1 Computational approaches for Drug-induced

2 liver injury (DILI) prediction: state of the art and

3 challenges

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12 ABSTRACT

- 13 Drug-induced liver injury (DILI) is one of the prevailing causes of fulminant hepatic failure. It is
- estimated that three idiosyncratic drug reactions out of four result in liver transplantation or death.
- Additionally, DILI is the most common reason for withdrawal of an approved drug from the market.
- 16 Therefore, the development of methods for the early identification of hepatotoxic drug candidates is of
- crucial importance. This review focuses on the current state of cheminformatics strategies being applied
- for the early *in silico* prediction of DILI. Herein, we discuss key issues associated with DILI modelling
- in terms of the data size, imbalance and quality, complexity of mechanisms, and the different levels of
- 20 hepatotoxicity to model going from general hepatotoxicity to the molecular initiating events of DILI.

INTRODUCTION

- 22 Drug-induced liver injury (DILI) refers to hepatotoxicity resulting from adverse reactions caused by
- drugs or their reactive metabolites and toxic chemical entities. DILI is a major concern as it is one of
- 24 the leading causes of acute liver failure in the world, accounting for more than 50% of cases in the US¹.
- Additionally, a recent study showed that DILI is responsible for more than 20% of the withdrawals of
- approved drugs from the market due to toxicity ²⁻⁴. This is an on-going problem, there have been at least
- eight withdrawals of drugs due to DILI from 1997 to 2016 alone: tolcapone, troyafloxacin,
- bromfenac, nefazodone, ximelagatran, lumiracoxib and sitaxentan⁵. Moreover, hepatotoxicity is also a
- 29 major reason for the failure of candidates in the drug discovery process⁶. These reasons underscore the
- 30 need for the accurate prediction of the risk of DILI for bioactive compounds. DILI itself is complex, it
- 31 comprises a broad set of effects which can be further characterised in several ways, either by the type
- of hepatotoxicity (physiological effect) or by whether the effect is dose-dependent or not.

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With regard to hepatotoxicity, three types or patterns may be observed. Firstly, hepatocellular injury which is the result of biochemical perturbations of the cell culminating in severe cellular malfunction or cell death, the latter resulting in formation of scaring tissue. It comprises steatosis, necrosis and cirrhosis and is characterised by the release of hepatocellular enzymes (e.g. alanine transferase (ALT) and aspartate transaminase (AST)). Secondly, cholestatic injury is the result of an impairment of the biliary system caused either by bile stasis (i.e. the accumulation of bile in the bile ducts), portal inflammation or proliferation or injury of bile ducts. It is usually characterised by elevated levels of alkaline phosphatase (ALP) and γ -glutamyl transpeptidase (GGT). Finally, mixed hepatocellular-cholestatic injury, which occurs rarely in other forms of acute liver disease, usually shows prominent hepatocyte necrosis and inflammation as well as marked bile stasis. It is characterised by the elevation of both ALT and ALP.

DILI itself may also be categorised into two subtypes. The first type, called intrinsic DILI (itDILI), is dose-dependent and is modulated by the presence of key compound substructures and its effects are reversed after discontinuation of drug administration. These reasons make it quite predictable⁷. The second type is idiosyncratic DILI (iDILI), which is very rare as it only occurs in 1:1,000 to 1:100,000 patients exposed to the drug⁸. iDILI is associated with poor prognosis and does not show any dose-response relationship. Because it is host-dependent^{9,10}, iDILI can be the result of either immunological effects (i.e. allergic reactions) or metabolic effects which makes it more unpredictable¹¹ and a considerable challenge for drug development and safety.

These problems emphasise the importance of the early detection of hepatotoxic compounds in the drug discovery process in order to reduce attrition rates and to increase drug safety. However, a major obstacle to the development of comprehensive tools for the early detection of iDILI is primarily the lacking predictivity of the existing animal studies and secondly its complexity, ranging from the variety of its effects but also from the diversity of factors affecting susceptibility to iDILI. Additionally, drug metabolism and pharmacokinetics (DMPK) aspects, including local and intracellular concentration, are difficult to evaluate and predict. Effects of iDILI include elevations in serum transaminases, jaundice, acute liver failure or chronic liver dysfunction. Factors affecting iDILI include age, gender, ethnicity, genetic polymorphism, use of other medication or pre-existing liver disease^{12,13}. Additionally, the development and mechanisms of iDILI are poorly understood making its early detection, and therefore its prediction, a challenge^{14,15}. A detailed summary of these mechanisms lies outside the scope of this review and the reader is referred to the works of Fraser et al. 16 and of Noureddin and Kaplowitz 17 for comprehensive information on DILI mechanisms. Nonetheless, a wide range of predictive models have been established for the prediction of DILI and can be divided among quantitative adverse outcome pathways (qAOPs)¹⁸, metabolomics¹⁹, cheminformatics ^{14,20}, pharmacokinetic-pharmacodynamics (PK-PD) modelling²¹, dynamical pathway modelling with ordinary differential equation (ODE) models²² and

multi-scale approaches modelling DILI with systems biology approaches²³.



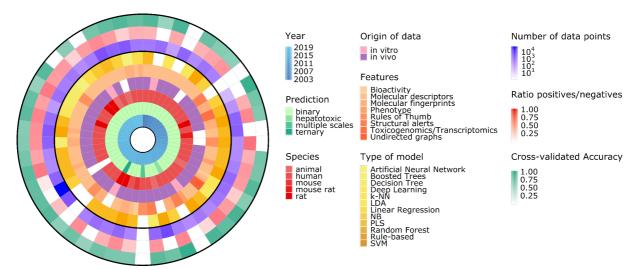


Figure 1: Visual summary of in silico models for liver toxicity prediction

The focus of this work is to characterise the application and scope of published cheminformatics models for DILI and to highlight their relevance, with a particular focus on machine learning.

APPROACHES TO PREDICTING DILI RISK

Better understanding of the underlying mechanisms of DILI, as well as better annotation of the risk associated with drug structures is key for the development of more accurate and valuable predictive models²⁰. Additionally there is no evidence that the mechanisms through which iDILI occurs are different than itDILI^{24,25}. Thus, the focus of DILI research has been to identify reported clinical cases of hepatotoxicity. For instance, such information was compiled by Ludwig and Axelsen²⁶, who created a list of 150 compounds associated with their adverse events. This compilation did not account for the difference(s) between itDILI and iDILI but was one of the first exhaustive lists of hepatotoxic drugs to link phenotypic outcomes in human.

A more recent study classified a list of 611 compounds using high content image screening (HCS) on human cells and compared the findings to conventional assays²⁷. The compounds were classified as either "severely", "moderately" or non-toxic and laid the foundation for the use of *in vitro* data as a surrogate for the prediction of clinical outcomes. Other sources of hepatotoxicity-related compounds come from medicines regulatory agencies and post-marketing data. For instance, Suzuki *et al.* compiled adjudicated cases of DILI reported from the literature resulting from drugs that had been suspended or withdrawn from the market²⁸ and Chen *et al.* annotated compounds based on information provided by the United States Food and Drug Administration (FDA)⁴. The first version of the latter organised compounds into three categories: no-DILI Concern compounds, for which no hepatotoxicity had been

observed, Less-DILI Concern, which caused only mild hepatotoxicity (i.e. steatosis, cholestasis and increase in liver aminotransferases) and Most-DILI concern, which were associated with severe hepatotoxicity⁴. In a later revision, called DILIRank²⁹, the data were curated based on causality evidence. This allowed for the separation of compounds for which association with hepatotoxicity was not supported by sufficient data and allowed for the creation of a new class of compounds (i.e. Ambiguous DILI Concern) consisting of the compounds of the Most and Less DILI Concern classes of the previous version of DILIRank for which no strong evidence of causality was observed.

Fourches *et al.* used text-mining approaches on the titles and abstracts of a collection of articles to identify 902 compounds associated with drug-induced liver effects³⁰. Based on these different approaches to annotate compounds, Kotsampasakou *et al.* aggregated the data from 9 datasets and applied extensive curation techniques³¹. Multiple datasets have been published³² either derived from clinical and/or post-marketing sources, from *in vitro/in vivo* experiments or aggregated from different types of sources (Table 1). However, the published data suffer from two major limitations: data size and imbalance in both the positive and negative DILI group compounds which would bias the outcome of the analysis.

LIMITED DATASET SIZES HAMPERS PROPER MODEL VALIDATION

As a consequence of the nature of the datasets described above, the majority of existing published models for DILI are binary classification models (Table 2). Of these, only one, by Cheng and Dixon, focused exclusively on the prediction of reported itDILI in humans using a set of 382 compounds related to 25 2D molecular descriptors selected with a Monte-Carlo regression algorithm⁷. The leave-10%-out cross-validated random forest model developed had very high specificity and reasonable sensitivity (0.90 and 0.78 respectively). Although similar performance was observed with the test set, its size was quite limited as it only included 23 positive compounds and 31 negatives. Similarly Cruz-Monteagudo *et al.* developed general hepatotoxicity binary classification models from a set of 74 compounds using Radial Distribution Function (RDF) descriptors³³. Even though the performance of the best performing model was consistent between the cross-validation and external validation sets (0.86 and 0.82 respectively), the validation set was small comprising only 13 hepatotoxic compounds and no negatives.

Table 1: Published classifications of drugs for DILI risk

Year		Reference	origin of data	number of compounds	endpoint
1983	26	Ludwig and Axelsen	Compilation of published data	150	Morphological endpoints
1999	34	Zimmerman	Compilation of published data	~500 hepatotoxic drugs	
2005	35	Guo et al.	Compilation of public data	175 drugs	0: no information about hepatotoxicity 1: no significant liver damage reported 2: multiple cases reports or significant injury 3: clear literature evidence of life-threatening hepatotoxicity
2006	27	O'Brien et al.	in vitro cell-based data	381 (42 +/102 ~/237 -)	Severely, moderately and non-toxic
2007	36–38	ToxCast	in vitro data	3799	Biochemical properties based on HTS assays, cell- based phenotypic assays, and genomic and

Year		Reference	origin of data	number of compounds	endpoint
				V .	metabolomic analyses of cells
2008	39	Xu et al.	Drug labels, expert opinion	344 (200 +/144 -)	Hepatotoxic and non-hepatotoxic
2010	40	Ekins et al.	Clinical data for hepatotoxicity	532 (272 +/ 260 -)	Based on Xu et al. 39
2010	30	Fourches et al.	Text mining	951 compounds	Liver effects in humans, rodents or non-rodents
2010	41	Greene et al.	Compilation of published data	1,266	Human or animal-only hepatotoxicity, weak or no evidence
2010	42	Rodgers et al.	FDA reports database	395 (76 +/ 319 –)	Compounds (not) associated with ALT or AST elevation or with combined score
2010	43,44	SIDER database	Compilation of public data	1,430 drugs	Association with 5,868 hepatotoxic side effects
2010	28	Suzuki <i>et al</i> .	Compilation of data from regulatory agencies	473 hepatotoxic drugs	Drugs causing overall injury, acute liver failure and suspended/withdrawn
2011	4	Chen et al.	FDA-approved labels	287 drugs	Most-DILI Concern Less-DILI Concern No DILI Concern
2011	45	Liew et al.	Micromedex reports of adverse reactions	1,274 compounds	T: transient and asymptomatic liver function abnormalities S: liver function abnormalities and hyperbilirubinemia S: hepatitis, jaundice and cholestasis H: fulminant hepatitis and liver failure S: fatality
2011	46	Liu et al.	SIDER database	888 drugs	Association with 13 hepatotoxic side effects
2011	47	Low et al.	in vivo toxicogenomics on rats	127 (53 +/74 –)	Hepatotoxic and non-hepatotoxic
2012	48	Sakatis et al.	Physician's Desk reference	223 (113 +/ 110 -)	Hepatotoxic and non-hepatotoxic
2013	49	Liver Toxicity Knowledge Base	FDA-approved labels	195 (113 +/ 82 –)	Most-DILI Concern No DILI Concern
2013	50	LiverTox	Compilation of published data	~1,200 hepatotoxic drugs, dietary supplements and herbal products	
2014	51	Zhu & Kruhlak	Post-marketing safety data	2,029 (662 +/ 1367 -)	Hepatotoxic and non-hepatotoxic
2016	52	DILIrank	FDA-approved labels	1036 drugs	Verified Most-DILI Concern Verified Less-DILI Concern Verified No DILI Concern Ambiguous DILI Concern
2016	53	Mulliner et al.	Compilation of public data	921 (519 +/ 402 -)	Hierarchical classification in 21 endpoints
2016	54,55	Tox21	In vitro data	~ 10,000	Biochemical properties based on HTS assays
2016		eTOX	in vitro and in vivo data	1947	In-life observations, gross necropsies, histopathology and laboratory values (e.g. clinical chemistry, haematology and urinalysis)
2017	31	Kotsampasakou et al.	Compilation of published data	966 (500 +/ 466 -)	Hepatotoxic and non-hepatotoxic
2018	56	Ai et al.	Zhu & Kruhlak ⁵¹ , FDA Orange Book	1,241 (683 +/ 558 -)	Hepatotoxic and non-hepatotoxic

Although the metrics indicate that Cheng and Dixon's and Cruz-Monteagudo *et al.*'s models performed well, one has to consider that a phenotypic readout such as general hepatotoxicity is the integrated result of many signalling pathways (e.g. oxidative stress and NRF2 pathway⁵⁷, unfolded protein response, DNA damage response and mitochondrial toxicity¹⁷). For each pathway, protein-protein interactions, as well as gene expression or gene and protein degradation could be disturbed, adding up to a multitude of different modes of actions by which a compound could induce toxicity. Thus, building general hepatotoxicity models from a rather small number of diverse compounds increases the difficulty to make reliable generalisations based on compound structures when considering all the possible toxicity modes of action that could be triggered. Xu *et al.* exemplified such a phenomenon and showed that an increase

of the size of the training set improved not only the accuracy of models but also reduced their variability⁵⁸. Additionally, the limited size of external test sets (Table 1) makes the interpretation of the validation of hepatotoxicity prediction models difficult since only a small fraction of hepatotoxicity mechanisms may be validated. The ideal validation set should comprise at least as many compounds as there are ways to disturb the processes involved in these pathways. However, the aggregation of such a dataset is, at this time, not possible. Nevertheless, sizes of both training sets and evaluation sets have been increasing (Table 2), notably through the aggregation and careful data curation of multiple datasets⁵⁹ but also through the United States Environmental Protection Agency's (EPA) ToxCast^{36–38} and the multi-agency Tox21^{54,55} open-data initiatives and the European eTOX^{60–63} and eTRANSAFE consortia. These consortia have gathered pharmaceuticals, data curators, modelers and software developers aiming at building a shared and mineable database of preclinical (eTOX) and clinical (eTRANSAFE) toxicity data to enable more effective read-across and predictive modelling of safety endpoints.

DATASET IMBALANCE

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The second limitation of published datasets is the imbalance of the validation sets (e.g. in ^{33,64–68} in Table 2). These datasets, where either only hepatotoxic compounds are represented or fewer than 10% of compounds are non-hepatotoxicants, do not allow for a proper estimation of the specificity of the models. From the perspective of the training set, the imbalance of the data has been a major challenge to overcome in the prediction of hepatotoxicity: we identified eight articles in which the ratio of nonhepatotoxic compounds considered represented less than 40% of the training set^{7,13,42,69–73}. The opposite trend was observed in six articles where hepatotoxic compounds represented less than 40% of the training set^{68,74–78}. Although building a robust model on an imbalanced dataset is possible, the performance decreases significantly when the number of individuals in the minority class approaches, or becomes, less than 10%. Whilst imbalanced sets affect the robustness of a model, they may better represent the distribution of compounds or dugs observed in real life. This is relevant for the work of Lu et al., who predicted the general hepatotoxicity of compounds based on the profiles of their predicted metabolites⁶⁹, where 64 hepatotoxic and 3,339 non-hepatotoxic compounds were considered – the minority class representing about 2% of the entire dataset. The strategies generally adopted to counteract the systematic prediction of compounds to belong to the majority class are (i) undersampling of the majority class, (ii) oversampling of the minority class⁷⁹, (iii) bagging, (iv) boosting, (v) cost-sensitive learning and (vi) hybrid methods^{80,81}. In their work, Lu et al. used the Synthetic Minority Oversampling Technique (SMOTE) algorithm⁷⁹ to correct for this data imbalance yielding a cross-validated balanced accuracy of 0.60 when predicting hepatotoxicity from predicted metabolites⁶⁹. The application of such meta-classifiers in the prediction of hepatotoxicity is quite recent since only five other works have used them since 2015^{13,72,77,82–84}. It is worth noting that a comparison of the behaviour of meta-classifiers has been performed on few selected imbalanced drug-induced cholestasis datasets⁸⁵. Bagging⁸⁶ has the worst

performance as it does not balance or weight the two classes, threshold selection performed better than bagging but gave lower sensitivity than when using stratified bagging, cost sensitive classifier or Meta-Cost⁸⁷. The authors emphasised the versatility of the stratified bagging technique despite its computational cost when extensive resampling has to be performed.

EARLY DILI PREDICTION STRATEGIES

Among the different *in silico* models that have been developed for the prediction of hepatotoxicity, four main groups of models can be identified based on the features, properties or data the prediction models are built upon: (i) structural alerts, (ii) rules of thumb, (iii) molecular descriptors and (iv) *in vitro* data.

These are described in detail below.

STRUCTURAL ALERTS

Structural alerts are specific substructures of molecules generally associated with hepatotoxicity. Structural alerts are generally developed by experts in toxicology who consider not only toxicological data but also the underlying mechanisms of toxicity, as well as chemical reactivity and biotransformation through metabolism.

One of the first approaches to determining such alerts for DILI utilised a four-stage process⁴¹. A dataset of 1,266 compounds associated with *in vivo* human DILI was aggregated from the literature. Candidate structural classes were derived from these compounds by experts through well-characterised and previously published relationships between compound structures and hepatoxicity. Then these classes were refined by the development of structure-activity relationships (SAR) for which sufficient evidence was available. Finally, the 38 structural alerts classes identified, such as tetracyclines and thiophenes, were validated against an in-house dataset from Pfizer consisting of 626 compounds (412 hepatotoxicants and 214 non-hepatotoxicants). The compounds were classified as either hepatotoxic for humans and/or animals or with weak or no evidence of hepatotoxicity. Although its sensitivity and accuracy were close to random (0.46 and 0.56 respectively) and its specificity quite reasonable (0.77), this approach was not designed for screening purposes. Nevertheless, it should be noted that alerts were prioritised based on their applicability to the Pfizer compound collection. Additionally, compounds that showed unambiguous toxicity during *in vitro* screening were not prioritised for *in vivo* studies, and thus were not considered in this study, potentially explaining the very low sensitivity.

In a second approach a set of 244 hepatotoxic compounds was aggregated from the literature and from failed clinical candidates and drugs withdrawn from the market⁸⁸. From these, 74 structural alerts were derived from mechanistic information, of which 56 were related to reactive and toxic metabolites metabolism. The remaining 18 alerts were based on high cut-off similarity queries, as no mechanistic information could be derived. The authors did not evaluate the predictive performance of these structural alerts but deployed them within the VERDI cheminformatics platform from Vertex pharmaceuticals.

In a third approach⁸⁹, a diverse set of 951 compounds was compiled through curation of the dataset from Fourches *et al.*³⁰. The protein binding potency of each compound was predicted and structural similarity-based clusters of compounds were identified. These categories were then manually curated and related to other well characterised structural alerts. Finally, each alert was thoroughly examined to derive a mechanistic hypothesis for the observed hepatotoxicity. In total 16 structural alerts were characterised. The authors did not validate such alerts on external datasets as their aim was to provide a scheme to identify mechanistically supported structural alerts.

Applying a similar process, Pizzo *et al.* compiled a dataset of 950 compounds of which 510 were hepatotoxicants and identified 13 structural alerts manually and 75 through automatic identification, 11 and 40 of which were respectively associated with hepatotoxicity⁹⁰. The authors then developed an expert-based decision tree based on these structural alerts to predict binary general hepatotoxicity. The model developed was subsequently validated against an external dataset of 101 compounds (69 hepatotoxicants), of which 41% could not be predicted as did not contain any structural alert. Although sensitivity and accuracy were satisfactory for such an approach (0.80 and 0.68 respectively) the model performed poorly in terms of specificity (0.33). Through thorough examination the authors derived a mechanistic hypothesis for the manually derived structural alerts. In addition to the β-lactam substructures, retinoids, oestrogen steroids identified by Hewitt *et al.*⁸⁹, the authors characterised N-containing heterocyclic aromatic compounds, sulphonamides, nucleoside analogues, tricyclic antidepressants, aromatic amines, macrolide antibiotics, anti-bacterial agents, cationic amphiphilic drugs to be mostly associated with hepatotoxicity and nitrosourea compounds not to be associated with hepatotoxicity.

Finally, aggregating DILI associated compounds from LiverTox⁵⁰ with literature findings, Liu *et al.* performed substructure searches using literature-based structural alerts⁹¹. Alerts were ranked by their probability of chance occurrence to classify compounds as being hepatotoxic, non-hepatotoxic, or possible hepatotoxic. This led to the identification of 12 statistically relevant alerts that, unfortunately, were not validated on an external set for prospective prediction. In addition to steroids that were already well characterised hepatotoxicants, sulphonamides, hydrazines, arylacetic acids, anilines, sulfinyls, acyclic bivalent sulphurs, acyclic diaryl ketones, halogen atoms bonded to a sp³ carbon, aminocyclopropyls, aminophenols and phenothiazines were identified as being toxic to the liver.

238 Table 2: Reported computational models for the prediction of DILI

year	ref.	endpoint	prediction	Descriptors	d	ata point	's (positive/negative)	methods	performance	type of data	species
2003	7	general hepatotoxicity	binary	Cerius2 2D molecular descriptors		382 54	(149/233) (23/31)	RF	CV: 0.85 Acc, 0.78 Sen, 0.90 Spe EV: 0.81 Acc, 0.70 Sen, 0.90 Spe	in vivo	human
2004	92	4 endpoints, general hepatotoxicity	binary	molecular electrostatic field			654	SIMCA	0.52 Acc	in vitro	human
2008	33	general hepatotoxicity	binary	radial distribution function		74 13	(33/41) (13/0)	LDA ANN one-level DT	CV: 0.86 Acc, 0.81 Sen, 0.90 Spe EV: 0.82 Acc CV: 0.78 Acc, 0.75 Sen, 0.80 Spe CV: 0.81 Acc, 0.76 Sen, 0.98 Spe	in vivo	human
2009	93	Liver disorders, jaundice and cholestasis, liver enzymes elevation, bile duct disorders	binary	molecular fragment descriptors	EV:	1044 - 18		4 commercial QSAR programs	CV: 0.32-0.47 Sen, 0.85-0.88 Spe EV: 0.89 Sen	in vivo	human
2010	42	AST level, ALT level Composite score	binary	MolconnZ topological descriptors and DRAGON molecular descriptors		190 210 188	(76/114) (84/126) (75/113)	k-NN	CV: 0.74-0.92 Acc, 0.60-0.88 Sen, 0.89-0.96 Spe	in vivo	human
2010	30	general hepatotoxicity	binary	P	EV:		(248/283)	SVM	CV: 0.62-0.68 Acc EV: 0.56-0.73 Acc		human rodents non-rodents
2010	40	general hepatotoxicity	binary	extended connectivity fingerprint with counts and bound diameter 6			(158/137) (114/123)	NB	CV:0.58 Acc, 0.53 Sen, 0.64 Spe EV: 0.60 Acc, 0.56 Sen, 0.67 Spe	in vivo	human
2010	41	general hepatotoxicity	binary	structural alerts	EV:		(412/214)	-	EV: 0.56 Acc, 0.46 Sen, 0.73 Spe	in vivo	human
2011	94	13 hepatopathology endpoints	binary	function class fingerprint with counts and bond diameter 6	CV: EV:			NB	CV: 0.93-0.99 Acc EV: 0.60-0.70 Acc	in vivo	human
2011	47	general hepatotoxicity	binary	toxicogenomics descriptors	CV:	127	(53/74)	RF, k-NN, SVM	CV: 0.69-0.76 Acc, 0.57-0.67 Sen, 0.77-0.84 Spe	in vivo	rats
2011	45	general hepatotoxicity	binary	PaDEL molecular descriptors	CV: EV:	1087 120	(654/433) (72/48)	SVM, NB, k-NN	CV: 0.64 Acc, 0.64 Sen, 0.63 Spe EV: 0.62 Acc, 0.62 Sen, 0.62 Spe	in vivo	human
2012	95	general hepatotoxicity, 3 hepatopathology endpoints	binary	ChemTree augmented atom pairs	CV: EV: 3 en	231-9	01 EV: 28-539	RF	EV: 0.64-0.81 Acc, 0.58-0.73 Sen, 0.71-0.88 Spe EV: 0.62-1.00 Acc, 0.75-1.00 Sen, 0.60-1.00 Spe		mouse rat
2013	78	general hepatotoxicity	binary	Log P and daily dose	CV: EV:		(116/48) (115/64)	Rule of 2	IV: 0.55 Acc, 0.36 Sen, 0.96 Spe EV: 0.51 Acc, 0.29 Sen, 0.91 Spe		human animal
2013	96	general hepatotoxicity	binary	Mold2 chemical descriptors	CV: EV:		(81/116) 28 (95-214/95-114)	RF	CV: 0.70 Acc, 0.58 Sen, 0.78 Sep EV: 0.62-0.69 Acc, 0.58-0.66 Sen, 0.66-0.72 Spe	in vivo	human
2014	97	general hepatotoxicity	binary	CDK, Dragon and MOE molecular descriptors and 8 cellular phenotypes	CV:	292	(156/136)	RF	CV:0.68-0.73 Acc, 0.71-0.73 Sen, 0.64-0.74 Spe	in vivo	human
2014	64	general hepatotoxicity	binary	E-dragon molecular descriptors	IV: EV:	23	(436/436) (54/162) (23/0)	SVM	CV: 0.83 Acc IV: 0.82 Acc, 0.87 Sen, 0.81 Spe EV: 0.74 Acc	in vivo	human
2015	70	hypertrophy, injury, proliferative lesions	binary	QuikProp physicochemical descriptors, PaDEL fingerprints and <i>in vitro</i> bioactivity data		677	(161/463) (101/463) (99/463)	LDA, NB, SVM, k-NN	CV: 0.62-0.84 BAcc, 0.27-0.77 Sen, 0.85-1.00 Spe	in vivo	animal
2015	58	general hepatotoxicity	binary	undirected graph recursive neural networks		475 198	(236/239) (114/84)	DL	CV: 0.88 Acc, 0.90 Sen, 0.87 Spe EV: 0.87 Acc, 0.83 Sen, 0.93 Spe	in vivo	human
2015	68	general hepatotoxicity	binary	PaDEL molecular descriptors	CV: EV:		(136/65) (83/8)	RF	CV: 0.79 Acc, 0.91 Sen, 0.54 Spe EV: 0.87 Acc, 0.90 Sen, 0.63 Spe	in vivo	human

year	ref.	endpoint	prediction	Descriptors	da	ta points	(positive/negative)	methods	performance	type of data	species
2015	65	7 hepatopathology endpoints, general hepatotoxicity	binary	ISIDA descriptors and in vivo endpoints	CV: EV:	414 10	(41-168) (9/1)	SVM, RF, ANN	QSAR CV: 0.58-71 BAcc Endpoints CV: 0.86-0.87 BAcc Endpoints EV: 0.90 Acc, 0.89 Sen, 1.00 Spe	in vitro in vivo	human
2015	91	general hepatotoxicity	hepatotoxic non-hepatotoxic possible hepatotoxic	structural alerts			178 185 242	-		in vivo	human
2016	74	general hepatotoxicity	binary	FP4 descriptors	EV:	336 84	(206/130) (51/33)	NB	CV: 0.94 Acc, 0.97 Sen, 0.89 Spe EV: 0.73 Acc, 0.73 Sen, 0.73 Spe	in vivo	human
2016	53	21 endpoints	binary	CATS, MOE, MDL, VolSurf+ physicochemical descriptors	CV: EV:	3712 221-26	9	SVM with GA	CV: 0.73-0.83 Acc EV: 0.38-0.64 Acc	in vivo	animal human
2016	90	general hepatotoxicity	binary	structural alerts	CV: EV:	950 202	(510/440) (137/65)	expert manual DT	CV: 0.81 Acc, 0.93 Sen, 0.67 Spe EV: 0.68 Acc, 0.80 Sen, 0.33 Spe	in vivo in vitro	human
2016	76	general hepatotoxicity	multiple scales	Log P, daily dose or Cmax, and formation of metabolites	IV:	192	(124/68)	-	IV: 0.47Acc, 0.38 Sen, 1.00 Spe	in vivo	human
2016	98	general hepatotoxicity	binary	CDK, Dragon and Mold2 molecular descriptors, HTS bioactivity data	CV:	233	(67-166)	RF	CV: 0.66-0.73 Acc, 0.62-0.77 Sen, 0.56-0.79 Spe	in vivo	mouse
2016	99	general hepatotoxicity	binary	FP4 and MACCS fingerprints	CV: IV: EV:	978 251 88	(571/407) (155/96) (59/29)	SVM, NB, k-NN, DT, RF	CV: 0.67-0.82 Acc, 0.92-0.96 Sen, 0.32-0.62 Spe IV: 0.60-0.66, 0.77-0.93 Sen, 0.24-0.34 Spe EV: 0.65-0.75 Acc, 0.81-0.93 Sen, 0.21-0.38 Spe	in vivo	human
2017	100	general hepatotoxicity No/Less/Most DILI	binary ternary	Mold2 molecular descriptors		451 721	(183/268) (183/270/268)	RF	CV: 0.73 Acc, 0.63 Sen, 0.53 Spe CV: 0.53 Acc	in vivo in vitro in vitro	rat rat human
2017	101	general hepatotoxicity	binary	PubChem fingerprints	CV: EV:	312 398	(180/132) (224/174)	RF, SVM	CV: 0.73-0.74 Acc EV: 0.61 Acc	in vivo	human
2017	102	general hepatotoxicity	binary	Log P and daily dose	IV:	568	(313/255)	Rule of 2	IV: 0.58 Acc, 0.80 Sen, 0.52 Spe	in vivo	human
2017	73	general hepatotoxicity	binary	CORAL descriptors	CV:	2029	(662/1367)	Monte Carlo optimization	CV: 0.83-0.87 Acc, 0.71-1.00 Sen, 0.85-0.87 Spe	in vivo	human
2017	69	general hepatotoxicity	binary	molecular descriptors	CV:	34023	(64/3339)	NB, Ensemble	CV: 0.78 BAcc, 0.74 Sen, 0.83 Spe CV: 0.60 BAcc, 0.70 Sen, 0.65 Spe		
2017	13	general hepatotoxicity	binary	MACCS public fingerprints, CDK and Mold2 molecular descriptors	CV:	1054	(122/932)	RF	CV: 0.77-0.84 Acc, 0.76-0.88 Sen, 0.73-0.80 Spe	in vivo	human
2017	66	general hepatotoxicity	binary	Mold2 descriptors	CV: EV:		(127/65) (14/6)	RF	CV: 0.80-0.84 Acc, 0.82-0.84 Sen, 0.70-0.75 Spe EV: 0.90 Acc, 1.00 Sen, 0.67 Spe	in vivo	human
2017	103	17 modes of actions	binary	Mold2 descriptors	CV: EV:	222 111	(155/178)	RF	CV: 0.70-0.76 Acc IV: 0.70-0.71 Acc	in vivo	human
2018	56	general hepatotoxicity	binary	CDK estate, MACCS, FP4, atom pairs fingerprints	CV: EV:	1241 286	(683/558) (221/65)	XGBoost, RF, SVM	CV: 0.63-0.70 Acc, 0.66-0.82 Sen, 0.41-0.63 Spe EV: 0.73-0.86 Acc, 0.72-0.89 Sen, 0.42-0.83 Spe	in vivo	human
2018	104	general hepatotoxicity	binary	PaDEL molecular descriptors and fingerprints	1731 IV: EV:	(980/7: 413 151	51) (270/143) (88/63)	SVM, k-NN, NB, DT, RF	IV: 0.62-0.80 Acc, 0.53-0.97 Sen,0.13-0.83 Spe EV: 0.66-0.83 Acc, 0.68-0.93 Sen, 0.54-0.70 Spe	in vivo	human
2018	75	general hepatotoxicity	binary	PaDEL descriptors	CV:	712	(444/268)	RF, ANN	CV: 0.80-0.90 Acc, 0.78-0.90 Sen, 0.81-0.90 Spe	in vivo	human
2018	105	general hepatotoxicity	binary	PaDEL molecular descriptors	CV: EV:	99 25	(48/51) (10/15)	k-NN with GA	CV: 0.76 Acc, 0.79 Sen, 0.74 Spe EV: 0.92 Acc, 0.90 Sen, 0.93 Spe	in vivo	rats
2018	77	general hepatotoxicity	binary	PaDEL molecular descriptors	CV:	575	(384/191)	DT, k-NN, SVM, ANN	CV: 0.53-0.98 Acc	in vivo	human

year	ref.	endpoint	prediction	Descriptors	data points (positive/negative)	methods	performance	type of data	species
2018	52	general hepatotoxicity	binary	maximum daily dose, LogP, Fraction of sp3 carbons	326 (163/163)	Expert manual DT	0.82 Acc, 0.79 Sen, 0.85 Spe	in vivo	human
2018	106	hepatocellular hypertrophy	binary	DRAGON molecular descriptors	405 (207/198) EV: 405 (218/187)	ANN, RF, SVM	EV: 0.68-0.76 Acc, 0.58-0.90 Sen, 0.46-0.84 Spe	in vivo	rats
2018	72	serum ALT level	binary	DRAGON molecular descriptors	CV: 176 (40/136)	LR	CV: 0.60 Acc, 0.65 Sen, 0.58 Spe EV: 0.60 Sen, 0.40-0.50Acc and Spe	in vivo	rats
2018	107	non-neoplasic proliferative lesions inflammatory liver changes degenerative lesions	binary and continuous	Adriana and GRIND2 molecular descriptors	332 (168/164) 258 (164/94) 246 (164/82)	PLS, RF	CV: 0.70 Sen, 0.69 Spe EV: 0.50 Sen, 0.62 Spe CV: 0.44 Sen, 0.84 Spe EV: 0.54 Sen, 0.76 Spe CV: 0.68 Sen, 0.55 Spe EV: 0.67 Sen, 0.59 Spe	in vitro in vivo	animal
2018	108	general hepatotoxicity	four categories		EV: 164 (116/48) EV: 192 (124/68)	Rule-based	EV: 0.62-0.72 Acc EV: 0.66-0.78 Acc	in vivo in vitro	human animal
2019	67	general hepatotoxicity 4 severity degrees 22 adverse events	binary	MOE molecular descriptors	CV: 2513 (1475-1720/1038) CV: 426-1180 (213-590/213-590) CV: 200-1104 (100-552/100-552) EV: 11-16/0	RF	CV: 0.69 Acc, 0.84 Sen, 0.51 Spe CV: 0.70-0.71 Acc, 0.71-0.77 Sen, 0.63-0.70 Spe CV: 0.67-0.78 Acc, 0.65-0.84 Sen, 0.63-0.81 Spe Tiered CV: 0.67 Acc EV: 0.81-0.82 Spe	in vivo	human
2019	109	general hepatotoxicity	binary	Marvin molecular descriptors	CV: 1254 (636/618) EV: 204 (125/79)	NB, k-NN, RF, ANN, Ensemble	CV: 0.60-78 Acc, 0.61-0.86 Sen, 0.40-0.76 Spe Ensemble CV: 0.78 Acc, 0.82 Sen, 0.75 Spe Ensemble EV: 0.73 Acc, 0.77 Sen, 0.66 Spe		animal human
2019	71	general hepatotoxicity	binary	PaDEL molecular fingerprints	CV: 1812 (453/1359) IV: 664 (166/498)	ANN, SVM, RF, k-NN, Ensemble	CV: 0.85-0.90 Acc, 0.71-0.86 Sen, 0.82-0.92 Spe IV: 0.82-0.89 Acc, 0.60-0.80 Sen, 0.83-0.93 Spe	in vivo	human
2019	83	biliary hyperplasia, fibrosis, and necrosis	binary	transcriptomic data	CV: 2324 (91/2333, 37/2287, 275/2049) EV: 341-376 (20/321, 22/354, 32/326)	DL, RF, SVM	CV: 0.48-0.89 MCC EV: 0.36-0.90 MCC	in vivo	rats
2019	110	general hepatotoxicity	binary	PaDEL molecular fingerprints and descriptors	450 (182/268)	LR, SVM, GBT, RF, Ensemble	CV: 0.77 Acc, 0.64 Sen, 0.86 Spe IV: 0.82 Acc, 0.65 Sen, 0.96 Spe	in vivo	human
2019	111	general hepatotoxicity	four categories		CV: 200 (79/56/47/18) IV: 21 EV: 7	Rule of Thumbs	CV: 41-80 Sen, 58-97 Spe	in vivo	human
2019		5 1	ternary	assays	96 (33/40/23)	NB	0.63 BAcc Binary: 0.86 Acc, 0.87 Sen, 0.85 Spe, [extremally gradient tree boosting 113] is an imple	in vivo	human

ANN: artificial neural network, DL: deep learning, DT: decision tree, GA: genetic algorithm, GBT: gradient-boosted trees (of which XGBoost [extremally gradient tree boosting¹¹³] is an implementation),

k-NN: k-nearest neighbours, LDA: latent Dirichlet allocation, LR: logistic regression, NB: naïve Bayes, PLS: partial least squares, RF: random forest, SVM: support vector machine, CV: cross-validation,

IV: internal validation, EV: external validation, Acc: accuracy, BAcc: balanced accuracy, Sen: sensitivity, Spe: specificity, MCC: Matthews correlation coefficient

Other studies on the development of quantitative structure-activity relationship (QSAR) models have also focused on the identification of molecular patterns related to hepatotoxicity. Structural fingerprints of compounds (e.g. Kletkota-Roth¹¹⁴ or extended connectivity fingerprints¹¹⁵) have been calculated for a training set. Association of the presence of one pattern with hepatotoxicity was evaluated either based on the feature importance of each bit of such fingerprints or on their frequency. The importance of fingerprint bits has been notably derived from extended connectivity fingerprints with a maximum diameter of 6 (ECFP6) using naïve Bayes models^{40,74} and a random forest ⁵⁶ with 12 different fingerprints. This analysis pointed not only to substructures associated with hepatotoxicity but also those associated with non-hepatotoxic compounds. Frequency focused determination of substructures of interest was performed either by determining the information gain of using such substructures or by using logistic regression, and deriving odds ratios and/or *p*-values associated with these moieties^{13,30,45,99,104,106}.

The real benefit of using structural alerts is that they may be associated with well characterised mechanisms (e.g. biotransformation to reactive metabolites or alteration in membrane structure integrity, adduction to proteins) and with specific organ level toxicity effects¹¹⁶. This reason makes them valuable when determining the toxicity of new drugs and postulating key mechanisms involved. In addition to expert-derived structural alerts, the identification of key substructures associated with DILI is of crucial importance since it allows for further research on, and understanding of, the associated underlying mechanisms.

Nevertheless, a key concept of applying structural alerts is that the absence of a matching alert for a compound is not proof of it not being hepatotoxic¹¹⁷. Moreover, the presence of structural alerts should not be seen as a clear indication of the DILI potential of a drug. To emphasise this, Stepan *et al.* retrospectively examined the 200 most prescribed and sold drugs in the US in 2009 and 68 other drugs that had been recalled or were associated with black box warning due to iDILI¹¹⁸. Although structural alerts were present in 78%-86% of hepatotoxic drugs, approximately half of the top 200 drugs for 2009 also contained one or more structural alerts, mitigating the use of alerts in for the screening of the toxicity of a compound. According to the authors, "the major differentiating factor appeared to be the daily dose", as drugs with high daily doses were mostly associated with toxicity.

RULES OF THUMB

To expand on Stepan *et al.*'s observation about daily dose, few rules of thumb based on two or three molecular features of compounds have been derived. Chen *et al.* identified that from a dataset of 164 US FDA-approved oral medications, a high risk of DILI was associated with lipophilic drugs (Log P \geq 2) given at high dosage (daily dose \geq 100 mg; odds ratio 14.05, *p*-value < 0.001)⁷⁸. This '*rule of two*' was validated using Greene *et al.*'s dataset of 179 oral medications⁴¹. Of the compounds being positive

for such a rule, 85% were associated with hepatotoxicity. However, this high positive predicted value was associated with very low sensitivity (0.29) but very high specificity (0.91), which overall gives an accuracy (0.51) close to that of a random prediction. When applying this 'rule of two' to five datasets^{29,39,41,48,51}, accounting for a total of 1,036 compounds, the authors noticed that the association between toxicity and high lipophilicity was statistically significant for only three of them (those of Chen et al., Greene et al. and Zhu et al.). Moreover they found that all compounds with a daily dose higher than 100 mg per day were significantly associated with DILI risk¹⁰². The authors also collected hepatic metabolism information for 398 drugs and observed that drugs, which are more than 50% metabolised in the liver, were more prone to be hepatotoxic (odds ratios between 1.80 and 2.67). Combining significant hepatic metabolism with high daily dose allowed for the correct identification of 78% of hepatotoxic compounds and 60% of non-hepatotoxicants, giving this prediction method an overall accuracy of 0.68. Factoring high lipophilicity with reactive metabolite (RM) formation and high daily dose for a dataset of 192 drugs, the authors were then able to develop a prediction method with a specificity of 1.00 but sensitivity of 0.38⁷⁶. The assessment of the association between daily dose, lipophilicity, RM formation and DILI risk by logistic regression analysis confirmed the significant importance of these features 119 and allowed for the development of a DILI score significantly correlated with the severity of liver injury in human for three different datasets^{4,28,41}.

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Another rule of thumb was derived by Leeson, who investigated the predictivity of physicochemical properties of compounds related to their dose⁵². More specifically, the differences between dose, lipophilicity and the fraction of sp³ hybridised carbons atoms (Fsp3) in relationship to whether drugs with the most and no DILI concern were acids, bases or neutral (from the Chen *et al.* dataset)⁴ were examined. As the mean Fsp3 values of bases, which were enriched in the non-hepatotoxicants class, are greater than for acids¹²⁰, the author was able to integrate Fsp3 to the '*rule of two*', yielding accurate predictions for 82% of compounds and with high and balanced sensitivity and specificity (0.79 and 0.85 respectively).

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Despite the simplicity of these rules of thumb that have high specificity, their major flaw is that their applicability is limited to the datasets they are built upon¹⁰². The datasets may have different causality assessment scales to derive DILI annotation which vary from one dataset to the other¹²¹, or reported hepatotoxicity evidence maybe is vague^{122,123}. This limitation of the data was stressed by Leeson who identified that among the 155 oral drugs belonging to the top 200 prescribed medications in the US in 2009 that were annotated by Chen *et al*⁴, 59% belonged to the Less DILI category, hence questioning the significance of such a class.

QUANTITATIVE STRUCTURE-ACTIVITY AND TOXICITY RELATIONSHIPS

Because the acquisition of some of the parameters mentioned above is only possible from in vitro and

in vivo studies QSAR or structure-toxicity relationship-based models have been developed using molecular properties to allow for the early screening of compounds for which no data exist. Examples of experimental properties which may not be available for models include metabolism activity, maximum daily dose or peak concentration in serum after drug administration (Cmax). There are several different types of cheminformatics model: models predicting general hepatotoxicity, histopathological phenotypes (e.g. increase in serum biomarkers, cholangitis) or specific modes of action mediated through protein-ligand interactions.

General hepatotoxicity

Derived from the first phenotypic observations of hepatotoxicity and used to provide a general estimation for compound prioritisation in drug discovery, QSAR models were first built using general binary DILI annotations. For instance, Cheng and Dixon developed one of the first hepatotoxicity QSAR models derived from molecular descriptors, without regard to dose-dependence. In addition to those descriptors, the similarities to the 382 compounds in the training set (149 hepatotoxicants and 233 non-hepatotoxicants) were also used as explanatory variables. Monte Carlo feature selection was applied to reduce the number of descriptors to 25, of which 6 were physicochemical properties. A random forest model was developed and validated on a test set of 54 compounds. Its performance was very encouraging with good accuracy, fair sensitivity and high specificity (0.81, 0.70 and 0.90 respectively). However, such an approach, with such a limited description of the molecular structure and similarity profiles to the training set, did not allow for extrapolation to other compound classes.

Since then, a wide variety of general QSAR models predicting hepatotoxicity have been derived using different types of molecular descriptors, molecular fingerprints and machine learning algorithms (see Table 2). The most recent work predicting general hepatotoxicity solely from molecular descriptors is from He *et al.* ¹⁰⁹. The authors combined a total of 14 datasets for which hepatotoxicity labels originated from animal and cell experiments, clinical reports, drug labels, medical monographs and the scientific literature. In addition, compounds that were classified by fewer than two of eight effective classifiers were discarded, allowing for the creation of a large, balanced and high-quality dataset of 1,254 compounds (636 positives and 638 negatives). Using a set of 85 physicochemical and topological properties an ensemble model based from the eight base classifiers was obtained with high and balanced performance evaluated with 10-fold cross-validation (sensitivity 0.82, specificity 0.75, accuracy 0.78 and balanced accuracy 0.78) and on an external test set of 204 compounds (sensitivity 0.77, specificity 0.66, accuracy 0.73 and balanced accuracy 0.72). To further validate their model to identify non-hepatotoxicants, the authors assembled a dataset of 312 negative compounds. Their classification ensemble model correctly predicted 215 of these compounds, giving a reasonable accuracy of 0.70. The relevance of building classification models from molecular descriptors alone, in comparison with

molecular fingerprints, was questioned by Li et al. 104. The relative performances of k-nearest neighbour

(k-NN), support vector machine (SVM), random forest (RF), naïve Bayesian (NB) and decision tree (DT) models built from seven PaDEL molecular fingerprints¹²⁴ and molecular descriptors were compared for a dataset of 980 DILI-positive and 751 DILI-negative compounds. Models based solely on molecular descriptors had the lowest average performance with low accuracy (0.62 to 0.73), specificity (0.13 to 0.70) and AUC (0.0.63 to 0.78). The combination of public MACCS fingerprints in an SVM yielded the best classification performance on an external test set of 88 hepatotoxicants and 63 non-hepatotoxicants (0.83 accuracy, 0.93 sensitivity, 0.68 specificity and 0.88 AUC) despite their limited dimensionality of 166 bits. Only one model, also developed with public MACCS fingerprints but using k-NN, had higher specificity than the previous one (0.70) but lower accuracy, sensitivity and AUC (0.76, 0.81 and 0.82 respectively). This emphasised the usefulness of ensemble models, which was the strategy used by Wu et al. 71, who combined four PaDEL molecular fingerprints with k-NN, RF, SVM and artificial neural network (ANN) base classifiers in consensus voting models and also identified the public MACCS fingerprints and SVM-based based classifier to perform well on an external test set of 166 positive and 498 negative compounds (0.75 sensitivity, 0.93 specificity, 0.88 accuracy and 0.70 Matthews correlation coefficient [MCC]). Their consensus models were based on the number of times a compound was predicted to be hepatotoxic by base classifiers. The best performing consensus model, which was that based on three positive predictions out of the 4 base classifiers, was selected (0.77 sensitivity, 0.97 specificity, 0.92 accuracy and 0.78 MCC respectively).

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Ai *et al.*⁵⁶ adopted the same strategy as Wu *et al.* but filtered out bits of the fingerprints that were correlated and did not apply them to the dataset (e.g. all molecules contain carbon atoms so this information was removed). The five best performing base classifiers in terms of AUC, which interestingly did not include any based on public MACCS fingerprints, were then combined in an ensemble model by averaging their predicted hepatotoxicity probability (0.84 accuracy, 0.87 sensitivity, 0.75 specificity and 0.90 AUC on the external test set).

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Wang *et al.*¹¹⁰ recently combined the Ai *et al.*'s approach with the work of He *et al.* by developing an ensemble model based on the eight PaDEL fingerprints that performed best on their dataset as well as an ensemble model based on seven simple molecular properties (ALogP, molecular weight and numbers of aromatic rings, hydrogen-bond donors, acceptors, rotatable bonds and rings). The five base classifiers used for both these ensemble models were random forest and boosting tree models. The average probabilities for each ensemble were then summed and the weighted average of the two (i.e. 0.7 for fingerprint-based and 0.3 for molecular property-based) were used to classify compounds. The performance of the model was comparable, although slightly lower, than that obtained by Ai *et al.* but specificity was very good (0.82 accuracy, 0.65 sensitivity, 0.96 specificity, 0.80 AUC).

Phenotypically-focused models

To compensate for the complexity of predicting general hepatotoxicity, models focused on finer phenotypes have been devised. In this sense, Myshkin et al. derived an ontology database of hepatotoxic pathology from human and animal publicly available toxicity data⁹⁵. This database was organised by the type of pathology and by organ substructure and function impairment. From this ontology, different toxicity datasets were identified among which were datasets related to liver necrosis, liver weight gain and liver steatosis, comprising of 300, 305 and 172 instances respectively. For each endpoint, random forest OSAR models were derived using augmented atom pairs¹²⁵. The best performing models were then evaluated on external test sets (490, 539 and 478 respectively). Results were encouraging with 0.63, 0.74 and 0.60 specificity for liver necrosis, weight gain and liver steatosis respectively, 0.87, 0.86 and 0.75 sensitivity, 0.66, 0.76 and 0.62 accuracy and 0.35, 0.51 and 0.23 Matthews correlation coefficient. The authors then characterised the applicability domain of their models based on a Tanimoto distance between compounds in the training and test set. The models were quite sensitive as sensitivity decreased for compounds in the 30-59% compound dissimilarity range. Interestingly, the model based on weight gain was very robust as sensitivity remained above 0.72 for the entire 30-99% range. It is worth mentioning that these three models performed better than a general hepatotoxicity model (0.58 sensitivity, 0.71 specificity, 0.64 accuracy and 0.29 Matthews correlation coefficient) which showed a high sensitivity of 0.82 for the 30-39% Tanimoto dissimilarity range, highlighting the relatively high diversity of compounds in the validation set.

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Another work by Takeshita focused on the prediction of alanine transferase (ALT) elevation in rats from repeated-dose toxicity studies⁷². Two logistic regression models, with seven and nine explanatory variables out of an initial 3,636 DRAGON molecular descriptors respectively ¹²⁶, were derived to classify 176 compounds. Compounds which had either a lowest observed effect level (LOEL) associated to ALT elevation, (40 positives and 136 negatives) or an elevation in ALT at a dose below 1000 mg/kg (23 strong and 153 weak compounds) were included. Because of the imbalance of their datasets, the authors used the SMOTE algorithm⁷⁹. Although classification performance on the training set was limited between toxic and non-toxic compounds (0.65 sensitivity, 0.581 specificity and 0.600 accuracy), the logistic model showed better discrimination between weak and strong compounds (0.78 sensitivity, 0.74 specificity and 0.75 accuracy). External validation on a dataset of 59 compounds (23 strong and 36 weak compounds) showed decreased performance (0.60 sensitivity and specificity and accuracy between 0.40 and 0.50). Nevertheless, the significant difference between 52 out of a set of 197 molecular descriptors from the training and test sets was observed by the authors, emphasising the need for applicability domain determination.

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Focusing only on *in vivo* hepatocellular hypertrophy in rats, Ambe *et al.* developed deep learning (DL), RF and SVM classification models¹⁰⁶. The authors collected rat toxicity data following chronic exposure

of more than 27 days from two sources. Models were trained on half of the data of the two datasets (173 and 251 compounds respectively) as well as on half of their combination (405 compounds) and respectively evaluated on their other halves. DL models were clearly overfitted to the data. Their ROC AUC was 1.00 and accuracy, sensitivity, specificity were 0.96 when evaluated on training set, but dropped when the test set was evaluated. However, the DL model based on the combined dataset did not show such behaviour with more equivalent performance between training and test set. This observation could be the combined result of the two-fold increase in the size of the dataset and the reduction of features from 433 and 417 to 385, corresponding to a decrease in dimensionality by 7.7% to 11.1%. The applicability domain of the models was determined using distance in the molecular space to the training set¹²⁷ and resulted in 19, 38 and 50 compounds lying outside for two test sets and their combined version respectively. Using a consensus model based on the majority principle, similar predictive performance was achieved. Of the 107 compounds incorrectly predicted by the consensus model, 78 were predicted incorrectly by all three models. These incorrectly predicted compounds were mostly false positives and the authors exemplified the case of flufenoxuron, a benzoylphenyl urea-based insecticide which is not a hepatocellular hypertrophy inducer in rats but is in mouse carcinogenicity studies. This highlighted the need for the development of models in other species not only for better prediction, but also translation between species and understanding of any species-specific mechanisms involved.

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Mulliner et al. investigated species specific effects by creating hierarchical seven endpoint hepatopathology trees for human and preclinical findings⁵³. An additional tree was developed for preclinical findings obtained at doses lower than 500 mg/kg in order to reduce the leverage of high dose toxicants during model development. The endpoints were organised in three different levels: general hepatotoxicity, morphological and clinical chemistry findings, hepatocellular and hepatobiliary injuries. A total of 3,712 compounds were aggregated with overall concordance between human and animal hepatotoxicity of 77%. Individual SVM classification models were developed for each endpoint using a genetic algorithm for feature selection. All human endpoints were reasonably well predicted with accuracies between 0.73 and 0.78 for internal validation. For preclinical endpoints, only general hepatotoxicity could be modelled confidently for toxicity above 500 mg/kg (ROC AUC of 0.73 and lower than 0.67 for others in internal validation). Conversely all endpoints with the exception of hepatobiliary injuries could be modelled for toxicity below such a threshold (accuracies between 0.75 and 0.83 in internal validation). An external validation on 269 proprietary compounds with 14 to 28-day rat study data showed decreased performance for all models (accuracies between 0.38 and 0.64 and ROC AUC between 0.51 and 0.68). The reduction in performance observed between internal and external validation for preclinical data was expected to be similar for human endpoints, more especially when applying these models on early research drug candidates which do not exhibit similar molecular properties as drugs.

460 A similar work by López-Massaguer et al. 107 relied on an ontology to classify compounds for three 461 endpoints as well as predict the LOEL of compounds from the eTOX database⁶¹. This database was 462 463 derived from multiple types of publicly available and confidential preclinical data, in multiple species, 464 for various administration routes and for different exposure times. Aggregating rat in vivo microscopy 465 and hepatopathology findings, the authors gathered 164, 94 and 82 positive compounds for the three 466 endpoints (i.e. degenerative lesions [DEG], inflammatory liver changes [INF] and non-neoplasic 467 proliferative lesions [PRO]). It is worth noting that the negative compounds that were selected had been 468 tested at concentrations higher than 1000 mg/kg and had no observed treatment-related and liver-related 469 histopathology findings (168, 164 and 164 for DEG, INF and PRO respectively). Sensitivities and 470 specificities of random forest classification models were balanced after both cross and external 471 validation for PRO (0.70 and 0.50 sensitivities and 0.69 and 0.62 specificities at cross and external 472 validation respectively) and DEG (0.68 and 0.67 sensitivities and 0.55 and 0.59 specificities at cross and 473 external validation respectively) while were unbalanced for INF (0.84 and 0.67 sensitivities and 0.44 474 and 0.54 specificities at cross and external validation respectively). Partial least square regression 475 models showed poor fit with low goodness-of-fit (ranging from 0.26 to 0.58), poor predictive 476 performance (Q² ranging from -0.84 to 0.07) and high standard deviation (ranging from 1 to 2 log units). 477 This work emphasised the possibility of stringent selection of negative compounds as well as 478 aggregation of multiple sources of data containing compounds with different routes of administration 479 and exposure times. 480 Relying on an hepatopathology-based ontology, as was carried out in the two previous approaches, Liu et al. introduced a severity grade in their hierarchical approach⁶⁷. The authors organised their 481 ontology into three levels: level 1 denoted general hepatotoxicity, level 2 corresponded to the severity 482 483 of the hepatotoxicity and level 3 associated with adverse events (e.g. acute liver failure, cholestasis or 484 AST elevation). A total of 2,017 compounds associated with 403 clinical grade 3 adverse events were collected from SIDER^{43,44} and LiverTox⁵⁰, amongst other, databases. Individual classification random 485 486 forest models were built for 22 endpoints. The level 1 classification model, predicting general 487 hepatotoxicity, showed good sensitivity and ROC AUC but low specificity (0.81, 0.75 and 0.50 respectively). Models based on DILI severity showed more balanced sensitivities and specificities (0.70-488 489 0.71 and 0.63-0.70 respectively) resulting in comparable or slightly higher ROC AUC (0.75-0.78). 490 Adverse events prediction models showed balanced sensitivity and specificity ranging from 0.65 to 0.83 491 and from 0.63 to 0.79 respectively, as well as reasonable accuracy (0.67-0.78) and a high ROC AUC 492 (0.71 to 0.87). The 27 models were integrated in a tiered prediction model with high sensitivity (0.82). 493 Because of the limited size of the external validation dataset, adverse events prediction at level 3 was a 494 qualitative assessment of the models. Nevertheless, ticrynafen, which had been withdrawn from the

market for association with hepatitis, was predicted by level 3 models to be associated with hepatitis,

acute hepatic failure, and hepatocellular injury.

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Prediction of Specific Modes of Action

Biological mechanism-focused models have been gaining increasing interest in recent years, under the auspices and needs of the ToxCast and Tox21 initiatives. An example is the work of Wu *et al.*¹⁰³, who integrated quantitative high-throughput screening bioassay activity data to develop 17 QSAR models. The profiles of mode of action (MOA) of drugs were predicted with a set of 777 2D molecular descriptors using random forest models. The accuracies of prediction models ranged between 0.63 and 0.67, which was quite encouraging considering the imbalance in the data. Nevertheless, when predicting general hepatotoxicity from the predicted MOA profiles, 5-fold cross-validation on a dataset of 222 compounds (155 hepatotoxicants and 178 non-hepatotoxicants with test set included) gave an accuracy of 0.76 and internal validation on 111 drugs gave accuracy of 0.70. This performance was higher than when using a standard QSAR model (accuracy of 0.66 for cross-validation). Interestingly, the general hepatotoxicity model derived from the top four performing MOA profiles prediction models had slightly higher accuracy on the internal validation set while slightly lower through cross-validation (0.71 and 0.70 respectively). These models could be regarded as underperforming as compared to recent general hepatotoxicity QSAR models, however, it should be noted that only a small number of MOAs were considered in this study with regard to the different mechanisms involved in DILI.

Some other studies on the prediction of MOA profiles have been more focused on specific phenotypes. For instance, an impairment of the function of export pumps and transport proteins in the liver would result in the progress of a cholestatic phenotype. The export pumps comprise the biliary salt export pump (BSEP), the breast cancer resistance protein (BCRP) and the P-glycoprotein (P-gp). The transport proteins are the organic-anion-transporting polypeptides (OATPs). OATPs are members of the solute carrier (SLC) family and transport organic anions. Few models have been developed to predict the inhibition of such proteins. A prospective analysis⁸² was carried out to identify OATP1B1 and OATP1B3 inhibitors out of DrugBank¹²⁸. This screening was based on a training dataset of 1,708 compounds (190 inhibitors and 1,518 non-inhibitors) for OATP1B1 and of 1,725 compounds (124 inhibitors and 1,601 non-inhibitors) for OATP1B3, respectively. An external test set containing 201 compounds for OATP1B1 (64 inhibitors and 137 non-inhibitors) and 209 compounds for OATP1B3 (40 inhibitors and 169 non-inhibitors) was used to assess the validity of the model along with 5-fold and 10fold cross-validation. Two random forests and four support vector machine classifiers, using MetaCost⁸⁷ as metaclassifier to deal with the imbalance of the dataset, were generated for each transporter. As the performance of the models was relatively equivalent – accuracy values and ROC AUC for the test set in the range of 0.81-0.86 and of 0.81-0.92, respectively - a consensus scoring approach was used, summing up the prediction scores of each classification model. The screening of DrugBank (6,279 compounds) resulted in the identification and biological testing of the 9 compounds with highest predicted probability of being OATPB1 and O1TPB3 dual inhibitors and 1 selective inhibitor of OATP1B3. Only the latter was incorrectly predicted, yielding an accuracy of 90% for OATP1B1 and

80% for OATP1B3, respectively.

To compare the prediction of an inhibitory effect of transport proteins to a phenotypic readout, the relative performance of meta classifiers on unbalanced datasets was studied for OATP1B1 and OATP1B3 inhibition, human cholestasis and animal cholestasis based on molecular descriptors^{85,129}. Although imbalance ratios between negatives and positives ranged from 2:1 to 20:1, the balanced accuracies of models with sensitivity higher than 0.5 ranged from 0.67 to 0.83 for OATPB1, 0.63 to 0.86 for OATP1B3 and 0.64 to 0.78 for human cholestasis on test set and from 0.53 to 0.65 for animal cholestasis. This emphasised the difficulty in predicting a phenotypic outcome solely from compound structure.

Other work focused on the prediction of BSEP and MRP4 inhibition from both statistical and structure-based approaches¹³⁰. In this study, 57 and 171 compounds along with inhibitory effect on MRP4 and BSEP were gathered respectively. Bayesian models were trained on simple molecular descriptors and either extended-connectivity fingerprints maximum diameter 6 (ECFP6) or functional-class fingerprints maximum diameter 6 (FCFP6). For MRP4, although the models performed well in terms of specificity, they did not show high sensitivity. Nevertheless, the MRP4 pharmacophore model built on 9 compounds was able to correctly classify 30 of the 42 actives in the test set and 22 of the 35 inactives, leading to a sensitivity of 0.71 and specificity of 0.63. The BSEP inhibition prediction model showed more balanced and higher performance (sensitivity of 0.82 and 0.77, specificity of 0.77 and 0.84 respectively) but the pharmacophore model had a higher selectivity whilst poor specificity of 0.37. The lower performance of the MRP4 classification model was probably due to the 3:1 ratio between active and inactive compounds in the training dataset and to the small size of the dataset comprising only 86 compounds. This work emphasised not only the usefulness of structure-based modelling when it comes to the prediction of inhibitory effects of compounds but also the requirement for well-balanced datasets.

This difficulty to predict a phenotypic outcome of a compound using an imbalanced dataset was tackled using metaclassifiers and considering the predicted inhibitory effect of compounds on transport proteins as descriptors⁸⁴. Cholestasis-focused data were aggregated by mining and manually curating the literature for human drug-induced cholestasis. A total of 578 compounds were identified, of which 131 were cholestasis positives and 447 were DILI negatives. A k-NN classifier with MetaCost metaclassifier for data imbalance correction was generated and evaluated through both 10-fold cross-validation and external testing on a dataset covering multiple levels of hepatotoxicity and including hepatobiliary injury⁵³. Inclusion of BSEP, BCRP, P-glycoprotein, and OATP1B1 and OATP1B3 inhibition predictions increased accuracy (0.66 to 0.70) and ROC AUC (0.66 to 0.73) of the model through 10-fold cross validation but decreased for the test set (0.61 to 0.56 and 0.62 to 0.58 respectively). The authors speculated that this was the result of a different class assignments between the training and test

sets and argued that almost 20% of the compounds in the external validation set had contradictory labels with the training set (71 out of 419 shared compounds). Nevertheless, the authors showed that accuracy and specificity reach their peak only after the inclusion of BSEP predictions, but that when only using BSEP predictions, the model showed a slight increase in accuracy and specificity of the model but decreased sensitivity. This suggested that BSEP inhibition conveys most, but not all, of the relevant information when modelling cholestasis.

An effort to merge multiple publicly available datasets was undertaken to apply the models obtained to other datasets and investigate how export pump and transporter inhibition correlate to general hepatotoxicity⁵⁹. In this work, the authors gathered nine previously published datasets for model training (966 compounds) and three datasets for validation (996 compounds). Three random forests classifiers were built using two sets of molecular descriptors to predict transporter inhibition^{82,131,132}. Accuracy and ROC AUC of the models ranged from 0.57 to 0.69 and from 0.59 to 0.73 respectively in spite of the heterogeneity of such a dataset, ranging from in vitro cell-based assay readouts to FDA reports and postmarketing safety data. Nevertheless, the introduction of BSEP, BCRP, P-glycoprotein, and OATP1B1 and OATP1B3 inhibition binary prediction as descriptors slightly decreased the model performance. The authors argued that this could be the result of mispredictions of such transporter inhibition models resulting in noise added to the feature matrix and that the inhibition of only one transporter would not alter the function of hepatocytes. With regards to such possible misclassifications, the use of a hard threshold at 10 µM to classify a compound as being an inhibitor can lead to misclassification of compounds with IC50 around such a threshold, thus artificially lowering the performance of the model. Additionally, such a threshold is not in accordance with the 300 µM value that was suggested to be used for BSEP inhibition¹³³. QSAR models modelling BSEP inhibition based on the latter threshold showed very good performance 134,135. Finally, the endpoint to be predicted denotes general phenotypic hepatotoxicity and correlates only with transporter inhibition which is associated mostly with cholestasis.

It should also be noted that the BSEP, BCRP, P-glycoprotein, and OATP1B1 and OATP1B3 do not represent the entirety of transporters. One could also cite the canicular and basolateral multidrug resistance-associated proteins (MRP1 to MRP6), the organic solute transporters (OSTα/OSTβ), the multidrug and toxin extrusion transporter 1 (MATE1), the ATP-binding cassette subfamily G member 5/8 (ABCG5/G8), the multidrug resistance protein 3 (MDR3), the ATPase-aminophospholipid transporter (ATP8B1), the sodium taurocolate co-transporting polypeptide (NTCP), the organic cation transporters 1 and 3 (OCT1/3), the organic anion transporters 2 and 7 (OAT2/7) and other organic anion transporting polypeptides (e.g. OATP2B1)¹³⁶. However, to date, very few inhibition data have been collected for these targets, making such a modelling exercise rather difficult if not unfeasible.

Finally, Peng et *al.* developed MOA prediction models in the context of steatosis¹³⁷. Data from 24 *in vitro* HTS assays from the ToxCast program were compiled. The agonistic and/or antagonistic activity toward six transcription factors (namely the pregnane X receptor [PXR], liver X receptor [LXR], aryl hydrocarbon receptor [AhR], nuclear factor (erythroid-derived 2)-like 2 [Nrf2], PPARα and PPARγ) were modelled using DRAGON molecular descriptors and random forest models. For each MOA, four models were developed based on different strategies in feature selection and class balancing (i.e. majority class undersampling or balanced bagging) and integrated in a consensus model. External validation of the consensus models showed very good performance for all MOAs (accuracy between 0.74 and 0.96) but for agonistic activity on PPARγ (accuracy of 0.66) for compounds in the applicability domains. A second validation was carried out by screening 90 chemicals with *in vitro* steatosis data (six positives, 84 negatives) without experimental data for the molecular initiating events (MIE) endpoints considered and gave perfect sensitivity and AUC of 0.72. This exemplified how modelling the MIE can be successfully integrated in a virtual screening strategy for identifying chemicals causing hepatic steatosis.

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GENERAL DISCUSSION

Predicting DILI is a vital task, but is fraught with difficulties and complexities brought about from the data available to model, the number and varieties of phenotypic endpoint and mechanisms and the requirements of the end user. In the last decade, many QSAR and few rule-of-thumb models have been developed for the prediction of DILI with the majority of them focused on classification of compounds based on general hepatotoxicity annotation (Table 2). The good performance of models that have been developed is very encouraging, highlighting that machine learning methods are able to cope with complexities of the datasets, even though the data is inherently variable, limited in size and imbalanced. This is even more exciting considering that hepatotoxicity is an umbrella term for many different and complex phenotypes that are the integrated result of various mechanisms, and in spite of the paucity of phenotypically- and mechanistically-based large datasets. It is worth noting that only one regression model correlating to the severity of clinical outcome has been published so far⁷⁶. The same applies to multinomial classification modelling: only one three-level DILI classification model has been published¹⁰⁰. Nevertheless, as no golden standard for DILI annotation has been established, each annotation uses its own criteria and sources to label compounds¹⁰², leading to contradictory hepatotoxicity labelling of compounds by different authors, thus making the integration of multiple datasets a difficult endeavour^{59,121}. This stresses the requirement for sensitive biomarkers able to accurately differentiate medical symptoms of DILI. However, the downside of using more complex machine learning algorithms is that they lack transparency and accountability.

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Additionally, differences in molecular similarity among datasets 77,99,104,109 as well as their evaluation

with different metrics makes fair comparison between models a challenge¹³⁸. Among molecular descriptors, there seems to be a growing trend in using molecular fingerprints only, rather than relying on physicochemical or topological descriptors, although simple rules of thumbs have been devised from them. To date only one study has used graph-based molecular structural encoding, thus avoiding the molecular descriptor calculation and selection step, combined with deep learning algorithms⁵⁸. Some other studies have focused on matched molecular pairs – i.e. molecules that are structurally very similar – with opposing hepatotoxicity annotations^{30,42,45,47}.

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Standard physicochemical and topological descriptors, as well as substructure-based fingerprints in QSAR models (structural alerts excluded), are poor predictors of the reactivity of the molecules and its relationship to the metabolism and hence generally do not perform well to predict DILI. In addition, the development of prediction models able to correctly predict toxicity cliffs (i.e. where a very small change in the structure of a molecule can alter activity enormously) is a challenging field. Tackling toxicity cliffs both through better data compilation and more detailed structure evaluation would definitely help better understanding the mechanisms underlying DILI. Hybrid models integrating molecular descriptors with *in vitro* data, whether being transcriptomics⁴⁷, cell-imaging⁹⁷ or bioactivity data^{65,70,137}, have also been developed to enrich the information content and interpretability of the models but with rather limited predictive performance. Only a few models have included in vivo pharmacokinetic processes, such as absorption and metabolism inhibition of CYP450 proteins, the formation of GSH adducts and protein covalent-binding data^{48,139}. Additionally, models focused on the determination of MIE and MOA show very good performance and are of critical importance for better understanding of DILI mechanisms. Yet, it is striking that no ensemble read-across approach, combining systems biology network analysis for the prediction of molecular targets¹⁴⁰, MIE or MOA along with transcriptomics^{141,142}, cell-imaging and metabolomics, has been devised to this date. Such an approach, similar to the DILIsym¹⁴³ systems toxicology strategy, could address the limitations of OSAR¹⁴⁴ such as the modelling of chemical mixtures or inorganic compounds (e.g. cisplatin) as well as enhance models developed this far with the prediction of the exposure. Furthermore, computational structure-based mechanistic hypothesising is very limited by the lack of three-dimensional structures of proteins at stake. Additionally, since dose is an important predictor for DILI, the prediction of the toxicological point of departure¹⁴⁵ (POD) is challenge to be addressed. Finally, the most difficult challenge is to address interspecies variability, and the concordance between human and animal toxicity 30,100,146 that initiatives, such as the eTRANSAFE consortium, focus on.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

REFERENCES

689	(1)		724		N (0 H (1 2000 (7 (6) 1012 1015
690	(1)	Holt, M.; Ju, C. Drug-Induced Liver Injury. In <i>Annals of Internal Medicine</i> ; 2010; Vol. 137, pp 3–27.	725		Not? Hepatology 2008 , 47 (6), 1813–1815.
691		Internal Medicine; 2010; Vol. 137, pp 3–27. https://doi.org/10.1007/978-3-642-00663-0 1.		(11)	https://doi.org/10.1002/hep.22332. Kaplowitz, N. Idiosyncratic Drug Hepatotoxicity. <i>Nat. Rev.</i>
692	(2)	Siramshetty, V. B.; Nickel, J.; Omieczynski, C.; Gohlke, B. O.;	727	(11)	Drug Discov. 2005 , 4 (6), 489–499.
693	(2)	Drwal, M. N.; Preissner, R. WITHDRAWN - A Resource for	728		https://doi.org/10.1038/nrd1750.
694		Withdrawn and Discontinued Drugs. <i>Nucleic Acids Res.</i> 2016 ,	720	(12)	George, N.; Chen, M.; Yuen, N.; Hunt, C. M.; Suzuki, A.
695		44 (D1), D1080–D1086. https://doi.org/10.1093/nar/gkv1192.	730	(12)	Interplay of Gender, Age and Drug Properties on Reporting
696	(3)	Fung, M.; Thornton, A.; Mybeck, K.; wu, J. H. H.; Hornbuckle,	731		Frequency of Drug-Induced Liver Injury. <i>Regul. Toxicol.</i>
697	(3)	K.; Muniz, E. Evaluation of the Characteristics of Safety	732		Pharmacol. 2018.
698		Withdrawal of Prescription Drugs from Worldwide	700	(13)	Zhu, XW.; Li, SJ. In Silico Prediction of Drug-Induced Liver
699		Pharmaceutical Markets-1960 to 1999. Ther. Innov. Regul. Sci.	734	(13)	Injury Based on Adverse Drug Reaction Reports. <i>Toxicol. Sci.</i>
700		2001 , <i>35</i> (1), 293–317.	735		2017 , <i>158</i> (2), 391–400. https://doi.org/10.1093/toxsci/kfx099.
701		https://doi.org/10.1177/009286150103500134.	726	(14)	Mosedale, M.; Watkins, P. B. Drug-Induced Liver Injury:
702	(4)	Chen, M.; Vijay, V.; Shi, Q.; Liu, Z.; Fang, H.; Tong, W. FDA-	737	(14)	Advances in Mechanistic Understanding That Will Inform Risk
703	(4)	Approved Drug Labeling for the Study of Drug-Induced Liver	738		Management. Clin. Pharmacol. Ther. 2017, 101 (4), 469–480.
704		Injury. <i>Drug Discov. Today</i> 2011 , <i>16</i> (15–16), 697–703.	739		https://doi.org/10.1002/cpt.564.
705		https://doi.org/10.1016/j.drudis.2011.05.007.	7.40	(15)	Alempijevic, T.; Zec, S.; Milosavljevic, T. Drug-Induced Liver
706	(5)	Babai, S.; Auclert, L.; Le-Louët, H. Safety Data and Withdrawal	741	(13)	Injury: Do We Know Everything? World Journal of
707	(5)	of Hepatotoxic Drugs. <i>Therapie</i> 2018.	742		Hepatology. 2017. https://doi.org/10.4254/wjh.v9.i10.491.
708		https://doi.org/10.1016/j.therap.2018.02.004.	7.40	(16)	Fraser, K.; Bruckner, D. M.; Dordick, J. S. Advancing
709	(6)	van Tonder, J. J.; Steenkamp, V.; Gulumi, M. Pre-Clinical	744	()	Predictive Hepatotoxicity at the Intersection of Experimental, in
710	(-)	Assessment of the Potential Intrinsic Hepatotoxicity of	745		Silico, and Artificial Intelligence Technologies. Chem. Res.
711		Candidate Drugs. In New Insights into Toxicity and Drug	746		Toxicol. 2018, 31 (6), 412–430.
712		Testing; InTech, 2013. https://doi.org/10.5772/54792.	747		https://doi.org/10.1021/acs.chemrestox.8b00054.
713	(7)	Cheng, A.; Dixon, S. L. In Silico Models for the Prediction of	748 ((17)	Noureddin, N.; Kaplowitz, N. Overview of Mechanisms of
714		Dose-Dependent Human Hepatotoxicity. J. Comput. Aided.	749		Drug-Induced Liver Injury (DILI) and Key Challenges in DILI
715		Mol. Des. 2003 , 17 (12), 811–823.	750		Research; 2018; Vol. 1990, pp 3–18.
716		https://doi.org/10.1023/B:JCAM.0000021834.50768.c6.	751		https://doi.org/10.1007/978-1-4939-7677-5_1.
717	(8)	Devarbhavi, H. An Update on Drug-Induced Liver Injury. J.	752 ((18)	Vinken, M. Adverse Outcome Pathways and Drug-Induced
718		Clin. Exp. Hepatol. 2012, 2 (3), 247–259.	753		Liver Injury Testing. Chem. Res. Toxicol. 2015, 28 (7), 1391-
719		https://doi.org/10.1016/j.jceh.2012.05.002.	754		1397. https://doi.org/10.1021/acs.chemrestox.5b00208.
720	(9)	Zimmerman, H. J. Drug-Induced Liver Disease. Clin. Liver Dis.	755 ((19)	O'Connell, T. M.; Watkins, P. B. The Application of
721		2000 , 4 (1), 73–96, vi. https://doi.org/10.1016/S1089-	756		Metabonomics to Predict Drug-Induced Liver Injury. Clin.
722		3261(05)70097-0.	757		Pharmacol. Ther. 2010 , 88 (3), 394–399.
723	(10)	Senior, J. R. What Is Idiosyncratic Hepatotoxicity? What Is It	758		https://doi.org/10.1038/clpt.2010.151.

1	(20)	Przybylak, K. R.; Cronin, M. T. D. In Silico Models for Drug-
2	(20)	Induced Liver Injury Current Status. Expert Opin. Drug
3		Metab. Toxicol. 2012, 8 (2), 201–217.
4		https://doi.org/10.1517/17425255.2012.648613.
5	(21)	Chan, R.; Benet, L. Z. Evaluation of the Relevance of DILI
6	(21)	Predictive Hypotheses in Early Drug Development: Review of:
7		In Vitro Methodologies vs. BDDCS Classification; 2018; Vol.
8		7. https://doi.org/10.1039/c8tx00016f.
9	(22)	Kuijper, I. A.; Yang, H.; Van De Water, B.; Beltman, J. B.
10	()	Unraveling Cellular Pathways Contributing to Drug-Induced
11		Liver Injury by Dynamical Modeling. Expert Opin. Drug
12		Metab. Toxicol. 2017 , 13 (1), 5–17.
13		https://doi.org/10.1080/17425255.2017.1234607.
14	(23)	Bhattacharya, S.; Shoda, L. K. M.; Zhang, Q.; Woods, C. G.;
15		Howell, B. A.; Siler, S. Q.; Woodhead, J. L.; Yang, Y.;
16		McMullen, P.; Watkins, P. B.; et al. Modeling Drug- and
17		Chemical-Induced Hepatotoxicity with Systems Biology
18		Approaches. Front. Physiol. 2012, 3 DEC (December), 1-18.
19		https://doi.org/10.3389/fphys.2012.00462.
20	(24)	Roth, R. A.; Ganey, P. E. Intrinsic versus Idiosyncratic Drug-
21		Induced HepatotoxicityTwo Villains or One? J. Pharmacol.
22		Exp. Ther. 2010 , 332 (3), 692–697.
23		https://doi.org/10.1124/jpet.109.162651.
24	(25)	Corsini, A.; Ganey, P.; Ju, C.; Kaplowitz, N.; Pessayre, D.;
25		Roth, R.; Watkins, P. B.; Albassam, M.; Liu, B.; Stancic, S.; et
26		al. Current Challenges and Controversies in Drug-Induced
27		Liver Injury. Drug Saf. 2012, 35 (12), 1099–1117.
28		https://doi.org/10.2165/11632970-000000000-00000.
29 30	(26)	Ludwig, J.; Axelsen, R. Drug Effects on the Liver - An Updated
31		Tabular Compilation of Drugs and Drug-Related Hepatic Diseases. <i>Dig. Dis. Sci.</i> 1983 . <i>28</i> (7), 651–666.
32		g (.),
33	(27)	https://doi.org/10.1007/BF01299927. O'Brien, P. J.; Irwin, W.; Diaz, D.; Howard-Cofield, E.; Krejsa,
34	(27)	C. M.; Slaughter, M. R.; Gao, B.; Kaludercic, N.; Angeline, A.;
35		Bernardi, P.; et al. High Concordance of Drug-Induced Human
36		Hepatotoxicity with in Vitro Cytotoxicity Measured in a Novel
37		Cell-Based Model Using High Content Screening. Arch.
38		Toxicol. 2006, 80 (9), 580–604. https://doi.org/10.1007/s00204-
39		006-0091-3.
40	(28)	Suzuki, A.; Andrade, R. J.; Bjornsson, E.; Lucena, M. I.; Lee,
41		W. M.; Yuen, N. A.; Hunt, C. M.; Freston, J. W. Drugs
42		Associated with Hepatotoxicity and Their Reporting Frequency
43		of Liver Adverse Events in VigiBase TM . Drug Saf. 2010, 33 (6),
44		503-522. https://doi.org/10.2165/11535340-000000000-
45		00000.
46	(29)	Chen, M.; Suzuki, A.; Thakkar, S.; Yu, K.; Hu, C.; Tong, W.
47		DILIrank: The Largest Reference Drug List Ranked by the Risk
48		for Developing Drug-Induced Liver Injury in Humans. Drug
49		Discov. Today 2016, 21 (4), 648–653.
50		https://doi.org/10.1016/j.drudis.2016.02.015.
51	(30)	Fourches, D.; Barnes, J. C.; Day, N. C.; Bradley, P.; Reed, J. Z.;
52 52		Tropsha, A. Cheminformatics Analysis of Assertions Mined
53 54		from Literature That Describe Drug-Induced Liver Injury in
54 55		Different Species. Chem. Res. Toxicol. 2010, 23 (1), 171–183.

https://doi.org/10.1021/tx900326k.

2017,

Toxicology

Kotsampasakou, E.; Montanari, F.; Ecker, G. F. Predicting

Drug-Induced Liver Injury: The Importance of Data Curation.

389

(June).

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58

(31)

59 https://doi.org/10.1016/j.tox.2017.06.003 60 (32) Luo, G.; Shen, Y.; Yang, L.; Lu, A.; Xiang, Z. A Review of 61 Drug-Induced Liver Injury Databases. Arch. Toxicol. 2017, 91 62 (9), 3039-3049. https://doi.org/10.1007/s00204-017-2024-8. 63 (33)Cruz-Monteagudo, M.; Cordeiro, M. N. D. S.; Borges, F. 64 Computational Chemistry Approach for the Early Detection of 65 Drug-Induced Idiosyncratic Liver Toxicity. J. Comput. Chem. 66 2008, 29 (4), 533-549. https://doi.org/10.1002/jcc.20812. 67 (34) Zimmerman, H. . Hepatotoxicity: The Adverse Effects of Drugs 68 and Other Chemicals on the Liver., 2nd ed.; Lippincott 69 Williams and Wilkins: Philadelphia, 1999 70 (35)Guo, J. J.; Wigle, P. R.; Lammers, K.; Vu, O. Comparison of 71 Potentially Hepatotoxic Drugs among Major US Drug 72 Compendia. Res. Soc. Adm. Pharm. 2005, 1 (3), 460-479. 73 https://doi.org/10.1016/j.sapharm.2005.06.005. 74 (36) Richard, A. M.; Judson, R. S.; Houck, K. A.; Grulke, C. M.; 75 Volarath, P.; Thillainadarajah, I.; Yang, C.; Rathman, J.; 76 Martin, M. T.; Wambaugh, J. F.; et al. ToxCast Chemical 77 Landscape: Paving the Road to 21st Century Toxicology. Chem. 78 Toxicol. 2016, 29 (8), 79 https://doi.org/10.1021/acs.chemrestox.6b00135. 80 (37)EPA's National Center for Computational Toxicology. ToxCast 81 Database (invitroDB) 82 https://epa.figshare.com/articles/ToxCast_Database_invitroDB 83 /6062623. https://doi.org/10.23645/epacomptox.6062623.v2. 84 (38) Dix, D. J.; Houck, K. A.; Martin, M. T.; Richard, A. M.; Setzer, 85 R. W.; Kavlock, R. J. The ToxCast Program for Prioritizing 86 Toxicity Testing of Environmental Chemicals. Toxicol. Sci. 87 2007, 95 (1), 5-12. https://doi.org/10.1093/toxsci/kfl103. 88 (39)Xu, J. J.; Henstock, P. V.; Dunn, M. C.; Smith, A. R.; Chabot, 89 J. R.; de Graaf, D. Cellular Imaging Predictions of Clinical 90 Drug-Induced Liver Injury. Toxicol. Sci. 2008, 105 (1), 97-105. 91 https://doi.org/10.1093/toxsci/kfn109. 92 (40)Ekins, S.; Williams, A. J.; Xu, J. J. A Predictive Ligand-Based 93 Bayesian Model for Human Drug-Induced Liver Injury. Drug 94 Metab. Dispos. 2010. 38 (12), 2302-2308. 95 https://doi.org/10.1124/dmd.110.035113. 96 (41) Greene, N.; Fisk, L.; Naven, R. T.; Note, R. R.; Patel, M. L.; 97 Pelletier, D. J. Developing Structure-Activity Relationships for 98 the Prediction of Hepatotoxicity. Chem. Res. Toxicol. 2010, 23 99 (7), 1215-1222. https://doi.org/10.1021/tx1000865. 100 (42) Rodgers, A. D.: Zhu, H.: Fourches, D.: Rusyn, I.: Tropsha, A. 101 Modeling Liver-Related Adverse Effects of Drugs Using 102 Neighbor Quantitative Structure-Activity 103 Relationship Method. Chem. Res. Toxicol. 2010, 23 (4), 724-104 732. https://doi.org/10.1021/tx900451r. 105 (43) Kuhn, M.; Campillos, M.; Letunic, I.; Jensen, L. J.; Bork, P. A 106 Side Effect Resource to Capture Phenotypic Effects of Drugs. 107 Mol Syst. Riol 2010 (343).6 108 https://doi.org/10.1038/msb.2009.98. 109 (44) Kuhn, M.; Letunic, I.; Jensen, L. J.; Bork, P. The SIDER 110 Database of Drugs and Side Effects. Nucleic Acids Res. 2016. 111 44 (D1), D1075–D1079. https://doi.org/10.1093/nar/gkv1075. 112 (45) Liew, C. Y.; Lim, Y. C.; Yap, C. W. Mixed Learning 113 Algorithms and Features Ensemble in Hepatotoxicity 114 Prediction. J. Comput. Aided. Mol. Des. 2011, 25 (9), 855-871.

https://doi.org/10.1007/s10822-011-9468-3.

Liu, Z.; Shi, Q.; Ding, D.; Kelly, R.; Fang, H.; Tong, W.

115

116

(46)

139-145.

1		Translating Clinical Findings into Knowledge in Drug Safety
2		Evaluation - Drug Induced Liver Injury Prediction System
3		(DILIps). PLoS Comput. Biol. 2011.
4	(47)	Low, Y.; Uehara, T.; Minowa, Y.; Yamada, H.; Ohno, Y.;
5	(,	Urushidani, T.; Sedykh, A.; Muratov, E.; Kuz'min, V.;
6		Fourches, D.; et al. Predicting Drug-Induced Hepatotoxicity
7		
8		Using QSAR and Toxicogenomics Approaches. Chem. Res.
9		Toxicol. 2011, 24 (8), 1251–1262.
		https://doi.org/10.1021/tx200148a.
10	(48)	Sakatis, M. Z.; Reese, M. J.; Harrell, A. W.; Taylor, M. A.;
11		Baines, I. A.; Chen, L.; Bloomer, J. C.; Yang, E. Y.; Ellens, H.
12		M.; Ambroso, J. L.; et al. Preclinical Strategy to Reduce Clinical
13		Hepatotoxicity Using in Vitro Bioactivation Data for >200
14		Compounds. Chem. Res. Toxicol. 2012, 25 (10), 2067-2082.
15		https://doi.org/10.1021/tx300075j.
16	(49)	Chen, M.; Zhang, J.; Wang, Y.; Liu, Z.; Kelly, R.; Zhou, G.;
17		Fang, H.; Borlak, J.; Tong, W. The Liver Toxicity Knowledge
18		Base: A Systems Approach to a Complex End Point. Clin.
19		Pharmacol. Ther. 2013.
20	(50)	
	(50)	Hoofnagle, J. H. LiverTox: A Website on Drug-Induced Liver
21		Injury. Drug-Induced Liver Dis. 2013, 725–732.
22		https://doi.org/10.1016/B978-0-12-387817-5.00040-6.
23	(51)	Zhu, X.; Kruhlak, N. L. Construction and Analysis of a Human
24		Hepatotoxicity Database Suitable for QSAR Modeling Using
25		Post-Market Safety Data. Toxicology 2014, 321 (1), 62-72.
26		https://doi.org/10.1016/j.tox.2014.03.009.
27	(52)	Leeson, P. D. Impact of Physicochemical Properties on Dose
28		and Hepatotoxicity of Oral Drugs. Chem. Res. Toxicol. 2018, 31
29		(6), 494–505. https://doi.org/10.1021/acs.chemrestox.8b00044.
30	(53)	Mulliner, D.; Schmidt, F.; Stolte, M.; Spirkl, H. P.; Czich, A.;
31	(55)	Amberg, A. Computational Models for Human and Animal
32		Hepatotoxicity with a Global Application Scope. <i>Chem. Res.</i>
33		
34		Toxicol. 2016, 29 (5), 757–767.
		https://doi.org/10.1021/acs.chemrestox.5b00465.
35	(54)	Huang, R. A Quantitative High-Throughput Screening Data
36		Analysis Pipeline for Activity Profiling; Zhu, H., Xia, M., Eds.;
37		Methods in Molecular Biology; Springer New York: New York,
38		NY, 2016; Vol. 1473, pp 111–122. https://doi.org/10.1007/978-
39		1-4939-6346-1_12.
40	(55)	Huang, R.; Xia, M.; Sakamuru, S.; Zhao, J.; Shahane, S. A.;
41		Attene-Ramos, M.; Zhao, T.; Austin, C. P.; Simeonov, A.
42		Modelling the Tox21 10 K Chemical Profiles for in Vivo
43		Toxicity Prediction and Mechanism Characterization. Nat.
44		Commun. 2016. https://doi.org/10.1038/ncomms10425.
45	(56)	Ai, H.; Chen, W.; Zhang, L.; Huang, L.; Yin, Z.; Hu, H.; Zhao,
46	(30)	Q.; Zhao, J.; Liu, H. Predicting Drug-Induced Liver Injury
47		
48		Using Ensemble Learning Methods and Molecular Fingerprints.
		<i>Toxicol.</i> Sci. 2018 , No. August, 1–8.
49		https://doi.org/10.1093/toxsci/kfy121.
50	(57)	Copple, I. M.; den Hollander, W.; Callegaro, G.; Mutter, F. E.;
51		Maggs, J. L.; Schofield, A. L.; Rainbow, L.; Fang, Y.;
52		Sutherland, J. J.; Ellis, E. C.; et al. Characterisation of the NRF2
53		Transcriptional Network and Its Response to Chemical Insult in
54		Primary Human Hepatocytes: Implications for Prediction of
55		Drug-Induced Liver Injury. Arch. Toxicol. 2019, 93 (2), 385–
56		399. https://doi.org/10.1007/s00204-018-2354-1.
57	(58)	Xu, Y.; Dai, Z.; Chen, F.; Gao, S.; Pei, J.; Lai, L. Deep Learning
58	(50)	
50		for Drug-Induced Liver Injury. J. Chem. Inf. Model. 2015, 55

- 59 (10), 2085-2093. https://doi.org/10.1021/acs.jcim.5b00238. 60 (59)Kotsampasakou, E.; Montanari, F.; Ecker, G. F. Predicting 61 Drug-Induced Liver Injury: The Importance of Data Curation. 62 Toxicology 2017, 389. 139-145. 63 https://doi.org/10.1016/j.tox.2017.06.003. 64 (60)Steger-Hartmann, T.; Pognan, F.; Sanz, F.; Diaz, C. A. In Silico 65 Prediction of in Vivo Toxicities (ETox)-The Innovative 66 Medicines Initiative Approach. Toxicol. Lett. 2009, 189 (2009), 67 S258. https://doi.org/10.1016/j.toxlet.2009.06.374. 68
- (61)Cases, M.; Briggs, K.; Steger-Hartmann, T.; Pognan, F.; Marc, 69 P.; Kleinöder, T.; Schwab, C. H.; Pastor, M.; Wichard, J.; Sanz, 70 F. The ETOX Data-Sharing Project to Advance in Silico Drug-71 Induced Toxicity Prediction. Int. J. Mol. Sci. 2014, 15 (11), 72 21136-21154. https://doi.org/10.3390/ijms151121136.
- 73 (62) Pognan, F. Detection, Elimination, Mitigation, and Prediction 74 of Drug-Induced Liver Injury in Drug Discovery. In Drug-75 Induced Liver Toxicity, Methods in Pharmacology and 76 Toxicology; Chen, M., Will, Y., Eds.; Springer, 2018; pp 21-77 43. https://doi.org/10.1007/978-1-4939-7677-5 2.
- 78 (63)Sanz, F.; Pognan, F.; Steger-Hartmann, T.; Díaz, C. Legacy 79 Data Sharing to Improve Drug Safety Assessment: The ETOX 80 Project. Nature Reviews Drug Discovery. October 13, 2017, pp 81 811-812. https://doi.org/10.1038/nrd.2017.177.
- 82 (64)Jiang, L.; He, Y.; Zhang, Y. Prediction of Hepatotoxicity of 83 Traditional Chinese Medicine Compounds by Support Vector 84 Machine Approach. Int. Conf. Syst. Biol. ISB 2014, No. 85 81173522, 27–30. https://doi.org/10.1109/ISB.2014.6990426.
- 86 (65) Muller, C.; Pekthong, D.; Alexandre, E.; Marcou, G.; Horvath, 87 D.; Richert, L.; Varnek, A. Prediction of Drug Induced Liver 88 Injury Using Molecular and Biological Descriptors. Comb. 89 Chem. High Throughput Screen. 2015, 18 (3), 315-322. 90 https://doi.org/10.2174/1386207318666150305144650.
- 91 (66)Zhao, P.; Liu, B.; Wang, C. Hepatotoxicity Evaluation of 92 Traditional Chinese Medicines Using a Computational 93 Molecular Model. Clin. Toxicol. 2017, 55 (9), 996-1000. 94 https://doi.org/10.1080/15563650.2017.1333123.
- 95 (67) Liu, L.; Fu, L.; Zhang, J. W.; Wei, H.; Ye, W. L.; Deng, Z. K.; 96 Zhang, L.; Cheng, Y.; Ouyang, D.; Cao, Q.; et al. Three-Level 97 Hepatotoxicity Prediction System Based on Adverse Hepatic 98 Effects. Mol. Pharm. 2019, 16 (1), 393-408. 99 https://doi.org/10.1021/acs.molpharmaceut.8b01048.
- 100 (68)Huang, S. H.; Tung, C. W.; Fülöp, F.; Li, J. H. Developing a 101 QSAR Model for Hepatotoxicity Screening of the Active 102 Compounds in Traditional Chinese Medicines. Food Chem. 103 Toxicol. 2015, 78, 71-77. 104 https://doi.org/10.1016/j.fct.2015.01.020.
- 105 Lu, Y.; Liu, L.; Lu, D.; Cai, Y.; Zheng, M.; Luo, X.; Jiang, H.; 106 Chen, K. Predicting Hepatotoxicity of Drug Metabolites Via an 107 Ensemble Approach Based on Support Vector Machine. Comb. 108 Chem. High Throughput Screen. 2017, 20 (10), 839-849. 109 https://doi.org/10.2174/1386207320666171121113255.
- 110 (70)Liu, J.: Mansouri, K.: Judson, R. S.: Martin, M. T.: Hong, H.: 111 Chen, M.; Xu, X.; Thomas, R. S.; Shah, I. Predicting 112 Hepatotoxicity Using ToxCast in Vitro Bioactivity and 113 Chemical Structure. Chem. Res. Toxicol. 2015, 28 (4), 738-751. 114 https://doi.org/10.1021/tx500501h. 115
 - (71) Wu, Q.; Cai, C.; Guo, P.; Chen, M.; Wu, X.; Zhou, J.; Luo, Y.; Zou, Y.; Liu, A.; Wang, Q.; et al. In Silico Identification and

1		Mechanism Exploration of Hepatotoxic Ingredients in
2		Traditional Chinese Medicine. Front. Pharmacol. 2019, 10
3		(May), 1–15. https://doi.org/10.3389/fphar.2019.00458.
4	(72)	Takeshita, J. ichi; Nakayama, H.; Kitsunai, Y.; Tanabe, M.; Oki,
5		H.; Sasaki, T.; Yoshinari, K. Discriminative Models Using
6		Molecular Descriptors for Predicting Increased Serum ALT
7		Levels in Repeated-Dose Toxicity Studies of Rats. Comput.
8		Toxicol. 2018 , 6, 64–70.
9		· · · · · · · · · · · · · · · · · · ·
10	(52)	https://doi.org/10.1016/j.comtox.2017.05.002.
11	(73)	Toropova, A. P.; Toropov, A. A. CORAL: Binary
		Classifications (Active/Inactive) for Drug-Induced Liver Injury.
12		Toxicol. Lett. 2017, 268, 51–57.
13		https://doi.org/10.1016/j.toxlet.2017.01.011.
14	(74)	Zhang, H.; Ding, L.; Zou, Y.; Hu, S. Q.; Huang, H. G.; Kong,
15		W. B.; Zhang, J. Predicting Drug-Induced Liver Injury in
16		Human with Naïve Bayes Classifier Approach. J. Comput.
17		Aided. Mol. Des. 2016, 30 (10), 889–898.
18		https://doi.org/10.1007/s10822-016-9972-6.
19	(75)	Schöning, V.; Krähenbühl, S.; Drewe, J. The Hepatotoxic
20		Potential of Protein Kinase Inhibitors Predicted with Random
21		Forest and Artificial Neural Networks. Toxicol. Lett. 2018, 299
22		(October), 145–148.
23		https://doi.org/10.1016/j.toxlet.2018.10.009.
24	(76)	Chen, M.; Borlak, J.; Tong, W. A Model to Predict Severity of
25	(70)	Drug-Induced Liver Injury in Humans. <i>Hepatology</i> 2016 , 64
26		(3), 931–940. https://doi.org/10.1002/hep.28678.
27	(77)	
28	(77)	Hammann, F.; Schöning, V.; Drewe, J. Prediction of Clinically
29		Relevant Drug-Induced Liver Injury from Structure Using
		Machine Learning. J. Appl. Toxicol. 2019, 39 (3), 412–419.
30		https://doi.org/10.1002/jat.3741.
31	(78)	Chen, M.; Borlak, J.; Tong, W. High Lipophilicity and High
32		Daily Dose of Oral Medications Are Associated with
33		Significant Risk for Drug-Induced Liver Injury. Hepatology
34		2013 , 58 (1), 388–396. https://doi.org/10.1002/hep.26208.
35	(79)	Chawla, N. V.; Bowyer, K. W.; Hall, L. O.; Kegelmeyer, W. P.
	(17)	
36	(12)	SMOTE: Synthetic Minority Over-Sampling Technique. J .
36	(12)	SMOTE: Synthetic Minority Over-Sampling Technique. <i>J. Artif. Intell. Res.</i> 2002 , <i>16</i> , 321–357.
	(12)	
37	(80)	Artif. Intell. Res. 2002, 16, 321–357.
37 38	,	Artif. Intell. Res. 2002 , 16, 321–357. https://doi.org/10.1613/jair.953.
37 38 39	,	Artif. Intell. Res. 2002, 16, 321–357. https://doi.org/10.1613/jair.953. Galar, M.; Fern, A.; Barrenechea, E.; Bustince, H. A Review of
37 38 39 40	,	Artif. Intell. Res. 2002, 16, 321–357. https://doi.org/10.1613/jair.953. Galar, M.; Fern, A.; Barrenechea, E.; Bustince, H. A Review of Ensembles for the Class Imbalance Problem. 2012, 42 (4), 463–
37 38 39 40 41	(80)	Artif. Intell. Res. 2002, 16, 321–357. https://doi.org/10.1613/jair.953. Galar, M.; Fern, A.; Barrenechea, E.; Bustince, H. A Review of Ensembles for the Class Imbalance Problem. 2012, 42 (4), 463–484.
37 38 39 40 41 42	(80)	Artif. Intell. Res. 2002, 16, 321–357. https://doi.org/10.1613/jair.953. Galar, M.; Fern, A.; Barrenechea, E.; Bustince, H. A Review of Ensembles for the Class Imbalance Problem. 2012, 42 (4), 463–484. Kotsiantis, S.; Kanellopoulos, D.; Pintelas, P. Handling Imbalanced Datasets: A Review. Science (80). 2006, 30 (1),
37 38 39 40 41 42 43	(80)	Artif. Intell. Res. 2002, 16, 321–357. https://doi.org/10.1613/jair.953. Galar, M.; Fern, A.; Barrenechea, E.; Bustince, H. A Review of Ensembles for the Class Imbalance Problem. 2012, 42 (4), 463–484. Kotsiantis, S.; Kanellopoulos, D.; Pintelas, P. Handling
37 38 39 40 41 42 43 44	(80)	Artif. Intell. Res. 2002, 16, 321–357. https://doi.org/10.1613/jair.953. Galar, M.; Fern, A.; Barrenechea, E.; Bustince, H. A Review of Ensembles for the Class Imbalance Problem. 2012, 42 (4), 463–484. Kotsiantis, S.; Kanellopoulos, D.; Pintelas, P. Handling Imbalanced Datasets: A Review. Science (80). 2006, 30 (1), 25–36. https://doi.org/10.1007/978-0-387-09823-4_45. Kotsampasakou, E.; Brenner, S.; Jäger, W.; Ecker, G. F.
37 38 39 40 41 42 43 44 45 46	(80)	Artif. Intell. Res. 2002, 16, 321–357. https://doi.org/10.1613/jair.953. Galar, M.; Fern, A.; Barrenechea, E.; Bustince, H. A Review of Ensembles for the Class Imbalance Problem. 2012, 42 (4), 463–484. Kotsiantis, S.; Kanellopoulos, D.; Pintelas, P. Handling Imbalanced Datasets: A Review. Science (80). 2006, 30 (1), 25–36. https://doi.org/10.1007/978-0-387-09823-4_45. Kotsampasakou, E.; Brenner, S.; Jäger, W.; Ecker, G. F. Identification of Novel Inhibitors of Organic Anion
37 38 39 40 41 42 43 44 45 46 47	(80)	Artif. Intell. Res. 2002, 16, 321–357. https://doi.org/10.1613/jair.953. Galar, M.; Fern, A.; Barrenechea, E.; Bustince, H. A Review of Ensembles for the Class Imbalance Problem. 2012, 42 (4), 463–484. Kotsiantis, S.; Kanellopoulos, D.; Pintelas, P. Handling Imbalanced Datasets: A Review. Science (80). 2006, 30 (1), 25–36. https://doi.org/10.1007/978-0-387-09823-4_45. Kotsampasakou, E.; Brenner, S.; Jäger, W.; Ecker, G. F. Identification of Novel Inhibitors of Organic Anion Transporting Polypeptides 1B1 and 1B3 (OATP1B1 and
37 38 39 40 41 42 43 44 45 46 47	(80)	Artif. Intell. Res. 2002, 16, 321–357. https://doi.org/10.1613/jair.953. Galar, M.; Fern, A.; Barrenechea, E.; Bustince, H. A Review of Ensembles for the Class Imbalance Problem. 2012, 42 (4), 463–484. Kotsiantis, S.; Kanellopoulos, D.; Pintelas, P. Handling Imbalanced Datasets: A Review. Science (80). 2006, 30 (1), 25–36. https://doi.org/10.1007/978-0-387-09823-4_45. Kotsampasakou, E.; Brenner, S.; Jäger, W.; Ecker, G. F. Identification of Novel Inhibitors of Organic Anion Transporting Polypeptides 1B1 and 1B3 (OATP1B1 and OATP1B3) Using a Consensus Vote of Six Classification
37 38 39 40 41 42 43 44 45 46 47 48 49	(80)	Artif. Intell. Res. 2002, 16, 321–357. https://doi.org/10.1613/jair.953. Galar, M.; Fern, A.; Barrenechea, E.; Bustince, H. A Review of Ensembles for the Class Imbalance Problem. 2012, 42 (4), 463–484. Kotsiantis, S.; Kanellopoulos, D.; Pintelas, P. Handling Imbalanced Datasets: A Review. Science (80). 2006, 30 (1), 25–36. https://doi.org/10.1007/978-0-387-09823-4_45. Kotsampasakou, E.; Brenner, S.; Jäger, W.; Ecker, G. F. Identification of Novel Inhibitors of Organic Anion Transporting Polypeptides 1B1 and 1B3 (OATP1B1 and OATP1B3) Using a Consensus Vote of Six Classification Models. Mol. Pharm. 2015, 12 (12), 4395–4404.
37 38 39 40 41 42 43 44 45 46 47 48 49 50	(80) (81) (82)	Artif. Intell. Res. 2002, 16, 321–357. https://doi.org/10.1613/jair.953. Galar, M.; Fern, A.; Barrenechea, E.; Bustince, H. A Review of Ensembles for the Class Imbalance Problem. 2012, 42 (4), 463–484. Kotsiantis, S.; Kanellopoulos, D.; Pintelas, P. Handling Imbalanced Datasets: A Review. Science (80). 2006, 30 (1), 25–36. https://doi.org/10.1007/978-0-387-09823-4_45. Kotsampasakou, E.; Brenner, S.; Jäger, W.; Ecker, G. F. Identification of Novel Inhibitors of Organic Anion Transporting Polypeptides 1B1 and 1B3 (OATP1B1 and OATP1B3) Using a Consensus Vote of Six Classification Models. Mol. Pharm. 2015, 12 (12), 4395–4404. https://doi.org/10.1021/acs.molpharmaceut.5b00583.
37 38 39 40 41 42 43 44 45 46 47 48 49 50 51	(80)	Artif. Intell. Res. 2002, 16, 321–357. https://doi.org/10.1613/jair.953. Galar, M.; Fern, A.; Barrenechea, E.; Bustince, H. A Review of Ensembles for the Class Imbalance Problem. 2012, 42 (4), 463–484. Kotsiantis, S.; Kanellopoulos, D.; Pintelas, P. Handling Imbalanced Datasets: A Review. Science (80). 2006, 30 (1), 25–36. https://doi.org/10.1007/978-0-387-09823-4_45. Kotsampasakou, E.; Brenner, S.; Jäger, W.; Ecker, G. F. Identification of Novel Inhibitors of Organic Anion Transporting Polypeptides 1B1 and 1B3 (OATP1B1 and OATP1B3) Using a Consensus Vote of Six Classification Models. Mol. Pharm. 2015, 12 (12), 4395–4404. https://doi.org/10.1021/acs.molpharmaceut.5b00583. Wang, H.; Liu, R.; Schyman, P.; Wallqvist, A. Deep Neural
37 38 39 40 41 42 43 44 45 46 47 48 49 50 51 52	(80) (81) (82)	Artif. Intell. Res. 2002, 16, 321–357. https://doi.org/10.1613/jair.953. Galar, M.; Fern, A.; Barrenechea, E.; Bustince, H. A Review of Ensembles for the Class Imbalance Problem. 2012, 42 (4), 463–484. Kotsiantis, S.; Kanellopoulos, D.; Pintelas, P. Handling Imbalanced Datasets: A Review. Science (80). 2006, 30 (1), 25–36. https://doi.org/10.1007/978-0-387-09823-4_45. Kotsampasakou, E.; Brenner, S.; Jäger, W.; Ecker, G. F. Identification of Novel Inhibitors of Organic Anion Transporting Polypeptides 1B1 and 1B3 (OATP1B1 and OATP1B3) Using a Consensus Vote of Six Classification Models. Mol. Pharm. 2015, 12 (12), 4395–4404. https://doi.org/10.1021/acs.molpharmaceut.5b00583. Wang, H.; Liu, R.; Schyman, P.; Wallqvist, A. Deep Neural Network Models for Predicting Chemically Induced Liver
37 38 39 40 41 42 43 44 45 46 47 48 49 50 51 52 53	(80) (81) (82)	Artif. Intell. Res. 2002, 16, 321–357. https://doi.org/10.1613/jair.953. Galar, M.; Fern, A.; Barrenechea, E.; Bustince, H. A Review of Ensembles for the Class Imbalance Problem. 2012, 42 (4), 463–484. Kotsiantis, S.; Kanellopoulos, D.; Pintelas, P. Handling Imbalanced Datasets: A Review. Science (80). 2006, 30 (1), 25–36. https://doi.org/10.1007/978-0-387-09823-4_45. Kotsampasakou, E.; Brenner, S.; Jäger, W.; Ecker, G. F. Identification of Novel Inhibitors of Organic Anion Transporting Polypeptides 1B1 and 1B3 (OATP1B1 and OATP1B3) Using a Consensus Vote of Six Classification Models. Mol. Pharm. 2015, 12 (12), 4395–4404. https://doi.org/10.1021/acs.molpharmaceut.5b00583. Wang, H.; Liu, R.; Schyman, P.; Wallqvist, A. Deep Neural Network Models for Predicting Chemically Induced Liver Toxicity Endpoints from Transcriptomic Responses. Front.
37 38 39 40 41 42 43 44 45 46 47 48 49 50 51 52 53 54	(80) (81) (82)	Artif. Intell. Res. 2002, 16, 321–357. https://doi.org/10.1613/jair.953. Galar, M.; Fern, A.; Barrenechea, E.; Bustince, H. A Review of Ensembles for the Class Imbalance Problem. 2012, 42 (4), 463–484. Kotsiantis, S.; Kanellopoulos, D.; Pintelas, P. Handling Imbalanced Datasets: A Review. Science (80). 2006, 30 (1), 25–36. https://doi.org/10.1007/978-0-387-09823-4_45. Kotsampasakou, E.; Brenner, S.; Jäger, W.; Ecker, G. F. Identification of Novel Inhibitors of Organic Anion Transporting Polypeptides 1B1 and 1B3 (OATP1B1 and OATP1B3) Using a Consensus Vote of Six Classification Models. Mol. Pharm. 2015, 12 (12), 4395–4404. https://doi.org/10.1021/acs.molpharmaceut.5b00583. Wang, H.; Liu, R.; Schyman, P.; Wallqvist, A. Deep Neural Network Models for Predicting Chemically Induced Liver Toxicity Endpoints from Transcriptomic Responses. Front. Pharmacol. 2019, 10 (FEB), 1–12.
37 38 39 40 41 42 43 44 45 46 47 48 49 50 51 52 53 54 55	(80) (81) (82)	Artif. Intell. Res. 2002, 16, 321–357. https://doi.org/10.1613/jair.953. Galar, M.; Fern, A.; Barrenechea, E.; Bustince, H. A Review of Ensembles for the Class Imbalance Problem. 2012, 42 (4), 463–484. Kotsiantis, S.; Kanellopoulos, D.; Pintelas, P. Handling Imbalanced Datasets: A Review. Science (80). 2006, 30 (1), 25–36. https://doi.org/10.1007/978-0-387-09823-4_45. Kotsampasakou, E.; Brenner, S.; Jäger, W.; Ecker, G. F. Identification of Novel Inhibitors of Organic Anion Transporting Polypeptides 1B1 and 1B3 (OATP1B1 and OATP1B3) Using a Consensus Vote of Six Classification Models. Mol. Pharm. 2015, 12 (12), 4395–4404. https://doi.org/10.1021/acs.molpharmaceut.5b00583. Wang, H.; Liu, R.; Schyman, P.; Wallqvist, A. Deep Neural Network Models for Predicting Chemically Induced Liver Toxicity Endpoints from Transcriptomic Responses. Front. Pharmacol. 2019, 10 (FEB), 1–12. https://doi.org/10.3389/fphar.2019.00042.
37 38 39 40 41 42 43 44 45 46 47 48 49 50 51 52 53 54 55 56	(80) (81) (82)	Artif. Intell. Res. 2002, 16, 321–357. https://doi.org/10.1613/jair.953. Galar, M.; Fern, A.; Barrenechea, E.; Bustince, H. A Review of Ensembles for the Class Imbalance Problem. 2012, 42 (4), 463–484. Kotsiantis, S.; Kanellopoulos, D.; Pintelas, P. Handling Imbalanced Datasets: A Review. Science (80). 2006, 30 (1), 25–36. https://doi.org/10.1007/978-0-387-09823-4_45. Kotsampasakou, E.; Brenner, S.; Jäger, W.; Ecker, G. F. Identification of Novel Inhibitors of Organic Anion Transporting Polypeptides 1B1 and 1B3 (OATP1B1 and OATP1B3) Using a Consensus Vote of Six Classification Models. Mol. Pharm. 2015, 12 (12), 4395–4404. https://doi.org/10.1021/acs.molpharmaceut.5b00583. Wang, H.; Liu, R.; Schyman, P.; Wallqvist, A. Deep Neural Network Models for Predicting Chemically Induced Liver Toxicity Endpoints from Transcriptomic Responses. Front. Pharmacol. 2019, 10 (FEB), 1–12. https://doi.org/10.3389/fphar.2019.00042. Kotsampasakou, E.; Ecker, G. F. Predicting Drug-Induced
37 38 39 40 41 42 43 44 45 46 47 48 49 50 51 52 53 54 55	(80) (81) (82)	Artif. Intell. Res. 2002, 16, 321–357. https://doi.org/10.1613/jair.953. Galar, M.; Fern, A.; Barrenechea, E.; Bustince, H. A Review of Ensembles for the Class Imbalance Problem. 2012, 42 (4), 463–484. Kotsiantis, S.; Kanellopoulos, D.; Pintelas, P. Handling Imbalanced Datasets: A Review. Science (80). 2006, 30 (1), 25–36. https://doi.org/10.1007/978-0-387-09823-4_45. Kotsampasakou, E.; Brenner, S.; Jäger, W.; Ecker, G. F. Identification of Novel Inhibitors of Organic Anion Transporting Polypeptides 1B1 and 1B3 (OATP1B1 and OATP1B3) Using a Consensus Vote of Six Classification Models. Mol. Pharm. 2015, 12 (12), 4395–4404. https://doi.org/10.1021/acs.molpharmaceut.5b00583. Wang, H.; Liu, R.; Schyman, P.; Wallqvist, A. Deep Neural Network Models for Predicting Chemically Induced Liver Toxicity Endpoints from Transcriptomic Responses. Front. Pharmacol. 2019, 10 (FEB), 1–12. https://doi.org/10.3389/fphar.2019.00042.

```
59
                  608-615. https://doi.org/10.1021/acs.jcim.6b00518.
 60
       (85)
                  Jain, S.; Kotsampasakou, E.; Ecker, G. F. Comparing the
 61
                  Performance of Meta-Classifiers-a Case Study on Selected
 62
                  Imbalanced Data Sets Relevant for Prediction of Liver Toxicity.
 63
                  J. Comput. Aided. Mol. Des. 2018, 32 (5), 583-590.
 64
                  https://doi.org/10.1007/s10822-018-0116-z.
 65
       (86)
                  Breiman, L. Bagging Predictions. Mach. Learn. 1996, 24 (2),
 66
 67
       (87)
                  Domingos, P. MetaCost: A General Method for Making
 68
                  Classifiers Cost-Sensitive. In Proceedings of the fifth ACM
 69
                  SIGKDD international conference on Knowledge discovery and
 70
                  data mining - KDD '99; ACM Press: New York, New York,
 71
                  USA.
                             1999
                                        Vol.
                                                 25.
                                                                  155-164
 72
                  https://doi.org/10.1145/312129.312220.
 73
        (88)
                  Egan, W. J.; Zlokarnik, G.; Grootenhuis, P. D. J. In Silico
 74
                  Prediction of Drug Safety: Despite Progress There Is Abundant
 75
                  Room for Improvement. Drug Discov. Today Technol. 2004, 1
 76
                  (4), 381-387. https://doi.org/10.1016/j.ddtec.2004.11.002.
 77
        (89)
                  Hewitt, M.; Enoch, S. J.; Madden, J. C.; Przybylak, K. R.;
 78
                  Cronin, M. T. D. Hepatotoxicity: A Scheme for Generating
 79
                  Chemical Categories for Read-across, Structural Alerts and
 80
                  Insights into Mechanism(s) of Action. Crit. Rev. Toxicol. 2013.
 81
                  43
                                                                  537-558
                                          (7),
 82
                  https://doi.org/10.3109/10408444.2013.811215.
 83
        (90)
                  Pizzo, F.; Lombardo, A.; Manganaro, A.; Benfenati, E. A New
 84
                  Structure-Activity Relationship (SAR) Model for Predicting
 85
                  Drug-Induced Liver Injury, Based on Statistical and Expert-
 86
                  Based Structural Alerts. Front. Pharmacol. 2016, 7 (NOV), 1-
 87
                  15. https://doi.org/10.3389/fphar.2016.00442.
 88
       (91)
                  Liu, R.; Yu, X.; Wallqvist, A. Data-Driven Identification of
 89
                  Structural Alerts for Mitigating the Risk of Drug-Induced
 90
                  Human Liver Injuries. J. Cheminform. 2015, 7 (1), 4.
 91
                  https://doi.org/10.1186/s13321-015-0053-y.
 92
        (92)
                  Clark, R. D.; Wolohan, P. R. N.; Hodgkin, E. E.; Kelly, J. H.;
 93
                  Sussman, N. L. Modelling in Vitro Hepatotoxicity Using
 94
                  Molecular Interaction Fields and SIMCA. J. Mol. Graph.
 95
                                2004.
                  Model.
                                             22
                                                                  487-497.
                                                       (6),
 96
                  https://doi.org/10.1016/j.jmgm.2004.03.009.
 97
       (93)
                  Matthews, E. J.; Ursem, C. J.; Kruhlak, N. L.; Benz, R. D.;
 98
                  Sabaté, D. A.; Yang, C.; Klopman, G.; Contrera, J. F.
 99
                  Identification of Structure-Activity Relationships for Adverse
100
                  Effects of Pharmaceuticals in Humans: Part B. Use of (Q)SAR
101
                  Systems for Early Detection of Drug-Induced Hepatobiliary and
102
                  Urinary Tract Toxicities. Regul. Toxicol. Pharmacol. 2009, 54
103
                  (1), 23-42. https://doi.org/10.1016/j.yrtph.2009.01.009.
104
       (94)
                  Liu, Z.; Shi, Q.; Ding, D.; Kelly, R.; Fang, H.; Tong, W.
105
                  Translating Clinical Findings into Knowledge in Drug Safety
106
                  Evaluation--Drug Induced Liver Injury Prediction System
107
                  (DILIps). PLoS Comput. Biol. 2011, 7 (12), e1002310.
108
                  https://doi.org/10.1371/journal.pcbi.1002310.
109
       (95)
                  Myshkin, E.; Brennan, R.; Khasanova, T.; Sitnik, T.;
110
                  Serebriyskaya, T.; Litvinova, E.; Guryanov, A.; Nikolsky, Y.;
111
                  Nikolskaya, T.; Bureeva, S. Prediction of Organ Toxicity
112
                  Endpoints by QSAR Modeling Based on Precise Chemical-
113
                  Histopathology Annotations. Chem. Biol. Drug Des. 2012, 80
114
                              406-416
                                               https://doi.org/10.1111/j.1747-
                  (3),
```

115

116

(96)

0285.2012.01411.x.

Chen, M.; Hong, H.; Fang, H.; Kelly, R.; Zhou, G.; Borlak, J.;

1		Tong W. Quantitativa Structura Activity Polationskin Models
		Tong, W. Quantitative Structure-Activity Relationship Models for Predicting Drug-Induced Liver Injury Based on FDA-
3		Approved Drug Labeling Annotation and Using a Large
4		Collection of Drugs. <i>Toxicol. Sci.</i> 2013 , <i>136</i> (1), 242–249.
2 3 4 5		https://doi.org/10.1093/toxsci/kft189.
6	(97)	Zhu, X. W.; Sedykh, A.; Liu, S. S. Hybrid in Silico Models for
7	(27)	Drug-Induced Liver Injury Using Chemical Descriptors and in
8		Vitro Cell-Imaging Information. J. Appl. Toxicol. 2014, 34 (3),
9		281–288. https://doi.org/10.1002/jat.2879.
10	(98)	Zhu, X. W.; Xin, Y. J.; Chen, Q. H. Chemical and in Vitro
11	(20)	Biological Information to Predict Mouse Liver Toxicity Using
12		Recursive Random Forests. SAR QSAR Environ. Res. 2016, 27
13		(7), 559–572.
14		https://doi.org/10.1080/1062936X.2016.1201142.
15	(99)	Zhang, C.; Cheng, F.; Li, W.; Liu, G.; Lee, P. W.; Tang, Y. In
16		Silico Prediction of Drug Induced Liver Toxicity Using
17		Substructure Pattern Recognition Method. Mol. Inform. 2016,
18		35 (3-4), 136-144. https://doi.org/10.1002/minf.201500055.
19	(100)	Hong, H.; Thakkar, S.; Chen, M.; Tong, W. Development of
20		Decision Forest Models for Prediction of Drug-Induced Liver
21		Injury in Humans Using A Large Set of FDA-Approved Drugs.
22		Sci. Rep. 2017, 7 (1), 17311. https://doi.org/10.1038/s41598-
23		017-17701-7.

- (101) Kim, E.; Nam, H. Prediction Models for Drug-Induced
 Hepatotoxicity by Using Weighted Molecular Fingerprints.
 BMC Bioinformatics 2017, 18 (Suppl 7).
 https://doi.org/10.1186/s12859-017-1638-4.
- 28 (102) McEuen, K.; Borlak, J.; Tong, W.; Chen, M. Associations of 29 Drug Lipophilicity and Extent of Metabolism with Drug-30 Induced Liver Injury. Int. J. Mol. Sci. 2017, 18 (7). https://doi.org/10.3390/ijms18071335.
- 32 (103)Wu, L.; Liu, Z.; Auerbach, S.; Huang, R.; Chen, M.; McEuen, 33 K.; Xu, J.; Fang, H.; Tong, W. Integrating Drug's Mode of 34 Action into Quantitative Structure-Activity Relationships for 35 Improved Prediction of Drug-Induced Liver Injury. J. Chem. 36 Inf. Model. 2017. 57 (4), 1000-1006 37 https://doi.org/10.1021/acs.jcim.6b00719.
- 38 (104) Li, X.; Chen, Y.; Song, X.; Zhang, Y.; Li, H.; Zhao, Y. The 39 Development and Application of in Silico Models for Drug 40 Induced Liver Injury. RSC Adv. 2018, 8 (15), 8101–8111. https://doi.org/10.1039/c7ra12957b.
- 42 (105) Papa, E.; Sangion, A.; Taboureau, O.; Gramatica, P.
 43 Quantitative Prediction of Rat Hepatotoxicity by Molecular
 44 Structure. Int. J. Quant. Struct. Relationships 2018, 3 (2), 49–
 45 60. https://doi.org/10.4018/ijqspr.2018070104.
- 46 (106) Ambe, K.; Ishihara, K.; Ochibe, T.; Ohya, K.; Tamura, S.;
 47 Inoue, K.; Yoshida, M.; Tohkin, M. In Silico Prediction of
 48 Chemical-Induced Hepatocellular Hypertrophy Using
 49 Molecular Descriptors. *Toxicol. Sci.* 2018, 162 (2), 667–675.
 50 https://doi.org/10.1093/toxsci/kfx287.
- 51 (107)López-Massaguer, O.; Pinto-Gil, K.; Sanz, F.; Amberg, A.; 52 Anger, L. T.; Stolte, M.; Ravagli, C.; Marc, P.; Pastor, M. 53 Generating Modeling Data from Repeat-Dose Toxicity Reports. 54 Toxicol. Sci. 2018. 162 (1),287-300. 55 https://doi.org/10.1093/toxsci/kfx254
- 56 (108) Chan, R.; Benet, L. Z. Evaluation of DILI Predictive
 Hypotheses in Early Drug Development. *Chem. Res. Toxicol.*2017, 30 (4), 1017–1029.

- $\begin{array}{lll} 59 & & \text{https://doi.org/10.1021/acs.chemrestox.7b00025.} \\ 60 & \text{(109)} & \text{He, S.; Ye, T.; Wang, R.; Zhang, C.; Zhang, X.; Sun, G.; Sun,} \\ 61 & & \text{X.} & \text{An In Silico Model for Predicting Drug-Induced} \\ \end{array}$
- 62 Hepatotoxicity. *Int. J. Mol. Sci.* **2019**, *20* (8), 1897. 63 https://doi.org/10.3390/ijms20081897.
- Wang, Y.; Xiao, Q.; Chen, P.; Wang, B. In Silico Prediction of
 Drug-Induced Liver Injury Based on Ensemble Classifier
 Method. *Int. J. Mol. Sci.* 2019, 20 (17), 4106.
 https://doi.org/10.3390/ijms20174106.
- 68 (111) Aleo, M. D.; Shah, F.; Allen, S.; Barton, H. A.; Costales, C.;
 69 Lazzaro, S.; Leung, L.; Nilson, A.; Obach, R. S.; Rodrigues, A.
 70 D.; et al. Moving Beyond Binary Predictions of Human Drug71 Induced Liver Injury (Dili) Towards Contrasting Relative Risk
 72 Potential. Chem. Res. Toxicol. 2019, No. Dili.
 73 https://doi.org/10.1021/acs.chemrestox.9b00262.
- 74 (112) Williams, D. P.; Lazic, S.; Foster, A. J.; Semenova, E.; Morgan,
 75 P. Predicting Drug-Induced Liver Injury with Bayesian
 76 Machine Learning. Chem. Res. Toxicol. 2019,
 77 acs.chemrestox.9b00264.
- 78 https://doi.org/10.1021/acs.chemrestox.9b00264.
- 79 (113) Chen, T.; Guestrin, C. XGBoost: A Scalable Tree Boosting 80 System. 2016. https://doi.org/10.1145/2939672.2939785.
- 81 (114) Klekota, J.; Roth, F. P. Chemical Substructures That Enrich for 82 Biological Activity. *Bioinformatics* **2008**, *24* (21), 2518–2525. https://doi.org/10.1093/bioinformatics/btn479.
- 84 (115) Rogers, D.; Hahn, M. Extended-Connectivity Fingerprints. *J*85 *Chem Inf Model.* **2010**, *50* (5), 742–754.

 86 https://doi.org/10.1021/ci100050t.
- 87 (116) Cronin, M. T. D.; Enoch, S. J.; Mellor, C. L.; Przybylak, K. R.; 88 Richarz, A. N.; Madden, J. C. In Silico Prediction of Organ Level Toxicity: Linking Chemistry to Adverse Effects. *Toxicol.* 90 *Res.* 2017, 33 (3), 173–182. https://doi.org/10.5487/TR.2017.33.3.173.
- 92 (117) Ellison, C. M.; Madden, J. C.; Judson, P.; Cronin, M. T. D.
 93 Using in Silico Tools in a Weight of Evidence Approach to Aid
 94 Toxicological Assessment. *Mol. Inform.* 2010, 29 (1–2), 97–
 110. https://doi.org/10.1002/minf.200900006.
- 96 (118) Stepan, A. F.; Walker, D. P.; Bauman, J.; Price, D. A.; Baillie, 97 T. A.; Kalgutkar, A. S.; Aleo, M. D. Structural Alert/Reactive 98 Metabolite Concept as Applied in Medicinal Chemistry to 99 Mitigate the Risk of Idiosyncratic Drug Toxicity: A Perspective 100 Based on the Critical Examination of Trends in the Top 200 101 Drugs Marketed in the United States. Chem. Res. Toxicol. 2011, 102 24 (9), 1345–1410. https://doi.org/10.1021/tx200168d.
- 103 (119) Yu, K.; Geng, X.; Chen, M.; Zhang, J.; Wang, B.; Ilic, K.; Tong, W. High Daily Dose and Being a Substrate of Cytochrome P450 Enzymes Are Two Important Predictors of Drug-Induced Liver Injury. Drug Metab. Dispos. 2014, 42 (4), 744–750. https://doi.org/10.1124/dmd.113.056267.
- 108 (120) Leeson, P. D.; St-Gallay, S. A.; Wenlock, M. C. Impact of Ion 109 Class and Time on Oral Drug Molecular Properties. 110 Medchemcomm 2011, 2 (2), 91–105. 111 https://doi.org/10.1039/c0md00157k.
- 112 (121) García-Cortés, M.; Lucena, M. I.; Pachkoria, K.; Borraz, Y.;
 113 Hidalgo, R.; Andrade, R. J. Evaluation of Naranjo Adverse
 114 Drug Reactions Probability Scale in Causality Assessment of
 115 Drug-Induced Liver Injury. Aliment. Pharmacol. Ther. 2008, 27
 116 (9), 780–789. https://doi.org/10.1111/j.1365-

1		2024 2000 02455
1 2	(122)	2036.2008.03655.x.
3	(122)	Björnsson, E. S.; Hoofnagle, J. H. Categorization of Drugs Implicated in Causing Liver Injury: Critical Assessment Based
4		on Published Case Reports. <i>Hepatology</i> 2016 , <i>63</i> (2), 590–603.
5		https://doi.org/10.1002/hep.28323.
6	(123)	Teschke, R.; Eickhoff, A.; Frenzel, C.; Wolff, A.; J., S. Drug
7	(123)	Induced Liver Injury: Accuracy of Diagnosis in Published
8		Reports. Ann. Hepatol. 2014, 13 (2), 248–255.
9	(124)	Yap, C. W. PaDEL-Descriptor: An Open Source Software to
10		Calculate Molecular Descriptors and Fingerprints. J. Comput.
11		Chem. 2011.
12	(125)	Adamson, G. W.; Lynch, M. F.; Town, W. G. Analysis of
13		Structural Characteristics of Chemical Compounds in a Large
14		Computer-Based File. Part II. Atom-Centred Fragments. J .
15		Chem. Soc. C Org. 1971, No. 3702, 3702.
16		https://doi.org/10.1039/j39710003702.
17	(126)	Mauri, A.; Consonni, V.; Pavan, M.; Todeschini, R. Dragon
18		Software: An Easy Approach to Molecular Descriptor
19		Calculations. <i>Match</i> 2006 , <i>56</i> (2), 237–248.
20	(127)	Tetko, I. V; Sushko, I.; Pandey, A. K.; Zhu, H.; Tropsha, A.;
21		Papa, E.; Todeschini, R.; Fourches, D.; Varnek, A. Critical
22 23		Assessment of QSAR Models of Environmental Toxicity
24	(120)	Against. Osiris 2008, 1733–1746.
25	(128)	Knox, C.; Law, V.; Jewison, T.; Liu, P.; Ly, S.; Frolkis, A.; Pon,
26		A.; Banco, K.; Mak, C.; Neveu, V.; et al. DrugBank 3.0: A
27		Comprehensive Resource for "omics" Research on Drugs. Nucleic Acids Res. 2011 , 39 (Database issue), D1035-41.
28		https://doi.org/10.1093/nar/gkq1126.
29	(129)	Jain, S.; Ecker, G. F. In Silico Approaches to Predict Drug-
30	()	Transporter Interaction Profiles: Data Mining, Model
31		Generation, and Link to Cholestasis. In Experimental
32		Cholestasis Research; Vinken, M., Ed.; Humana Press: New
33		York, 2019; pp 383-396. https://doi.org/10.1007/978-1-4939-
34		9420-5_26.
35	(130)	Welch, M. A.; Kock, K.; Urban, T. J.; Brouwer, K. L. R.;
36		Swaan, P. W. Toward Predicting Drug-Induced Liver Injury:
37		Parallel Computational Approaches to Identify Multidrug
38		Resistance Protein 4 and Bile Salt Export Pump Inhibitors.
39		Drug Metab. Dispos. 2015 , 43 (5), 725–734.
40		https://doi.org/10.1124/dmd.114.062539.
41 42	(131)	Montanari, F.; Pinto, M.; Khunweeraphong, N.; Wlcek, K.;
42		Sohail, M. I.; Noeske, T.; Boyer, S.; Chiba, P.; Stieger, B.;
44		Kuchler, K.; et al. Flagging Drugs That Inhibit the Bile Salt
45		Export Pump. <i>Mol. Pharm.</i> 2016 , <i>13</i> (1), 163–171. https://doi.org/10.1021/acs.molpharmaceut.5b00594.
46	(132)	Montanari, F.; Zdrazil, B.; Digles, D.; Ecker, G. F. Selectivity
47	(132)	Profiling of BCRP versus P-Gp Inhibition: From Automated
48		Collection of Polypharmacology Data to Multi-Label Learning.
49		J. Cheminform. 2016, 8 (1), 1–13.
50		https://doi.org/10.1186/s13321-016-0121-y.
51	(133)	Dawson, S.; Stahl, S.; Paul, N.; Barber, J.; Kenna, J. G. In Vitro
52		Inhibition of the Bile Salt Export Pump Correlates with Risk of
53		Cholestatic Drug-Induced Liver Injury in Humans. Drug
54		Metab. Dispos. 2012 , 40 (1), 130–138.
55		https://doi.org/10.1124/dmd.111.040758.
56	(134)	$Xi,L.;Yao,J.;Wei,Y.;Wu,X.;Yao,X.;Liu,H.;Li,S.\ The\ in$
57		Silico Identification of Human Bile Salt Export Pump
58		(ABCB11) Inhibitors Associated with Cholestatic Drug-

59		Induced Liver Injury. Mol. Biosyst. 2017, 13 (2), 417–424.
60		https://doi.org/10.1039/c6mb00744a.
61	(135)	Warner, D. J.; Chen, H.; Cantin, LD.; Kenna, J. G.; Stahl, S.;
62	()	Walker, C. L.; Noeske, T. Mitigating the Inhibition of Human
63		Bile Salt Export Pump by Drugs: Opportunities Provided by
64		Physicochemical Property Modulation, In Silico Modeling, and
65		Structural Modification. Drug Metab. Dispos. 2012, 40 (12),
66		2332–2341. https://doi.org/10.1124/dmd.112.047068.
67	(136)	Pauli-Magnus, C.; Meier, P. J. Hepatocellular Transporters and
68		Cholestasis. J. Clin. Gastroenterol. 2005, 39 (4 SUPPL.), 103-
69		110.
70	(137)	Gadaleta, D.; Manganelli, S.; Roncaglioni, A.; Toma, C.;
71		Benfenati, E.; Mombelli, E. QSAR Modeling of ToxCast
72		Assays Relevant to the Molecular Initiating Events of AOPs
73		Leading to Hepatic Steatosis. J. Chem. Inf. Model. 2018, 58 (8),
74		1501-1517. https://doi.org/10.1021/acs.jcim.8b00297.
75	(138)	Mellor, C. L.; Marchese Robinson, R. L.; Benigni, R.; Ebbrell,
76		D.; Enoch, S. J.; Firman, J. W.; Madden, J. C.; Pawar, G.; Yang,
77		C.; Cronin, M. T. D. Molecular Fingerprint-Derived Similarity
78 7 8		Measures for Toxicological Read-across: Recommendations for
79		Optimal Use. Regul. Toxicol. Pharmacol. 2019, 101 (October
80		2018), 121–134. https://doi.org/10.1016/j.yrtph.2018.11.002.
81	(139)	Liu, Y. Incorporation of Absorption and Metabolism into Liver
82		Toxicity Prediction for Phytochemicals: A Tiered in Silico
83 84		QSAR Approach. Food Chem. Toxicol. 2018, 118 (April), 409–
85	(1.40)	415. https://doi.org/10.1016/j.fct.2018.05.039.
86	(140)	Peng, Y.; Wu, Z.; Yang, H.; Cai, Y.; Liu, G.; Li, W.; Tang, Y.
87		Insights into Mechanisms and Severity of Drug-Induced Liver
88		Injury via Computational Systems Toxicology Approach. Toxicol. Lett. 2019, 312 (April), 22–33.
89		https://doi.org/10.1016/j.toxlet.2019.05.005.
90	(141)	Su, R.; Wu, H.; Xu, B.; Liu, X.; Wei, L. Developing a Multi-
91	,	Dose Computational Model for Drug-Induced Hepatotoxicity
92		Prediction Based on Toxicogenomics Data. IEEE/ACM Trans.
93		Comput. Biol. Bioinforma. 2018, 5963 (c).
94		https://doi.org/10.1109/TCBB.2018.2858756.
95	(142)	Wu, Y.; Wang, G. Machine Learning Based Toxicity
96		Prediction: From Chemical Structural Description to
97		Transcriptome Analysis. Int. J. Mol. Sci. 2018, 19 (8), 2358.
98		https://doi.org/10.3390/ijms19082358.
99	(143)	Battista, C.; Howell, B. A.; Siler, S. Q.; Watkins, P. B. An
100		Introduction to DILIsym® Software, a Mechanistic
101		Mathematical Representation of Drug-Induced Liver Injury;
102		2018; pp 101–121. https://doi.org/10.1007/978-1-4939-7677-
103 104	(1.14)	5_6.
104	(144)	Cherkasov, A.; Muratov, E. N.; Fourches, D.; Varnek, A.;
105		Baskin, I. I.; Cronin, M.; Dearden, J.; Gramatica, P.; Martin, Y.
107		C.; Todeschini, R.; et al. QSAR Modeling: Where Have You Been? Where Are You Going To? <i>J. Med. Chem.</i> 2014 , <i>57</i> (12),
108		4977–5010. https://doi.org/10.1021/jm4004285.
109	(145)	Wang, D. Infer the in Vivo Point of Departure with ToxCast in
110	\ ")	Vitro Assay Data Using a Robust Learning Approach. Arch.
111		Toxicol. 2018.
112	(146)	Olson, H.; Betton, G.; Robinson, D.; Thomas, K.; Monro, A.;
113		Kolaja, G.; Lilly, P.; Sanders, J.; Sipes, G.; Bracken, W.; et al.
114		Concordance of the Toxicity of Pharmaceuticals in Humans and
115		

in Animals. Regul. Toxicol. Pharmacol. 2000, 32 (1), 56-67.

https://doi.org/10.1006/rtph.2000.1399.

115