstetricans* metamorphs in 2010 and, in that year we also saw the highest proportional mortality, with almost half of all *A. obstetricans* individuals encountered being dead. It is possible that the sharp increase in prevalence of infection in 2010 that we detected in both *B. spinosus* and *R. temporaria* was a direct result of the observed increase in the number of infected *A. obstetricans* in that year. However, there was no significant association between the number of deceased *A. obstetricans* mortalities seen each year and the prevalence of infection in either species. Further, the prevalence of infection in both *B. spinosus* and *R. temporaria* continued to increase the following year (Fig.1a), despite a decrease in the abundance of *A. obstetricans*. This contraindicates the hypothesis that the density of *A. obstetricans* is the sole driver of force of infection upon sympatric species.

To assess the impact of climate change on onset of spring, the optimal time period over which mean air temperature is most closely related to time of spring thaw was estimated (supplementary Fig. 1). For 2007-2015, mean temperature over days 83-153 (a 70 day period centred on day 118 of the year) was found to be the best predictor of onset of spring, explaining 73% of the variance. For every 1 degree increase in the mean temperature over this time, onset of spring is estimated to be 10 days earlier (Fig. 2c).

We then used the LARS-WG weather generator as a downscaling technique to generate local-scale climate scenarios for the site, based on projections from global climate models from the CMIP5 multi-model ensemble under the representative concentration pathway (RCP) 8.5 (greenhouse gas emissions continue rising over 21st Century) (27) and modelled using local weather data (Fig. 2c). To capture uncertainty in the CMIP5 climate projections we selected two global climate models, GISS-E2 and HadGEM2, with low and high climate sensitivities respectively, which thus predict lesser and greater amounts of warming for the region. The predictions all indicate that an early onset of spring in the western Pyrenees will become commonplace by the 2050s due to global warming (Fig. 2d). Over days 83-153 of the year, the mean daily temperature is projected to be around 5.5°C by GISS-E2 and 5.8°C by HadGEM2 in the 2050s (medians of 100 years plausible weather generated by LARS-WG) (Fig. 2d). This corresponds, under the current relationship, with an onset of spring around day 130-133, comparable to that seen in 2011. By the 2090s, the median GISS-E2 scenario for this time is 6.5°C, potentially indicating an onset of spring around day 122. The median HadGEM2 scenario is 7.8°C; since this is out of the range of observed temperatures (with the rest of the year also being substantially warmer) we make no estimated projection of onset of spring under this scenario. LARS-WG output across the winter months for the 2090s for HadGEM2 suggests that daily mean temperature will rarely drop below 0°C (daily minimum temperature may be below freezing for 3 months, compared to currently around 6 months), which may lead to a short, punctuated, or non-existent duration of ice cover (supplementary Fig. 2). If the trends that we have established here continue, increases in amphibian activity periods are predicted to correspond to increases in infection levels across these species, which could have profound effects on populations of hosts we currently consider to be resistant or tolerant of infection.

The mechanisms underpinning the seasonal forcing of infection that we observe are unknown. A broad range of biotic and abiotic factors are influenced by seasonality, and it is probable that many factors acting on both the host and the pathogen are driving the seasonal prevalence of infection in the amphibians studied. On one hand, abiotic drivers include increased temperature volatility causing a reduction in temperature acclimation of host resistance to infection (28), and patterns of disease are known to be also exacerbated as hosts are moved outside of their temperature norms (9). On the other hand, biotic factors such as the density and species complement of aquatic microfauna present at our research site are known to modulate the risk of infection (29). Aquatic microfauna have seasonal

peaks and dips in abundance throughout the spring and summer corresponding to the local availability of nutrients (30), therefore, changes in seasonality may affect the abundance of the microfauna, impacting the removal of *Bd* zoospores and force of infection. When complex biotic and abiotic drivers such as these interact, nonlinear responses are expected and the uncertainty associated with predicting future trends in this, and other montane, systems, will likely be high. However, regardless of the underlying mechanisms, our findings robustly show that seasonality drives synchronous infection dynamics in *B. spinosus* and *R. temporaria*, despite previous research indicating a clear difference in susceptibility to *Bd* infection between these species (14, 31). More broadly, we have shown that failing to include environmental information may undermine our understanding of how pathogens spread and persist within host communities. Identifying differences in infection levels across species under different environmental conditions is therefore central to understanding disease ecology within multi-host communities, especially where changes in climate are predicted to exacerbate the impact of emerging infectious disease leading to further losses of biodiversity.

## Methods

The study was conducted at Lac Arlet (Longitude: 0°36'54.12"W, Latitude: 42°50'24.20"N) in the Pyrenean National Park between 2007 and 2014 (with dates of thaw up to 2015). This lake sits at an altitude of 1986 m and encompasses an area of 2.7ha (Fig. 3).

### *Prevalence and intensity of Bd infection*

All three species of amphibian present at Lac Arlet were sampled: emerging metamorphs of *B. spinosus* and *R. temporaria* and two life stages of *A. obstetricans*, over-wintered (OW) tadpoles and recent metamorphs; OW tadpoles are those which have spent at least one winter in the lake before completing metamorphosis. To sample the animals, sterile rayon-tipped swabs (MWE medical wire) were gently rotated (10 times per individual) over the mouthparts of OW tadpoles, and used to swab the hind legs, feet and pelvic patch (five swipes per area, with the swab rotated between each area) of each recent metamorph. All swabs were stored in dry tubes at 4°C until processing. Sampling of tadpoles was conducted each year in July. Metamorphs were sampled in August during the height of metamorphic emergence. In 2007, toe clips were taken from metamorphic *A. obstetricans* instead of skin

swabs; a 2-3mm clip was cut from a single hind toe using a sterile scalpel blade. These toe clips were fixed in 70% ethanol.

We followed the protocol of Boyle et al. (32), to quantify *Bd* prevalence and intensity of infection, as assessed by quantitative PCR (qPCR). To avoid inhibition, all extractions were diluted 1:10 prior to qPCR; therefore results were multiplied by 10 in order to determine the true value. We defined infection intensity as the number of *Bd* zoospore genomic equivalents (GE) per swab. All samples were run in duplicate, and a sample was assigned a positive reading if both wells amplified and an average estimate of 0.1GE or above was produced when comparing the sample to the curve generated by the standards. Samples were repeated up to three times if only one well amplified, after which time they were assigned a negative reading if both wells failed to amplify.

### ***Population counts***

Counts of newly emerged live and dead metamorphic *A. obstetricans* were conducted twice during each metamorph emergence period each year from 2010 to 2014. Searches were carried out around the entire circumference of the lake, from the shallows of the lakes (<0.5 metres in depth) to a 1.5m distance from the water's edge onto dry land, including under movable rocks. Any deceased metamorphs found were removed so as not to be counted a second time on subsequent visits. We assumed that live animals would either disperse from the water's edge within a few days of emergence or die, and would therefore not be counted again in subsequent live estimates. Approximate counts (<100; 100-1000; >1000) were made for both newly emerged *R. temporaria* and *B. spinous* metamorphs each year, during the above surveying time. This measure was also applied to *A. obstetricans* OW larvae by scanning a 2m area of the water, from the water's edge.

### ***Diagnosis of chytridiomycosis in R. temporaria***

Owing to the high prevalence of *Bd* that we observed infecting *R. temporaria*, we decided to investigate whether individuals were suffering chytridiomycosis due to *Bd* infection, something which has not been shown before. Recently deceased, fresh *R. temporaria* metamorphs found around the edge of the lake were collected and fixed in 10% neutral buffered formalin. After fixation, the lower half of the each body (including pelvic region) and the front legs were processed for histopathological examination. Three levels per section were prepared, using a standard Haematoxylin & Eosin stain. Histological sections were

examined microscopically to determine the presence or absence of the disease chytridiomycosis.

#### *Determining length of season*

Lac Arlet water temperature was measured every half an hour throughout the study period using a data logger (HOBO Water Temperature Pro v2 Data Logger - U22-001). The logger was positioned two meters from the shore, approximately half a meter below the water surface, attached to a large rock with non-perishable wire. For each year, the date of the onset of spring and the date of the presumed end of the amphibian active season was recorded. The onset of spring was defined as the first day of the year (00:00 – 23:59) with a mean water temperature above 1°C, which remained so until the winter. The end of the active season was defined as the first day in the second half of the year when the mean water temperature dropped below 5°C, as European amphibians will often enter hibernation around this temperature (33). This allowed the length of the active season (total number of possible ‘active days’ for amphibians) to be calculated by subtracting the Julian date of the onset of spring from the Julian date of the end of season.

#### *Establishing the relationship between air temperature and lake thaw (onset of spring)*

To project the impact of rising temperatures, we first quantified a linear relationship between air temperature and time of thaw, which corresponds to onset of spring for this ecological system. This is consistent with findings from other studies showing air temperature to be highly informative for thawing of many lakes (34), even if temperatures are measured some distance from the lake. To establish this relationship for this site, daily weather data (maximum and minimum temperatures, precipitation) were obtained from the nearest meteorological station to Lac Arlet, Canfranc Los Arañones, 13km south-east of Lac Arlet. This is situated at 1160 metres above sea level, 826 metres below Lac Arlet. Hence, a correction of -5.29 °C was applied to the air temperatures to account for tropospheric temperature decrease with altitude, in accordance with previous studies (35). These adjusted temperature data along with daily precipitation for 1995-2015 were also used as baseline input for LARS-WG.

To find the optimal time period over which these air temperatures are most predictive for ice thawing time, and to quantify that relationship, linear regressions were performed between air temperature and thaw dates for 2007-2015 inclusive. Mean daily air temperature was calculated over time periods of lengths varying from 21-81 days in 10-day increments,

centred on days 30-170 of the year. The time period with the best predictive power for onset of spring, as judged by R-squared value and standard model checking plots of the linear regression, was chosen. Analysis was performed in R version 3.2.3.

We used the LARS-WG weather generator as a downscaling technique (36) to generate local-scale climate scenarios, based on climate projections from global climate models (GCMs) from the CMIP5 multi-model ensemble used in the latest IPCC Assessment Report 5 (AR5) (27). To capture uncertainty in the CMIP5 climate projections we selected two GCMs with low, (GISS-E2), and high, (HadGEM2), climate sensitivities (37), which thus predict lesser and greater degrees of warming for this region. This allowed us to quantify uncertainty in predictions of the onset of spring under climate change. We generated 100 years of daily plausible weather for the periods 2050s and 2090s under Representative Concentration Pathway (RCP) 8.5. The mean temperatures across the time period ascertained as most predictive for lake thaw were calculated from each of the 100 years of synthetic daily weather, for each GCM and time period combination.

### *Statistical analysis*

All statistical analyses were carried out using the statistical software package 'R', version 3.2.3. All *Bd* DNA values (GE) were rounded to the nearest whole number and treated as count data. Any value of 0.1 to 0.9 was assigned a value of 1. Negative binomial regression models (function `glm.nb` from the R-package MASS) were used to look for any differences in the intensity of infection (GE values) between years and species. Likelihood ratio tests were used to assess the significance of predictor variables and of differences between factor levels within predictors. Where more than three factor levels remained significant, Tukey post-hoc tests (function `glht` from the package multcomp) were applied to allow pairwise comparisons. Fisher's Exact Test was used to compare differences in the prevalence of infection in each of *A. obstetricans* metamorphs and tadpoles, over the years 2007 (2008 for tadpoles) to 2014; in *B. spinosus* compared to *R. temporaria* over the years 2008 to 2012 and between each of *B. spinosus* and *R. temporaria* compared to *A. obstetricans* metamorphs over the years 2008-2014 (2008 -2012 for *B. spinosus*). Pearson Correlation tests were used to determine whether there was a correlation between the prevalence of infection in both *B. spinosus* and *R. temporaria* and, 1) the mean infection intensity (GE) in *A. obstetricans* metamorphs, 2) the number of dead *A. obstetricans* metamorphs encountered each year.

Using the temperature data we were able to determine the date of the spring onset for all years apart from 2010, due to a failure of the datalogger resulting in missing data. However, a strong association was seen between the onset of spring and the date at which the first *A. obstetricans* metamorph was seen in the years excluding 2010 ( $t = 7.724$ ;  $p = 0.005$ , adjusted  $R^2 = 0.94$ ). We used this strong association to predict the onset of spring in 2010 and included the predicted value in all further analyses. The onset of season (mean Julian days=150, SD =20) varied more than the end of season (mean Julian days=302, SD =4), therefore the onset of spring is the measure which primarily dictates season length. For this reason, we used spring onset as a proxy to assess changes in season length. Generalized Linear Models (GLM) using a binomial response (logistic regression) were used to determine if there was a relationship between the prevalence of infection in all three species and the onset of spring each year. Likelihood ratio tests were used to assess the significance of these effects. A generalized adjusted  $R^2$  was calculated to assess the predictive power of each model (38).

## Acknowledgments

FC was funded by a CASE studentship from NERC, JH was funded by a BBSRC studentship from the Imperial College Grantham Institute, JB was funded by an Institute of Zoology research fellowship, AAC was part-funded by a Royal Society Wolfson Research Merit award, MCF and TWJG were funded by the NERC award NE/E006701/1 and the Biodiversa project RACE: Risk Assessment of Chytridiomycosis to European Amphibian Biodiversity.

## References

1. Harvell CD, Mitchell CE, Ward JR, Altizer S, Dobson AP, Ostfeld RS, et al. Ecology - Climate warming and disease risks for terrestrial and marine biota. *Science*. 2002;296(5576):2158-62.
2. Epstein P. The ecology of climate change and infectious diseases: comment. *Ecology*. 2010;91(3):925-8.
3. Rodo X, Pascual M, Doblas-Reyes FJ, Gershunov A, Stone DA, Giorgi F, et al. Climate change and infectious diseases: Can we meet the needs for better prediction? *Climatic Change*. 2013;118(3-4):625-40.
4. Stuart SN, Chanson JS, Cox NA, Young BE, Rodrigues AS, Fischman DL, et al. Status and trends of amphibian declines and extinctions worldwide. *Science*. 2004;306(5702):1783-6.

5. Fisher MC, Garner TWJ, Walker SF. Global Emergence of *Batrachochytrium dendrobatidis* and Amphibian Chytridiomycosis in Space, Time, and Host. Annual Review of Microbiology. 2009;63:291-310.
6. Pounds AJ, Bustamante MR, Coloma LA, Consuegra JA, Fogden MPL, Foster PN, et al. Widespread amphibian extinctions from epidemic disease driven by global warming. Nature. 2006;439:161-7.
7. Bosch J, Carrascal LM, Duran L, Walker S, Fisher MC. Climate change and outbreaks of amphibian chytridiomycosis in a montane area of Central Spain; is there a link? P R Soc B. 2007;274(1607):253-60.
8. Voyles J, Johnson LR, Briggs CJ, Cashins SD, Alford RA, Berger L, et al. Temperature alters reproductive life history patterns in *Batrachochytrium dendrobatidis*, a lethal pathogen associated with the global loss of amphibians. Ecol Evol. 2012;2(9):2241-9.
9. Ribas L, Li MS, Doddington BJ, Robert J, Seidel JA, Kroll JS, et al. Expression Profiling the Temperature-Dependent Amphibian Response to Infection by *Batrachochytrium dendrobatidis*. Plos One. 2009;4(12):-.
10. Garner TWJ, Rowcliffe JM, Fisher MC. Climate change, chytridiomycosis or condition: an experimental test of amphibian survival. Global Change Biol. 2011;17(2):667-75.
11. Rohr JR, Raffel TR. Linking global climate and temperature variability to widespread amphibian declines putatively caused by disease. P Natl Acad Sci USA. 2010;107(18):8269-74.
12. Rohr JR, Raffel TR, Romansic JM, McCallum H, Hudson PJ. Evaluating the links between climate, disease spread, and amphibian declines. P Natl Acad Sci USA. 2008;105(45):17436-41.
13. Olson DH, Aanensen DM, Ronnenberg KL, Powell CI, Walker SF, Bielby J, et al. Mapping the Global Emergence of *Batrachochytrium dendrobatidis*, the Amphibian Chytrid Fungus. Plos One. 2013;8(2).
14. Balaz V, Voros J, Civis P, Vojar J, Hettyey A, Sos E, et al. Assessing Risk and Guidance on Monitoring of *Batrachochytrium dendrobatidis* in Europe through Identification of Taxonomic Selectivity of Infection. Conservation Biology. 2014;28(1):213-23.
15. Bielby J, Bovero S, Angelini C, Favelli M, Gazzaniga E, Perkins M, et al. Geographic and taxonomic variation in *Batrachochytrium dendrobatidis* infection and transmission within a highly endemic amphibian community. Divers Distrib. 2013;19(9):1153-63.

16. Doddington BJ, Bosch J, Oliver JA, Grassly NC, Garcia G, Schmidt BR, et al. Context-dependent amphibian host population response to an invading pathogen. *Ecology*. 2013;94(8):1795-804.
17. Bosch J, Martínez-Solano I, García-París M. Evidence of a chytrid fungus infection involved in the decline of the common midwife toad (*Alytes obstetricans*) in protected areas of central Spain. *Biol Conserv*. 2001;97:331-7.
18. Whiles MR, Lips KR, Pringle CM, Kilham SS, Bixby RJ, Brenes R, et al. The effects of amphibian population declines on the structure and function of Neotropical stream ecosystems. *Front Ecol Environ*. 2006;4(1):27-34.
19. Briggs CJ, Knapp RA, Vredenburg VT. Enzootic and epizootic dynamics of the chytrid fungal pathogen of amphibians. *P Natl Acad Sci USA*. 2010;107(21):9695-700.
20. Walker SF, Bosch J, Gomez V, Garner TWJ, Cunningham AA, Schmeller DS, et al. Factors driving pathogenicity vs. prevalence of amphibian panzootic chytridiomycosis in Iberia. *Ecol Lett*. 2010;13(3):372-82.
21. Recuero E, Canestrelli D, Voros J, Szabo K, Poyarkov NA, Arntzen JW, et al. Multilocus species tree analyses resolve the radiation of the widespread *Bufo bufo* species group (Anura, Bufonidae). *Mol Phylogenet Evol*. 2012;62(1):71-86.
22. Garner TW, Walker S, Bosch J, Leech S, Rowcliffe M, Cunningham AA, et al. Life history trade-offs influence mortality associated with the amphibian pathogen *Batrachochytrium dendrobatidis*. *Oikos*. 2009(118):783-91.
23. Bielby J, Fisher MC, Clare FC, Rosa GM, Garner TWJ. Host species vary in infection probability, sub-lethal effects, and costs of immune response when exposed to an amphibian parasite. *Sci Rep-Uk*. 2015;5.
24. Briggs CJ, Vredenburg VT, Knapp RA, Rachowicz LJ. Investigating the population-level effects of chytridiomycosis: An emerging infectious disease of amphibians. *Ecology*. 2005;86(12):3149-59.
25. Bradley GA, Rosen PC, Sredl MJ, Jones TR, Longcore JE. Chytridiomycosis in native Arizona frogs. *J Wildlife Dis*. 2002;38(1):206-12.
26. Rachowicz LJ, Vredenburg VT. Transmission of *Batrachochytrium dendrobatidis* within and between amphibian life stages. *Dis Aquat Organ*. 2004;61(1-2):75-83.
27. Stocker T. Climate change 2013: the physical science basis: Working Group I contribution to the Fifth assessment report of the Intergovernmental Panel on Climate Change.: Cambridge University Press; 2014.

28. Raffel TR, Romansic JM, Halstead NT, McMahon TA, Venesky MD, Rohr JR. Disease and thermal acclimation in a more variable and unpredictable climate. *Nat Clim Change*. 2013;3(2):146-51.
29. Schmeller DS, Blooi M, Martel A, Garner TWJ, Fisher MC, Azemar F, et al. Microscopic Aquatic Predators Strongly Affect Infection Dynamics of a Globally Emerged Pathogen. *Curr Biol*. 2014;24(2):176-80.
30. Scheffer M, Rinaldi S, Kuznetsov YA, vanNes EH. Seasonal dynamics of *Daphnia* and algae explained as a periodically forced predator-prey system. *Oikos*. 1997;80(3):519-32.
31. Bosch J, Rincon PA. Chytridiomycosis-mediated expansion of *Bufo bufo* in a montane area of Central Spain: an indirect effect of the disease. *Divers Distrib*. 2008;14(4):637-43.
32. Boyle DG, Boyle DB, Olsen V, Morgan JAT, Hyatt AD. Rapid quantitative detection of chytridiomycosis (*Batrachochytrium dendrobatidis*) in amphibian samples using real-time Taqman PCR assay. *Dis Aquat Organ*. 2004;60(2):141-8.
33. Reading CJ. The effect of winter temperatures on the timing of breeding activity in the common toad *Bufo bufo*. *Oecologia*. 1998;117(4):469-75.
34. Thompson R, Ventura M, Camarero L. On the climate and weather of mountain and sub-arctic lakes in Europe and their susceptibility to future climate change. *Freshwater Biol*. 2009;54(12):2433-51.
35. Thompson R, Price D, Cameron N, Jones V, Bigler C, Rosen P, et al. Quantitative calibration of remote mountain-lake sediments as climatic recorders of air temperature and ice-cover duration. *Arct Antarct Alp Res*. 2005;37(4):626-35.
36. Semenov MA, Stratonovitch P. Use of multi-model ensembles from global climate models for assessment of climate change impacts. *Clim Res*. 2010;41(1):1-14.
37. Semenov MA, Stratonovitch P. Adapting wheat ideotypes for climate change: accounting for uncertainties in CMIP5 climate projections. *Clim Res*. 2015;65:123-39.
38. Heinzl H, Mittlbock M. Pseudo R-squared measures for Poisson regression models with over- or underdispersion. *Comput Stat Data An*. 2003;44(1-2):253-71.

**Supplementary Information:** see appended supplement

402 **Table 1:** The timing, in Julian days, of the start of spring and the end of season

403

Year	Start of spring	End of season	Days of activity
2008	162	302	140
2009	150	NA	NA
2010	137	298	161
2011	120	298	178
2012	145	302	157
2013	181	308	127
2014	160	309	149

404

**Table 2:** Visual estimates of amphibian abundance + <100; ++100-1000; +++ >1000

<b>Year</b>	<b><i>Ao</i> OW tadpoles</b>	<b><i>Rt</i> Mets</b>	<b><i>Bb</i> Mets</b>
<b>2008</b>	+++	+++	+++
<b>2009</b>	+++	+++	+++
<b>2010</b>	+++	+++	++
<b>2011</b>	+++	+++	++
<b>2012</b>	++	+++	+
<b>2013</b>	+	+++	0
<b>2014</b>	+	+++	0

## 409    **Figure Legends**

410    **Figure 1 a.** Temporal change in prevalence of infection for *Alytes obstetricans* (Ao), *Bufo*  
411    *spinosus* (Bs) and *Rana temporaria* (Rt); **b.** Temporal change in intensity of infection; **c.**  
412    Seasonal changes in water temperature and timing of spring onset in Lac Arlet; **d.**  
413    Relationship between spring onset and the prevalence of *Bd* infection across all species

414    **Figure 2 a.** Histology slide showing a section from the hind legs of a deceased *R. temporaria*  
415    metamorph demonstrating clear evidence of the disease chytridiomycosis. The two long  
416    arrows point to two of many sporangia full of zoospores, and the two short arrows point to  
417    empty sporangia cases (once zoospores have burst out), both embedded with the upper skin  
418    layers; **b.** Counts of live and dead *A. obstetricans* metamorphs over time with the number of  
419    dead (black) and alive (white) *A. obstetricans* metamorphs encountered. No 'alive' counts  
420    were made for years 2008 - 2009; **c.** Relationship between air temperature and the onset of  
421    spring at Lac Arlet; **d.** Future predictions of air temperatures at Lac Arlet

422    **Figure 3 a.** Lac Arlet showing the position of temperature datalogger (red arrow). **b.** Mass  
423    mortalities of midwife toads *Alytes obstetricans* caused by *Batrachochytrium dendrobatidis*  
424    lineage *BdGPL* at Lac Arlet