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Differential phenotypic expression of a novel *PDHA1* mutation in a female monozygotic twin pair

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Abstract

Pyruvate dehydrogenase complex (PDC) deficiency caused by mutations in the X-linked *PDHA1* gene has a broad clinical presentation, and the pattern of X-chromosome inactivation has been proposed as a major factor contributing to its variable expressivity in heterozygous females. Here, we report the first set of monozygotic twin females with PDC deficiency, caused by a novel, de novo heterozygous missense mutation in exon 11 of *PDHA1* (NM_000284.3: c.1100A>T). Both twins presented in infancy with a similar clinical phenotype including developmental delay, episodes of hypotonia or encephalopathy, epilepsy, and slowly progressive motor impairment due to pyramidal, extrapyramidal, and cerebellar involvement. However, they exhibited clear differences in disease severity that correlated well with residual PDC activities (approximately 60% and 20% of mean control values, respectively) and levels of immunoreactive E1 α subunit in cultured skin fibroblasts. To address whether the observed clinical and biochemical differences could be explained by the pattern of X-chromosome inactivation, we undertook an androgen receptor assay in peripheral blood. In the less severely affected twin, a significant bias in the relative activity of the two X chromosomes with a ratio of approximately 75:25 was detected, while the ratio was close to 50:50 in the other twin. Although it may be difficult to extrapolate these results to other tissues, our observation provides further support to the hypothesis that the pattern of X-chromosome inactivation may influence the phenotypic expression of the same mutation in heterozygous females and broadens the clinical and genetic spectrum of PDC deficiency.

Abbreviations

E1 α	α Subunit of the E1 component of the pyruvate dehydrogenase complex
E1 β	β Subunit of the E1 component of the pyruvate dehydrogenase complex
GTCS	Generalised tonic-clonic seizures

PDC	Pyruvate dehydrogenase complex
ULN	Upper limit of normal
WES	Whole-exome sequencing
XCI	X-chromosome inactivation

Introduction

The pyruvate dehydrogenase complex (PDC) is a large mitochondrial multienzyme complex that catalyses the oxidative decarboxylation of pyruvate to acetyl-CoA, a rate-limiting step for the aerobic oxidation of glucose in the brain and other tissues. PDC contains multiple copies of three catalytic components (E1 or pyruvate dehydrogenase, E2 or dihydrolipoamide acetyltransferase, and E3 or dihydrolipoamide dehydrogenase) and the non-catalytic E3 binding protein. E1 is a thiamine diphosphate-dependent enzyme formed by two α and two β subunits (abbreviated E1 α and E1 β), whereas E2 and E3 consist of a single type of polypeptide chain. PDC activity is modulated by phosphorylation and dephosphorylation of

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Robert D. S. Pitceathly and Michael G. Hanna contributed equally to the study.

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three serine residues of E1 α performed by two enzymes, pyruvate dehydrogenase kinase (PDK) and phosphatase (PDP), which are also linked to the complex. All components of PDC are encoded by autosomal genes with the exception of E1 α , encoded by the *PDHA1* gene in the X chromosome (De Meirleir et al. 2016; Patel et al. 2014).

PDC deficiency represents a common cause of primary lactic acidosis and neurological disease in infancy and early childhood, with more than 400 cases reported to date (Sperl et al. 2015). Although mutations affecting E1 β , E2, E3, and E3 binding protein and the regulatory enzyme PDP have been described, most cases are caused by mutations affecting E1 α (Patel et al. 2012; Sperl et al. 2015). The clinical spectrum of PDC-E1 α deficiency is broad. In males, three main presentations are recognised: (a) neonatal lactic acidosis and encephalopathy, sometimes associated with brain malformations; (b) infantile or childhood-onset Leigh or Leigh-like syndrome; and (c) a childhood-onset milder/relapsing neurological disorder that often includes ataxia, dystonia, and peripheral neuropathy. Heterozygous females appear to have a distinct clinical presentation that frequently includes dysmorphic features and microcephaly, especially in neonatal forms, in addition to moderate or severe psychomotor delay, spastic di/quadriplegia, and epilepsy. Brain imaging may reveal cortical/subcortical atrophy, dilated ventricles, cysts, and corpus callosum agenesis. Lactic acidosis may be present (Barnerias et al. 2010; De Meirleir et al. 2016; DeBrosse et al. 2012; Imbard et al. 2011; Lissens et al. 2000; Quintana et al. 2010).

That males are hemizygous and all females reported thus far are heterozygous for *PDHA1* mutations partly explains the clinical differences between sexes (Brown et al. 1994; Dahl 1995; Sperl et al. 2015). However, phenotypic variability among females with the same or functionally equivalent mutations also exists, and the pattern of X-chromosome inactivation (XCI) has been proposed as an important factor contributing to this variability (Brown et al. 1994; Dahl 1995; Dahl et al. 1992; Matthews et al. 1993). Here, we report for the first time female monozygotic twins with PDC-E1 α deficiency, caused by a novel missense mutation in exon 11 of *PDHA1*. Both twins presented with a similar, primarily neurological phenotype but showed clear differences in disease severity, residual PDC activities, and XCI ratios in blood. This observation broadens the clinical and genetic spectrum of PDC-E1 α deficiency and provides support to the hypothesis that the pattern of XCI is a major determinant of the phenotype in heterozygous females.

Methods

The female twins were clinically evaluated in the Myopathy clinic and the Highly Specialised Service for Mitochondrial Disease clinic of the National Hospital for Neurology and

Neurosurgery, London, UK. The study was granted ethical approval by the University College London (UCL) and UCL Hospital Joint Research Office (REC 09/H0716/76). Laboratory, neurophysiological, and imaging studies and tissue biopsies were performed using standard diagnostic protocols. Spectrophotometric assays of mitochondrial respiratory chain enzyme complexes in homogenised snap-frozen muscle samples were performed as previously described (Hargreaves et al. 2002). Total genomic DNA extracts from peripheral blood leukocytes and skeletal muscle biopsy were obtained for diagnostic purposes and used for research following informed consent. Diagnostic testing for point mutations and large-scale rearrangements of the mitochondrial DNA (mtDNA) was performed by restriction enzyme analysis, long-range PCR and Southern blotting on DNA extracted from skeletal muscle (Hammans et al. 1991; Muqit et al. 2008). For additional methods on molecular genetic studies, see the online Supplementary material.

Fibroblasts studies

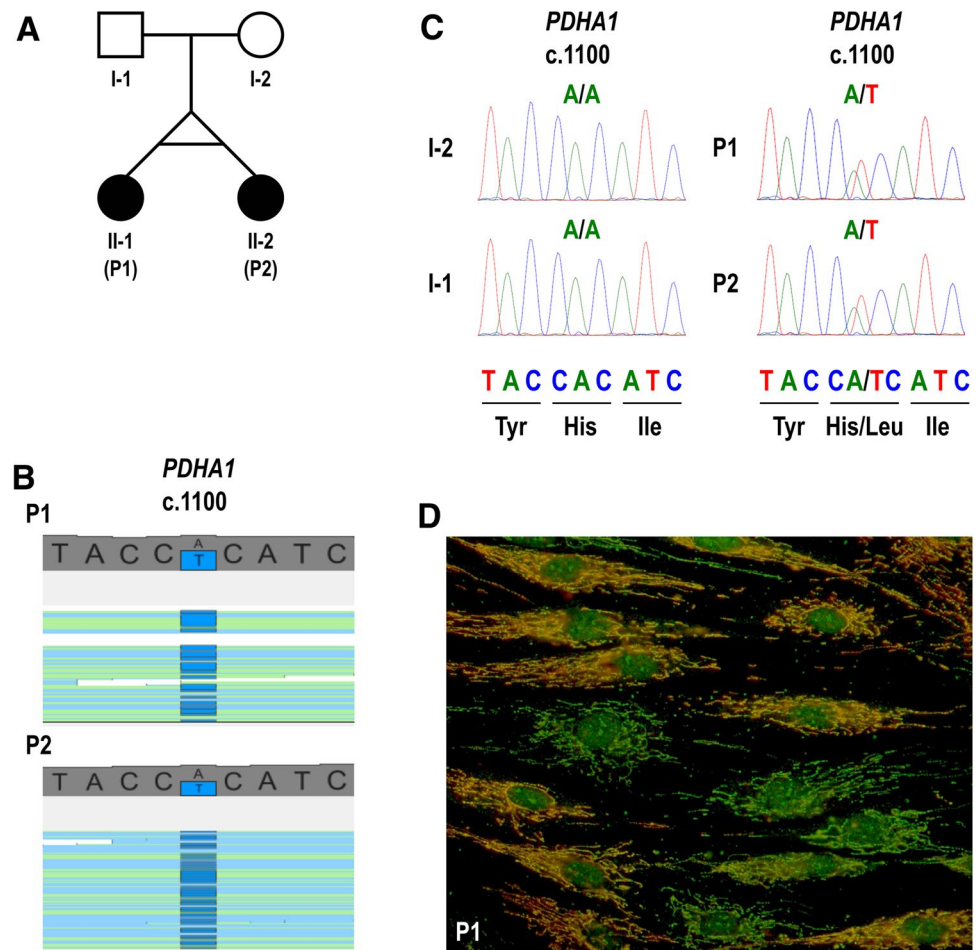
Skin biopsies were performed following informed consent. Fibroblasts from the patients were established from skin explants. All cells were maintained at 37 °C and 5% CO₂ under humidified conditions and cultured in high glucose Dulbecco's Modified Eagle medium (Invitrogen) supplemented with 10% foetal bovine serum (Biowest), penicillin/streptomycin, and 0.05 mg/ml uridine. Overall PDC activity was measured after maximal activation with dichloroacetate, using [1-¹⁴C]-pyruvate as substrate as described previously (Wicking et al. 1986). In this assay, the activity of normal control fibroblasts is 0.6–0.9 of ¹⁴CO₂ produced/mg protein/min. For immunocytochemical analysis, cultured fibroblasts were fixed and stained with antibodies to the PDC E1 α subunit and E2 enzyme as described previously (Lib et al. 2002). The anti-E1 α antibody was detected with a secondary antibody labelled with Alexa Fluor 594 (red). The anti-E2 antibody was labelled directly with Alexa Fluor 488 (green).

Results

Case reports

The female monozygotic twins were born to non-consanguineous parents by caesarean section for preeclampsia at 31 weeks of gestation (Fig. 1a). The second twin (P2) was briefly ventilated after delivery but there were no other major complications and both twins were discharged from the neonatal unit after 6 weeks. In subsequent months, they were noted to have mild global developmental delay and, at age 15 months, both became febrile and hypotonic 10 days after a vaccination, necessitating hospitalisation of

Fig. 1 Family pedigree, DNA sequencing, and immunocytochemistry. **a** Family pedigree. **b** Visualisation of the heterozygous variant NM_000284.3 (*PDHA1*): c.1100A>T (p.His367Leu) on whole-exome sequencing. **c** Sanger sequencing electropherograms of nucleotide NM_000284.3 (*PDHA1*) c.1100. The heterozygous variant c.1100A>T is present in both twins and absent in their parents, confirming that the variant is de novo. **d** Mosaic expression of PDC E1 α subunit in cultured fibroblasts from P1. Merged images of fibroblasts labelled with anti-E1 α antibody (red) and anti-E2 antibody (green). Yellow cells express both the E2 component of the PDC (encoded in chromosome 11) and the E1 α subunit, indicating that the active X chromosome in these cells contains the wild-type *PDHA1* allele. Green cells express the E2 component but are deficient in E1 α subunit, indicating that the active X chromosome in these cells contains the mutant *PDHA1* allele



P2. They lost developmental milestones after the episode, although subsequently gradually recovered. At age 3 years, P2 became confused and lethargic during a febrile illness and was admitted to the hospital again. This second episode prompted referral to a specialist clinic and, at age 5 years, both twins were diagnosed with Leigh syndrome on the basis of increased CSF lactate concentrations and abnormal neuroimaging (signal changes in the globi pallidi and peritrigonal white matter).

P2 continued to have episodes of acute encephalopathy triggered by infections, from which she recovered well. However, learning difficulties and impaired motor skills, coordination, and gait became apparent during the first decade of life, and repeated examinations revealed dysarthria, dysmetria, and dystonic posturing in the upper limbs, along with pyramidal tract signs in the lower limbs. She attended a special needs school and required a wheelchair for outdoor mobility during the teenage years. In adolescence, she suffered from three generalised tonic–clonic seizures (GTCS) during an encephalopathic episode and subsequently developed recurrent GTCS and seizures with impaired awareness and automatisms. Partial control of seizures was achieved with antiepileptic drug polytherapy. In her early

20s, following two further encephalopathic episodes with seizures, she developed behavioural changes and her overall motor function deteriorated. Coenzyme Q₁₀ and quetiapine treatment was initiated at that time and symptoms slowly improved.

Aged 29 years, P2 was being treated with lamotrigine, levetiracetam, midazolam (as required), quetiapine, riboflavin, thiamine, L-carnitine, and coenzyme Q₁₀. On examination, she was hypomimic and had moderate dysarthria and laterocollis to the left. Upgaze was markedly restricted, horizontal saccades were slow, and smooth pursuit was broken horizontally and vertically. Cranial nerve examination was otherwise normal. Tone was increased in limbs and there was mild weakness (MRC grade 4–4+/5) distally in the upper limbs and proximally in the lower limbs. Deep tendon reflexes were all present (2/5) and plantar responses were extensor. She had a mild upper limb postural tremor and intention tremor with dysmetria, with an emphasis on the right side. There was also dystonic posturing and mild-to-moderate bradykinesia of the upper and lower limbs with left-sided predominance. Pinprick sensation, vibration sense, and joint position sense were normal. Romberg's test was negative and she had a narrow-based, dystonic–spastic gait.

P1 had similar, but milder symptoms and clinical course compared to her sister. Her second and third encephalopathic episodes occurred at ages 14 and 19 years in the context of acute infections and she recovered completely. She developed seizures with impaired awareness and GTCS in her mid and late teenage years, respectively, but good seizure control was achieved with carbamazepine monotherapy. As with her sister, she had mild learning difficulties and, after the seizures started, began complaining of memory problems and perseverative behaviour. Motor function was retained in the upper limbs, but there were walking difficulties from early childhood. Examinations from the age of 10 revealed mild dysmetria and dystonia in the upper limbs and pyramidal tract signs in the lower limbs. She used a wheelchair outdoors since her early 20s.

Aged 29 years, P1 was being treated with carbamazepine, riboflavin, thiamine, L-carnitine, and coenzyme Q₁₀. On examination, she had moderate dysarthria and mild laterocollis to the left. Upgaze was moderately restricted, saccades were mildly slowed, and smooth pursuit was broken horizontally and vertically. Cranial nerve examination was otherwise unremarkable. Tone was slightly increased in her lower limbs, but muscle strength was intact. Deep tendon reflexes were all present (3/5) and plantar responses were extensor. There was mild postural and intention tremor with mild dysmetria and dysdiadochokinesia in both upper limbs. There was also mild dystonic posturing of the upper limbs and mild bradykinesia of the left upper and lower limb. Pinprick sensation, vibration sense, and joint position sense were normal. Romberg's test was negative. Gait was narrow based and mildly spastic.

Diagnostic studies

Following the initial clinical diagnosis at age 5 years, the twins were reinvestigated 5 years later and again in their 20s to determine their precise biochemical and genetic aetiology. At age 10 years, CSF and plasma lactate concentrations were increased (Supplementary Table 1). Electroretinography and visual evoked potentials were normal. Brain MRI scans of both twins were described as showing bilateral signal changes in the globus pallidum, peritrigonal white matter, and cerebellum. A cyst of the velum interpositum was also observed in P1. Needle muscle biopsy revealed type 2 fibre atrophy in P1 and slightly increased amounts of fine lipid droplets within the muscle fibres in the two twins. Histochemical staining for mitochondrial enzymes was normal. Skin fibroblasts were obtained for biochemical analysis, but failed to grow in culture.

Between the ages of 17 and 28 years, routine blood tests were repeatedly normal except for mildly raised creatine kinase (190–210 IU/L; upper limit of normal 140 IU/L) and plasma lactate (Supplementary Table 1). Plasma ammonia,

amino acids, and acyl-carnitines and urine organic acids were all within normal limits. White cell ubiquinone levels and lysosomal enzymes activities were normal. ECG and echocardiography showed no abnormalities. Brain MRI scans revealed bilateral lesions in the basal ganglia, peritrigonal white matter, and cerebellum that were more evident in P2 than P1 (Supplementary Fig. 1). Interictal EEG recordings revealed no signs of epileptiform activity. Brainstem evoked responses were normal in P1 but poorly formed in P2, suggesting either VIII cranial nerve or brainstem involvement. Further neuro-otological evaluation of both twins revealed impairment of ocular smooth pursuit, optokinetic responses, and vestibulo-ocular reflex suppression, consistent with cerebellar dysfunction. In P2, nerve conduction studies showed mildly decreased conduction velocities in the upper and lower limbs and mildly reduced sensory amplitudes in the lower limbs. Similar but milder changes with a patchy distribution were observed in P1 (Supplementary Tables 2 and 3). Quadriceps muscle biopsy of P1 at age 23 years only revealed features of denervation with reinnervation. The activity of mitochondrial respiratory chain complexes I, II + III, and IV was normal.

Genetic and biochemical studies

Based on the phenotype and MRI features suggestive of Leigh syndrome, P1 was screened for common point mutations (m.3243A>G, m.8344A>G, and m.8993T>G/C) and large-scale rearrangements of the mtDNA extracted from muscle, which resulted negative. Whole-exome sequencing (WES) was subsequently performed in both patients. Their exomes were 93% concordant, supporting monozygosity of the twin pair. Analysis focused on nonsynonymous, splice-site and coding indel variants with a minor allele frequency <0.1% in the Exome Aggregation Consortium dataset (ExAC; <http://exac.broadinstitute.org/>). From a total of 359 (P1) and 480 (P2) variants that met these filtering criteria, nine of them were in genes listed in MitoCarta, an inventory of genes encoding proteins with probable mitochondrial localisation (Supplementary Table 4) (Calvo et al. 2016). The nine candidate variants were individually reviewed and classified on the basis of gene-associated phenotypes and mode of inheritance (Supplementary Table 5). Only the heterozygous missense change NM_000284.3 (*PDHA1*): c.1100A>T (p.His367Leu) affected a gene known to cause lactic acidosis and/or Leigh syndrome, and was therefore considered the most plausible disease-causing variant (Fig. 1b). The presence of this heterozygous variant was confirmed by Sanger sequencing in both twins, but it was not detected in their parents, indicating that it had arisen de novo (Fig. 1c).

The c.1100A>T variant, located in exon 11 of *PDHA1*, is not reported in the Genome Aggregation Database

(gnomAD; <https://gnomad.broadinstitute.org/>), but affects the same nucleotide and amino acid as c.1100A>C (p.His367Pro), which is registered in ClinVar as likely pathogenic based on two independent submissions (ClinVar accession number RCV000493915.1). c.1100A>T (p.His367Leu) is predicted as being deleterious by most of the pathogenicity prediction tools used (Supplementary Table 6) and affects an evolutionarily conserved histidine residue (Supplementary Fig. 2). Analysis of the structural changes caused by the p.His367Leu substitution shows a potential alteration of the conformation of the C-terminus of E1 α and its interaction with E1 β within the E1 $\alpha_2\beta_2$ heterotetramer. This is so because the side chain of p.His367 is involved in a direct hydrogen bond with the C-terminal carboxylate of p.Ser390 in the same E1 α subunit; this carboxylate, in turn, hydrogen bonds to the side chain of p.Tyr131 in the E1 β subunit (Fig. 2). The first hydrogen bond cannot exist in the p.His367Leu variant and the second one is likely to be weakened in the face of competing interactions with water. In addition, the hydrophobic side chain of p.Leu367 would tend to associate with the side chain of p.Leu137 in the E1 β subunit (see Supplementary Figs. 3, 4 and 5 for details).

To confirm the functional impact of the variant, we measured the overall activity of PDC in cultured fibroblasts, which was moderately reduced in P1 and markedly reduced in P2 (Table 1). Immunocytochemical staining of E1 α in fibroblasts revealed a mosaic pattern that appeared to correlate well with the residual activity of the enzyme in each case (Fig. 1d and Table 1). To address whether differences in disease severity between the two twins could be explained by differences in the pattern of XCI, we determined this

Table 1 Pyruvate dehydrogenase complex activity in cultured fibroblasts and X chromosome inactivation pattern in DNA extracted from peripheral blood

Patient	PDC activity ^a (nmol/mg protein/ min)	Immunoreactive E1 α	XCI pattern ^b
P1	0.46	Moderate reduction	76:24
P2	0.17	Marked reduction	55:45

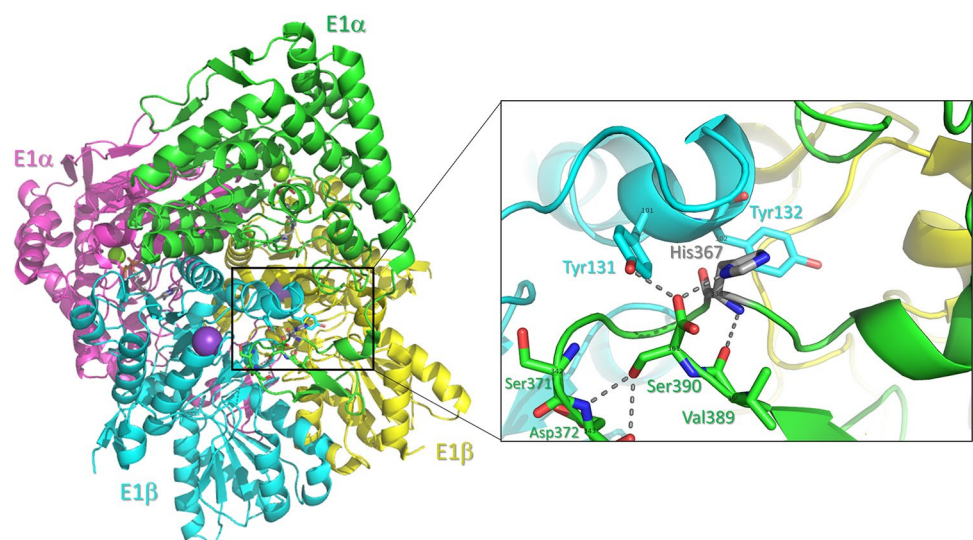
E1 α E1 α subunit of the E1 component of the pyruvate dehydrogenase complex, PDC pyruvate dehydrogenase complex, XCI X chromosome inactivation

^aRange in normal controls: 0.6–0.9 nmol/mg protein/min

^bRatio of the two PCR-amplified CAG repeat alleles within the inactive X chromosomes

pattern in DNA extracted from peripheral blood by assessing the methylation status of two CCGG sites located < 100 bp upstream a polymorphic CAG repeat within the androgen receptor (AR) gene. After restriction digestion using the methylation-sensitive HpaII enzyme, the CAG repeats on the inactive (methylated, undigested) X chromosomes were PCR amplified. Peak heights (signal intensity) of the resulting PCR products were determined by fragment analysis and the relative proportion of the two inactive X chromosome CAG alleles was calculated (see supplementary methods for details). Results were consistent with a skewed pattern of XCI in P1 (> 75–80% inactivation of the same allele) (Minks et al. 2008), but not in P2 (Table 1 and Supplementary Table 7). Analysis of CAG repeat sizes in the mother indicated that the maternal allele was preferentially active in both twins.

Fig. 2 Three-dimensional structure of human pyruvate dehydrogenase (E1 component) of PDC. The four subunits are arranged tetrahedrally as a dimer of dimers and the tetramer possesses a 2-fold symmetry axis. The inset shows the location of p.His367 (grey sticks) at a loop position of E1 α that is found at the interface with E1 β . Hydrogen bonds are displayed as dashed lines. The small-sized labels on top of C-alpha carbons denote the PDB numbering scheme (as per entry 1N14)



Discussion

This report describes the first set of female monozygotic twins with PDC-E1 α deficiency. Monozygosity was established on the basis of physical similarities and concordance of WES data. Both twins presented with a primarily neurological disorder that included mild developmental delay and learning difficulties, episodes of hypotonia or encephalopathy triggered by vaccination and infections from early childhood, and later onset of epilepsy and signs of pyramidal, extrapyramidal, and cerebellar dysfunction leading to slowly progressive motor impairment. Brain MRI revealed lesions similar to those seen in Leigh syndrome that were more symmetrically distributed in P2 than in P1. They had no dysmorphic features and no history of lactic acidosis, although their plasma lactate levels were often mildly increased and associated with a lactate:pyruvate ratio < 20. This clinical picture was consistent with previous descriptions of heterozygous females with PDC-E1 α deficiency, but the impossibility to perform enzymatic assays at age 10 years due to failure of fibroblast culture caused a significant delay in the diagnosis. The molecular diagnosis was ultimately achieved using WES and the pathogenicity of the newly identified de novo mutation supported by biochemical and immunocytochemical studies.

PDC-E1 α deficiency has considerable allelic heterogeneity with over 100 mutations, mostly missense mutations and small in-frame and frameshift indels, identified throughout the entire coding sequence of *PDHA1* except exon 2 (Imbard et al. 2011; Lissens et al. 2000; Patel et al. 2012; Quintana et al. 2010). Missense mutations are more common than indels and tend to localise to exons 3–9, whereas most indels are localised to exons 10–11. Hemizygous males, in whom the defective enzyme is expressed in all cells and the residual PDC activity depends on the effect of the mutation, frequently have missense mutations that allow the production of a partially functioning enzyme. In contrast, heterozygous females, in whom the defective enzyme is only expressed in a proportion of cells, can tolerate more severe mutations and have frameshift mutations more often than males. Heterozygous females harbouring missense mutations may be asymptomatic or have milder clinical phenotypes and survive into the second or third decades of life, as illustrated by the present and other cases (Cameron et al. 2004; Fujii et al. 1994; Lissens et al. 1999; van Dongen et al. 2015; Willemsen et al. 2006). However, they can also be severely affected (Cameron et al. 2004; Patel et al. 2012; Quintana et al. 2010; Willemsen et al. 2006), which likely reflects the differential effect of each mutation on E1 α structure and function (Imbard et al. 2011; Korotchkina et al. 2004).

In addition, heterozygous females can exhibit variable expression of the same missense mutation probably due to interindividual differences in the pattern of XCI (Brown et al. 1994; Dahl 1995; Dahl et al. 1992; De Meirleir et al. 1993; Fujii et al. 1996; Otero et al. 1998).

The novel missense mutation we detected in exon 11 of *PDHA1* (p.His367Leu) resides close to the C-terminal region of E1 α , where other point mutations (e.g. p.Arg378Cys, p.Arg378His, and p.Trp383*) have been previously identified in female patients (Barnerias et al. 2010; Chun et al. 1995; Fujii et al. 1996; Willemsen et al. 2006). Structural analysis of p.His367Leu indicates a potential deleterious effect of this amino acid substitution on the conformation of the C-terminus of E1 α and its interaction with E1 β . These changes result in E1 $\alpha_2\beta_2$ heterotetramer instability in neighbouring regions that may lead to increased degradation of the unassembled subunits. This possibility is consistent with the observation of reduced steady-state levels of immunoreactive E1 α in a proportion of cultured fibroblasts from both twins. A similar pathogenic mechanism has been proposed for the recurrent mutation p.Arg378His (Korotchkina et al. 2004), which severely impairs E1 function in vitro and is associated with reduced or undetectable amounts of E1 α and E1 β in patient-derived fibroblasts despite normal levels of E1 α mRNA (Hansen et al. 1991; Tripatara et al. 1999; Wexler et al. 1997). E1 α deletion mutants lacking one to four of the C-terminal amino acids (including p.Ser390, which forms a hydrogen bond with p.His367) (Korotchkina et al. 2004) also show decreased amounts of E1 α and E1 β despite normal transcription and mitochondrial import of E1 α , supporting that the integrity of the C-terminus of E1 α is essential for the stability or assembly of the E1 enzyme (Seyda et al. 2000).

Although monozygotic, the twins exhibited different disease severity with P1 being less severely affected than P2. This clinical observation correlated well with residual PDC activities in cultured skin fibroblasts, approximately 60% of mean control values in P1 and 20% in P2, and the corresponding reduction of immunoreactive E1 α . Analysis of WES data did not reveal mutations involving other components of the PDC in either twin. Therefore, we considered the possibility of constitutional unbalanced XCI favouring the wild-type allele in P1 and/or the mutant allele in P2 as the mechanism underlying their phenotypic differences. Using the AR assay in DNA extracted from peripheral blood, we detected a XCI ratio close to 50:50 in P2, indicating similar degree of activation of each X chromosome. In P1, in contrast, there was a significant bias in the relative activity of the two X chromosomes with a ratio of approximately 75:25. Based on these results and the fact that the proportion of normal and E1 α -deficient fibroblasts appear to remain constant in culture (Brown et al. 1990), it is possible that XCI skewing was also present in fibroblasts

from P1, favouring the expression of wild-type E1 α and, consequently, of fully functional PDC. It is well known that skewed XCI with ratios $\geq 80:20$ biased towards the expression of the wild-type allele can lead to normal PDC activities in fibroblast from female patients (Brown and Brown 1993; Matthews et al. 1994; Willemsen et al. 2006). However, XCI ratios of 70:30–80:20 have also been associated with PDC activities of 60–65% of mean control values, both determined in fibroblasts (Brown and Brown 1993), similar to what we observed in P1 in different tissues.

We believe that differences in the pattern of XCI in the twins is the most plausible explanation for the observed phenotypic differences, and the results of the AR assay in blood provide indirect support for this hypothesis. A good level of concordance in the XCI pattern between haematopoietic tissues and brain has been demonstrated in some female individuals (Bittel et al. 2008). If this were the case in P1, skewed XCI biased towards the wild-type allele in brain might account for her milder neurological disease. Nevertheless, due to inter- and intra-tissue variations in the pattern of XCI, we cannot exclude other scenarios such as skewed XCI favouring the mutant allele in fibroblasts or brain in P2, or variations in the XCI mosaic across different neuroanatomical regions in P1. The latter possibility has been observed in post-mortem brain tissue from one individual with Rett syndrome (Gibson et al. 2005), an X-linked neurodevelopmental disorder that mostly affects females, as well as in genetically engineered female mice expressing fluorescent reporters that allow visualisation of XCI at cellular resolution (Wu et al. 2014). Of note, lesion distribution on brain MRI was patchier in P1 than in P2, which might reflect topographic fluctuations in the pattern of XCI in P1, although we cannot discount the influence of other epigenetic or environmental factors on disease expression.

Distinct expression of X-linked disorders in monozygotic twin females has been widely reported in the literature (Bennett et al. 2008; Costa et al. 1997; De Gregorio et al. 2005; Johnston-MacAnanny et al. 2011; Tiberio 1994; Watkiss et al. 1994; Willemsen et al. 2000). However, with a few exceptions (Devys et al. 1992; Ishii et al. 2001; Mittal et al. 2011), most reported twin pairs exhibited discordant phenotypes, i.e. only one twin was clinically affected and the other twin was asymptomatic, with or without paraclinical evidence of heterozygous carrier status (e.g. reduced α -galactosidase activity in Fabry disease or factor VIII procoagulant activity in haemophilia A) (Bennett et al. 2008; Redonnet-Vernhet et al. 1996). In contrast, both members of the twin pair described here were clinically affected. PDC plays a pivotal role in energy production from glucose in all metabolically active tissues, especially in the brain, where it operates at high levels of activity even during fasting states (Siess et al. 1971). Consequently, even relatively mild reductions in PDC activity in heterozygous females or

mosaic individuals may have deleterious effects on normal brain development and function (Coughlin et al. 2010; Ridout et al. 2008). This most likely explains why a significant number of heterozygous females with PDC-E1 α deficiency manifest a severe neurological phenotype and the co-occurrence of the disease in both twins.

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Data availability statement Data sharing not applicable to this article as no datasets were generated or analysed during the current study.

Compliance with ethical standards

Conflict of interest The authors have no conflict of interest to declare.

Ethical approval All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki Declaration and its later amendments or comparable ethical standards.

Informed consent Informed consent was obtained from all individual participants included in the study.

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References


- Barnerias C, Saudubray JM, Touati G, De Lonlay P, Dulac O, Ponsot G, Marsac C, Brivet M, Desguerre I (2010) Pyruvate dehydrogenase complex deficiency: four neurological phenotypes with differing pathogenesis. *Dev Med Child Neurol* 52:e1–e9. <https://doi.org/10.1111/j.1469-8749.2009.03541.x>
- Bennett CM, Boye E, Neufeld EJ (2008) Female monozygotic twins discordant for hemophilia A due to nonrandom X-chromosome inactivation. *Am J Hematol* 83:778–780. <https://doi.org/10.1002/ajh.21219>
- Bittel DC, Theodoro MF, Kibiryeva N, Fischer W, Talebizadeh Z, Butler MG (2008) Comparison of X-chromosome inactivation patterns in multiple tissues from human females. *J Med Genet* 45:309–313. <https://doi.org/10.1136/jmg.2007.055244>
- Brown RM, Brown GK (1993) X chromosome inactivation and the diagnosis of X linked disease in females. *J Med Genet* 30:177–184. <https://doi.org/10.1136/jmg.30.3.177>
- Brown RM, Fraser NJ, Brown GK (1990) Differential methylation of the hypervariable locus DXS255 on active and inactive X chromosomes correlates with the expression of a human X-linked

- gene. *Genomics* 7:215–221. [https://doi.org/10.1016/0888-7543\(90\)90543-4](https://doi.org/10.1016/0888-7543(90)90543-4)
- Brown GK, Otero LJ, LeGris M, Brown RM (1994) Pyruvate dehydrogenase deficiency. *J Med Genet* 31:875–879. <https://doi.org/10.1136/jmg.31.11.875>
- Calvo SE, Clauser KR, Mootha VK (2016) MitoCarta2.0: an updated inventory of mammalian mitochondrial proteins. *Nucleic Acids Res* 44:D1251–D1257. <https://doi.org/10.1093/nar/gkv1003>
- Cameron JM, Levandovskiy V, Mackay N, Tein I, Robinson BH (2004) Deficiency of pyruvate dehydrogenase caused by novel and known mutations in the E1alpha subunit. *Am J Med Genet A* 131:59–66. <https://doi.org/10.1002/ajmg.a.30287>
- Chun K, MacKay N, Petrova-Benedict R, Federico A, Fois A, Cole DE, Robertson E, Robinson BH (1995) Mutations in the X-linked E1 alpha subunit of pyruvate dehydrogenase: exon skipping, insertion of duplicate sequence, and missense mutations leading to the deficiency of the pyruvate dehydrogenase complex. *Am J Hum Genet* 56:558–569
- Costa T, Greer W, Rysiecki G, Buncic JR, Ray PN (1997) Monozygotic twins discordant for Aicardi syndrome. *J Med Genet* 34:688–691. <https://doi.org/10.1136/jmg.34.8.688>
- Coughlin CR 2nd, Krantz ID, Schmitt ES, Zhang S, Wong LJ, Kerr DS, Ganesh J (2010) Somatic mosaicism for PDHA1 mutation in a male with pyruvate dehydrogenase complex deficiency. *Mol Genet Metab* 100:296–299. <https://doi.org/10.1016/j.ymgme.2010.04.004>
- Dahl HH (1995) Pyruvate dehydrogenase E1 alpha deficiency: males and females differ yet again. *Am J Hum Genet* 56:553–557
- Dahl HH, Hansen LL, Brown RM, Danks DM, Rogers JG, Brown GK (1992) X-linked pyruvate dehydrogenase E1 alpha subunit deficiency in heterozygous females: variable manifestation of the same mutation. *J Inherit Metab Dis* 15:835–847. <https://doi.org/10.1007/BF01800219>
- De Gregorio L, Jinnah HA, Harris JC, Nyhan WL, Schretlen DJ, Trombley LM, O'Neill JP (2005) Lesch-nyhan disease in a female with a clinically normal monozygotic twin. *Mol Genet Metab* 85:70–77. <https://doi.org/10.1016/j.ymgme.2004.11.009>
- De Meirleir L, Lissens W, Denis R, Wayenberg JL, Michotte A, Brucher JM, Vamos E, Gerlo E, Liebaers I (1993) Pyruvate dehydrogenase deficiency: clinical and biochemical diagnosis. *Pediatr Neurol* 9:216–220. [https://doi.org/10.1016/0887-8994\(93\)90088-T](https://doi.org/10.1016/0887-8994(93)90088-T)
- De Meirleir LJ, Garcia-Cazorla A, Brivet M (2016) Disorders of pyruvate metabolism and the tricarboxylic acid cycle. In: Saudubray JM, Baumgartner MR, Walter J (eds) *Inborn metabolic diseases: diagnosis and treatment*, 6th edn. Springer, Berlin Heidelberg, pp 187–199. https://doi.org/10.1007/978-3-662-49771-5_11
- DeBrosse SD, Okajima K, Zhang S, Nakouzi G, Schmotzer CL, Lusk-Kopp M, Frohnapfel MB, Grahame G, Kerr DS (2012) Spectrum of neurological and survival outcomes in pyruvate dehydrogenase complex (PDC) deficiency: lack of correlation with genotype. *Mol Genet Metab* 107:394–402. <https://doi.org/10.1016/j.ymgme.2012.09.001>
- Devys D, Biancalana V, Rousseau F, Boue J, Mandel JL, Oberle I (1992) Analysis of full fragile X mutations in fetal tissues and monozygotic twins indicate that abnormal methylation and somatic heterogeneity are established early in development. *Am J Med Genet* 43:208–216. <https://doi.org/10.1002/ajmg.1320430134>
- Fujii T, Van Coster RN, Old SE, Medori R, Winter S, Gubits RM, Matthews PM, Brown RM, Brown GK, Dahl HH et al (1994) Pyruvate dehydrogenase deficiency: molecular basis for intrafamilial heterogeneity. *Ann Neurol* 36:83–89. <https://doi.org/10.1002/ana.410360116>
- Fujii T, Garcia Alvarez MB, Sheu KF, Kranz-Eble PJ, De Vivo DC (1996) Pyruvate dehydrogenase deficiency: the relation of the E1 alpha mutation to the E1 beta subunit deficiency. *Pediatr Neurol* 14:328–334. [https://doi.org/10.1016/0887-8994\(96\)00058-6](https://doi.org/10.1016/0887-8994(96)00058-6)
- Gibson JH, Williamson SL, Arbuckle S, Christodoulou J (2005) X chromosome inactivation patterns in brain in Rett syndrome: implications for the disease phenotype. *Brain Dev* 27:266–270. <https://doi.org/10.1016/j.braindev.2004.07.002>
- Hamman SR, Sweeney MG, Brockington M, Morgan-Hughes JA, Harding AE (1991) Mitochondrial encephalopathies: molecular genetic diagnosis from blood samples. *Lancet* 337:1311–1313. [https://doi.org/10.1016/0140-6736\(91\)92981-7](https://doi.org/10.1016/0140-6736(91)92981-7)
- Hansen LL, Brown GK, Kirby DM, Dahl HH (1991) Characterization of the mutations in three patients with pyruvate dehydrogenase E1 alpha deficiency. *J Inherit Metab Dis* 14:140–151. <https://doi.org/10.1007/BF01800586>
- Hargreaves P, Rahman S, Guthrie P, Taanman JW, Leonard JV, Land JM, Heales SJ (2002) Diagnostic value of succinate ubiquinone reductase activity in the identification of patients with mitochondrial DNA depletion. *J Inherit Metab Dis* 25:7–16. <https://doi.org/10.1023/A:1015104910239>
- Imbard A, Boutron A, Vequaud C, Zater M, de Lonlay P, de Baulny HO, Barnerias C, Mine M, Marsac C, Saudubray JM, Brivet M (2011) Molecular characterization of 82 patients with pyruvate dehydrogenase complex deficiency. Structural implications of novel amino acid substitutions in E1 protein. *Mol Genet Metab* 104:507–516. <https://doi.org/10.1016/j.ymgme.2011.08.008>
- Ishii T, Makita Y, Ogawa A, Amamiya S, Yamamoto M, Miyamoto A, Oki J (2001) The role of different X-inactivation pattern on the variable clinical phenotype with Rett syndrome. *Brain Dev* 23(Suppl 1):S161–S164. [https://doi.org/10.1016/S0387-7604\(01\)00344-8](https://doi.org/10.1016/S0387-7604(01)00344-8)
- Johnston-MacAnanny EB, Koty P, Pettenati M, Brady M, Yalcinkaya TM, Schmidt DW (2011) The first case described: monozygotic twin sisters with the fragile X premutation but with a different phenotype for premature ovarian failure. *Fertil Steril* 95(2431):e13–e15. <https://doi.org/10.1016/j.fertnstert.2011.01.039>
- Korotchkina LG, Ciszak EM, Patel MS (2004) Function of several critical amino acids in human pyruvate dehydrogenase revealed by its structure. *Arch Biochem Biophys* 429:171–179. <https://doi.org/10.1016/j.abb.2004.06.027>
- Lib MY, Brown RM, Brown GK, Marusich MF, Capaldi RA (2002) Detection of pyruvate dehydrogenase E1 alpha-subunit deficiencies in females by immunohistochemical demonstration of mosaicism in cultured fibroblasts. *J Histochem Cytochem* 50:877–884. <https://doi.org/10.1177/002215540205000701>
- Lissens W, Vreken P, Barth PG, Wijburg FA, Ruitenbeek W, Wanders RJ, Seneca S, Liebaers I, De Meirleir L (1999) Cerebral palsy and pyruvate dehydrogenase deficiency: identification of two new mutations in the E1alpha gene. *Eur J Pediatr* 158:853–857. <https://doi.org/10.1007/s004310051222>
- Lissens W, De Meirleir L, Seneca S, Liebaers I, Brown GK, Brown RM, Ito M, Naito E, Kuroda Y, Kerr DS, Wexler ID, Patel MS, Robinson BH, Seyda A (2000) Mutations in the X-linked pyruvate dehydrogenase (E1) alpha subunit gene (PDHA1) in patients with a pyruvate dehydrogenase complex deficiency. *Hum Mut* 15:209–219. [https://doi.org/10.1002/\(SICI\)1098-1004\(200003\)15:3<3c209:AID-HUMU1%3e3.0.CO;2-K](https://doi.org/10.1002/(SICI)1098-1004(200003)15:3<3c209:AID-HUMU1%3e3.0.CO;2-K)
- Matthews PM, Brown RM, Otero L, Marchington D, Leonard JV, Brown GK (1993) Neurodevelopmental abnormalities and lactic acidosis in a girl with a 20-bp deletion in the X-linked pyruvate dehydrogenase E1 alpha subunit gene. *Neurology* 43:2025–2030. <https://doi.org/10.1212/wnl.43.10.2025>
- Matthews PM, Brown RM, Otero LJ, Marchington DR, LeGris M, Howes R, Meadows LS, Shevell M, Scriver CR, Brown GK (1994) Pyruvate dehydrogenase deficiency: Clinical presentation

- and molecular genetic characterization of five new patients. *Brain* 117(Pt 3):435–443. <https://doi.org/10.1093/brain/117.3.435>
- Minks J, Robinson WP, Brown CJ (2008) A skewed view of X chromosome inactivation. *J Clin Invest* 118:20–23. <https://doi.org/10.1172/JCI34470>
- Mittal K, Kabra M, Juyal R, Bk T (2011) De novo deletion in MECP2 in a monozygotic twin pair: a case report. *BMC Med Genet* 12:113. <https://doi.org/10.1186/1471-2350-12-113>
- Muqit MM, Lerner AJ, Sweeney MG, Sewry C, Stinton VJ, Davis MB, Healy DG, Payne SJ, Chotai K, Wood NW, Lane RJ (2008) Multiple mitochondrial DNA deletions in monozygotic twins with OPMD. *J Neurol Neurosurg Psychiatry* 79:68–71. <https://doi.org/10.1136/jnnp.2006.112250>
- Otero LJ, Brown RM, Brown GK (1998) Arginine 302 mutations in the pyruvate dehydrogenase E1 α subunit gene: identification of further patients and in vitro demonstration of pathogenicity. *Hum Mut* 12:114–121. [https://doi.org/10.1002/\(SICI\)1098-1004\(1998\)12:2%3c114:AID-HUMU6%3e3.0.CO;2-%23](https://doi.org/10.1002/(SICI)1098-1004(1998)12:2%3c114:AID-HUMU6%3e3.0.CO;2-%23)
- Patel KP, O'Brien TW, Subramony SH, Shuster J, Stacpoole PW (2012) The spectrum of pyruvate dehydrogenase complex deficiency: clinical, biochemical and genetic features in 371 patients. *Mol Genet Metab* 105:34–43. <https://doi.org/10.1016/j.ymgme.2011.09.032>
- Patel MS, Nemeria NS, Furey W, Jordan F (2014) The pyruvate dehydrogenase complexes: structure-based function and regulation. *J Biol Chem* 289:16615–16623. <https://doi.org/10.1074/jbc.R114.563148>
- Quintana E, Gort L, Busquets C, Navarro-Sastre A, Lissens W, Moliner S, Lluch M, Vilaseca MA, De Meirleir L, Ribes A, Briones P, Group PDHW (2010) Mutational study in the PDHA1 gene of 40 patients suspected of pyruvate dehydrogenase complex deficiency. *Clin Genet* 77:474–482. <https://doi.org/10.1111/j.1399-0004.2009.01313.x>
- Redonnet-Vernhet I, Ploos van Amstel JK, Jansen RP, Wevers RA, Salvayre R, Levade T (1996) Uneven X inactivation in a female monozygotic twin pair with Fabry disease and discordant expression of a novel mutation in the alpha-galactosidase A gene. *J Med Genet* 33:682–688. <https://doi.org/10.1136/jmg.33.8.682>
- Ridout CK, Brown RM, Walter JH, Brown GK (2008) Somatic mosaicism for a PDHA1 mutation in a female with pyruvate dehydrogenase deficiency. *Hum Genet* 124:187–193. <https://doi.org/10.1007/s00439-008-0538-0>
- Seyda A, McEachern G, Haas R, Robinson BH (2000) Sequential deletion of C-terminal amino acids of the E α component of the pyruvate dehydrogenase (PDH) complex leads to reduced steady-state levels of functional E α β β tetramers: implications for patients with PDH deficiency. *Hum Mol Genet* 9:1041–1048. <https://doi.org/10.1093/hmg/9.7.1041>
- Siess E, Wittmann J, Wieland O (1971) Interconversion and kinetic properties of pyruvate dehydrogenase from brain. *Hoppe Seylers Z Physiol Chem* 352:447–452. <https://doi.org/10.1515/bchm2.1971.352.1.447>
- Sperl W, Fleuren L, Freisinger P, Haack TB, Ribes A, Feichtinger RG, Rodenburg RJ, Zimmermann FA, Koch J, Rivera I, Prokisch H, Smeitink JA, Mayr JA (2015) The spectrum of pyruvate oxidation defects in the diagnosis of mitochondrial disorders. *J Inher Metab Dis* 38:391–403. <https://doi.org/10.1007/s10545-014-9787-3>
- Tiberio G (1994) MZ female twins discordant for X-linked diseases: a review. *Acta Genet Med Gemellol (Roma)* 43:207–214. <https://doi.org/10.1017/S000156600001963>
- Tripatara A, Korotchkina LG, Patel MS (1999) Characterization of point mutations in patients with pyruvate dehydrogenase deficiency: role of methionine-181, proline-188, and arginine-349 in the alpha subunit. *Arch Biochem Biophys* 367:39–50. <https://doi.org/10.1006/abbi.1999.1231>
- van Dongen S, Brown RM, Brown GK, Thorburn DR, Boneh A (2015) Thiamine-Responsive and Non-responsive Patients with PDHC-E1 Deficiency: a Retrospective Assessment. *JIMD Rep* 15:13–27. https://doi.org/10.1007/8904_2014_293
- Watkiss E, Webb T, Rysiecki G, Girdler N, Hewett E, Bunday S (1994) X inactivation patterns in female monozygotic twins and their families. *J Med Genet* 31:754–757
- Wexler ID, Hemalatha SG, McConnell J, Buist NR, Dahl HH, Berry SA, Cederbaum SD, Patel MS, Kerr DS (1997) Outcome of pyruvate dehydrogenase deficiency treated with ketogenic diets. Studies in patients with identical mutations. *Neurology* 49:1655–1661. <https://doi.org/10.1212/wnl.49.6.1655>
- Wicking CA, Scholem RD, Hunt SM, Brown GK (1986) Immunochemical analysis of normal and mutant forms of human pyruvate dehydrogenase. *Biochem J* 239:89–96. <https://doi.org/10.1042/bj2390089>
- Willemsen R, Olmer R, De Diego Otero Y, Oostra BA (2000) Twin sisters, monozygotic with the fragile X mutation, but with a different phenotype. *J Med Genet* 37:603–604. <https://doi.org/10.1136/jmg.37.8.603>
- Willemsen M, Rodenburg RJ, Teszas A, van den Heuvel L, Kosztolanyi G, Morava E (2006) Females with PDHA1 gene mutations: a diagnostic challenge. *Mitochondrion* 6:155–159. <https://doi.org/10.1016/j.mito.2006.03.001>
- Wu H, Luo J, Yu H, Rattner A, Mo A, Wang Y, Smallwood PM, Erlanger B, Wheelan SJ, Nathans J (2014) Cellular resolution maps of X chromosome inactivation: implications for neural development, function, and disease. *Neuron* 81:103–119. <https://doi.org/10.1016/j.neuron.2013.10.051>

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