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Can Skeletal Surface Area Predict *in vivo* Foot Surface Area?

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1 **Abstract**

2 The surface area of feet in contact with the ground is a key morphological feature that
3 influences animal locomotion. Underfoot pressures (and consequently stresses experienced
4 by the foot), as well as stability of an animal during locomotion, depend on the size and
5 shape of this area. Here we tested whether the area of a skeletal foot could predict *in vivo*
6 soft tissue foot surface area. Computed tomography scans of 29 extant tetrapods (covering
7 mammals, reptiles, birds and amphibians) were used to produce models of both the soft
8 tissues and the bones of their feet. Soft tissue models were oriented to a horizontal plane,
9 and their outlines projected onto a surface to produce two-dimensional silhouettes.
10 Silhouettes of skeletal models were generated either from bones in CT pose or with all
11 autopodial bones aligned to the horizontal plane. Areas of these projections were calculated
12 using alpha shapes (mathematical tight-fitting outline). Under-foot area of soft tissue was
13 approximately 1.67 times that of skeletal tissue area (~2 times for manus, ~1.6 times for
14 pes, if analyzed separately). This relationship between skeletal foot area and soft tissue
15 area, while variable in some of our study taxa, could provide information about the size of
16 the organisms responsible for fossil trackways, suggest what size of tracks might be
17 expected from potential trackmakers known only from skeletal remains, and aid in soft
18 tissue reconstruction of skeletal remains for biomechanical modelling.

19 **Key Words**

20 Locomotion, ichnology, biomechanics, anatomy

21

22 **Introduction**

23

24 The surface area of tetrapod autopodia (feet) reflects several important biomechanical
25 factors, including body mass (McMahon, 1975), habitat (Blackburn et al., 1999), speed
26 (Segal et al., 2004), and bipedal or quadrupedal locomotory habits (Snyder, 1962). Foot
27 surface area is determined by autopodial morphology and posture (Hildebrand, 1980; Full
28 et al., 2002), and, in conjunction with the body mass and locomotory mode of an animal,
29 determines underfoot pressure (Miller et al., 2008; Michilsens et al., 2009;
30 Panagiotopoulou et al., 2012; Qian et al., 2013; Panagiotopoulou et al., 2016).

31 For very large animals, such as rhinoceroses and elephants, foot surface area needs to be
32 large, as a method of reducing underfoot pressure and avoiding injury to the foot, as well as
33 avoiding sinking on soft ground (Falkingham et al., 2011a). However, foot contact area
34 does not appear to scale isometrically with mass. Larger animals often have smaller foot
35 contact areas than would be expected, and the relationship between foot contact area and
36 mass differs between unguligrade, digitigrade and plantigrade animals (Michelsens et al.,
37 2009; Chi and Roth, 2010). Large animals must compensate for their size with other
38 mechanisms, such as fatty footpads, in order to reduce stress (Panagiotopoulou et al.,
39 2012). Presumably the extinct sauropod dinosaurs, many times larger than extant elephants
40 (Bates et al., 2016) used similar compensatory adaptations (Platt and Hasiotis, 2006).

41 Foot surface area is also reflective of an animal's posture and limb use (Biewener, 1989),
42 with bipedal animals requiring feet large enough to support their body weight with half as

43 many limbs as their quadrupedal counterparts (Gatesy and Biewener, 1991), and, in the
44 case of birds, in a huge range of environments and ecological niches with different
45 demands (Alexander, 2004). An animal's balance (e.g. keeping the body's centre of mass
46 (CoM) close to the centre of pressure of feet-- influenced by foot area) is also of vital
47 importance, as the stability of an animal during locomotion is vital to its ability to catch
48 prey, escape predators, migrate effectively, and avoid injury when overexerting itself and
49 when moving on unstable ground (Hodgins and Raibert, 1991; Patla, 2003; Geyer et al.,
50 2006; Birn-Jeffery et al., 2014).

51 Foot surface area appears to correlate with relative speed during certain forms of
52 locomotion. Body mass has a direct effect on maximum running speed, especially notable
53 in large animals, as speed scales with body mass up to moderate sizes and then declines
54 (Garland, 1983; Bejan and Marden, 2006), and the duration of foot contact with the ground
55 also scales with body mass (Farley et al., 1993). The position and number of toes also tends
56 to be a specialisation for terrestrial running, with a reduced number of toes present in both
57 horses and ostriches (among other cursorial taxa; Coombs, 1978), reducing foot weight, a
58 useful adaptation because heavier feet necessitate more energy usage to recover from a
59 stride (Snyder, 1962; McGuigan and Wilson, 2003; Schaller, et al., 2011). Peak plantar
60 pressure and speed are demonstrably linked in humans (Rosenbaum et al., 1994; Segal et
61 al., 2004; Pataky et al., 2008) and ostriches (Schaller, et al., 2011); however, this link has
62 not been fully explored in other terrestrial animals, especially quadrupeds.

63 Large feet have a potentially conflicting relationship with speed in that they will be more
64 massive and thus have greater inertia, making them more difficult to swing quickly through
65 the air (Taylor et al., 1974; Fedak et al., 1982; Kilbourne and Hoffman, 2013; Kilbourne
66 and Carrier, 2016). Nonetheless, it is important that foot surface area and underfoot
67 pressures evolve to allow an organism's locomotion to be energy-efficient and its posture
68 stable, while enabling sufficient bursts of speed if necessary. In other words, the surface
69 area of the autopodia should be subject to selective pressures in the same manner as any
70 other part of the locomotor system.

71 Foot surface area is also potentially influenced by Allen's rule (Allen, 1877; Allee and
72 Schmidt, 1937), which supposes that warm-blooded animals in cold climates will tend to
73 have smaller feet than their relatives in warmer clines (Blackburn et al., 1999). This may or
74 may not be due to causal links (i.e. natural selection) to either reduce surface area exposed
75 to the cold, or be a reflection of adaptations in warmer climates to increase surface area to
76 promote heat dissipation. This 'rule' may conflict with constraints imposed by keeping
77 pressures low (i.e. foot areas large) to avoid sinking into soft substrates such as snow or
78 sand. Allen's rule also potentially conflicts with the outcome of Bergmann's rule – the
79 contentious but broadly supported tendency for ectotherms to be larger in colder climates
80 (Clarke, 2017). Therefore, colder conditions will tend to correlate with increased body
81 mass, implying a larger foot surface area while simultaneously selecting for smaller feet.

82 Some animals exhibit notable disparity in the size of fore- and hind-feet, which is apparent
83 in their foot surface area: a condition known as heteropody. A previous study (Henderson,
84 2006) demonstrated that the ratio of fore- and hind-foot surface areas, in its subject
85 animals, could match CoM position, e.g. an elephant has 40%/60% relative fore- vs. hind-
86 foot surface area, and a CoM of 40% of the distance from the glenoid to the acetabulum. It
87 would seem logical to assume that animals spread their body weight relatively evenly over
88 their feet, in order to reduce maximum pressure, excess tissue or substrate stress and strain
89 (Cheung et al., 2005), and to prevent sinking when walking across compliant substrates

90 (Falkingham et al., 2011a). However, this assumption runs contrary to pressure
91 experiments showing higher mean peak pressures in elephant forelimbs (Panagiotopoulou
92 et al., 2012). It is therefore worth exploring a possible correlation of the relative sizes of an
93 animal's manus and pes, and CoM with both observations in mind, and worth considering
94 possible implications of such a correlation across Tetrapoda.

95 Heteropody is a common occurrence in some extinct animals, such as sauropod dinosaurs,
96 as indicated by trace fossil evidence (Lockley et al., 1994; Henderson, 2006). Preserved
97 trackways from these dinosaurs indicate that often their fore- and hind-feet impressions
98 differ in depth (Falkingham et al., 2011b; Falkingham et al., 2012), implying differential
99 underfoot pressures. Determining foot surface area in these animals can be complex,
100 however, and attribution of specific trackmakers to trackways is notoriously difficult
101 (Farlow, 1992; Clack, 1997; Falkingham, 2014a), partly because matching impressions of
102 fully fleshed feet to skeletal remains would require accurate methods of predicting skeletal
103 to skin foot morphology, which is currently difficult and largely speculative (Jannel et al.,
104 2019). Indeed, matching the tracks of extant animals to the correct species is often not
105 straightforward – as illustrated by the existence of field guides produced to help
106 fieldworkers with this problem (e.g. Bang and Dahlstrøm, 2001).

107 For terrestrial and arboreal fauna, the substrate underfoot can have a noticeable effect on
108 locomotion, and the way the foot moves in a step. Both substrate and autopodial tissue will
109 be compressible to varying degrees, slightly altering foot contact area during stance
110 (Gatesy, et al., 1999; Gatesy, 2003; Falkingham and Gatesy, 2014; Gatesy and Falkingham,
111 2017).

112 Palaeobiologists must rely on soft tissue data from extant animals to infer many facets of
113 the morphology of extinct animals (Witmer, 1995), because preservation of soft tissues is
114 rare and only partial details about muscle and tendon structures can be inferred from the
115 skeletal elements they interacted with. In this way, a study of the relationship of flesh and
116 skeletal foot surface area should help to fill gaps in our understanding of the anatomy of
117 extinct animals' feet, as well as the interaction of foot structure and CoM, and would be
118 particularly valuable for linking fossil trackways and supposed trackmakers. Here we aim
119 to test whether skin and skeletal surface area are correlated across Tetrapoda, and if so, if
120 their correlation is strong enough to make it a useful tool in the study of fossils and
121 trackways.

122

123 **Materials & Methods**

124 In order to compare skeletal and fully fleshed foot anatomy in extant animals, computed
125 tomography (CT) scans of cadaveric autopodia from 29 species of tetrapod (one specimen
126 of each except for *Crocodylus moreletii* and *Osteolaemus teraspis* – see supplementary
127 material), covering amphibians, reptiles, birds, and mammals, were analysed. The sex of
128 individuals was unknown, and all but *Crocodylus niloticus* were adults. All specimens were
129 museum or zoo-donated specimens whose cause of death was unrelated to this study (and
130 generally unknown).

131 MeVisLab (Heckel et al., 2009) was used to segment the scans into separate 3D models
132 (OBJ format meshes) of the soft tissue and skeletal elements. The resultant meshes were
133 then imported into Autodesk Maya 2018, where they were cleaned, aligned and re-posed to

134 the horizontal plane (figure 1). The aligned meshes were then processed using MatLab
135 (Mathworks Inc. Natick, MA, USA), where they were ‘flattened’ by setting the vertical
136 component of each vertex to 0. This flattening produced 2D ‘silhouettes’ of the models,
137 either as soft tissue of the foot or its skeleton, from which area was calculated using an
138 alpha shape (see below).

139 Skin models were oriented and posed so that only areas of the feet that would touch the
140 ground during locomotion would be used upon flattening the models, and any parts of the
141 models that extended past this area were removed (figure 1B). The extent of the soles of the
142 feet were, for the most part, obvious from visible anatomy. In addition, from *in vivo*
143 biplanar fluoroscopy studies, X-ray images, and photographs *in situ*, we made educated
144 estimates of accurate positions for taxa (Astley and Roberts, 2014; Bonnan, et al., 2016;
145 Kambic et al., 2015; Panagiotopoulou, et al., 2016). For a more repeatable approach (Pose
146 2, see below), parts of the skin model extending past the functional foot area (the unguals
147 for unguligrade animals, the digits for digitigrade animals, and the entire sole of the foot for
148 plantigrade animals and semi-digitigrade animals, so that the full extent of fatty foot pads
149 were accounted for) were removed where present.

150 However, since these models were taken from CT scans, without the full weight of the
151 animal deforming the foot underneath, the true shape of the foot during stance for many of
152 these animals may have been slightly different, due to compliant soft tissues (Alexander, et
153 al., 1986; Gatesy, 2003). This is especially significant for those animals with large fatty
154 foot pads such as *Elephas* and *Ceratotherium*, and less significant for the majority of
155 ungulates, whose hooves are stiff, and more resistant to deformation (Hinterhofer, et al.,
156 2000; Hutchinson, et al., 2011).

157 Skeletal models were posed in one of two ways. Firstly (Pose 1), matching the pose of skin
158 models (Figure 1B-D), secondly (Pose 2), with all bones aligned to the horizontal (Figure
159 1E-F). For the latter pose, models were cropped proximal to the digits for digitigrade
160 animals, proximal to the unguals for unguligrade animals, proximal to the tarsals/carpals
161 for plantigrade animals.

162 For large, semi-digitigrade/subunguligrade animals (*Elephas maximus*, *Ceratotherium*
163 *simum*, and *Hippopotamus amphibius*), proximal foot elements are raised off the ground,
164 supported by fatty foot pads, increasing foot contact area. Therefore using only the
165 phalanges, as for other digitigrade animals, would severely underestimate contact area. To
166 explore this ambiguity, skeletal outlines were generated from just the digits (Pose 2a), the
167 digits plus metatarsals (Pose 2b), and with the entire foot skeleton (Pose 2c). This analysis
168 was designed to be more objective and repeatable in determining skin from skeletal surface
169 area, particularly, in extinct animals, where knowledge of *in vivo* foot posture may be
170 lacking.

171 Results for area where left and right forefeet or hindfeet were available were averaged
172 (mean), as were area results for animals with multiple specimens, and *Camelus*, where both
173 feet were unassigned as forefeet or hindfeet.

174 It should be noted that our 29 animals studied include several ungulates, possessing large,
175 keratinous hooves, much harder and stiffer than most other tissues categorised under ‘soft
176 tissues’ in this study. While ungulate hooves have properties that distinguish them from
177 other soft tissues, and take longer to decompose than softer tissues, they are also distinct
178 from skeletal tissue, and are rarely preserved, especially in fossils (Pollitt, 2004; Saitta, et

179 al., 2017). In terms of comparisons between skeletal and fossil remains and the overall foot
180 structure of living animals, hooves clearly are an important part of a living ungulate's foot
181 structure, and their ability to locomote; thus being able to predict their size from skeletal
182 remains is as much of a part of the goal of this study as predicting the areas of softer tissues
183 (Warner, et al., 2013). In this sense, the term 'soft tissue' as used in this study refers to
184 'non-skeletal tissue', with the hardness of these tissues largely irrelevant.

185 Initially, we attempted to calculate the 2D convex hull (a shape made by joining the
186 outermost data points in a simplified representation of the data (see figure 1C-D, in green))
187 of each silhouette, but found via pose tests using bird feet that this method was extremely
188 sensitive to pose, particularly whether the digits were laterally spread or not
189 (Supplementary material 1). Instead, 2D, tight-fitting alpha shapes (where the outermost
190 data points were joined in a shape that most closely fits the silhouette's true shape (figure
191 1C-D, in pink)) were produced for each silhouette, and the area of these alpha shapes
192 calculated. The alphaShape command in MatLab uses an 'alpha value' to determine the
193 maximum distance between edge points to bridge (a sufficiently large 'alpha value' will
194 produce a convex hull). We used the automatically determined alpha value for each alpha
195 shape, which is calculated based on the density of vertices in the model, as this produces
196 the tightest fitting single shape for any given set of points. We set the hole threshold to be
197 extremely large (larger than the foot as a whole) to remove any holes from the interior of
198 the alpha shape. The surface area of the skeleton's alpha shape as a percentage of the skin's
199 shape was then used to compare each organism.

200 The dataset was then run through PGLS (phylogenetic generalised least squares) regression
201 analyses to assess the significance of the relationship between the variables, and how much
202 impact common ancestry between the animals studied affected the results (Blomberg et al.,
203 2012; Felsenstein, 1985). This was accomplished using Mesquite (Maddison and
204 Maddison, 2001) to draw three simple trees (manually compiled "consensus" phylogenies
205 based on the most recent and broadly accepted phylogenies at the time of writing, within
206 which the placement of Carnivora, Cetartiodactyla and Perissodactyla in relation to each
207 other, was the only major point of contention (Gauthier et al., 1988; Nery et al., 2012; Prum
208 et al., 2015)) connecting the organisms involved in this study. We then applied the Grafen
209 method (Grafen, 1989) of branch length estimation to the trees, and ran PGLS via the Ape
210 (Paradis et al., 2004), Geiger (Harmon et al., 2008), Nlme (Bliese, 2006) and Phytools
211 (Revell, 2012) packages in R. Results for forefeet, hindfeet, and all feet were each tested.
212 The influence of body mass was also tested using PGLS, in order to determine whether
213 phylogeny, body mass, or a combination of both factors had a significant effect on the
214 relationship between skin and skeletal foot surface area. P values <0.05 were considered
215 significant. Body masses were taken from scan metadata where possible, or estimated from
216 the literature (e.g. Dunning Jr, 1992) where such metadata were not available
217 (Supplementary material 1).

218 Skin surface area was plotted against skeletal surface area for all analyses, using the entire
219 data set, and then broken up into smaller groups: unguligrade, digitigrade, plantigrade,
220 terrestrial, semi-aquatic, erect posture, sprawling posture, mammals, and birds. The plots
221 were framed in terms of the predictability of skeletal area from skin area, to emphasise
222 potential utility for trackmaker identification from fossils. However, these data are intended
223 to be interpretable both ways, and the prediction of *in vivo* surface area from skeletal
224 remains is of equal utility. For the purposes of these analyses, the digitigrade (Pose 2a) and
225 plantigrade (Pose 2c) poses of semi-digitigrade/subunguligrade (*sensu* Carrano, 1997)

226 animals were added to their respective groups, whereas Pose 2b was used for the remaining
 227 groups, as it represents an intermediate pose. Semi-aquatic included amphibians,
 228 crocodylians and hippopotamuses, terrestrial did not include birds except for *Dromaius*
 229 *novaehollandiae*, and sprawling (here meaning non-erect) posture included amphibians,
 230 lepidosaurs and crocodylians, although crocodylians use a range of limb postures spanning
 231 the sprawling-to-erect continuum (Gatesy, 1991; Reilly and Elias, 1998).

232

233 **Results**

234 For the Pose 1 analysis (approximate life position), projected foot skeleton surface area as a
 235 percentage of projected fully fleshed foot surface area (Figure 2, above cladogram) was an
 236 average of 56% (both mean and median) for all organisms measured (three amphibians,
 237 four crocodylians, seven birds, and fourteen mammals), with means of 49% for amphibians
 238 (53% median), 47% for crocodylians (48% median), 68% for birds (67% median), and 55%
 239 for mammals (54% median) with an average standard deviation of 13%. Extremely similar
 240 results were found with bones oriented as in Pose 2. The smallest percentages of skeletal
 241 vs. fleshed surface area observed were in *Equus* species (*Equus quagga* at 34%, *Equus*
 242 *ferus caballus* 38%), *Giraffa camelopardalis* (38%), *Crocodylus niloticus* (38%), and
 243 *Cryptobranchus alleganiensis* (39%). However, besides *Equus* and *Giraffa*, other ungulates
 244 did not stand out as having particularly low skeletal areas relative to skin areas.
 245 Carnivorans had proportionately high skeletal calculated area. The highest skeletal areas
 246 relative to skin areas (as seen from the underside, and in two dimensions) were *Coturnix*
 247 *coturnix* at 83%, followed by *Panthera leo persica* and *Ceratotherium simum*, at 81% and
 248 73%, respectively.

249 Where skeletal models were set flat (Pose 2), all unguligrade animals expressed lower
 250 skeletal area compared to skin surface area, compared with Pose 1 (Figure 2). The zebra
 251 stood out most with just 22% skeletal representation.

252 *Elephas*, *Hippopotamus*, and *Ceratotherium* showed considerable variability depending on
 253 which foot bones (Pose a/b/c) were used to predict skeletal area: *Hippopotamus*
 254 (37/76/100%), *Ceratotherium* (31/74/98%), *Elephas* (17/42/68%). 100% skeletal surface
 255 area representation in the hippopotamus clearly suggests that treating these animals as
 256 plantigrade does not yield results representative of these animals' foot morphology, or
 257 indeed results that are useful for predictive purposes, especially given the steep
 258 (subvertical) angle at which these animals position their feet *in situ*.

259 Carnivorans, particularly cats, typically do not have their digits extended fully when
 260 walking or standing, as such relative skeletal area calculated from Pose 2 (eg. *Panthera*
 261 93%, *Vulpes* 92%) generally produces higher relative skeletal areas than the more life-like
 262 Pose 1 (eg. *Panthera* 81%, *Vulpes* 70%).

263 Overall, mammalian data were highly variable (47% range from maximal to minimal
 264 values in Pose 1, over 80% range in Pose 2). Given that mammalian species dominated our
 265 study sample (then birds, then crocodylians), perhaps with more data the variability within
 266 other groups would increase to comparable levels. However, that mammalian feet have
 267 unusually high morphological disparity compared to other taxa in our sample, is reflective
 268 of their unusually high morphological disparity in terms of body size, foot anatomy, and
 269 posture compared to other groups (Kubo et al., 2019).

270 Bird and crocodilian data were more consistent than mammals (25% range for birds in all
271 analyses, 18% range for crocodilians). *Dromaius*, which was morphologically and
272 functionally distinct from the other birds in the study in terms of being large and flightless,
273 fell neatly within the range for birds.

274 Raw numbers for projected skeleton and projected skin surface area, calculated from Pose
275 1, were plotted as a log graph, and a power trendline fitted (Figure 3). This plot, despite the
276 variation seen in Figure 2, showed a strongly positive correlation ($R^2 = 0.99$, p value <0.05)
277 in 'Pose 1' between skin and skeletal foot surface area. This correlation can be described
278 with the equation $y = 0.59x^{0.99}$ (where y = skeletal foot surface area and x = foot skin
279 surface area). This skin and skeletal foot surface area's scaling relationship was close to
280 isometry (slope of 1.0). Soft tissue surface area may therefore be predicted, on average, as
281 approximately 1.67 times skeletal surface area. There were very few outlying animals,
282 indeed, *Elephas* and *Ceratotherium* were the only animals that diverged notably from the
283 linear trendline. If the three largest animals were removed from the data set, or the three
284 smallest, the strength of the correlation was unaffected, but soft tissue area predictions from
285 skeletal area decreased (Supplementary Material 1). If both groups were removed, the
286 predicted value decreased further.

287 When the forelimb and hindlimb results were calculated separately, the equations differed
288 noticeably ($y = 0.52x^{0.99}$ and $y = 0.64x^{0.98}$ respectively); although the difference in slope
289 was not statistically significant, and R^2 values remained ~ 0.99 (Figure 3). However, soft
290 tissue area was ~ 2 times skeletal area in the forelimb, but only ~ 1.56 times in the hindlimb.
291 See Table 1 for full list of formulae and R^2 values, rounded to two significant figures (and
292 see Supplementary Material 1 for slope uncertainties for all poses, and for all limbs,
293 forelimbs, and hindlimbs.).

294 For all flat pose analyses (Pose 2), heavier animals remained the outliers, with *Elephas*,
295 *Hippopotamus*, and *Ceratotherium* diverging most from the trendline (Figure 4). Similar to
296 the Pose 1 analysis, Pose 2b suggested high predictability, with soft tissue as approximately
297 1.67 times skeletal surface area. Regressions for Pose 1 and Pose 2b were statistically
298 similar. The analysis treating semi-digitigrade/sub-unguligrade as plantigrade (Pose 2c)
299 suggested soft tissue as approximately 2.04 times skeletal surface area, and semi-
300 digitigrade as digitigrade (Pose 2a) resulted in soft tissue as 1.05 times skeletal surface
301 area. Interestingly, the hindlimbs-only regression for Pose 2b was significantly different
302 from its equivalent with both fore- and hindlimbs and forelimbs-only (Table 1).

303 PGLS results (e.g. for all feet, in 'Pose 1', with Carnivora and Perissodactyla in a single
304 clade) produced a correlation of -0.171 between the predictor and the intercept, and a
305 Pagel's lambda value ~ 1 , with an adjusted R^2 of 0.92 (t-statistic 18.06, residual S.E. 12005,
306 29 DF (26 residual)). Similar results were found when running the same tests on fore-and
307 hind-feet separately, with the other two phylogenetic tree arrangements. When skeletal
308 elements were laid flat, variable adjusted R^2 , Pagel's lambda (though all ~ 1), and t-statistics
309 were found, with higher standard error (15686.49 SE (28 DF (26 residual))) in Pose 2a)
310 (Supplementary material 1). Despite these variations, this still suggests that phylogeny is
311 not the main driver of the correlations found.

312 Separate regressions for unguligrade, digitigrade, plantigrade, terrestrial, semi-aquatic,
313 erect posture, sprawling posture, birds and mammals, all showed strong correlations (Table
314 2, Supplementary material 1 and 2). Equations for all the analyses varied, with opposing
315 regressions (e.g. sprawling versus erect posture, or terrestrial versus semi-aquatic)

316 statistically different from each other (Table 2, equations and R^2 values rounded to two
317 significant figures). Although R^2 values suggest high correlations for these regressions, the
318 lack of data points in each of them (particularly those with the highest R^2 values) suggests
319 their predictive value is relatively low at present. There are potentially functional reasons
320 why, for example, sprawling animals, semi-aquatic animals, and birds would have stronger
321 correlations and more predictable foot morphologies, but the lower scores in groups with
322 more data points suggests high correlation in groups with few data points may be an
323 artefact, and should be viewed with caution.

324 Body mass had no significant effect on relative skin/skeletal areas. This was unsurprising
325 because *Ceratotherium* results indicated more skeletal representation than other large
326 animals such as *Elephas*, and percentage of skeletal vs. non-skeletal (skin) area results for
327 small animals did not appear to skew towards either obviously high or low skeletal
328 representation (Supplementary Material 1).

329

330 **Discussion**

331 Projected skeletal surface area as a percentage of projected skin surface area varied
332 between the organisms studied, most notably in mammals, which yielded both the lowest
333 and second highest values (Figure 2). Bird feet are all similarly digitigrade in their posture
334 and are largely made up of skeleton (with three major digits and consistent phalangeal
335 numbers), skin, and connective tissue, so their more consistent percentages are not
336 surprising considering that some of the mammals in this dataset had hooves, fatty footpads,
337 and a wide range of foot anatomies and postures (from plantigrade to unguligrade). PGLS
338 results suggested that the correlation between skin and skeletal foot surface area in all
339 poses, as well as being very strong, still held with phylogeny taken into account. This
340 suggestion was supported by Figures 3 and 4.

341 *Equus* and *Giraffa* stood out in this dataset for having an especially low relative skeletal
342 surface area. All extant horses have one toe with a large, keratinous hoof (Bowker et al.,
343 1998), so this was perhaps to be expected. Giraffes also have relatively small feet and
344 gracile legs compared to other animals of similar size, and a combination of high body
345 mass and high running speeds, which contribute to an overall unique morphology (van
346 Sittert et al., 2015). Pose 2 resulted in a lower relative skeletal area across unguligrade
347 animals, though none as extreme as either *Equus* species. By focusing on ungual bones, it
348 became clear that the keratinous sheath that forms the hoof dominates the ‘silhouettes’,
349 with skeletal tissue only represented by the very tip of the toe, so this is to be expected.
350 Non-unguligrade ungulates: *Ceratotherium*, *Hippopotamus*, *Camelus dromedaries*, and
351 *Vicugna pacos*, did not yield similar results to unguligrade ungulates, and varied
352 significantly from this group, as well as from each other.

353 For *Crocodylus niloticus*, the fact that Crocodylia have relatively thin, long, digital bones,
354 somewhat similar to human phalanges, that converge to form a surprisingly robust foot,
355 could have some effect (Ferraro and Binetti, 2014). Furthermore, joint range of motion
356 studies have suggested an unusual wrist function and resultant manus posture in
357 crocodylians favouring rigidity, which could affect potential foot contact area (Hutson and
358 Hutson, 2014). This rigidity could potentially aid in swimming, with the stiff foot acting in
359 a flipper-like fashion to push through water efficiently, which smaller crocodylians tend to
360 rely upon (Seebacher, et al., 2003). Furthermore, the *Crocodylus niloticus* specimen used

361 was the only juvenile in this study, and its phalanges were small and spaced far apart in
362 some cases, so this result could be an artefact of ontogeny, or the quality of the models
363 used. Further studies on the effect of ontogeny on skeleton to skin surface area ratio could
364 elucidate this further. Indeed, in future studies consideration should be given to levels of
365 ossification of manus and pes bones. For example, our *Cryptobranchus* CT scan was
366 missing wrist bones on all feet when segmented because these elements were cartilaginous
367 in the specimen scanned, and were indistinguishable from soft tissue. Such ossification is
368 likely to vary across species, and across ontogeny.

369 At the other extreme, where skeletal surface area was high (most closely approaching
370 projected skin surface area), several birds (most notably *Coturnix*, *Accipiter nisus*, and
371 *Alectoris chukar*) along with carnivorans and *Ceratotherium* (as well as *Hippopotamus* in
372 Pose 2b and 2c) stand out the most. For birds, this is understandable considering their
373 relative lack of musculature and fat in their feet. For carnivorans this could be explained by
374 their claws, extending beyond the main body of the foot, by the resting position of their
375 digits *in vivo*, and by their footpads, for which stiffness scales directly with body mass,
376 while foot contact area lags behind (Chi and Roth, 2010). This scaling allows carnivorans
377 to maintain relatively small feet that are light enough to be moved quickly (Kilbourne and
378 Carrier, 2016; Kilbourne and Hoffman, 2013).

379 Body mass seemed to have little general effect on the relationship between skin and
380 skeletal foot surface area. Previous studies have found a scaling relationship between body
381 mass and foot contact area not significantly different from isometry (Michilsens et al.,
382 2009), implying that the ratio of skeleton to soft tissue in the foot was not affected by this
383 scaling effect. The scaling relationship between the ratio of skin to skeletal foot surface
384 area was at best trivially different from isometry— a sensible result given that the variables
385 are two facets of the same structure (i.e. the manus or pes), and therefore their structure and
386 development are intrinsically linked. Despite this result, the largest animals in our dataset
387 were the most outlying (much less so when plotted logarithmically (Figure 3)). It is notable
388 that these largest animals, namely, *Elephas*, *Ceratotherium*, and *Hippopotamus*, were also
389 the only semi-digitigrade/subunguligrade animals in our data. These animals both had the
390 largest feet in the study and possess fatty foot pads to reduce loads on their individual toes
391 and spread out underfoot pressure due to their large body masses (Hutchinson et al., 2011;
392 Regnault et al., 2013). The divergence of these data appears to be influenced by their foot
393 posture as well as their large size, with the adaptation of a semi-digitigrade posture
394 potentially occurring specifically to support their large body weights.

395 It may be worth considering that beyond a certain weight threshold, specialised foot
396 morphologies are necessary for weight support and locomotion, and thus successively
397 heavier animals may have more disparate soft tissue structure and foot posture adaptations
398 to cope with increased load (Hutchinson, et al., 2011). This has implications for the
399 inherent predictability of our methods for very large extinct animals, such as sauropod
400 dinosaurs, especially where foot posture is loosely inferred and little information about soft
401 tissue structure is available. Follow-up studies on semi-digitigrade foot postures and how
402 they support loads differently to other foot postures, as well as similar studies to this, using
403 additional heavy and semi-digitigrade animals, would increase understanding of this
404 variation of foot form and function. Contrary to the semi-digitigrade animals in our study,
405 the giraffe, an unguligrade animal, was the largest other tetrapod (<1500kg vs. 3000+kg in
406 larger individuals of the semi-unguligrade taxa), and deviated little from trendlines.

407 The strength of the correlation between skin and skeletal foot surface area, despite
408 variations seen in Figure 2, implied sufficient reliability to predict one from the other
409 (Figure 3).. Despite this, birds only appeared above the trendline (Figure 3). Perhaps a
410 more accurate correlation could be achieved for birds alone with a larger avian dataset
411 (with a wider range of foot sizes), which would allow more accurate predictions of bird
412 foot surface area, and of foot surface area for animals with similar pedal anatomy to birds
413 (such as non-avian theropod dinosaurs). Although our main results could be refined with a
414 much larger tetrapod data set, it appears that foot surface area can be predicted from foot
415 skeletal surface area, with soft tissue generally predictable as approximately 1.67 times
416 skeletal foot surface area, as demonstrated in Poses 1 and 2b. However, when analyzed
417 separately, manus and pes presented differing ratios, with soft tissue surface area of the
418 former being predicted as ~ 2 times skeletal area, but just ~ 1.56 times for the pes. This
419 correlation could potentially be used to estimate skeletal foot surface area of animals from
420 their footprints, and its inverse used to predict skin-on-foot surface area of extinct animals
421 from their skeletons, and even of cadavers from skeletons, with potential forensic
422 applications.

423 For Pose 2, *Elephas*, *Ceratotherium*, and *Hippopotamus* were tested in three different
424 poses. Their foot anatomy is unusual in that they have a foot posture with most foot
425 elements far off the ground, but also have fatty pads which give them a large foot surface
426 area. With this in mind, all foot elements being in line with the horizontal plane, as in Pose
427 2c, is highly unrealistic. Pose 2a is perhaps more realistic than 2c, but assumes fewer foot
428 elements are supportive during stance than is accurate *in vivo*. The most representative
429 position for semi-digitigrade would arguably be Pose 1, as this did not force these animals
430 into an unrealistic foot posture. However, both Pose 1, and Pose 2b both result in the same
431 1.67 times skeletal surface area value, and Pose 2b's intermediate nature tests a pose in
432 between digitigrade and plantigrade. Pose 2b then, is perhaps the best repeatable method.
433 If, despite this, our other methods were chosen to predict foot surface area, skin surface
434 area would be equal to 1.05 times skeletal surface area for Pose 2a, and 2.04 times skeletal
435 surface area for Pose 2c. The variability in these analyses does reveal that altering the
436 results of the largest animals in the study alters the equation used. Therefore, perhaps this
437 method would be best applied to smaller and non-semi-digitigrade animals. However,
438 variation in area results is to be expected when fundamentally changing the number of
439 skeletal elements in an analysis.

440 Where data were divided into smaller groups for analysis, strong correlations were found in
441 results for plantigrade animals, semi-aquatic animals, sprawling posture, and birds (Table
442 2). Selective pressures potentially could drive a need for similar foot anatomy across these
443 groups, and therefore predictable foot structures, such as adaptations for perching,
444 swimming, and supporting body weight when feet are not directly under the body. Yet
445 considering that these groups were also the groups with the fewest data points, we cannot
446 draw any definitive conclusions from these results.

447 In terms of methods used, we found that convex hulls are highly sensitive to foot pose,
448 such as the size of inter-digital angles (Supplementary material 1), a result consistent with
449 previous findings (Cholewo and Love, 1999). This could be the cause of wide error
450 margins if these hulls were used for predictive purposes. This is especially relevant in re-
451 posed foot models, where inter-digital angles are manipulated to resemble *in vivo*
452 arrangements, and in animals that have long, thin digits, such as crocodylians. Alpha shapes

453 produced more consistent, ‘tight-fitting’ outlines for area calculation, a much more accurate
454 measure of the real scope of foot surface area for these models.

455 Inevitably, models derived from CT scans, such as those we used, ignore certain *in vivo*
456 factors such as foot deformation during contact with the ground. While we attempted to
457 stick closely to the *in situ* positions of feet (Pose 1), and aimed for a more objective
458 iteration of our analysis by laying bones flat to remove subjectivity (Pose 2), deformation is
459 a very difficult issue to control for. Collection of the data needed to account for this would
460 require advanced *in vivo* imaging techniques such as biplanar fluoroscopy (i.e. “XROMM”;
461 Brainerd et al., 2010; Gatesy et al., 2010); however, such techniques remain limited in the
462 size of potential subjects (e.g. Panagiotopoulou et al., 2016) and can be expensive and
463 time-consuming to conduct. Despite this issue, deformation of the foot should generally not
464 be significant enough that it should diminish the usefulness of this study or the
465 predictability of the methods employed here, as even in soft footpads, foot contact area
466 does not maintain constant stress with body mass, and larger body mass can lead to
467 increased foot stiffness (Chi and Roth, 2010). Combining this methodology with XROMM
468 data for elephants and other animals with large, fatty foot pads, however, would be
469 advantageous in determining the overall effect of deformation on the predictability of these
470 methods and on foot surface area in general, as this particular aspect of foot anatomy is the
471 most prone to deformation with body weight, due to its high compliance (Hutchinson, et
472 al., 2011). Overall, CT scans are a reliable resource for studies like these, and their utility in
473 determining foot surface area could potentially contribute to future studies on animal
474 locomotion and posture if used in conjunction with *in vivo* loading, centre of mass and
475 pressure data. However, as in this study, where quality of the models varied, results could
476 potentially be limited by the fidelity of the scans available, and therefore, more scans
477 available for each animal to have the option to pick and choose the most complete and
478 highest quality, as well as more computing power and high-end software, would be a boon
479 to future studies.

480 Most studies concerning underfoot areas and pressures have focused on humans and other
481 primates. Adaptations for arboreal locomotion have resulted in large functional differences
482 between the forelimb and hindlimb in primates (Schmitt and Hanna, 2004). Such
483 differences, would make them an interesting subject for a follow-up study.

484 Assigning specific trackmakers to fossilised trackways is a difficult task (Falkingham,
485 2014b). It is our hope that these results could be used to constrain potential trackmaker
486 identity. However, as an extrapolation from a bivariate plot, with a number of variables
487 unaccounted for such as soft tissue and substrate compliance, the applications of figure 3
488 and its predictions are currently limited, and such identifications of trackmakers must be
489 undertaken cautiously.

490 When predicting the skeletal surface area of the feet of extinct animals, and identifying
491 trackmakers, the many complexities of footprint formation must be taken into account. The
492 shape of footprints is determined not only by foot anatomy, but also dynamics of the limbs,
493 and substrate consistency (Falkingham, 2014a; Minter et al., 2007; Padian and Olsen,
494 1984). Underfoot pressures (Hatala et al., 2013), centre of mass position (Castanera et al.,
495 2013), and style of locomotion (Hatala et al., 2016) all contribute to variations in limb
496 dynamics, and consequently the morphology of a track. Given that foot size and shape is
497 the focus of this study, the findings herein concern matters of critical importance to
498 footprint formation and trackmaker identification, relating as they do to both anatomy and
499 dynamics.

500 When trying to model footprint formation and dynamics of extinct animals, centre of mass
501 and underfoot pressures of the animals in question are determining factors. When
502 considering these factors, the difference between manus and pes size and pressure is of
503 great importance. Disparity between the cranial and caudal parts of the body is especially
504 notable as previous biomechanical models have often underestimated mass in the cranial
505 half of the body (See discussion in Allen et al., 2009). Simply put, taking into account the
506 differences between soft tissue area in manus and pes could make a notable difference in
507 estimations of underfoot pressures and simulations of footprint formation. As an example,
508 when the skeletal remains of *Plateosaurus engelhardti* feet were laid flat, and their skin
509 areas predicted from alpha hulls, estimated manus skin area was 32% of pes area when
510 using the 1.67 multiplier from combined analyses, and 40% of pes area using the separate
511 multipliers (2 for manus, 1.6 for pes). Using body mass and centre of mass calculations
512 from Allen et al. (2013), these results predicted manus underfoot pressure of 80% pes
513 pressure when combined, and 64% when separate (Supplementary Material 1). This effect
514 should also be considered in the inverse when considering trackmaker anatomy from fossil
515 footprints. In this way, this method is a useful tool to consider in digital reconstruction and
516 trackmaker identification.

517 **Conclusions**

518 The surface areas of the skin of the foot *in situ* and of the foot's skeletal components are
519 strongly correlated and thus should be predictable in terrestrial tetrapods. Skin surface area
520 was approximately 1.67 times that of skeletal surface area (~2 times for manus, ~1.6 times
521 for pes, if analysed separately). This trend was not affected by body mass and showed little
522 evidence of being strongly affected by phylogeny. This predictability has potential in
523 aiding with estimating the size and possible species of trackmakers in the fossil record,
524 both by estimating the size of skeletal feet using footprints, and by estimating foot size, and
525 therefore potential footprint size, from fossil feet.

526

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535

536 **Author Contributions**

537 Research and analysis was conducted by ECS. Manuscript and figures by ECS, with
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540

541 **References**

- 542 Alexander, R. M., 2004. Bipedal animals, and their differences from humans. *J. Anat.* 204,
543 321-330.
- 544 Alexander, R. M., Bennet, M. B., Ker, R. F., 1986. Mechanical properties and function of
545 the paw pads of some mammals. *J. Zool. Lond.* 209, 405-419.
- 546 Allee, W.C., Schmidt, K., 1937. *Ecological animal geography*. John Wiley and Sons.
- 547 Allen, J.A., 1877. The influence of physical conditions in the genesis of species. *Radic.*
548 *Rev.* 1, 108–140.
- 549 Allen, V., Bates, K. T., Li, Z., Hutchinson, J. R., 2013. Linking the Evolution of Body
550 Shape and Locomotor Biomechanics in Bird-Line Archosaurs. *Nature* 497, 104.
- 551 Allen, V., Paxton, H., Hitchinson, J. R., 2009. Variation in Centre of Mass Estimates for
552 Extant Sauropsids and Its Importance for Reconstructing Inertial Properties of
553 Extinct Archosaurs. *Anat. Rec.* 292, 1442-1461.
- 554 Astley, H. C., Roberts, T. J., 2014. The mechanics of elastic loading and recoil in anuran
555 jumping. *J. Exp. Biol.* 217, 4372-4328.
- 556 Bates, K.T., Mannion, P.D., Falkingham, P.L., Brusatte, S.L., Hutchinson, J.R., Otero, A.,
557 Sellers, W.I., Sullivan, C., Stevens, K.A., Allen, V., 2016. Temporal and
558 phylogenetic evolution of the sauropod dinosaur body plan. *R. Soc. Open Sci.* 3,
559 150636.
- 560 Bejan, A., Marden, J.H., 2006. Unifying constructal theory for scale effects in running,
561 swimming and flying. *J. Exp. Biol.* 209, 238–248.
- 562 Biewener, A.A., 1989. Mammalian terrestrial locomotion and size. *Bioscience* 39, 776–
563 783.
- 564 Birn-Jeffery, A.V., Hubicki, C.M., Blum, Y., Renjewski, D., Hurst, J.W., Daley, M.A.,
565 2014. Don't break a leg: running birds from quail to ostrich prioritise leg safety and
566 economy on uneven terrain. *J. Exp. Biol.* 217, 3786–3796.
- 567 Blackburn, T.M., Gaston, K.J., Loder, N., 1999. Geographic gradients in body size: a
568 clarification of Bergmann's rule. *Divers. Distrib.* 5, 165–174.
- 569 Bliese, P., 2006. Multilevel Modeling in R (2.2)—A Brief Introduction to R, the multilevel
570 package and the nlme package. October.
- 571 Blomberg, S.P., Lefevre, J.G., Wells, J.A., Waterhouse, M., 2012. Independent contrasts
572 and PGLS regression estimators are equivalent. *Syst. Biol.* 61, 382–391.
- 573 Bonnan, M. F., Shulman, J., Varadharajan, R., Gilbert, C., Wilkes, M., Horner, A.,
574 Brainerd, E., 2016. Forelimb kinematics of rats using XROMM, with implications
575 for small eutherians and their fossil relatives. *PloS one* 11.
- 576 Bowker, R.M., Van Wulfen, K.K., Springer, S.E., Linder, K.E., 1998. Functional anatomy
577 of the cartilage of the distal phalanx and digital cushion in the equine foot and a
578 hemodynamic flow hypothesis of energy dissipation. *Am. J. Vet. Res.* 59, 961–968.
- 579 Brainerd, E.L., Baier, D.B., Gatesy, S.M., Hedrick, T.L., Metzger, K.A., Gilbert, S.L.,
580 Crisco, J.J., 2010. X-ray reconstruction of moving morphology (XROMM):
581 precision, accuracy and applications in comparative biomechanics research. *J. Exp.*
582 *Zool. Part Ecol. Integr. Physiol.* 313, 262–279.
- 583 Carrano, M. T., 1997. Morphological indicators of foot posture in mammals: a
584 statistical and biomechanical analysis. *Zool. J. Linnean Soc.* 121, 77-104.
- 585 Castanera, D., Vila, B., Razzolini, N.L., Falkingham, P.L., Canudo, J.I., Manning, P.L.,
586 Galobart, À., 2013. Manus track preservation bias as a key factor for assessing
587 trackmaker identity and quadrupedalism in basal ornithopods. *PLoS One* 8, e54177.
- 588 Cheung, J.T.M., Zhang, M., Leung, A.K.-L., Fan, Y.-B., 2005. Three-dimensional finite
589 element analysis of the foot during standing—a material sensitivity study. *J.*
590 *Biomech.* 38, 1045–1054.

- 591 Chi, K.J., Roth, V.L., 2010. Scaling and mechanics of carnivoran footpads reveal the
592 principles of footpad design. *J. R. Soc. Interface* 7, 1145–1155.
- 593 Cholewo, T.J., Love, S., 1999. Gamut boundary determination using alpha-shapes, in:
594 Color and Imaging Conference. Society for Imaging Science and Technology, pp.
595 200–204.
- 596 Clack, J.A., 1997. Devonian tetrapod trackways and trackmakers; a review of the fossils
597 and footprints. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* 130, 227–250.
- 598 Clarke, K.A., 2017. Principles of thermal ecology. Oxford University Press.
- 599 Coombs Jr, W. P., 1978. Theoretical aspects of cursorial adaptations in dinosaurs.
600 *Quarterly Rev. Biol.* 53, 393–418.
- 601 Dunning Jr, J.B., 1992. CRC handbook of avian body masses. CRC press.
- 602 Falkingham, P.L., 2014. Interpreting ecology and behaviour from the vertebrate fossil track
603 record. *J. Zool.* 292, 222–228.
- 604 Falkingham, P.L., Bates, K.T., Mannion, P.D., 2012. Temporal and palaeoenvironmental
605 distribution of manus-and pes-dominated sauropod trackways. *J. Geol. Soc.* 169,
606 365–370.
- 607 Falkingham, P.L., Bates, K.T., Margetts, L., Manning, P.L., 2011a. The ‘Goldilocks’
608 effect: preservation bias in vertebrate track assemblages. *J. R. Soc. Interface* 8,
609 1142–1154.
- 610 Falkingham, P.L., Bates, K.T., Margetts, L., Manning, P.L., 2011b. Simulating sauropod
611 manus-only trackway formation using finite-element analysis. *Biol. Lett.* 7, 142–
612 145.
- 613 Falkingham, P. L., 2014a. Interpreting Ecology and Behaviour from the Vertebrate
614 Fossil Track Record. *J. Zool.* 292, 222–228.
- 615 Falkingham, P.L., Gatesy, S. M., 2014b. The birth of a dinosaur footprint: Subsurface 3D
616 motion reconstruction and discrete element simulation reveal track ontogeny. *PNAS*
617 111, 18279–18284.
- 618 Farley, C.T., Glasheen, J., McMahon, T.A., 1993. Running springs: speed and animal size.
619 *J. Exp. Biol.* 185, 71–86.
- 620 Farlow, J.O., 1992. Sauropod tracks and trackmakers: integrating the ichnological and
621 skeletal records. *Zubia* 10.
- 622 Fedak, M.A., Heglund, N.C., Taylor, C.R., 1982. Energetics and mechanics of terrestrial
623 locomotion. II. Kinetic energy changes of the limbs and body as a function of speed
624 and body size in birds and mammals. *J. Exp. Biol.* 97, 23–40.
- 625 Felsenstein, J., 1985. Phylogenies and the comparative method. *Am. Nat.* 125, 1–15.
- 626 Ferraro, J.V., Binetti, K.M., 2014. American alligator proximal pedal phalanges resemble
627 human finger bones: Diagnostic criteria for forensic investigators. *Forensic Sci. Int.*
628 240, 151–e1.
- 629 Full, R.J., Kubow, T., Schmitt, J., Holmes, P., Koditschek, D., 2002. Quantifying dynamic
630 stability and maneuverability in legged locomotion. *Integr. Comp. Biol.* 42, 149–
631 157.
- 632 Garland, T., 1983. The relation between maximal running speed and body mass in
633 terrestrial mammals. *J. Zool.* 199, 157–170.
- 634 Gatesy, S. M., 1991. Hind limbs movements of the American alligator (*Alligator*
635 *mississippiensis*) and postural grades. *J. Zool.* 224, 577–588.
- 636 Gatesy, S.M., Baier, D.B., Jenkins, F.A., Dial, K.P., 2010. Scientific rotoscoping: a
637 morphology-based method of 3-D motion analysis and visualization. *J. Exp. Zool.*
638 *Part Ecol. Integr. Physiol.* 313, 244–261.
- 639 Gatesy, S.M., Biewener, A.A., 1991. Bipedal locomotion: effects of speed, size and limb
640 posture in birds and humans. *J. Zool.* 224, 127–147.

- 641 Gatesy, S. M., 2003. Direct and Indirect Track Features: What Sediment Did a Dinosaur
642 Touch? *Ichnos* 10, 91-98.
- 643 Gatesy, S.M., Falkingham, P.L., 2017. Neither bones nor feet: track morphological
644 variation and ‘preservation quality.’ *J. Vertebr. Paleontol.* e1314298.
- 645 Gatesy, S.M., Middleton, K. M., Jenkins Jr, F. A., Shubin, N. H., 1999. Three-dimensional
646 preservation of foot movements in Triassic theropod dinosaurs. *Nature* 399, 141–
647 144.
- 648 Gauthier, J., Kluge, A.G., Rowe, T., 1988. Amniote phylogeny and the importance of
649 fossils. *Cladistics* 4, 105–209.
- 650 Geyer, H., Seyfarth, A., Blickhan, R., 2006. Compliant leg behaviour explains basic
651 dynamics of walking and running. *Proc. R. Soc. Lond. B Biol. Sci.* 273, 2861–2867.
- 652 Grafen, A., 1989. The phylogenetic regression. *Philos. Trans. R. Soc. Lond. B. Biol. Sci.*
653 326, 119–157.
- 654 Harmon, L.J., Weir, J.T., Brock, C.D., Glor, R.E., Challenger, W., 2008. GEIGER:
655 investigating evolutionary radiations. *Bioinformatics* 24, 129–131.
- 656 Hatala, K.G., Dingwall, H.L., Wunderlich, R.E., Richmond, B.G., 2013. The relationship
657 between plantar pressure and footprint shape. *J. Hum. Evol.* 65, 21–28.
- 658 Hatala, K.G., Wunderlich, R.E., Dingwall, H.L., Richmond, B.G., 2016. Interpreting
659 locomotor biomechanics from the morphology of human footprints. *J. Hum. Evol.*
660 90, 38–48.
- 661 Heckel, F., Schwier, M., Peitgen, H.-O., 2009. Object-oriented application development
662 with MeVisLab and Python. *GI Jahrestag.* 154, 1338–51.
- 663 Henderson, D.M., 2006. Burly gaits: centers of mass, stability, and the trackways of
664 sauropod dinosaurs. *J. Vertebr. Paleontol.* 26, 907–921.
- 665 Hildebrand, M., 1980. The adaptive significance of tetrapod gait selection. *Am. Zool.* 20,
666 255–267.
- 667 Hinterhofer, C. H., Stanek, C. H., Haider, H., 2000. The effect of flat horseshoes, raised
668 heels and lowered heels on the biomechanics of the equine hoof assessed by finite
669 element analysis (FEA). *J. Vet. Med.* 47, 73-82.
- 670 Hodgins, J.K., Raibert, M.N., 1991. Adjusting step length for rough terrain locomotion.
671 *IEEE Trans. Robot. Autom.* 7, 289–298.
- 672 Hutchinson, J.R., Delmer, C., Miller, C.E., Hildebrandt, T., Pitsillides, A.A., Boyde, A.,
673 2011. From flat foot to fat foot: structure, ontogeny, function, and evolution of
674 elephant “sixth toes.” *Science* 334, 1699–1703.
- 675 Hutson, J.D., Hutson, K.N., 2014. A repeated-measures analysis of the effects of soft
676 tissues on wrist range of motion in the extant phylogenetic bracket of dinosaurs:
677 implications for the functional origins of an automatic wrist folding mechanism in
678 Crocodylia. *The Anatomical Record* 297, 1228-1249
- 679 Jackson, S.J., Whyte, M.A., Romano, M., 2009. Laboratory-controlled simulations of
680 dinosaur footprints in sand: a key to understanding vertebrate track formation and
681 preservation. *Palaios* 24, 222–238.
- 682 Jannel, A., Nair, J. P., Panagiotopoulou, O., Romilio, A., Salisbury, S. W., 2019. ‘Keep Your
683 Feet on the Ground’: Simulated Range of Motion and Hind Foot Posture of the
684 Middle Jurassic Sauropod *Rhoetosaurus brownei* and its Implications for
685 Sauropod Biology. *J. Morph.*
- 686 Kambic, R. E., Roberts, T. J., Gatesy, S. M., 2015. Guineafowl with a twist: asymmetric
687 limb control in steady bipedal locomotion. *J. Exp. Biol.* 218, 3836-3844.
- 688 Kilbourne, B.M., Carrier, D.R., 2016. Manipulated Changes in Limb Mass and Rotational
689 Inertia in Trotting Dogs (*Canis lupus familiaris*) and Their Effect on Limb
690 Kinematics. *J. Exp. Zool. Part Ecol. Genet. Physiol.* 325, 665–674.

- 691 Kilbourne, B.M., Hoffman, L.C., 2013. Scale effects between body size and limb design in
692 quadrupedal mammals. PLoS One 8, e78392.
- 693 Knaust, D., Hauschke, N., 2004. Trace fossils versus pseudofossils in Lower Triassic playa
694 deposits, Germany. Palaeogeogr. Palaeoclimatol. Palaeoecol. 215, 87–97.
- 695 Kubo, T., Sakamoto, M., Meade, A., Venditti, C., 2019. Transitions Between Foot
696 Postures are Associated with Elevated Rates of Body Size Evolution in
697 Mammals. PNAS. 116, 2618-2623.
- 698 Li, R., Lockley, M.G., Makovicky, P.J., Matsukawa, M., Norell, M.A., Harris, J.D., Liu,
699 M., 2008. Behavioral and faunal implications of Early Cretaceous deinonychosaur
700 trackways from China. Naturwissenschaften 95, 185–191.
- 701 Lockley, M.G., Farlow, J.O., Meyer, C.A., 1994. *Brontopodus* and *Parabrontopodus*
702 ichnogen. nov. and the significance of wide-and narrow-gauge sauropod trackways.
703 Gaia 10, 135–145.
- 704 Lockley, M.G., Xing, L., 2015. Flattened fossil footprints: implications for paleobiology.
705 Palaeogeogr. Palaeoclimatol. Palaeoecol. 426, 85–94.
- 706 Maddison, W.P., Maddison, D., 2001. Mesquite: a modular system for evolutionary
707 analysis.
- 708 McGuigan, M. R., Wilson, A. M., 2003. The effect of gait and digital flexor muscle
709 activation on limb compliance in the forelimb of the horse *Equus caballus*. J. Exp.
710 Biol. 206, 1325-1336.
- 711 McMahon, T.A., 1975. Using body size to understand the structural design of animals:
712 quadrupedal locomotion. J. Appl. Physiol. 39, 619–627.
- 713 Michilsens, F., Aerts, P., Van Damme, R., D’Août, K., 2009. Scaling of plantar pressures
714 in mammals. J. Zool. 279, 236–242.
- 715 Miller, C.E., Basu, C., Fritsch, G., Hildebrandt, T., Hutchinson, J.R., 2008. Ontogenetic
716 scaling of foot musculoskeletal anatomy in elephants. J. R. Soc. Interface 5, 465–
717 475.
- 718 Minter, N. J., Braddy, S. J., Davis, R. B., 2007. Between a Rock and a Hard Place:
719 Arthropod Trackways and Ichnotaxonomy. Lethaia 40, 365-375.
- 720 Morse, S.A., Bennett, M.R., Liutkus-Pierce, C., Thackeray, F., McClymont, J., Savage, R.,
721 Crompton, R.H., 2013. Holocene footprints in Namibia: the influence of substrate
722 on footprint variability. Am. J. Phys. Anthropol. 151, 265–279.
- 723 Nery, M.F., González, D.J., Hoffmann, F.G., Opazo, J.C., 2012. Resolution of the
724 laurasiatherian phylogeny: evidence from genomic data. Mol. Phylogenet. Evol. 64,
725 685–689.
- 726 Padian, K., Olsen, P. E., 1984. The Fossil Trackway *Pteraichnus*: Not Pterosaurian but
727 Crocodylian. J. Palaeontol. 58, 178-184.
- 728 Panagiotopoulou, O., Pataky, T.C., Day, M., Hensman, M.C., Hensman, S., Hutchinson,
729 J.R., Clemente, C.J., 2016. Foot pressure distributions during walking in African
730 elephants (*Loxodonta africana*). Open Sci. 3, 160203.
- 731 Panagiotopoulou, O., Pataky, T.C., Hill, Z., Hutchinson, J.R., 2012. Statistical parametric
732 mapping of the regional distribution and ontogenetic scaling of foot pressures
733 during walking in Asian elephants (*Elephas maximus*). J. Exp. Biol. 215, 1584–
734 1593.
- 735 Panagiotopoulou, O., Rankin, J. W., Gatesy, S. M., Hutchinson, J. R., 2016. A preliminary
736 case study of the effect of shoe-wearing on the biomechanics of a horse’s foot.
737 PeerJ 4, e2164.
- 738 Paradis, E., Claude, J., Strimmer, K., 2004. APE: analyses of phylogenetics and evolution
739 in R language. Bioinformatics 20, 289–290.

- 740 Pataky, T.C., Caravaggi, P., Savage, R., Parker, D., Goulermas, J.Y., Sellers, W.I.,
741 Crompton, R.H., 2008. New insights into the plantar pressure correlates of walking
742 speed using pedobarographic statistical parametric mapping (pSPM). *J. Biomech.*
743 41, 1987–1994.
- 744 Patla, A.E., 2003. Strategies for dynamic stability during adaptive human locomotion.
745 *IEEE Eng. Med. Biol. Mag.* 22, 48–52.
- 746 Platt, B.F., Hasiotis, S.T., 2006. Newly discovered sauropod dinosaur tracks with skin and
747 foot-pad impressions from the Upper Jurassic Morrison Formation, Bighorn Basin,
748 Wyoming, USA. *Palaios* 21, 249–261.
- 749 Pollitt, C. C., 2004. Anatomy and physiology of the inner hoof wall. *Clinical Techniques in*
750 *Equine Practice* 3, 1, 3-21
- 751 Prum, R.O., Berv, J.S., Dornburg, A., Field, D.J., Townsend, J.P., Lemmon, E.M.,
752 Lemmon, A.R., 2015. A comprehensive phylogeny of birds (Aves) using targeted
753 next-generation DNA sequencing. *Nature* 526, 569–573.
- 754 Qian, Z., Ren, L., Ding, Y., Hutchinson, J.R., Ren, L., 2013. A dynamic finite element
755 analysis of human foot complex in the sagittal plane during level walking. *PLoS One*
756 8, e79424.
- 757 Reilly, S. M., Elias, J. A., 1998. Locomotion in *Alligator mississippiensis*: kinematic
758 effects of speed and posture and their relevance to the sprawling-to-erect
759 paradigm. *J. Exp. Biol* 201, 2559-2574.
- 760 Regnault, S., Hermes, R., Hildebrandt, T., Hutchinson, J., Weller, R., 2013.
761 Osteopathology in the feet of rhinoceroses: lesion type and distribution. *J. Zoo*
762 *Wildl. Med.* 44, 918–927.
- 763 Revell, L.J., 2012. phytools: an R package for phylogenetic comparative biology (and other
764 things). *Methods Ecol. Evol.* 3, 217–223.
- 765 Rosenbaum, D., Hautmann, S., Gold, M., Claes, L., 1994. Effects of walking speed on
766 plantar pressure patterns and hindfoot angular motion. *Gait Posture* 2, 191–197.
- 767 Saitta, E. T., Rogers, C. , Brooker, R. A., Abbott, G. D., Kumar, S. , O'Reilly, S. S.,
768 Donohoe, P. , Dutta, S. , Summons, R. E., Vinther, J., 2017. Low fossilization
769 potential of keratin protein revealed by experimental taphonomy. *Palaeontology*, 60:
770 547-556.
- 771 Schaller, N. U., D'Août, K., Villa, R., Herkner, B., Aerts, P., 2011. Toe function and
772 dynamic pressure distribution in ostrich locomotion. *J. Exp. Biol.* 214, 1123-1130.
- 773 Schmitt, D., Hanna, J.B., 2004. Substrate alters forelimb to hindlimb peak force ratios in
774 primates. *J. Hum. Evol.* 46, 237–252.
- 775 Seebacher, F., Elsworth, P. G., Franklin, C. F. Ontogenetic changes of swimming
776 kinematics in a semi-aquatic reptile (*Crocodylus porosus*). *Aus. J. Zool* 51, 15-24.
- 777 Segal, A., Rohr, E., Orendurff, M., Shofer, J., O'Brien, M., Sangeorzan, B., 2004. The
778 effect of walking speed on peak plantar pressure. *Foot Ankle Int.* 25, 926–933.
- 779 Snyder, R.C., 1962. Quadrupedal and bipedal locomotion of lizards. *Copeia* 1952, 64–70.
- 780 Taylor, C.R., Shkolnik, A., Dmi'el, R., Baharav, D., Borut, A., 1974. Running in cheetahs,
781 gazelles, and goats: energy cost and limb configuration. *Am. J. Physiol. Content*
782 227, 848–850.
- 783 Van Sittert, S., Skinner, J., Mitchell, G., 2015. Scaling of the appendicular skeleton of the
784 giraffe (*Giraffa camelopardalis*). *J. Morphol.* 276, 503–516.
- 785 Warner SE, Pickering P, Panagiotopoulou O, Pfau T, Ren L, Hutchinson JR., 2013. Size-
786 Related Changes in Foot Impact Mechanics in Hoofed Mammals. *PLoS ONE* 8(1):
787 e54784

788 Witmer, L.M., Thomason, J.J., 1995. The extant phylogenetic bracket and the importance
 789 of reconstructing soft tissues in fossils. *Funct. Morphol. Vertebr. Paleontol.* 1, 19–
 790 33.
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793 Tables

794 Table 1 – Regressions and Confidence Intervals for Main Analyses

Analysis	Linear Regression	Linear R ²	Log Regression	Log R ²	95% CI	P value
Pose 1 - All limbs	$y = 0.51x + 146.71$	R ² = 0.94	$y = 0.59x^{0.99}$	R ² = 0.99	1.922 ± 0.06186	<2.2E-16
Pose 1 - Forelimbs	$y = 0.45x + 641.27$	R ² = 0.92	$y = 0.52x^{0.99}$	R ² = 0.99	1.916 ± 7.887E-02	3.27E-15
Pose 1 - Hindlimbs	$y = 0.59x - 292.02$	R ² = 0.97	$y = 0.64x^{0.98}$	R ² = 0.99	1.9229 ± 0.0632	<2E-16
Pose 2a -All limbs	$y = 0.20x + 1303.8$	R ² = 0.82	$y = 0.87x^{0.91}$	R ² = 0.99	3.9266 ± 0.3584	1.93E-11
Pose 2a -Forelimbs	$y = 0.21x + 1345.4$	R ² = 0.85	$y = 0.69x^{0.93}$	R ² = 0.97	3.9614 ± 0.3954	8.68E-09
Pose 2a -Hindlimbs	$y = 0.19x + 1177.4$	R ² = 0.79	$y = 1.06x^{0.89}$	R ² = 0.96	4.1603 ± 0.4157	2.08E-10
Pose 2b - All limbs	$y = 0.48x + 436.75$	R ² = 0.87	$y = 0.58x^{0.98}$	R ² = 0.97	1.856 ± 0.1199	5.98E-15
Pose 2b - Forelimbs	$y = 0.52x + 410.47$	R ² = 0.89	$y = 0.47x^{0.10}$	R ² = 0.97	1.7074 ± 0.1388	3.39E-10
Pose 2b - Hindlimbs	$y = 0.44x + 535.85$	R ² = 0.89	$y = 0.71x^{0.96}$	R ² = 0.97	2.029 ± 0.139	4.83E-14
Pose 2c - All limbs	$y = 0.74x - 700.51$	R ² = 0.93	$y = 0.49x^{1.00}$	R ² = 0.97	1.279 ± 6.225E-02	<2.2E-16
Pose 2c - Forelimbs	$y = 0.79x - 1120.2$	R ² = 0.95	$y = 0.40x^{1.02}$	R ² = 0.97	1.211 ± 6.473E-02	3.03E-13
Pose 2c - Hindlimbs	$y = 0.69x - 228.13$	R ² = 0.92	$y = 0.57x^{0.99}$	R ² = 0.97	1.333 ± 7.677E-02	8.04E-16

795

796 Table 2 – Regressions and Confidence Intervals for Analysis Subgroups

Analysis	Linear Regression	Linear R ²	Log Regression	Log R ²	95% CI	P value
Unguligrade	$y = 0.36x - 593.56$	R ² = 0.95	$y = 0.27x^{1.01}$	R ² = 0.97	2.6121 ± 0.2903	0.000844
Digitigrade	$y = 0.19x + 1823.1$	R ² = 0.83	$y = 2.02x^{0.84}$	R ² = 0.97	4.336 ± 0.537	2.02E-06
Plantigrade	$y = 0.74x + 1128.3$	R ² = 0.96	$y = 0.35x^{1.06}$	R ² = 0.99	1.29686 ± 0.08747	1.25E-07
Terrestrial	$y = 0.45x + 491.99$	R ² = 0.91	$y = 0.68x^{0.96}$	R ² = 0.91	1.9998 ± 0.1769	4.25E-08
Semi-aquatic	$y = 0.77x + 408.03$	R ² = 1.00	$y = 0.42x^{1.02}$	R ² = 0.99	1.30129 ± 0.02233	4.26E-09
Erect Posture	$y = 0.48x + 588.49$	R ² = 0.89	$y = 0.94x^{0.93}$	R ² = 0.95	1.8517 ± 0.1486	1.37E-10
Sprawling Posture	$y = 0.51x - 19.70$	R ² = 0.99	$y = 0.50x^{0.99}$	R ² = 1.00	1.96139 ± 0.06779	1.13E-07
Birds	$y = 0.59x + 32.25$	R ² = 1.00	$y = 0.87x^{0.96}$	R ² = 0.99	1.69386 ± 0.01636	1.59E-09
Mammals	$y = 0.48x + 903.78$	R ² = 0.87	$y = 0.57x^{0.98}$	R ² = 0.91	1.8353 ± 0.2018	9.87E-07

797

798 Figure Legends

799 Figure 1 - Projected area calculated from 3D models. A) *Hippopotamus* Left forelimb, soft tissue and bones
800 reconstructed from CT data. B) The soft tissue was cropped at a point representative of the area that would
801 contact the ground during life. The bones were cropped based on the same posterior extent (pose 1). C) The
802 alpha shape (pink) and the convex hull (green) were used to determine underfoot area of the bones alone and
803 D) the soft tissue. E) Bones were laid flat for a more repeatable approach (pose 2). Where semi-digitigrade
804 animals were treated as digitigrade (pose 2a) only bones in pink were used, where semi-digitigrade animals
805 were treated as intermediate between digitigrade and plantigrade (pose 2b), blue and pink bones were used,
806 and where semi-digitigrade animals were treated as plantigrade, all bones including those in green were used.
807 F) Alpha shapes for poses 2a-c, where pink is 2a, blue is 2b, and green is 2c. G-K) Distinctive foot
808 morphologies in the data set. Scale bar = 10cm for all but G, where scale bar = 1cm.

809
810 Figure 2 – Bar graph showing projected skin surface area as a percentage of projected skeletal surface area
811 across all specimens in A) Pose 1, with phylogeny for context, and B) Pose 2 (for elephant, rhino, and hippo,
812 main bar represents Pose 2b and additional bars show poses 2a and 2c). Silhouettes from Phylopic. Mammalia
813 data are in purple, Aves data in red, Crocodylia data in green, Lepidosauria data in blue, and Lissamphibia in
814 yellow.

815
816 Figure 3 – Log₁₀ plots for projected skin surface area against projected skeletal surface area in A) Pose 1, for
817 all limbs, B) For Pose 1, for forelimbs, C) For Pose 1, for hindlimbs, Silhouettes from Phylopic. All numbers
818 rounded to two significant figures. Mammalia data are in purple, Aves data in red, Crocodylia data in green,
819 Lepidosauria data in blue, and Lissamphibia in yellow.

820
821 Figure 4 - Log₁₀ plots for projected skin surface area against projected skeletal surface area for A) Pose 2a, all
822 limbs, B) Pose 2b, all limbs, and C) For Pose 2c, all limbs. Silhouettes from Phylopic. All numbers rounded
823 to two significant figures. Mammalia data are in purple, Aves data in red, Crocodylia data in green,
824 Lepidosauria data in blue, and Lissamphibia in yellow.

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828 Supplementary Figure Legends

829 Supplementary material 1: Supplementary tables – Additional data including p-values for all analysis,
830 calculated soft-tissue and skeletal areas, approximate body masses for all animals, data for analyses with
831 smallest and largest taxa removed, and demonstration of utility using *Plateosaurus engelhardti*.

832
833 Supplementary material 2: Supplementary graphs – Plots for projected skin surface area against projected
834 skeletal surface area in Pose 1 and Pose 2, presented as sub-groups by phylogeny and ecology.

835
836 Supplementary material 3: Supplementary outlines – Top-down projections of models used in study, showing
837 alpha shapes and convex hulls.

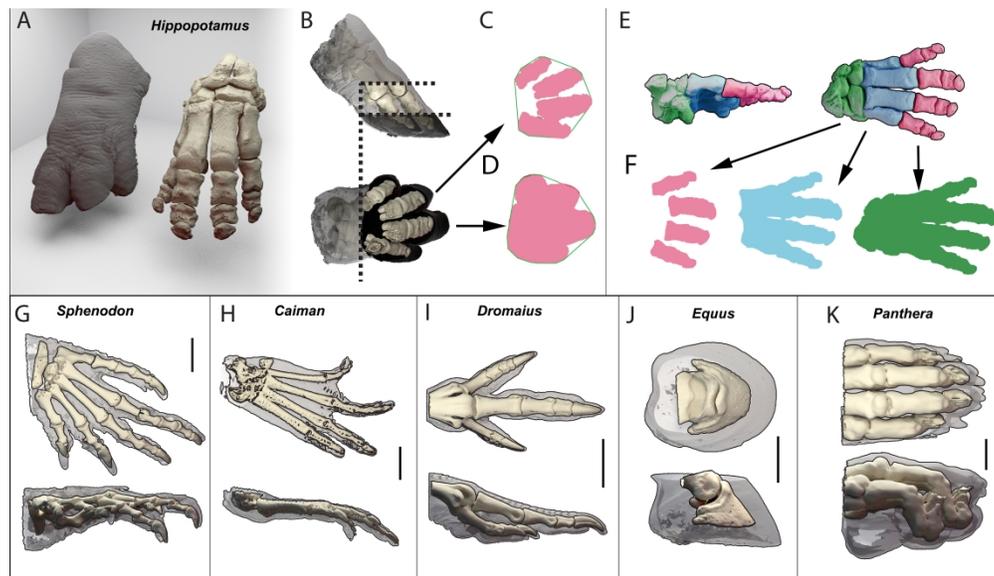


Figure 1 - Projected area calculated from 3D models. A) Hippopotamus Left forelimb, soft tissue and bones reconstructed from CT data. B) The soft tissue was cropped at a point representative of the area that would contact the ground during life. The bones were cropped based on the same posterior extent (pose 1). C) The alpha shape (pink) and the convex hull (green) were used to determine underfoot area of the bones alone and D) the soft tissue. E) Bones were laid flat for a more repeatable approach (pose 2). Where semi-digitigrade animals were treated as digitigrade (pose 2a) only bones in pink used, where semi-digitigrade animals were treated as intermediate between digitigrade and plantigrade (pose 2b), blue and pink bones were used, and where semi-digitigrade animals were treated as plantigrade, all bones including those in green were used. F) Alpha shapes for poses 2a-c, where pink is 2a, blue is 2b, and green is 2c. G-K) Distinctive foot morphologies in the data set. Scale bar = 10cm for all but G, where scale bar = 1cm.

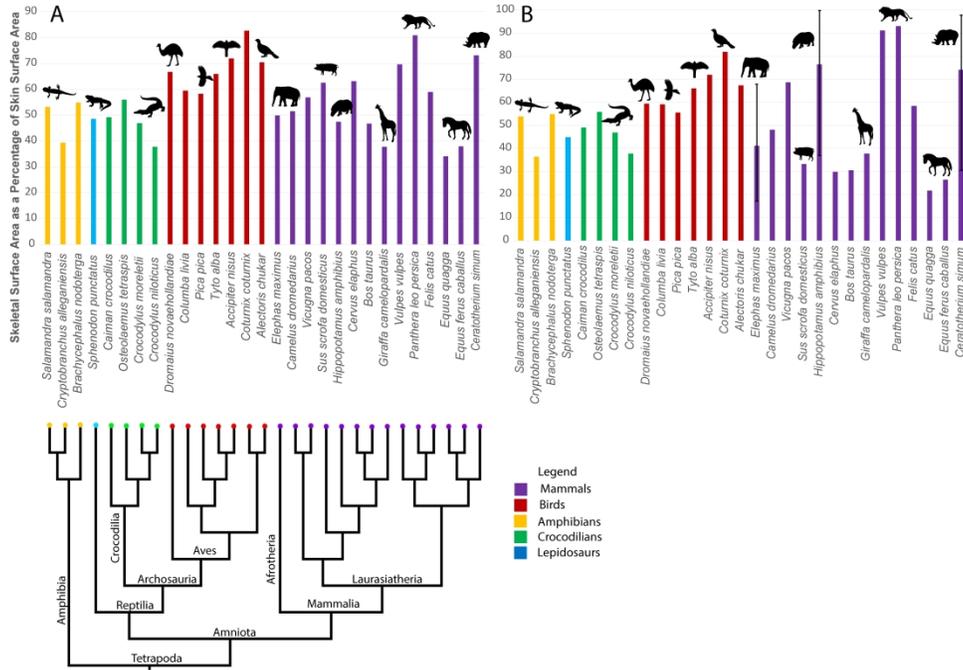


Figure 2 – Bar graph showing projected skin surface area as a percentage of projected skeletal surface area across all specimens in A) Pose 1, with phylogeny for context, and B) Pose 2 (for elephant, rhino, and hippo, main bar represents Pose 2b and additional bars show poses 2a and 2c). Silhouettes from Phylopic. Mammalia data are in purple, Aves data in red, Crocodylia data in green, Lepidosauria data in blue, and Lissamphibia in yellow.

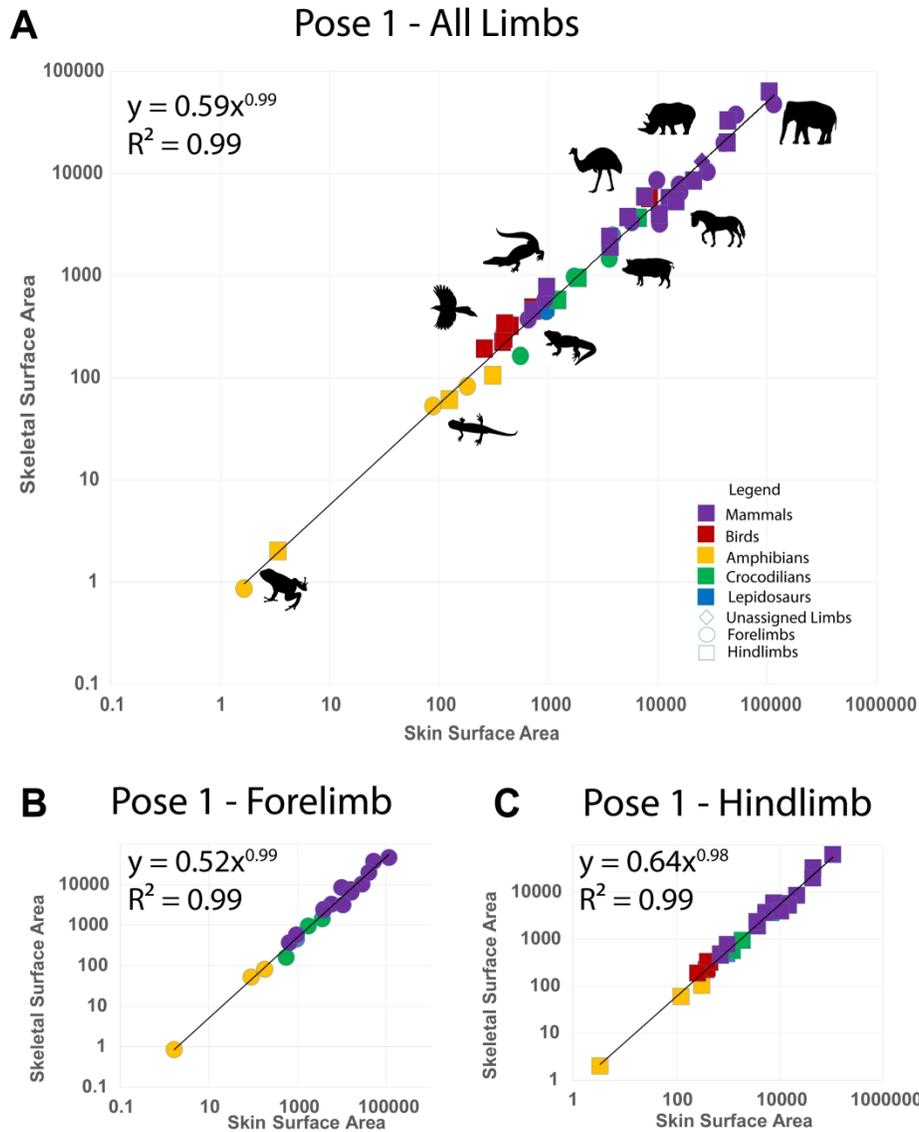


Figure 3 – Log₁₀ plots for projected skin surface area against projected skeletal surface area in A) Pose 1, for all limbs, B) For Pose 1, for forelimbs, C) For Pose 1, for hindlimbs, Silhouettes from Phylopic. All numbers rounded to two significant figures. Mammalia data are in purple, Aves data in red, Crocodylia data in green, Lepidosauria data in blue, and Lissamphibia in yellow.

Poses 2a-2c - All Limbs

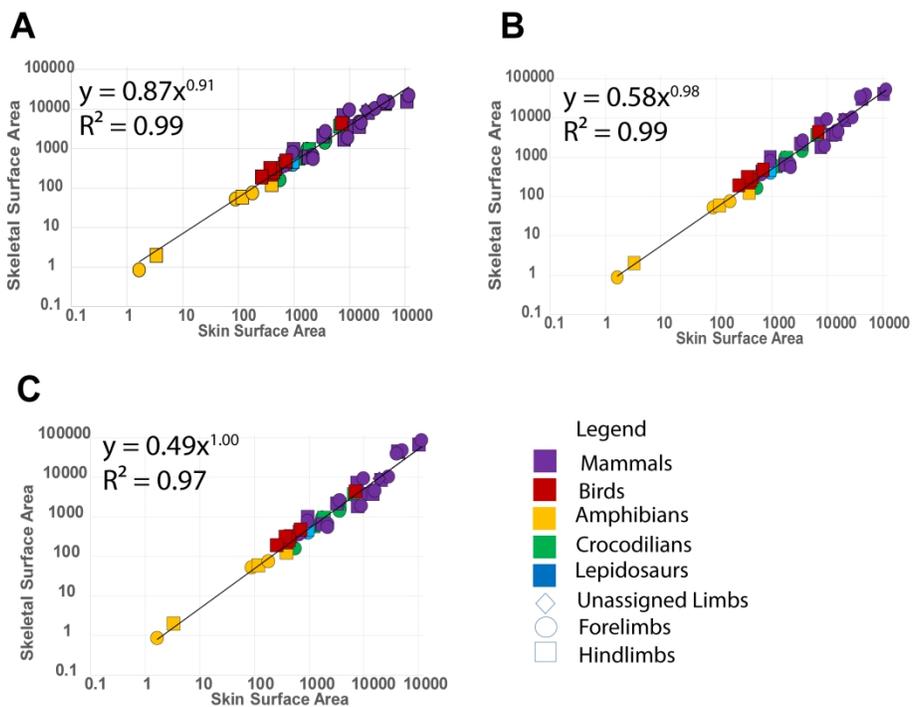


Figure 4 - Log10 plots for projected skin surface area against projected skeletal surface area for A) Pose 2a, all limbs, B) Pose 2b, all limbs, and C) For Pose 2c, all limbs. Silhouettes from Phylopic. All numbers rounded to two significant figures. Mammalia data are in purple, Aves data in red, Crocodylia data in green, Lepidosauria data in blue, and Lissamphibia in yellow.

Supplementary Tables

Supplementary Table 1 – Phylogenetic Comparative Tests for All Limbs in All Poses

Analysis	Adjusted R ²	PIC			PGLS			T value	P value
		CI	SE	P value	CI	SE	P value		
Pose 1	0.92	2.24	0.124	2.20E-16	2,24	0.124	18.0588	<0.0001	
Pose 2a	0.5228	3.3875	0.6019	5.67E-06	2.713	0.263	10.12195	<0.0001	
Pose 2b	0.6601	2.0175	0.2711	5.28E-08	1.7171	0.086	19.86243	<0.0001	
Pose 2c	0.8483	1.45E+00	0.1156	8.79E-13	1.044	0.08	12.97394	0.00E+00	

Pagel's Lambda

	Pagel's Lambda		
	Combined Data	Forelimb	Hindlimb
Pose 1	1.027319	1.045933	1.030825
All Limbs	Pose 2a	Pose 2b	Pose 2c
	1.017103	1.030825	-0.87799

Supplementary Table 2 – Area (mm²) Measurements for All Animals and Proportions of Skeleton to Skin Surface Area (%)

Pose 1				
Specimen	Fore/Hind Foot	Skin SA	Skel SA	Skeleton as % of Skin
<i>Salamandra salamandra</i>	Forefoot	88.93501	52.14457	58.6322122
<i>Salamandra salamandra</i>	Hindfoot	124.8898	59.64688	47.75962093
<i>Cryptobranchus alleganiensis</i>	Forefoot	181.4453	82.04254	45.21613433
<i>Cryptobranchus alleganiensis</i>	Hindfoot	311.3837	103.9794	33.39268462
<i>Brachycephalus nodoterga</i>	Forefoot	1.6515	0.852027	51.59112909
<i>Brachycephalus nodoterga</i>	Hindfoot	3.39012	1.973128	58.20229633
<i>Sphenodon punctatus</i>	Forefoot	962.9668	447.0096	46.42004335
<i>Sphenodon punctatus</i>	Hindfoot	960.4319	487.3412	50.74188493
<i>Crocodylus niloticus</i>	Forefoot	553.7884	162.1849	29.28643561
<i>Crocodylus niloticus</i>	Hindfoot	1228.612	566.4358	46.10373211
<i>Osteolaemus teraspis</i>	Forefoot	1733.117	962.1605	55.51618621
<i>Osteolaemus teraspis</i>	Hindfoot	3678.328	2070.685	56.29419196
<i>Caiman crocodilus</i>	Hindfoot	1902.971	935.316	49.15029816
<i>Crocodylus moreletii</i>	Forefoot	3619.647	1447.981	40.00337567
<i>Crocodylus moreletii</i>	Hindfoot	6721.266	3618.094	53.83053494
<i>Alectoris chukar</i>	Hindfoot	451.7428	318.1106	70.4185217
<i>Tyto alba</i>	Hindfoot	721.4122	475.5846	65.92411232
<i>Pica pica</i>	Hindfoot	382.6398	222.9701	58.2715291
<i>Columba livia</i>	Hindfoot	397.7637	236.5437	59.46839319
<i>Coturnix coturnix</i>	Hindfoot	404.1557	334.2137	82.69428892

<i>Accipiter nisus</i>	Hindfoot	262.7824	189.0737	71.95065971
<i>Dromaius novaehollandiae</i>	Hindfoot	8524.232	5689.942	66.75019735
<i>Bos taurus</i>	Forefoot	15663.52	7659.069	48.8974879
<i>Bos taurus</i>	Hindfoot	12739.92	5669.063	44.49841375
<i>Elephas maximus</i>	Forefoot	115297.7	47094.3	40.84583773
<i>Elephas maximus</i>	Hindfoot	106205	62562.12	58.90696806
<i>Ceratotherium simum</i>	Forefoot	52322.45	37586.9	71.8370514
<i>Ceratotherium simum</i>	Hindfoot	43938.84	32640.53	74.28627909
<i>Vicugna pacos</i>	Forefoot	3879.717	2447.911	63.09507459
<i>Vicugna pacos</i>	Hindfoot	3737.41	1889.661	50.56071099
<i>Giraffa camelopardalis</i>	Forefoot	28591.02	10324.47	36.11087691
<i>Giraffa camelopardalis</i>	Hindfoot	21393.04	8422.208	39.36892218
<i>Panthera leo persica</i>	Forefoot	9849.389	8485.304	86.15055843
<i>Panthera leo persica</i>	Hindfoot	7690.173	5819.748	75.6777249
<i>Felis catus</i>	Forefoot	651.6308	367.1313	56.34038863
<i>Felis catus</i>	Hindfoot	724.9928	446.6624	61.6092115
<i>Equus ferus caballus</i>	Forefoot	16103.96	6521.98	40.49922225
<i>Equus ferus caballus</i>	Hindfoot	14886.19	5258.705	35.32606867
<i>Sus scrofa</i>	Forefoot	5833.796	3301.937	56.60015442
<i>Sus scrofa</i>	Hindfoot	5410.751	3703.087	68.43944159
<i>Cervus elaphus</i>	Forefoot	3876.212	2398.473	61.87673137
<i>Cervus elaphus</i>	Hindfoot	3644.912	2343.958	64.30766863
<i>Equus quagga</i>	Forefoot	10510.49	3188.5	30.33636481
<i>Equus quagga</i>	Hindfoot	10438.59	3927.968	37.62927945
<i>Camelus dromedarius</i>	Unassigned	25222.78	12990.49	51.50299004
<i>Vulpes vulpes</i>	Forefoot	939.0155	575.1637	61.25178197
<i>Vulpes vulpes</i>	Hindfoot	974.4242	759.2161	77.91433447
<i>Hippopotamus amphibius</i>	Forefoot	40556.15	19879.08	49.01619162
<i>Hippopotamus amphibius</i>	Hindfoot	43485.7	19909.79	45.78468328
Pose 2				
<i>Salamandra salamandra</i>	Forefoot	88.93501	52.14457	58.6322122
<i>Salamandra salamandra</i>	Hindfoot	118.7929	58.1717	48.9690014
<i>Cryptobranchus alleganiensis</i>	Forefoot	179.4845	75.75179	42.2052092
<i>Cryptobranchus alleganiensis</i>	Hindfoot	398.9009	121.8758	30.55289108
<i>Brachycephalus nodoterga</i>	Forefoot	1.6515	0.852027	51.59112909
<i>Brachycephalus nodoterga</i>	Hindfoot	3.39012	1.973128	58.20229633
<i>Sphenodon punctatus</i>	Forefoot	962.9668	394.6228	40.97989334
<i>Sphenodon punctatus</i>	Hindfoot	960.432	467.5052	48.67655561
<i>Crocodylus niloticus</i>	Forefoot	553.7884	162.1849	29.28643561
<i>Crocodylus niloticus</i>	Hindfoot	1228.612	566.4358	46.10373211
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<i>Crocodylus moreletii</i>	Forefoot	3619.647	1447.981	40.00337567
<i>Crocodylus moreletii</i>	Hindfoot	6721.266	3618.094	53.83053494
<i>Alectoris chukar</i>	Hindfoot	463.5517	312.3395	67.37963874
<i>Tyto alba</i>	Hindfoot	721.4122	475.5846	65.92411232

<i>Pica pica</i>	Hindfoot	398.4393	221.1307	55.49920666
<i>Columba livia</i>	Hindfoot	430.617	254.0651	59.00024677
<i>Coturnix coturnix</i>	Hindfoot	374.0761	306.0736	81.82121905
<i>Accipiter nisus</i>	Hindfoot	262.7824	189.0737	71.95065971
<i>Dromaius novaehollandiae</i>	Hindfoot	7189.013	4273.903	59.45048029
<i>Bos taurus</i>	Forefoot	14860.38	4672.811	31.44475656
<i>Bos taurus</i>	Hindfoot	12400.11	3656.876	29.49067084
<i>Elephas maximus</i>	Forefoot pose 2a	115297.7	21888.71	18.98452046
<i>Elephas maximus</i>	Hindfoot pose 2a	106205	16361.32	15.40542458
<i>Elephas maximus</i>	Forefoot pose 2b	115297.7	53085.99	46.04255484
<i>Elephas maximus</i>	Hindfoot pose 2b	106205	39665.72	37.34827271
<i>Elephas maximus</i>	Forefoot pose 2c	115297.7	85872.49	74.47894594
<i>Elephas maximus</i>	Hindfoot pose 2c	106205	64990.32	61.19330515
<i>Ceratotherium simum</i>	Forefoot pose 2a	40263.47	15929.85	39.56403613
<i>Ceratotherium simum</i>	Hindfoot pose 2a	43571.67	14885.36	34.1629407
<i>Ceratotherium simum</i>	Forefoot pose 2b	50319.26	38994.05	77.49327689
<i>Ceratotherium simum</i>	Hindfoot pose 2b	43938.84	31147.23	70.88768228
<i>Ceratotherium simum</i>	Forefoot pose 2c	40263.47	40068.14	99.51486813
<i>Ceratotherium simum</i>	Hindfoot pose 2c	43571.67	43695.43	100.2840375
<i>Vicugna pacos</i>	Forefoot	3651.553	2680.183	73.39842765
<i>Vicugna pacos</i>	Hindfoot	3349.815	2141.799	63.93783478
<i>Giraffa camelopardalis</i>	Forefoot	28591.02	10324.47	36.11087691
<i>Giraffa camelopardalis</i>	Hindfoot	21393.04	8422.208	39.36892218
<i>Panthera leo persica</i>	Forefoot	9849.389	9416.026	95.60010391
<i>Panthera leo persica</i>	Hindfoot	7690.173	6969.753	90.63193541
<i>Felis catus</i>	Forefoot	651.6308	367.1313	56.34038863
<i>Felis catus</i>	Hindfoot	680.9717	412.0175	60.50435475
<i>Equus ferus caballus</i>	Forefoot	16103.96	4560.854	28.3213136
<i>Equus ferus caballus</i>	Hindfoot	14886.19	3679.188	24.71545112
<i>Sus scrofa</i>	Forefoot	2182.029	665.3783	30.49356342
<i>Sus scrofa</i>	Hindfoot	1730.437	621.656	35.92479041
<i>Cervus elaphus</i>	Forefoot	2213.147	556.7199	25.15511885
<i>Cervus elaphus</i>	Hindfoot	1835.489	631.2443	34.39107695
<i>Equus quagga</i>	Forefoot	9146.338	1911.856	20.902962
<i>Equus quagga</i>	Hindfoot	7881.42	1775.487	22.5275036
<i>Camelus dromedarius</i>	Unassigned	19383.7	9322.263	48.09331236
<i>Vulpes vulpes</i>	Forefoot	939.0155	789.1365	84.03871265
<i>Vulpes vulpes</i>	Hindfoot	974.4242	958.9808	98.41512652
<i>Hippopotamus amphibius</i>	Forefoot pose 2a	40263.47	15929.85	39.56403613
<i>Hippopotamus amphibius</i>	Hindfoot pose 2a	43571.67	14885.36	34.1629407
<i>Hippopotamus amphibius</i>	Forefoot pose 2b	40263.47	34742	86.28665173
<i>Hippopotamus amphibius</i>	Hindfoot pose 2b	43571.67	29026.14	66.61700047
<i>Hippopotamus amphibius</i>	Forefoot pose 2c	40263.47	40068.14	99.51486813
<i>Hippopotamus amphibius</i>	Hindfoot pose 2c	43571.67	43695.43	100.2840375

Supplementary Table 3 - Body Mass for Each Subject Animal, Source of Data, and F and p Values for GLS with Body Mass as a Predictor of Correlatory Power for All Poses

Species	Body Mass (g)	Source				
<i>Salamandra salamandra</i>	19.1	Encyclopedia of Life				
<i>Cryptobranchus alleganiensis</i>	358	Encyclopedia of Life				
<i>Brachycephalus nodoterga</i>	1	Pires Jr et al, 2005 (Toxicon, vol. 45, issue 1, 73-79)				
<i>Sphenodon punctatus</i>	700	Animal Diversity Web				
<i>Caiman crocodilus</i>	2174	Hutchinson metadata (Crocbase)				
<i>Osteolaemus tetraspis</i>	7820	Hutchinson metadata (Cocbase)				
<i>Crocodylus moreletii</i>	14150	Hutchinson metadata (Crocbase)				
<i>Crocodylus niloticus</i>	1336	Hutchinson metadata (Crocbase)				
<i>Dromaius novaehollandiae</i>	34200	CRC Handbook of Avian Body Masses				
<i>Columba livia</i>	358.7	Encyclopedia of Life				
<i>Pica pica</i>	151.3865	Encyclopedia of Life				
<i>Tyto alba</i>	520	Animal Diversity Web				
<i>Accipiter nisus</i>	237.5	CRC Handbook of Avian Body Masses				
<i>Coturnix coturnix</i>	112.5	Encyclopedia of Life				
<i>Alectoris chukar</i>	503.5	CRC Handbook of Avian Body Masses				
<i>Elephas maximus</i>	3269794.34	Pantheria				
<i>Camelus dromedarius</i>	492714.47	Pantheria				
<i>Vicugna pacos</i>	64900	Pantheria				
<i>Sus scrofa domesticus</i>	84471.54	Pantheria				
<i>Hippopotamus amphibius</i>	1536310.4	Pantheria				
<i>Cervus elaphus</i>	240867.13	Pantheria				
<i>Bos taurus</i>	618642.42	Pantheria				
<i>Giraffa camelopardalis</i>	964654.73	Pantheria				
<i>Vulpes vulpes</i>	4820.36	Pantheria				
<i>Panthera leo persica</i>	158623.93	Pantheria				
<i>Felis catus</i>	2884.8	Pantheria				
<i>Equus quagga</i>	400000	Pantheria				
<i>Equus ferus caballus</i>	403598.53	Pantheria				
<i>Ceratotherium simum</i>	2285939.43	Pantheria				
	Pose 1			Pose 2a	Pose 2b	Pose 2c
Body Mass GLS	Combined Data	Forelimb	Hindlimb	Combined Data	Combined Data	Combined Data
F-Statistic	0.6473	0.3169	1.0615	4.8346	0.0615	0.01384
p-value	0.4287	0.5813	0.8062	0.0374	0.8062	0.9073

Supplementary Table 4 – Slope Uncertainties for all Poses and Combinations of Limbs

	All Limbs 1	Forelimbs 1	Hindlimbs 1	All Limbs 2a	Forelimbs 2a	Hindlimbs 2a
Slope	1.83	2.05	1.66	3.82	3.74	4.08
Uncertainty (Slope)	0.07	0.14	0.05	0.28	0.42	0.42
Correlation Coefficient (R²)	0.94	0.92	0.97	0.80	0.82	0.79
F Statistic	700.03	217.89	1006.50	182.37	80.86	95.99
Regression of Sum of Squares	2.62E+10	1.33E+10	1.27E+10	2.16E+10	1.13E+10	1.03E+10
Y-Intercept	551.14	-127.88	756.10	-2637.39	-2776.37	-2629.16
Uncertainty (Y-Intercept)	992.47	2047.32	742.55	1908.16	3271.67	2369.49
Standard Error for Y Estimate	6114.66	7825.46	3557.13	10892.56	11822.08	10379.59
Degrees of Freedom	47.00	18.00	26.00	47.00	18.00	26.00
Residual Sum of Squares	1.76E+09	1.10E+09	3.29E+08	5.58E+09	2.52E+09	2.80E+09
	All Limbs 2b	Forelimbs 2b	Hindlimbs 2b	All Limbs 2c	Forelimbs 2c	Hindlimbs 2c
Slope	1.83	1.71	2.03	1.27	1.23	1.32
Uncertainty (Slope)	0.10	0.14	0.14	0.05	0.07	0.08
Correlation Coefficient (R²)	0.89	0.89	0.89	0.93	0.95	0.92
F Statistic	367.33	151.26	213.21	648.43	345.79	289.64
Regression of Sum of Squares	2.47E+10	1.29E+10	1.17E+10	2.54E+10	1.31E+10	1.21E+10
Y-Intercept	618.26	900.74	43.67	1652.98	1986.94	1142.07
Uncertainty (Y-Intercept)	1325.83	2363.94	1571.50	986.25	1534.95	1335.15
Standard Error for Y Estimate	8205.80	9233.32	7419.28	6255.62	6162.94	6452.82
Degrees of Freedom	47.00	18.00	26.00	47.00	18.00	26.00
Residual Sum of Squares	3.16E+09	1.53E+09	1.43E+09	1.84E+09	6.84E+08	1.08E+09

Supplementary Table 5 – List of Taxa Used with Common Names and Latin Names

Latin Name	Common Name
<i>Salamandra salamandra</i>	Salamandra
<i>Cryptobranchus alleganiensis</i>	Hellbender
<i>Brachycephalus nodoterga</i>	Saddleback Toad
<i>Sphenodon punctatus</i>	Tuatara
<i>Caiman crocodilus</i>	Nile Crocodile
<i>Osteolaemus tetraspis</i>	Dwarf Crocodile
<i>Crocodylus moreletii</i>	Spectacled Caiman
<i>Crocodylus niloticus</i>	Morelet's Crocodile
<i>Dromaius novaehollandiae</i>	Chukar
<i>Columba livia</i>	Barn Owl

<i>Pica pica</i>	Magpie
<i>Tyto alba</i>	Pigeon
<i>Accipiter nisus</i>	Quail
<i>Coturnix coturnix</i>	Sparrowhawk
<i>Alectoris chukar</i>	Emu
<i>Elephas maximus</i>	Cow
<i>Camelus dromedarius</i>	Elephant
<i>Vicugna pacos</i>	Rhinoceros
<i>Sus scrofa domesticus</i>	Alpaca
<i>Hippopotamus amphibius</i>	Giraffe
<i>Cervus elaphus</i>	Lion
<i>Bos taurus</i>	Cat
<i>Giraffa camelopardalis</i>	Horse
<i>Vulpes vulpes</i>	Pig
<i>Panthera leo persica</i>	Deer
<i>Felis catus</i>	Zebra
<i>Equus quagga</i>	Camel
<i>Equus ferus caballus</i>	Fox
<i>Ceratotherium simum</i>	Hippopotamus

Supplementary Table 6 – Examples of Results with Large and Small Animals Removed

	R Squared	Equation	Multiplier
Original Data	0.9877	$y=0.5901x+0.9865$	1.671751
Without Largest	0.9848	$y=0.6225x+0.9777$	1.569478
Without Smallest	0.9754	$y=0.7582x+0.9592$	1.265102
Without Largest and Smallest	0.9636	$y=0.969x+0.9257$	0.955315

Supplementary Table 7 – Example of Study Utility Using *Plateosaurus engelhardti*

<i>Plateosaurus</i>	Skeleton	Skin (Combined Estimate)	Skin (Manus and Pes Distinct)
Manus Area	0.0194	0.032398	0.0388
Pes Area	0.0605	0.101035	0.0968
Manus as % of Pes	32.0661157	32.0661157	40.08264

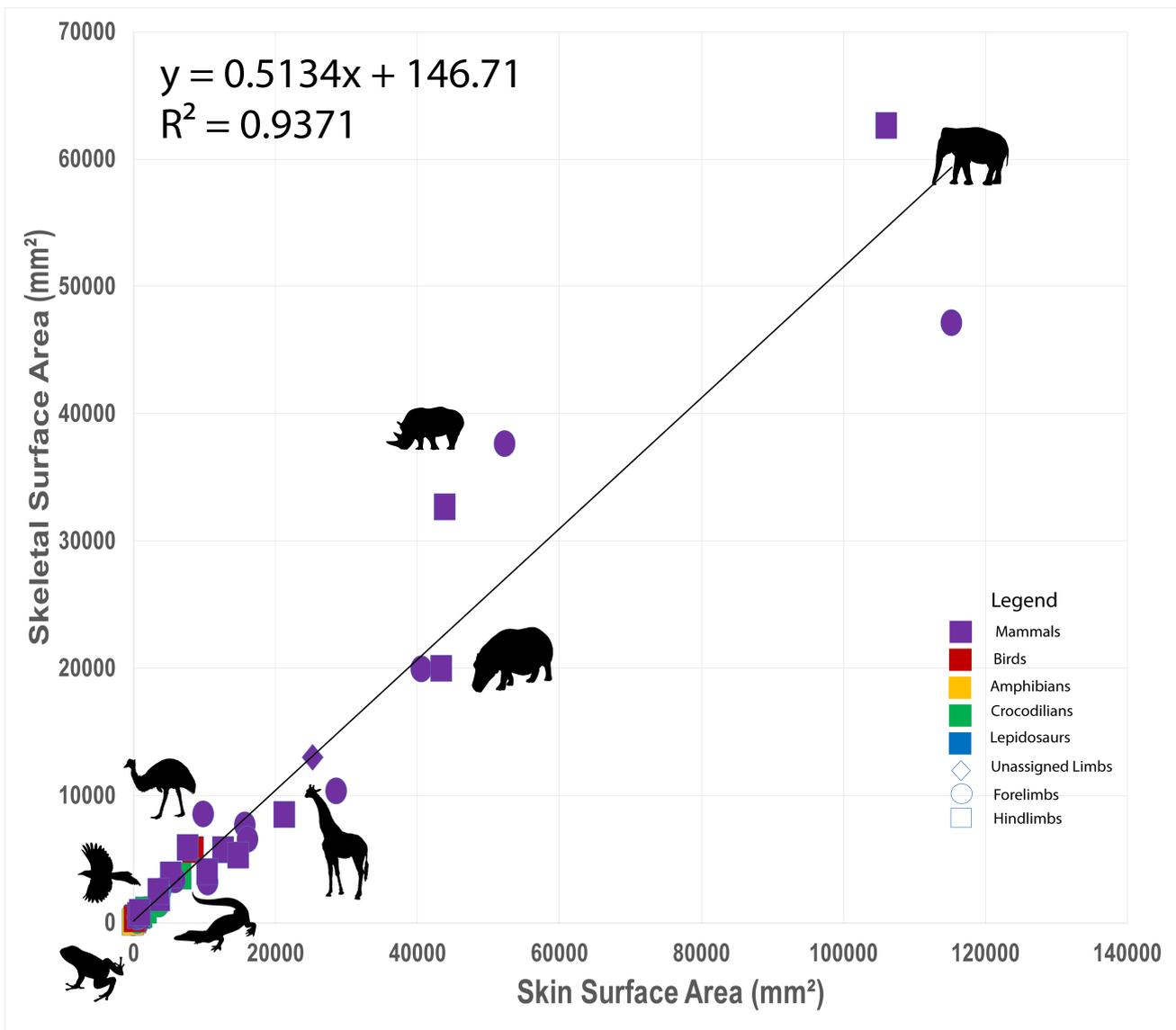
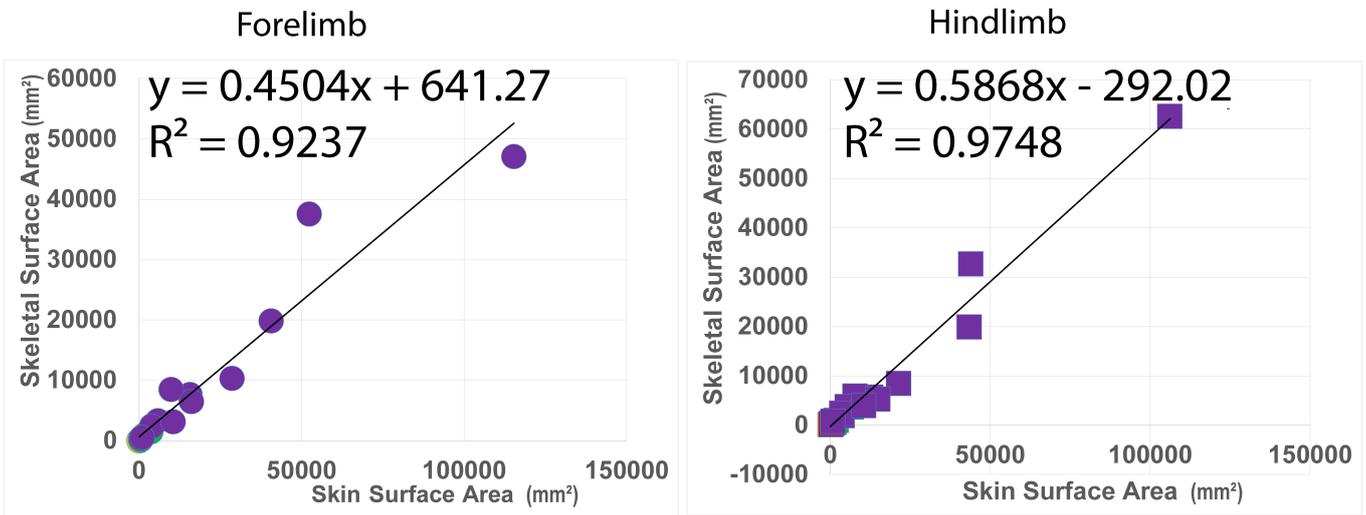
Plateosaurus

Body Mass (N)	7384
CoM (%GAD)	20.43
Manus Load	1508.5512
Pes Load	5875.4488
Combined	
Manus Pressure	46563.09649
Pes Pressure	58152.6085
Separate	
Manus Pressure	38880.18557

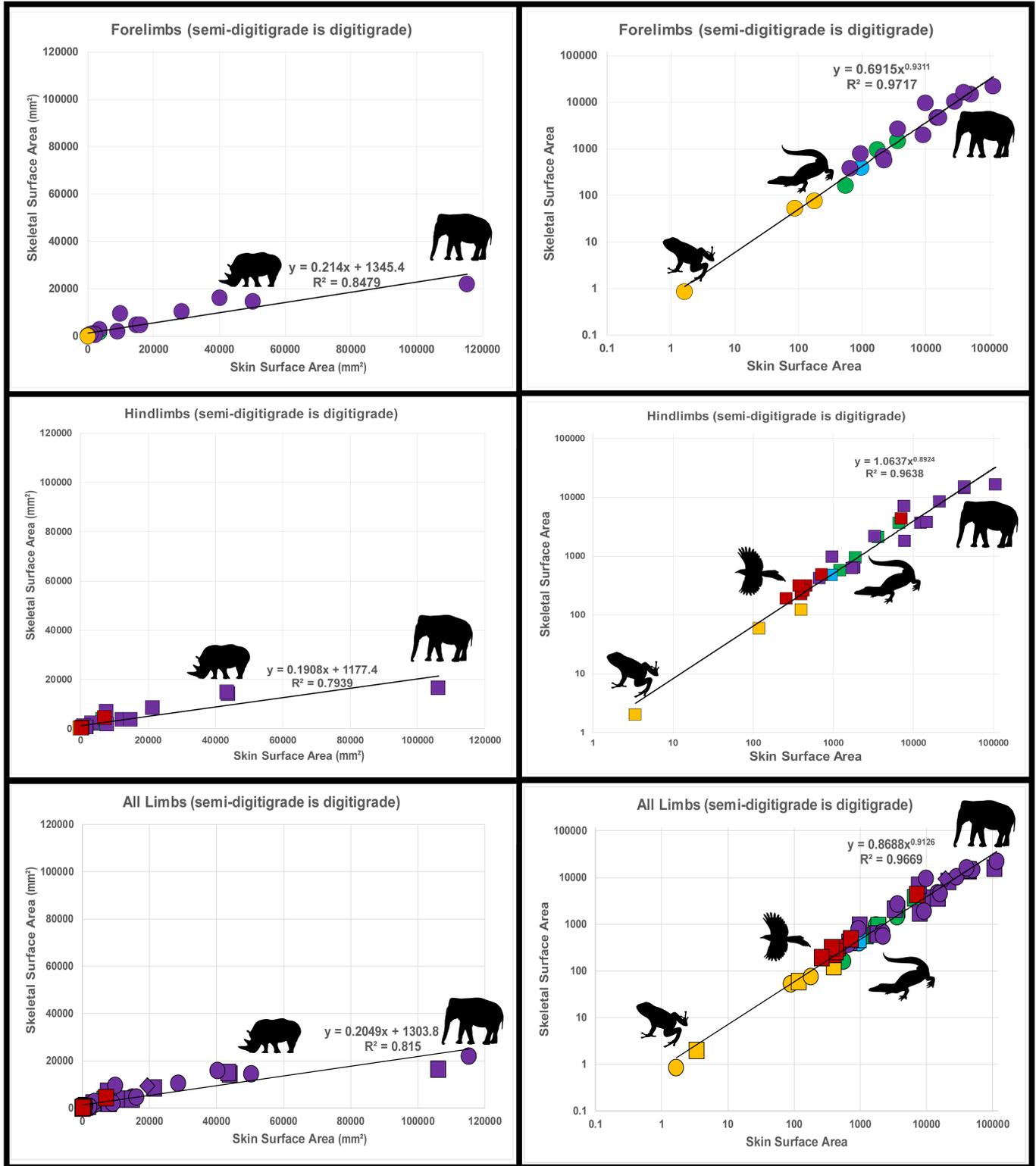
	Pes Pressure		60696.78512	
Skeleton	Area	Load	Pressure	
Manus		0.0194	1508.5512	77760.37
Pes		0.0605	5875.4488	97114.86
Manus as % of Pes		32.0661157	25.67550584	80.07052
Combined (Skin)	Area	Load	Pressure	
Manus		0.032398	1508.5512	46563.1
Pes		0.101035	5875.4488	58152.61
Manus as % of Pes		32.0661157	25.67550584	80.07052
Separate (Skin)	Area	Load	Pressure	
Manus		0.0388	1508.5512	38880.19
Pes		0.0968	5875.4488	60696.79
Manus as % of Pes		40.08264463	25.67550584	64.05642

Supplementary References

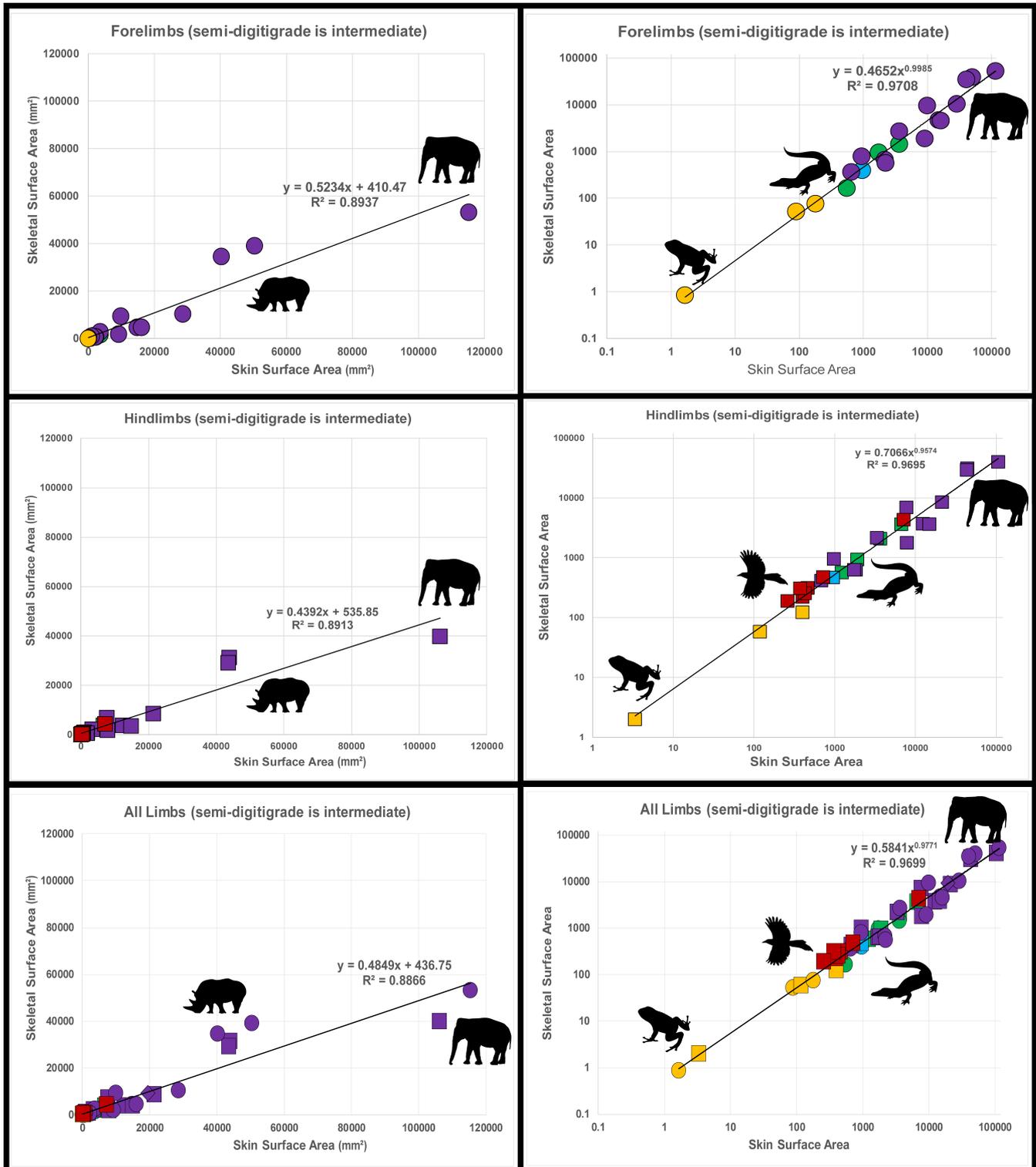
- Myers, P., Espinosa, R., Parr, C.S., Jones, T., Hammond, G.S. and Dewey, T.A., 2006. The animal diversity web. *Accessed October, 12(2006)*, p.2.
- Dunning Jr, John B. *CRC Handbook of Avian Body Masses*. CRC press, 1992.
- Hutchinson, J. R.. *Crocbase*. DOI 10.17605/OSF.IO/X38NH
- Jones, Kate E., Jon Bielby, Marcel Cardillo, Susanne A. Fritz, Justin O'Dell, C. David L. Orme, Kamran Safi, Wes Sechrest, Elizabeth H. Boakes, and Chris Carbone. "PanTHERIA: A Species-Level Database of Life History, Ecology, and Geography of Extant and Recently Extinct Mammals." *Ecology* 90, no. 9 (2009): 2648–2648.
- Pires Jr, Osmindo R., Antonio Sebben, Elisabeth F. Schwartz, Rodrigo AV Morales, Carlos Bloch Jr, and Carlos A. Schwartz. "Further Report of the Occurrence of Tetrodotoxin and New Analogues in the Anuran Family Brachycephalidae." *Toxicon* 45, no. 1 (2005): 73–79.
- Wilson, Edward O. "The Encyclopedia of Life." *Trends in Ecology & Evolution* 18, no. 2 (2003): 77–80.



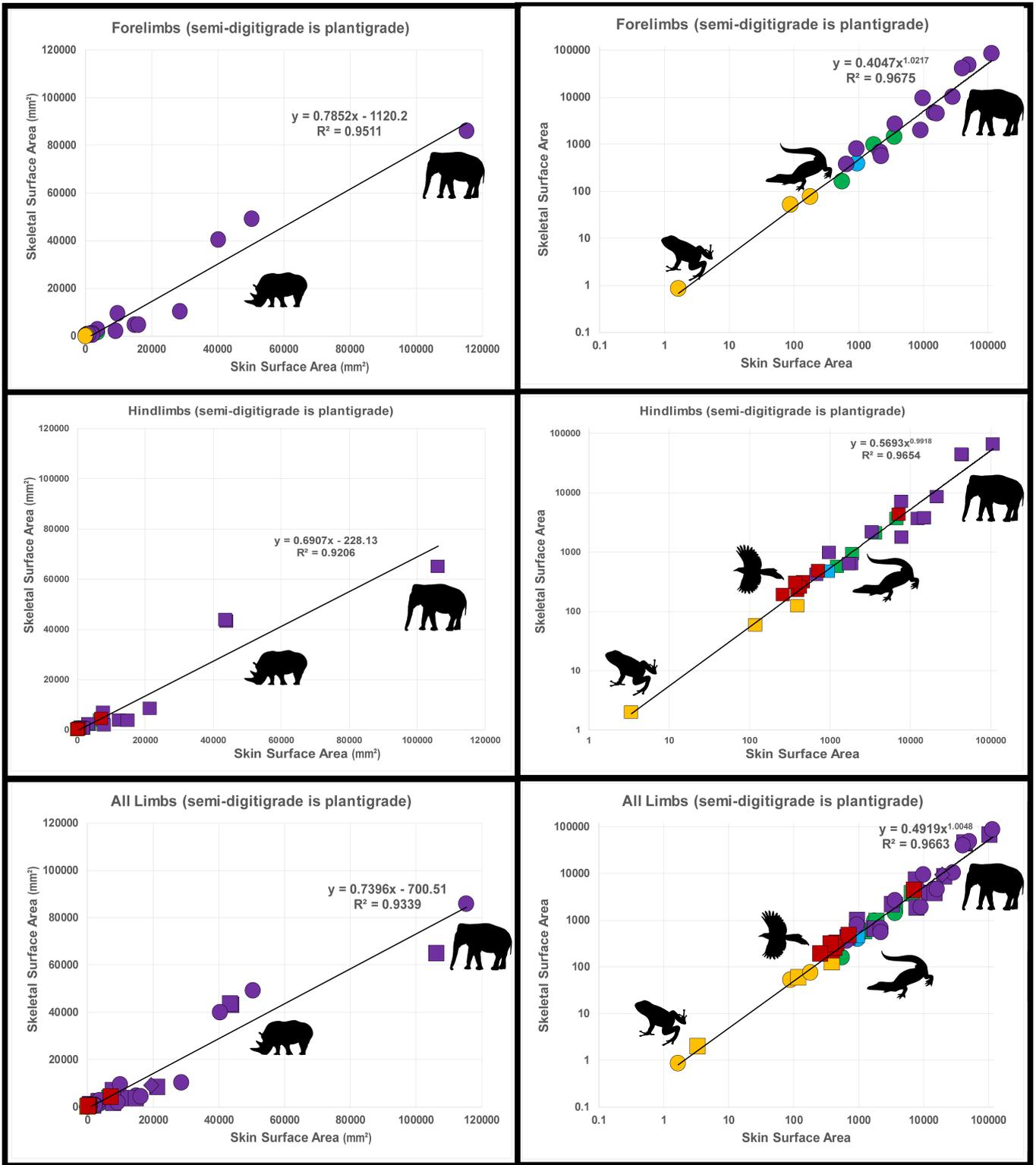
Linear plots for projected skin surface area against projected skeletal surface area in pose 1, for forelimbs, hindlimbs, and all limbs. Silhouettes from Phylopic.



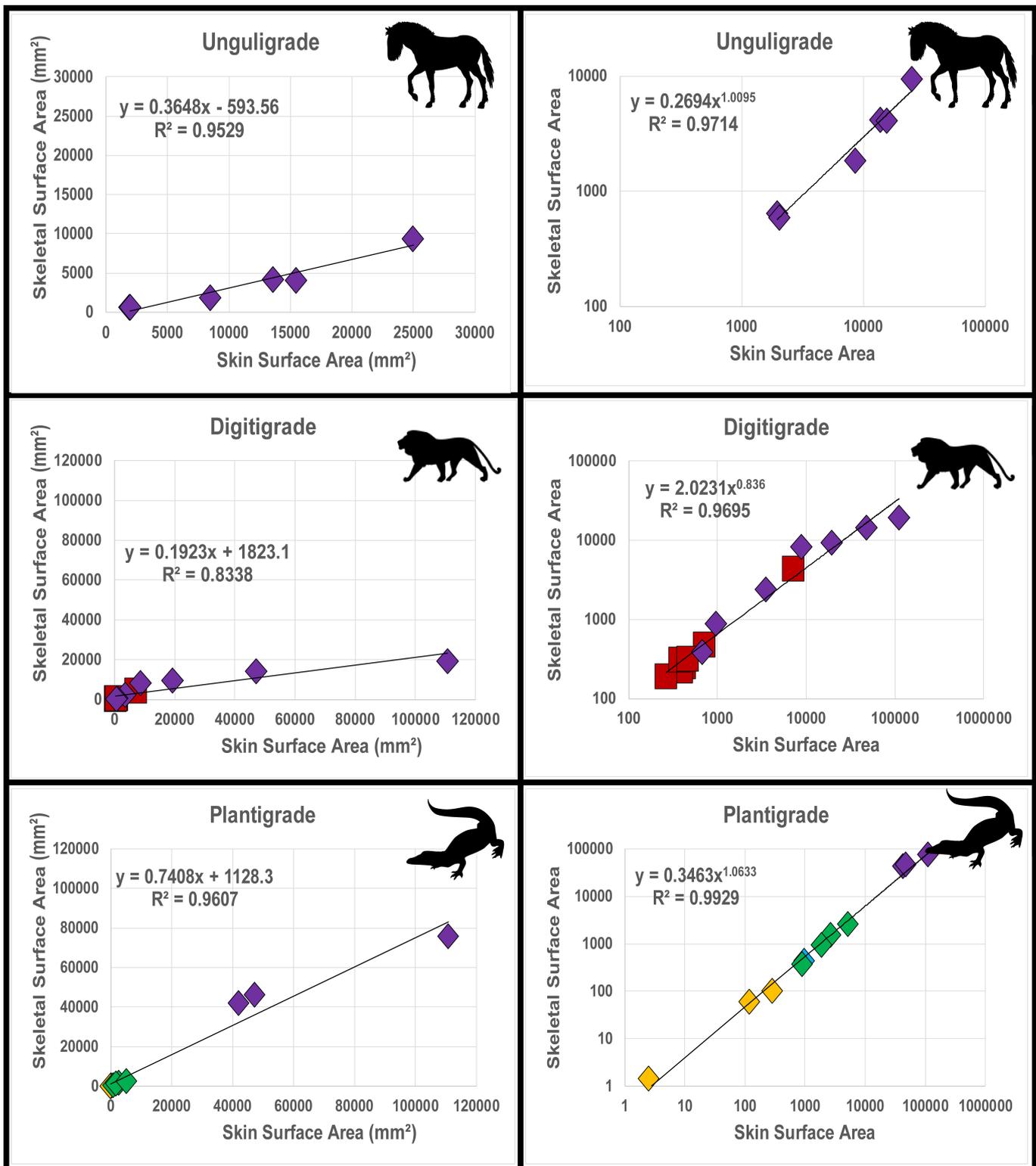
Linear and Log10-transformed plots for projected skin surface area against projected skeletal surface area, in pose 2a, for forelimbs, hindlimbs, and all limbs. Silhouettes from Phylopic.



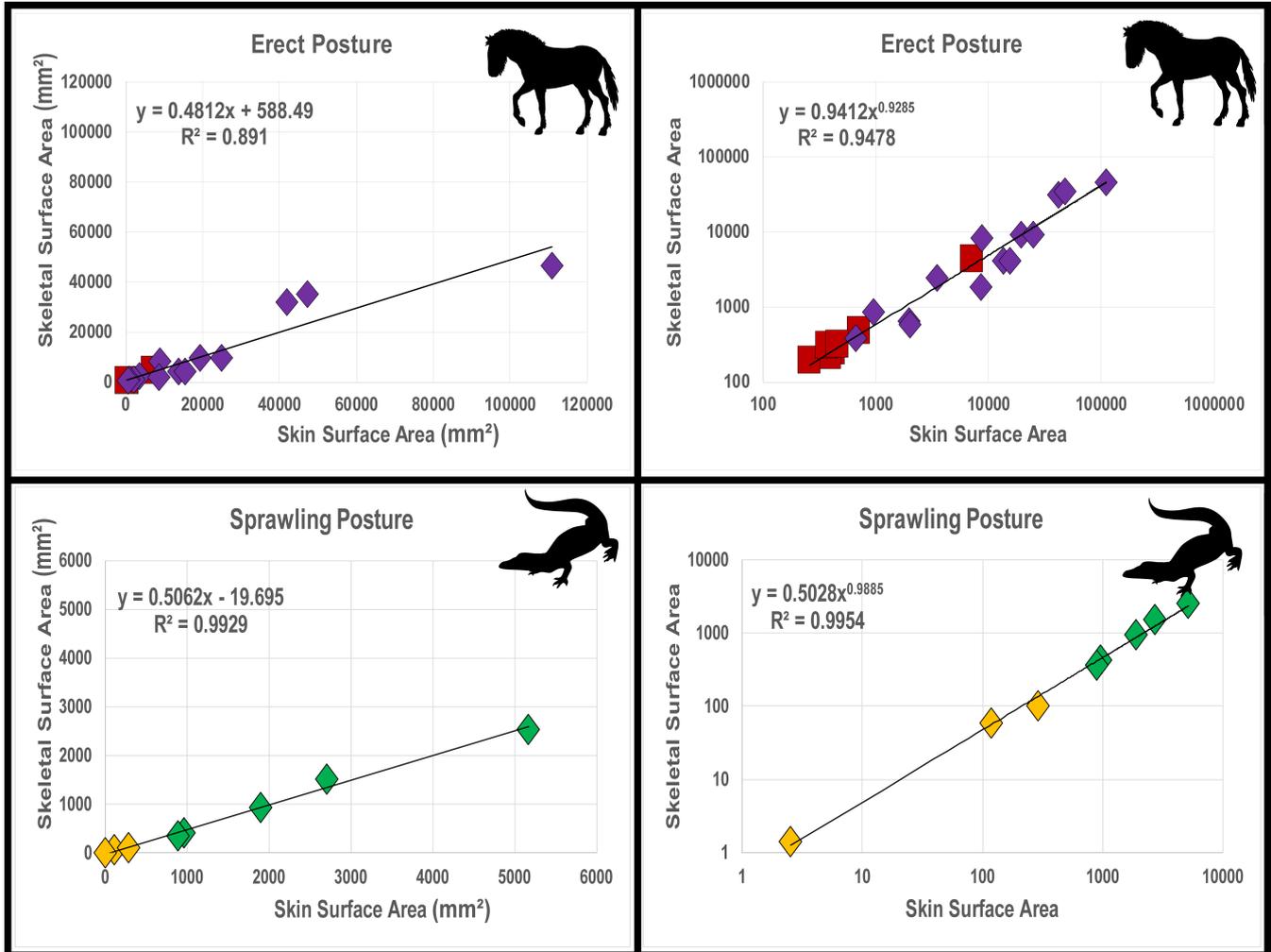
Linear and Log10-transformed plots for projected skin surface area against projected skeletal surface area, in pose 2b, for forelimbs, hindlimbs, and all limbs. Silhouettes from Phylopic.



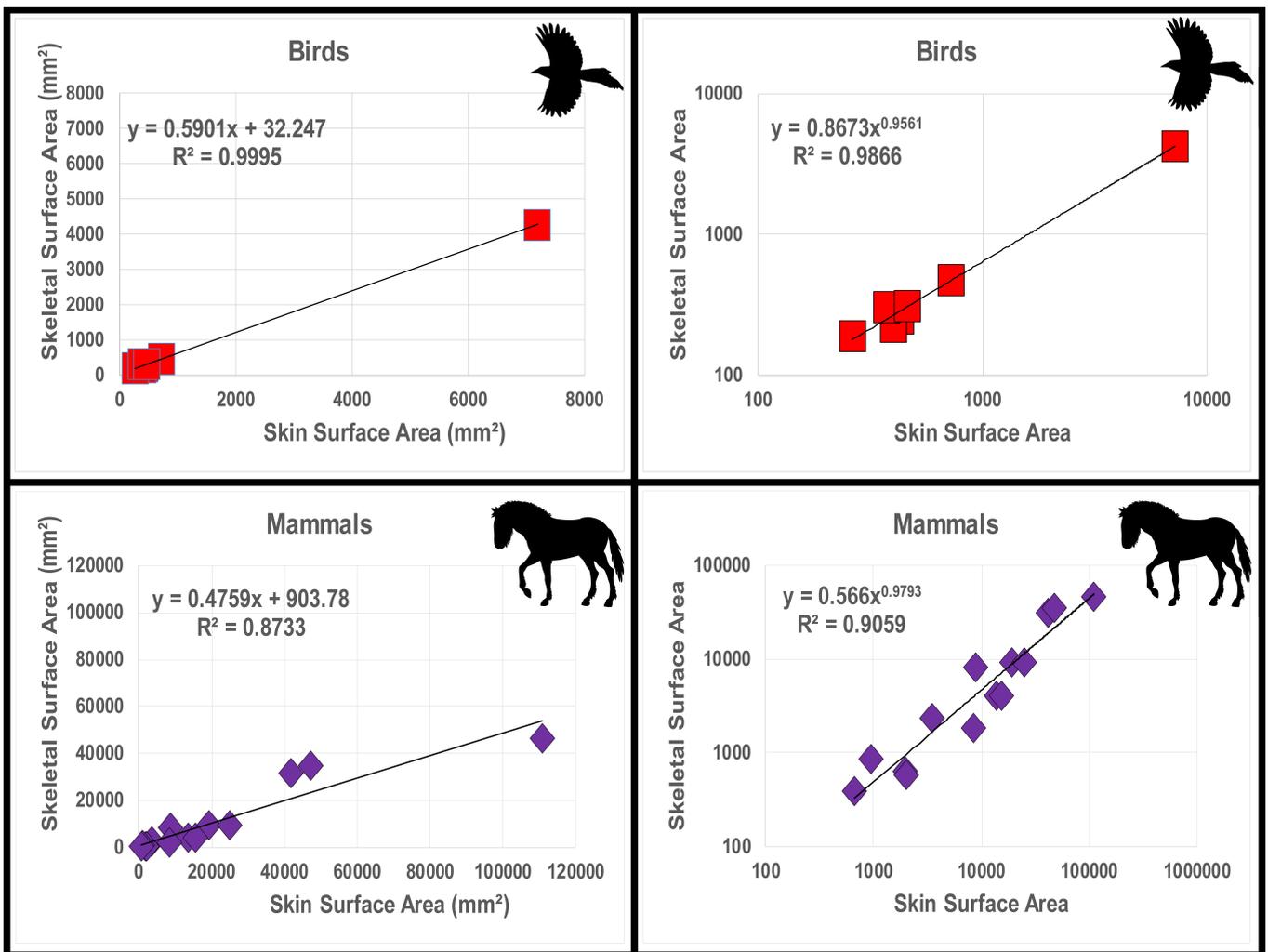
Linear and Log10-transformed plots for projected skin surface area against projected skeletal surface area, in pose 2c, for forelimbs, hindlimbs, and all limbs. Silhouettes from Phylopic.



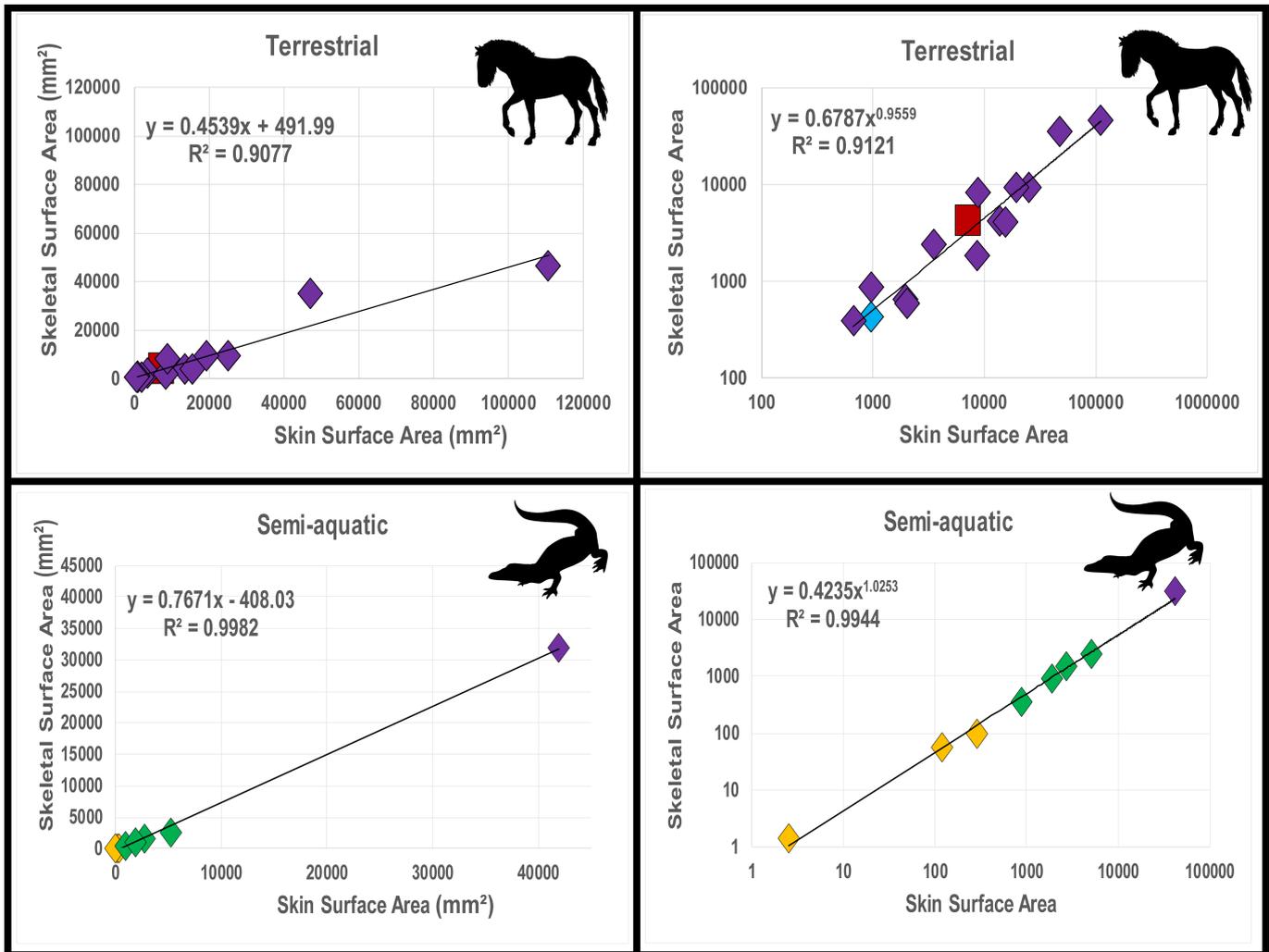
Linear and Log₁₀-transformed plots for locomotor mode sub-analysis of projected skin surface area against projected skeletal surface area, in pose 2. Silhouettes from Phylopic.



Linear and Log10-transformed plots for posture sub-analysis of projected skin surface area against projected skeletal surface area, in pose 2. Silhouettes from Phylopic.



Linear and Log10-transformed plots for clade-based sub-analysis of projected skin surface area against projected skeletal surface area, in pose 2. Silhouettes from Phylopic.



Linear and Log10-transformed plots for ecological sub-analysis of projected skin surface area against projected skeletal surface area, in pose 2. Silhouettes from Phylopic.

Supplemental Data

Presented here are the alpha shape outlines generated via matlab. Outlines are presented for skin surface area and skeletal area in pose 1 (approximate life position), and skeletal outlines for pose 2 (bones laid flat on the horizontal plane).

In some cases (e.g. many crocodylians), pose 1 and pose 2 were identical, as the foot bones are horizontal in both poses.

Large digitigrade/sub-unguligrade animals (Elephant, Hippo, and Rhino) which in life walk on a large fatty pad beneath the foot, had skeletal areas calculated in Pose 2 from just the digits (Pose 2a, as digitigrade), the digits and metatarsals/metacarpals (Pose2b, intermediate) and from the entire Pes/Manus (Pose 2c, as plantigrade).

All units are in mm, except the Tuatara where units are in 0.1mm.

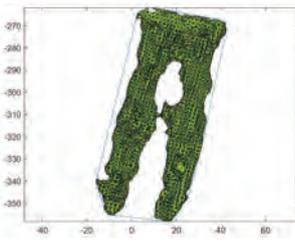
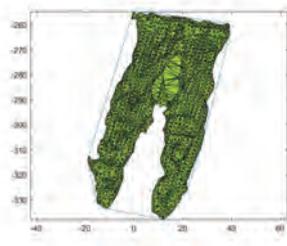
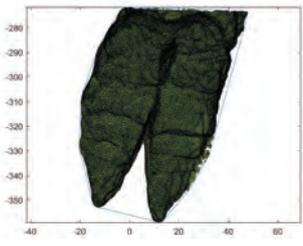
Alpaca

Soft-tissue area

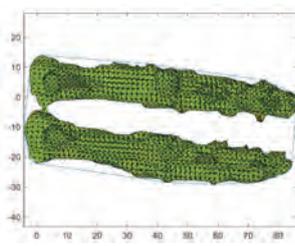
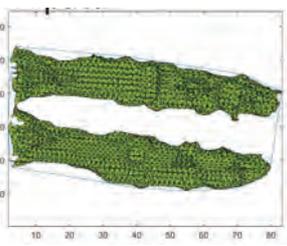
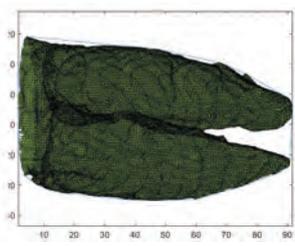
Skeletal area
(pose 1)

Skeletal area
(pose 2)

Manus

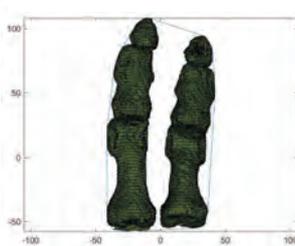
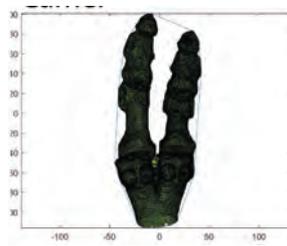
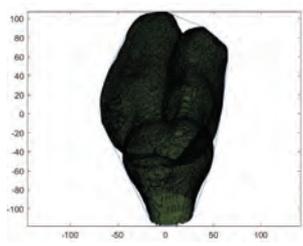


Pes

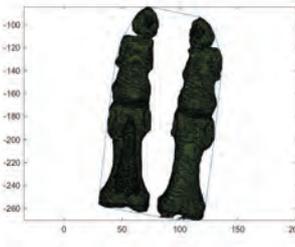
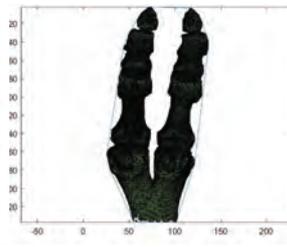
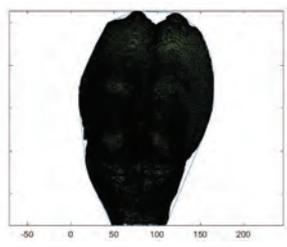


Camel

Unassigned

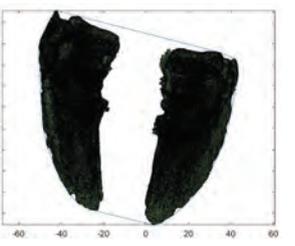
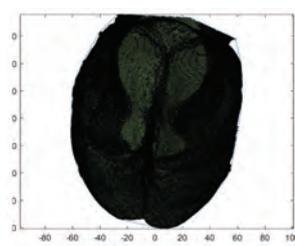


Unassigned

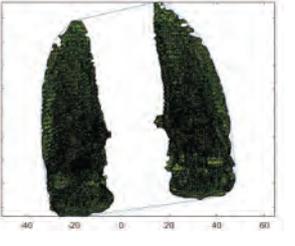


Cow

Manus



Pes



Soft-tissue area

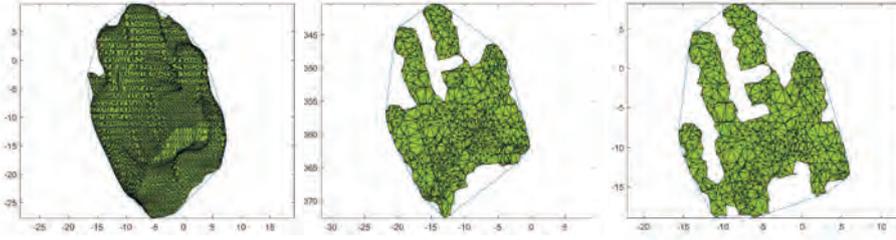
Skeletal area
(pose 2)

Cat

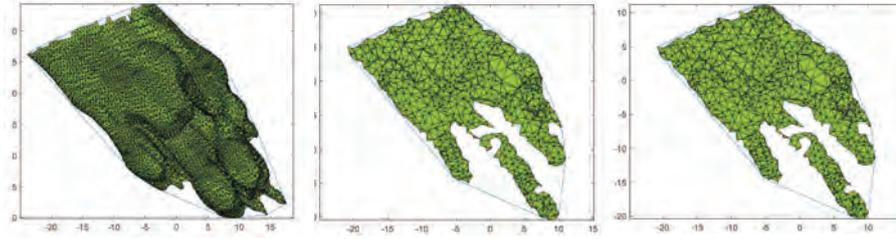
Left Manus



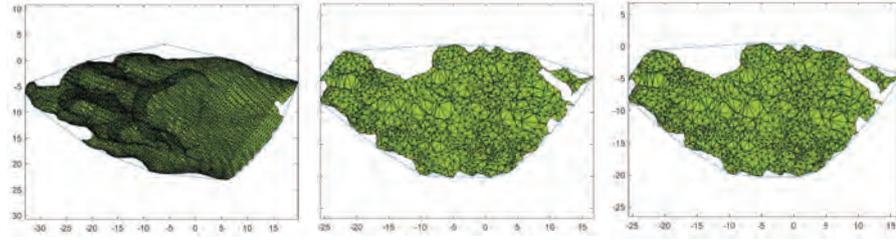
Left Pes



Right Manus



Right Pes



Deer

Manus



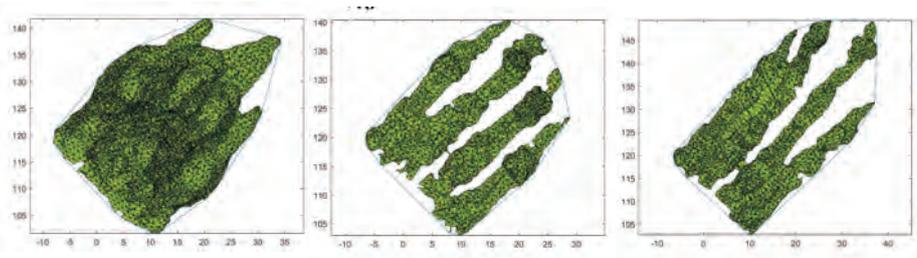
Pes



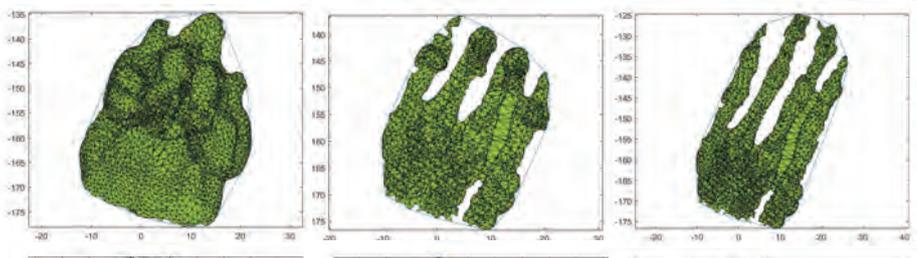
Soft-tissue area

Fox

Left Manus



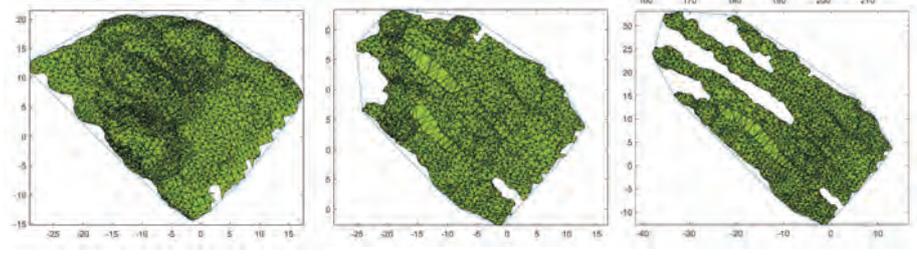
Left Pes



Right Manus

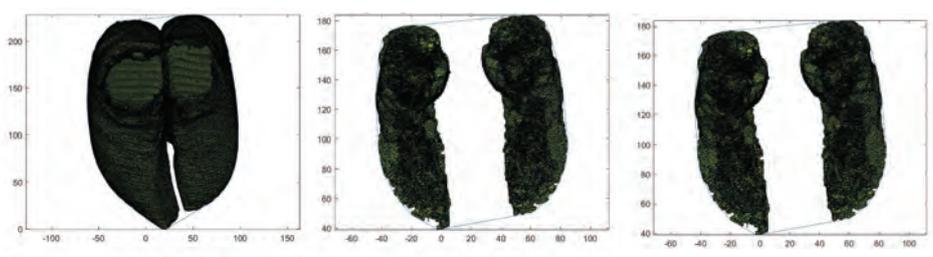


Right Pes

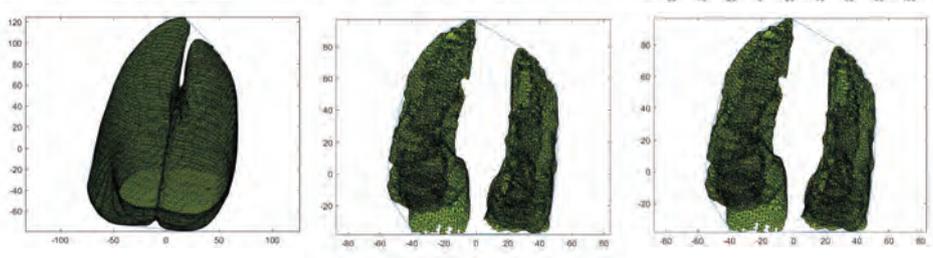


Giraffe

Manus



Pes

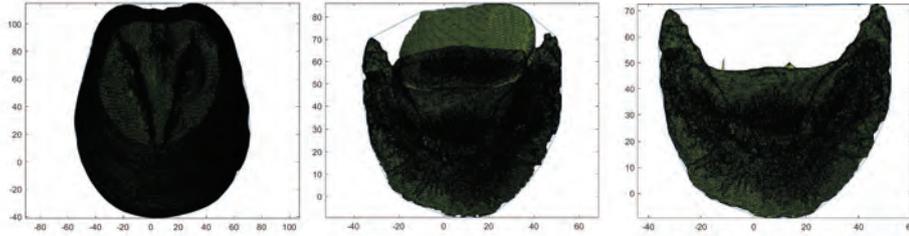


Soft-tissue area

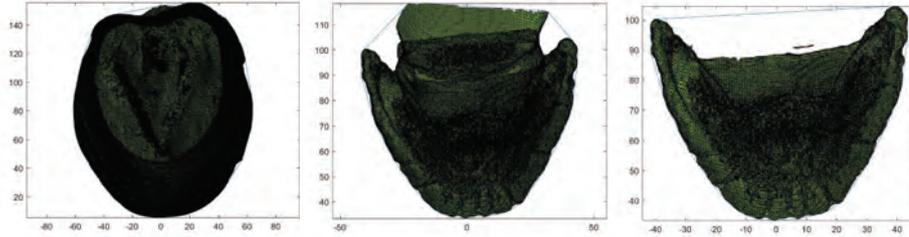
Skeletal area
(pose 2)

Horse

Manus

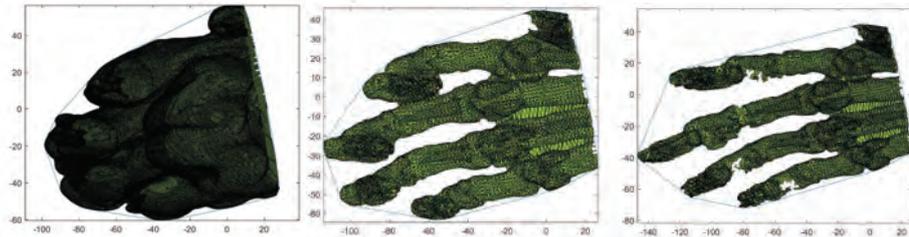


Pes

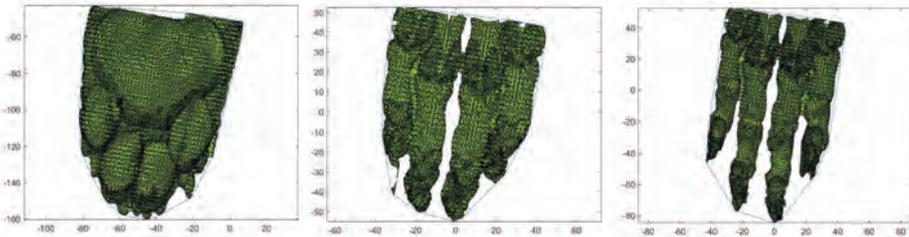


Lion

Manus

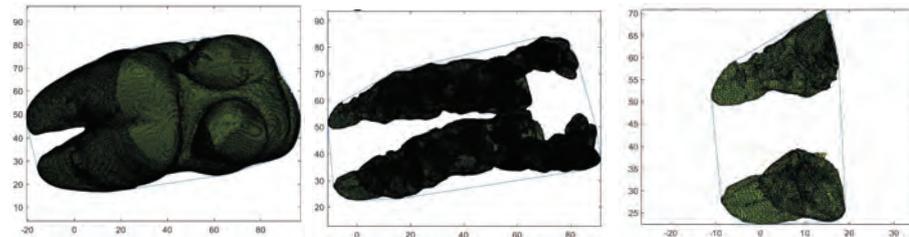


Pes

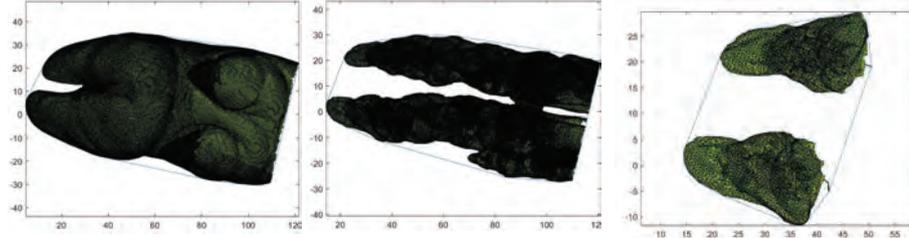


Pig

Manus



Pes



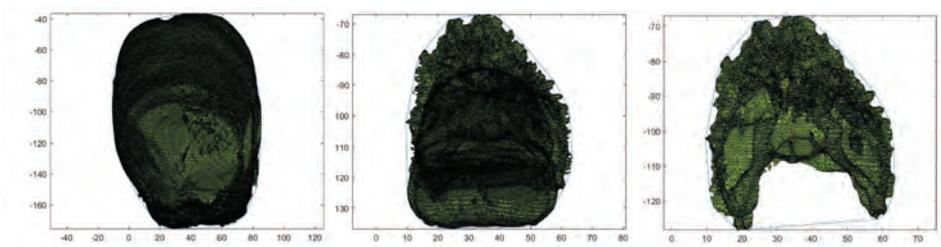
Soft-tissue area

Skeletal area
(pose 1)

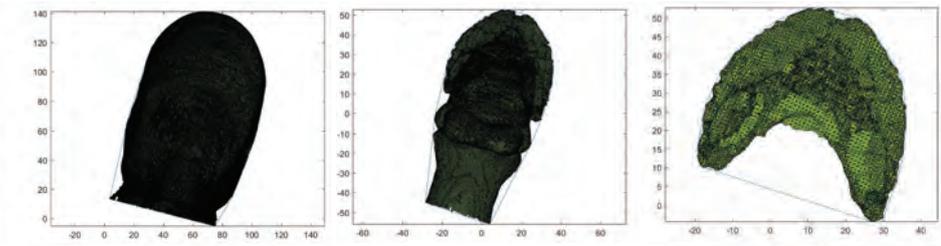
Skeletal area
(pose 2)

Zebra

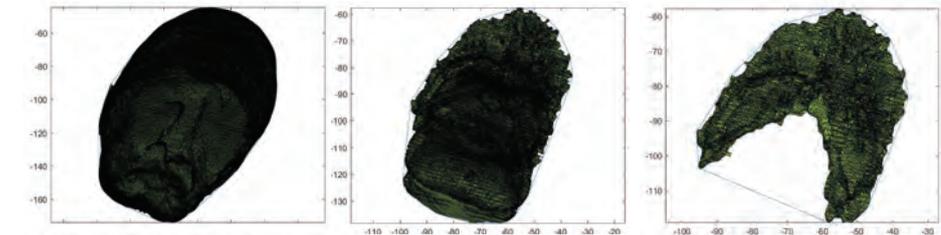
Left Manus



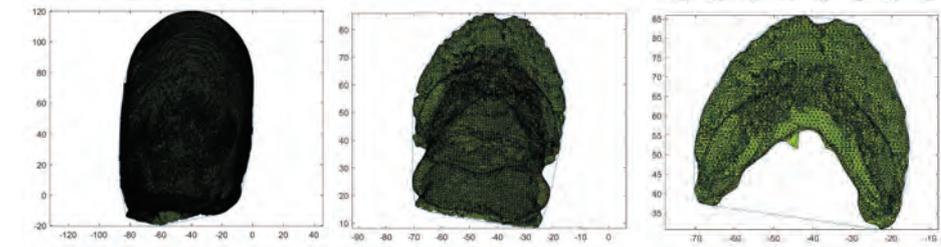
Left Pes



Right Manus



Right Pes



Soft-tissue area

Skeletal area
(pose 1)

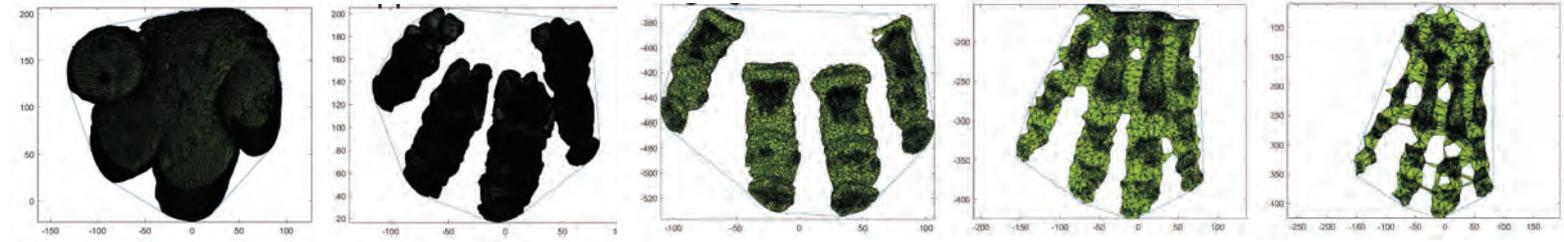
Skeletal area
(pose 2a)

Skeletal area
(pose 2b)

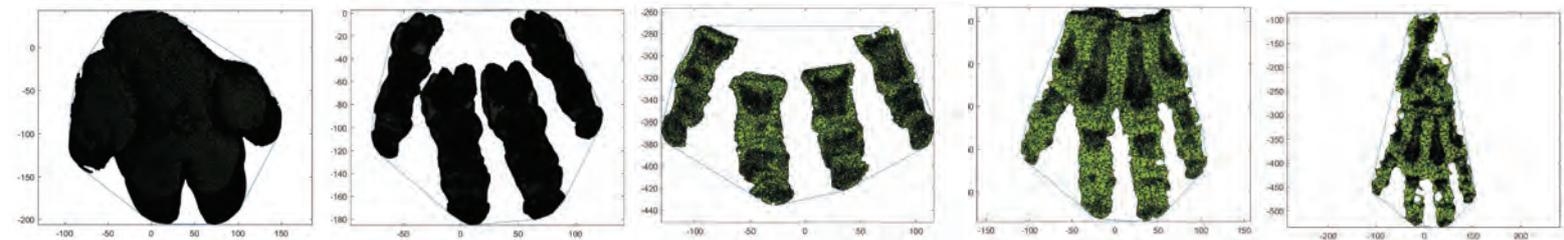
Skeletal area
(pose 2c)

Hippo

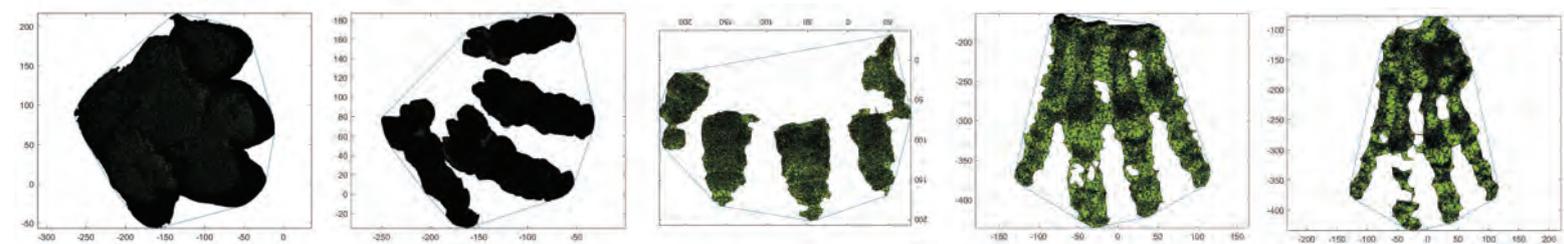
Left Manus



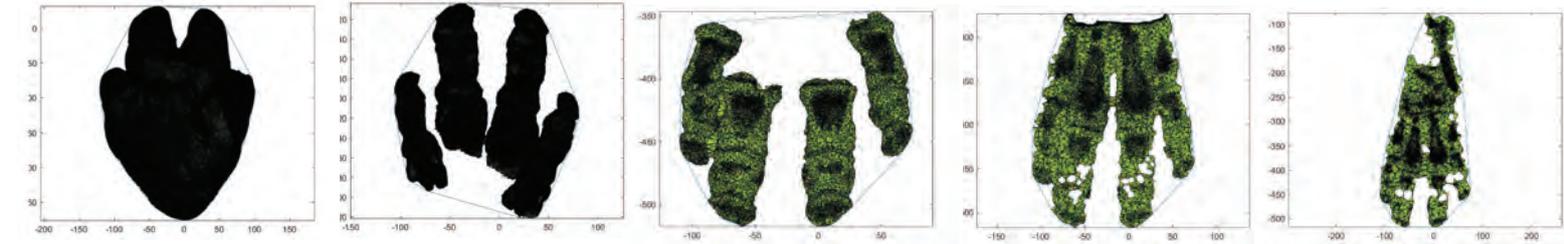
Left Pes



Right Manus



Right Pes



Soft-tissue area

Skeletal area
(pose 1)

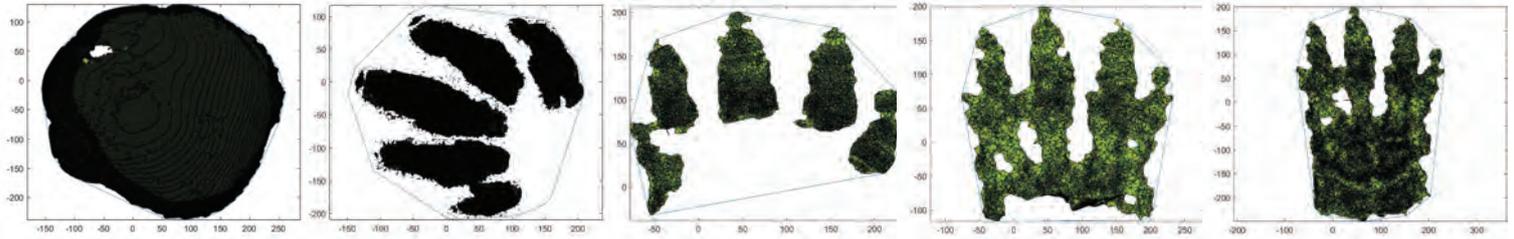
Skeletal area
(pose 2a)

Skeletal area
(pose 2b)

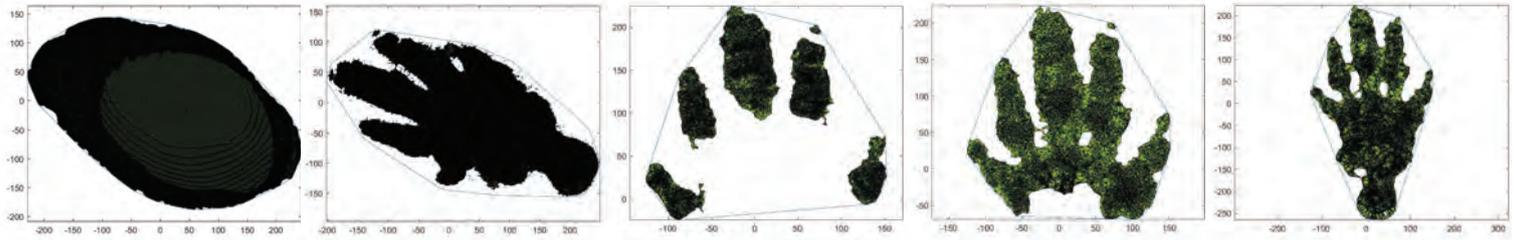
Skeletal area
(pose 2c)

Elephant

Manus

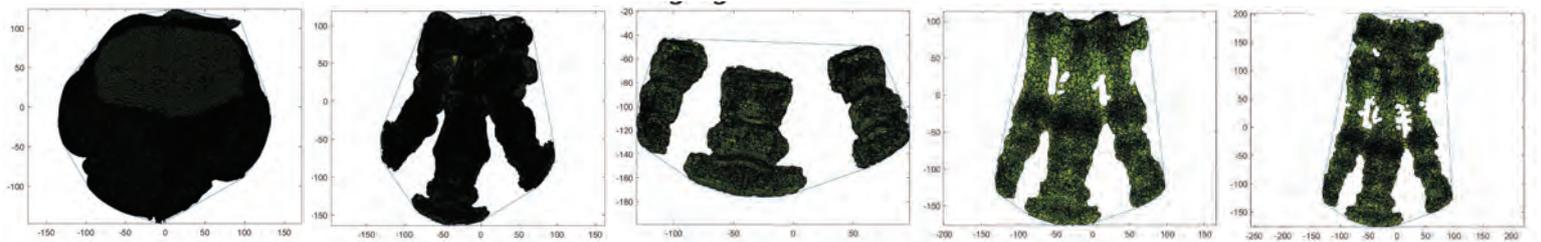


Pes

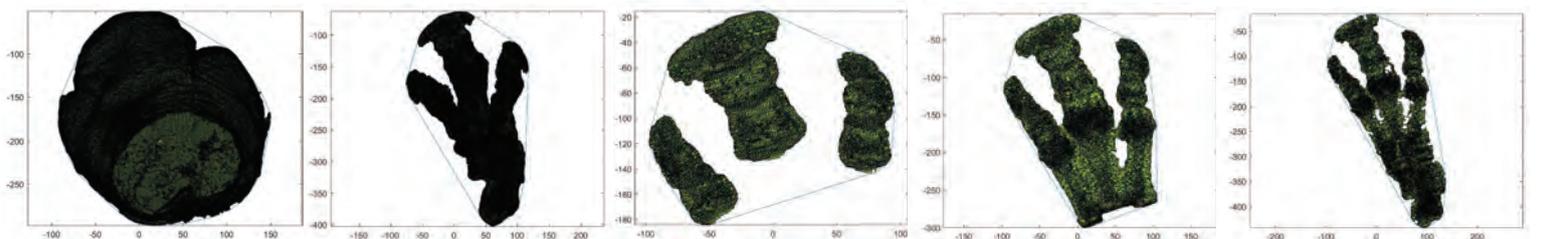


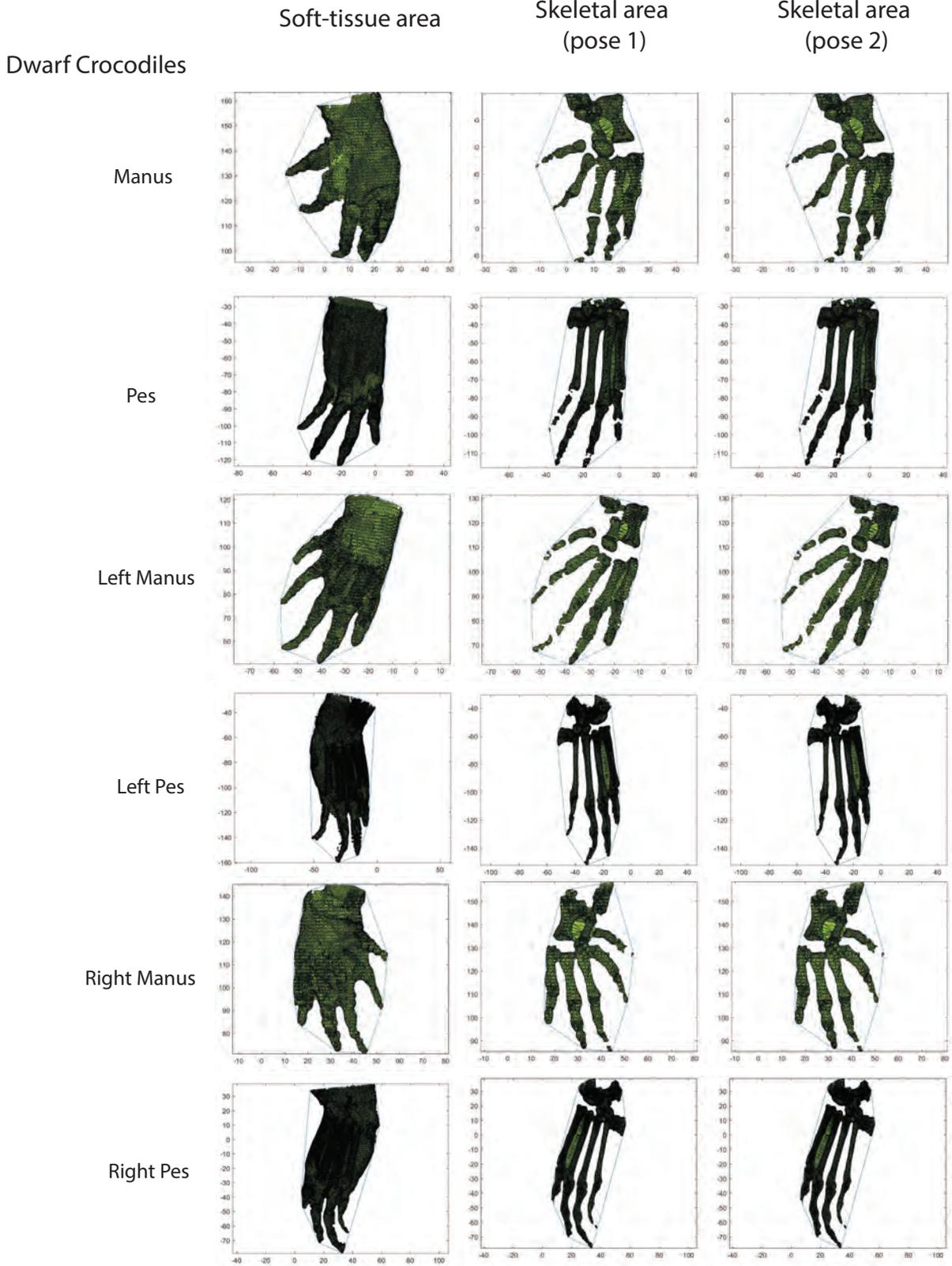
Rhino

Manus



Pes





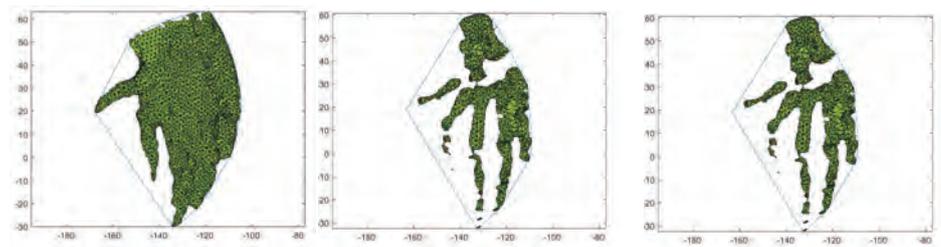
Soft-tissue area

Skeletal area
(pose 1)

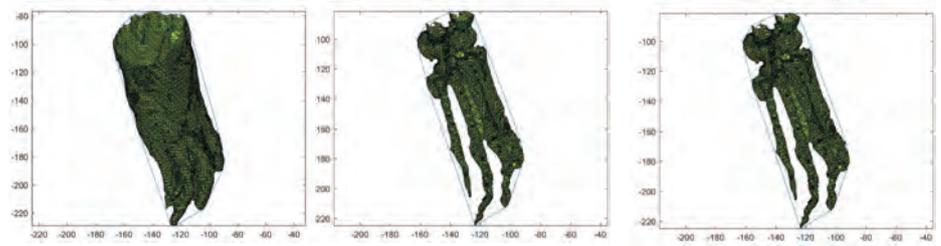
Skeletal area
(pose 2)

Morlet's Crocodiles

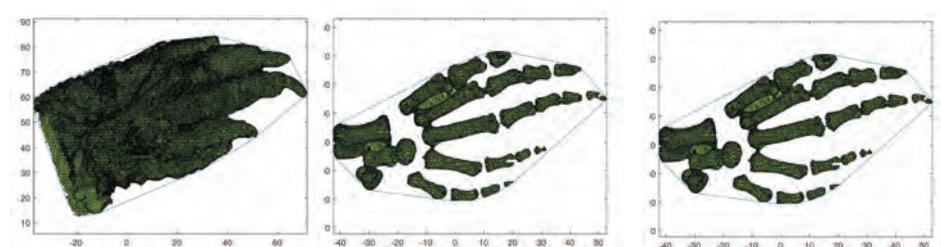
Left Manus



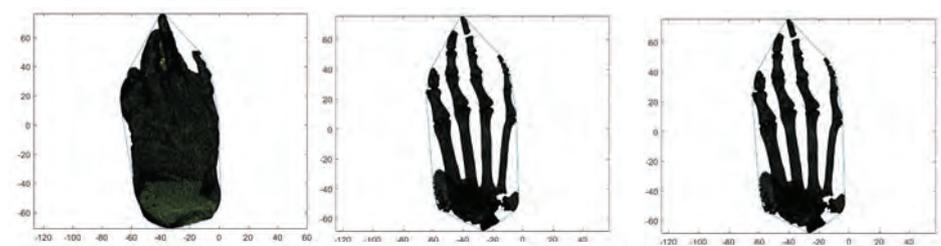
Left Pes



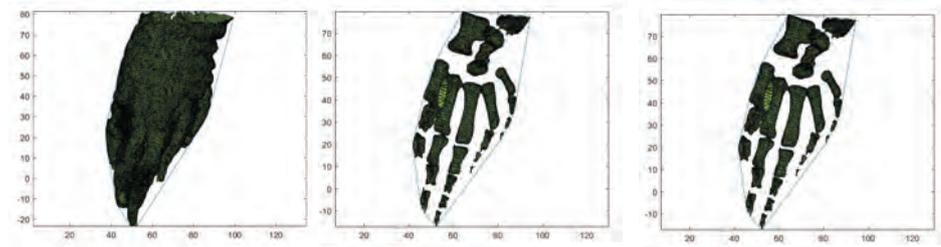
Right Manus



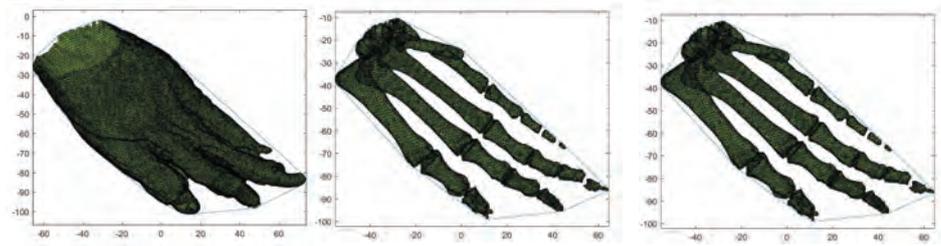
Right Pes



Right Manus



Right Pes



Nile Crocodile

Soft-tissue area

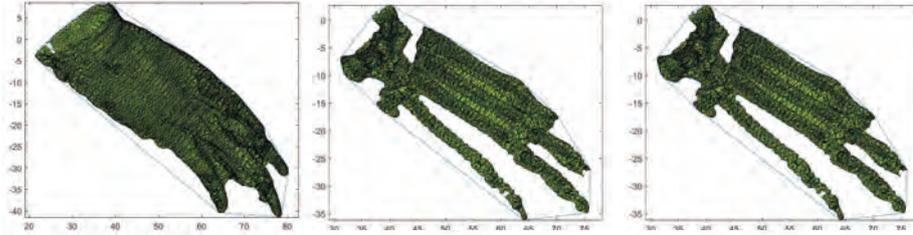
Skeletal area
(pose 1)

Skeletal area
(pose 2)

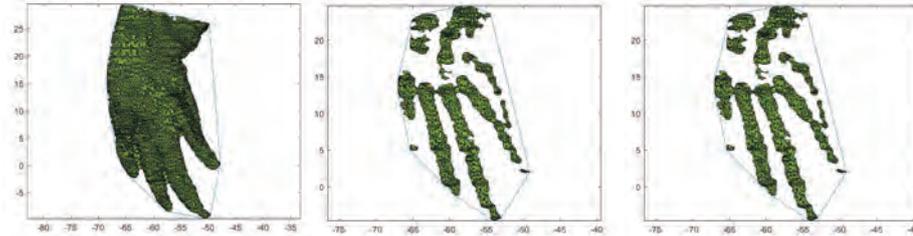
Left Manus



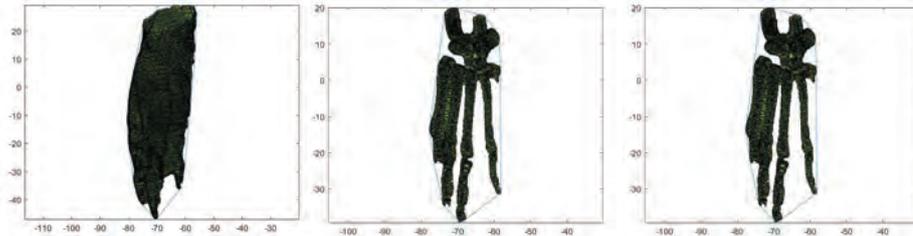
Left Pes



Right Manus

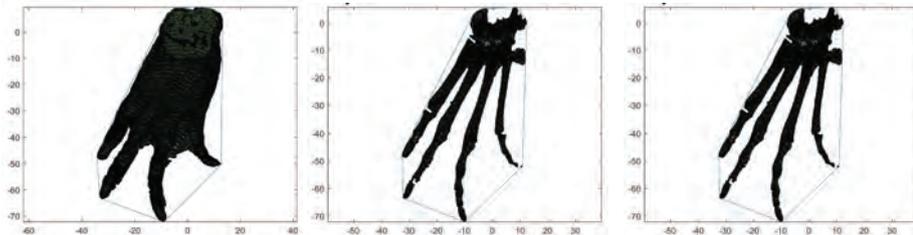


Right Pes



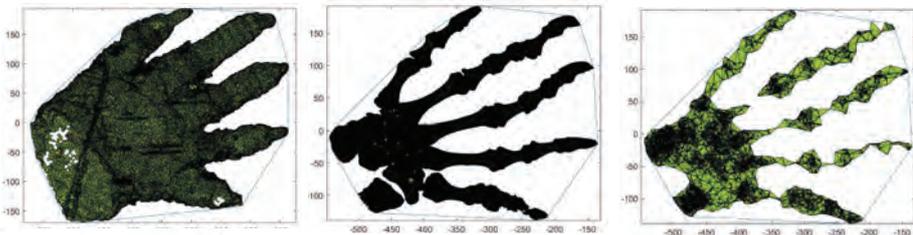
Spectacled Caiman

Left Pes

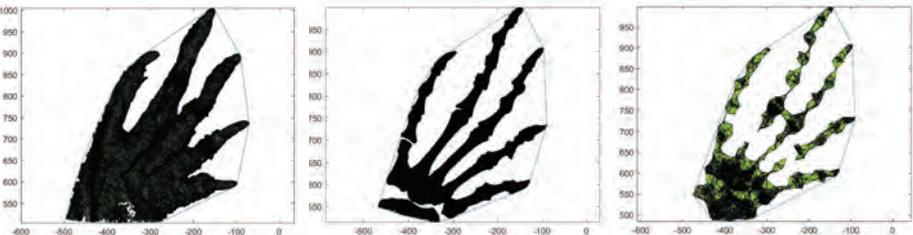


Tuatara

Manus



Pes



Amphibians

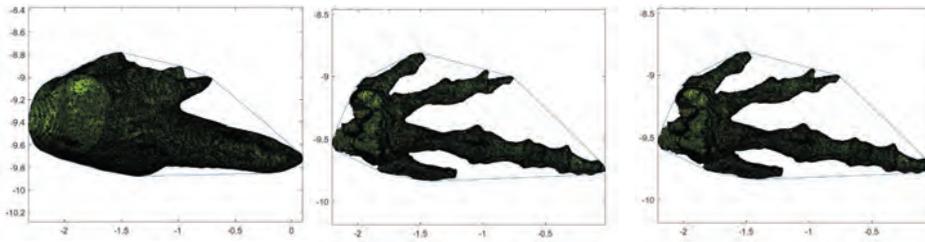
Frog

Soft-tissue area

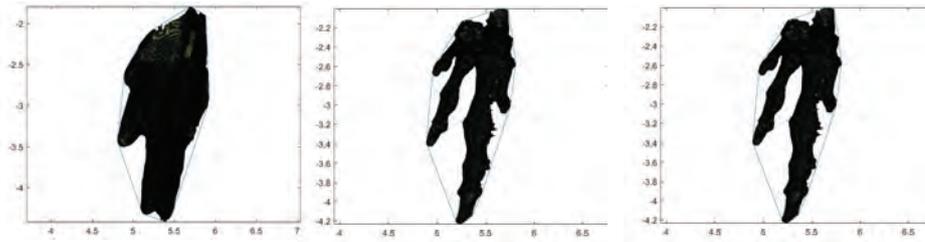
Skeletal area
(pose 1)

Skeletal area
(pose 2)

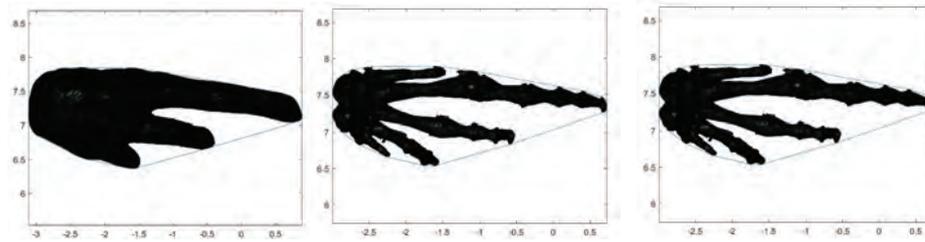
Left Manus



Right Manus

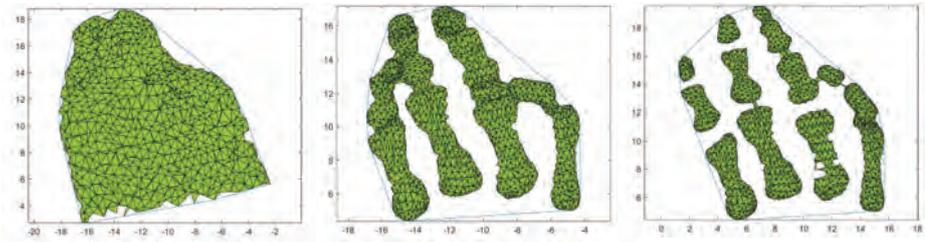


Right Pes

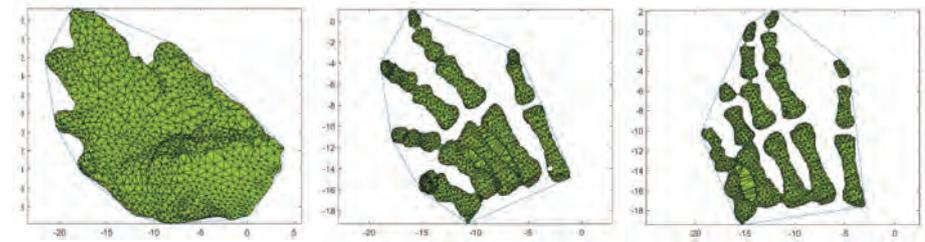


Hellbender

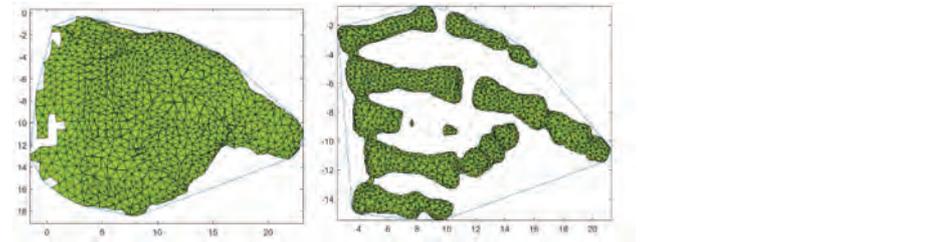
Left Manus



Left Pes



Right Pes



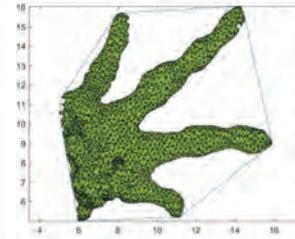
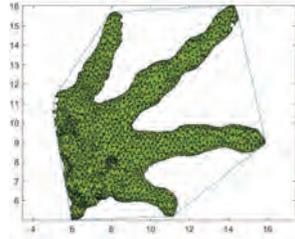
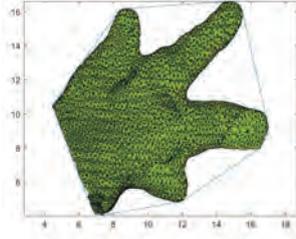
Fire Salamander

Soft-tissue area

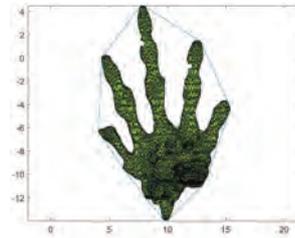
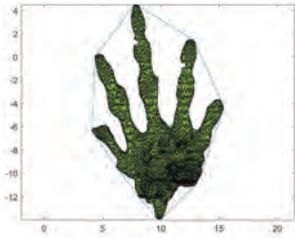
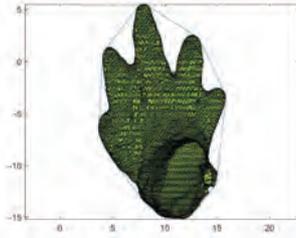
Skeletal area
(pose 1)

Skeletal area
(pose 2)

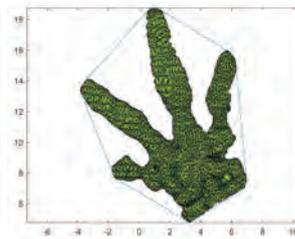
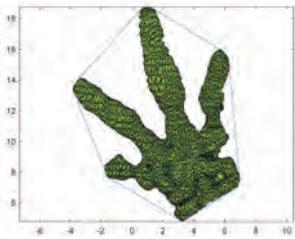
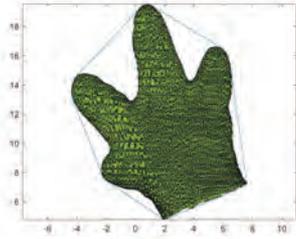
Left Manus



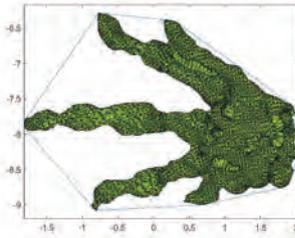
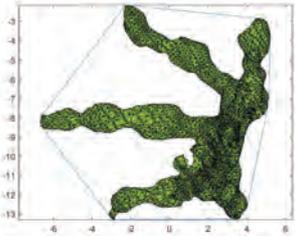
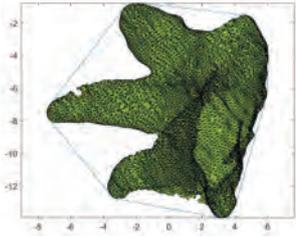
Left Pes



Right Manus



Right Pes



Birds

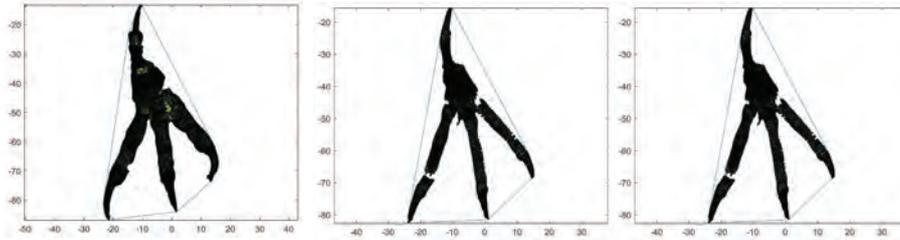
Barn Owl

Soft-tissue area

Skeletal area
(pose 1)

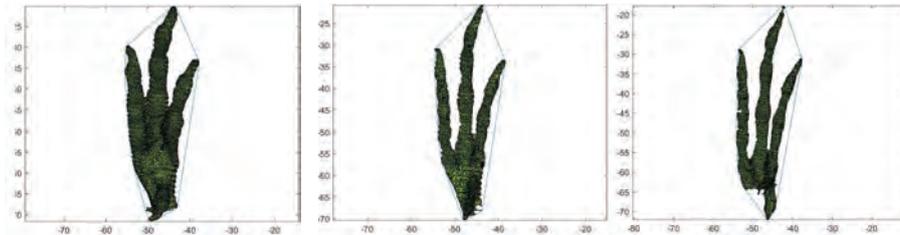
Skeletal area
(pose 2)

Pes

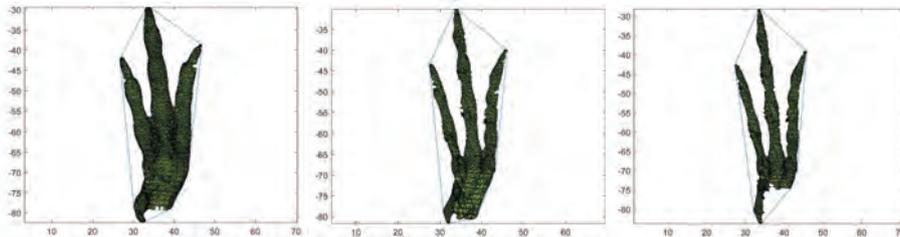


Chukar

Left Pes

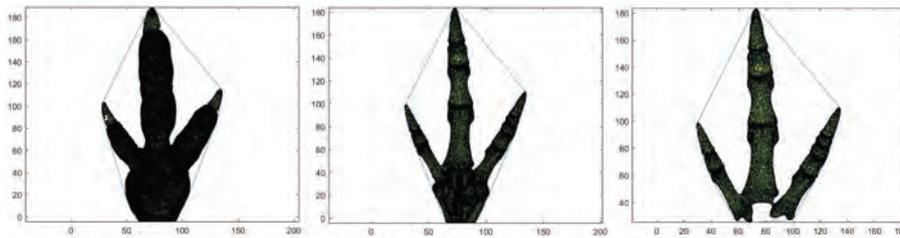


Right Pes



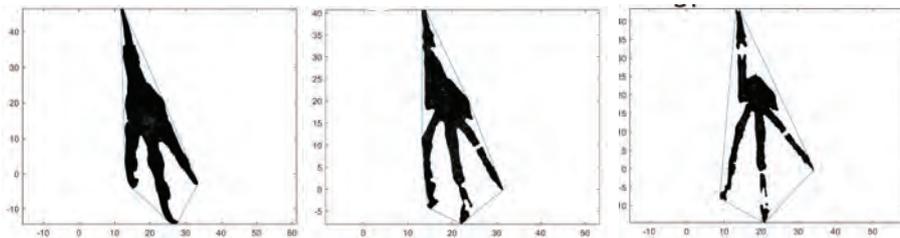
Emu

Pes



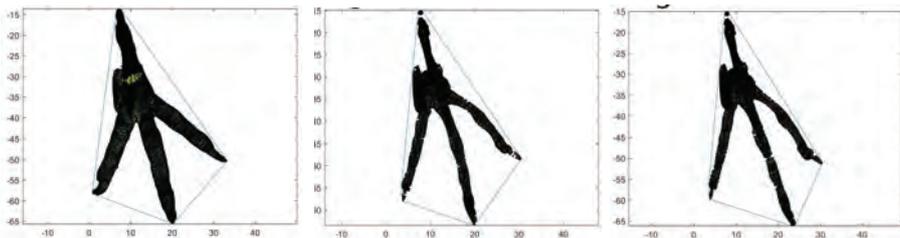
Magpie

Pes



Pigeon

Pes



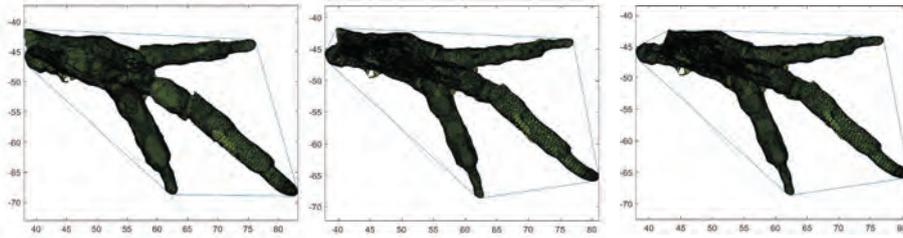
Quail

Soft-tissue area

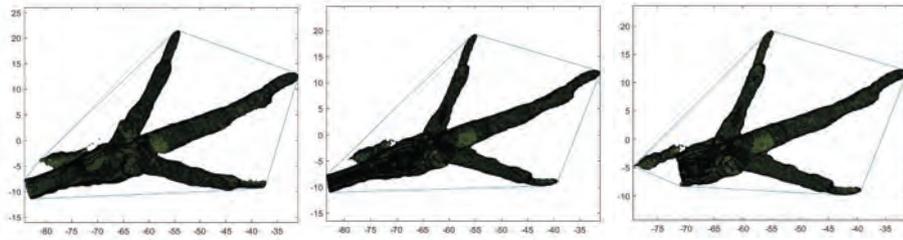
Skeletal area
(pose 1)

Skeletal area
(pose 2)

Left Pes



Right Pes



Sparrowhawk

Pes

