

Review

Light-based technologies for management of COVID-19 pandemic crisis

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Highlights

- Light-based technologies currently are cost-effective and widely available in market
- Photons can be used to inactivate SARS-CoV-2 in air, liquids and on surfaces
- Phototherapy can also be used as an adjuvant to modulate the host immune system.
- Light-based solutions can significantly contribute to mitigate the impacts of COVID-19 pandemic

Abstract

The global dissemination of the novel coronavirus disease (COVID-19) has accelerated the need for the implementation of effective antimicrobial strategies to target the causative agent SARS-CoV-2. Light-based technologies have a demonstrable broad range of activity over standard chemotherapeutic antimicrobials and conventional disinfectants, negligible emergence of resistance, and the capability to modulate the host immune response. This perspective article identifies the benefits, challenges, and pitfalls of repurposing light-based strategies to combat the emergence of COVID-19 pandemic.

Keywords: photoinactivation; ultraviolet; photodynamic; photobiomodulation; germicidal; virucidal; photobiology.

1. Introduction

The pandemic spread of the novel coronavirus disease (COVID-19), caused by the SARS-CoV-2 virus, is a red-alert global health threat [1,2]. In December 2019, COVID-19 expanded from Wuhan throughout China and was then exported throughout the world [1-4]. So far, more than 1 million people have been diagnosed with COVID-19 infection, and many more are expected to be diagnosed within the coming months [5,6]. As the epidemic evolves, national and global organizations are facing an urgent need to coordinate and combat this unprecedented large-scale public health crisis [6].

The epidemiological features of COVID-19 (*i.e.*, severity, full spectrum of disease, transmissibility) have not been fully dissected [7]. The consensus is that the risk for severe acute disease symptoms and death is higher among the elderly and the immunocompromised [8-10]. In severe cases, infected patients need to be transferred to intensive care units for tracheal intubation [11]. This phenomenon is particularly worrisome because it can overwhelm healthcare facilities during the epidemic peak [10-13].

The spread and persistence of SARS-CoV-2 in diverse environments, such as healthcare, community, and residential areas, underlines the urgency for developing effective decontamination approaches as the pandemic crisis evolves [14]. A successful disinfection strategy coupled with additional infection-prevention countermeasures may substantially reduce transmissibility amongst asymptomatic carriers, a feature that is considered pivotal in the rapid dissemination of SARS-CoV-2. New light-mediated disinfection protocols are currently validated in hospitals and healthcare facilities for surface, air, and water as well as personal protective equipment (PPE), including eyewear, N95 respirators, and masks. Additionally, photobiomodulation, a light-based anti-inflammatory therapy, may have direct palliative effects on patients suffering from the so-called ‘cytokine storm’ associated with severe COVID-19. This review examines the potential of light-based technologies to prevent COVID-19 infection and control its dissemination by direct viral inactivation and to treat COVID-19 by modulating the host immune system. The direct antimicrobial actions of solar and UV radiation, photodynamic therapy, antimicrobial blue light, and ultrafast pulsed lasers for disinfection or *in vivo* use are considered, and the application of photobiomodulation to stimulate the host to mount an anti-viral response is discussed.

2. SARS-CoV-2 stability outside the human body

SARS-CoV-2 is highly infectious [15] and transmission occurs through contaminated air, water, and surfaces, which plays a pivotal role in its unbridled dissemination. A recent study by van Doremalen and colleagues investigated the stability of SARS-CoV-2 in aerosols and on inanimate surfaces (*e.g.*, glass, metal, plastic, or cardboard) that can act as important transmission vectors [16]. Their findings suggest that aerosol and fomite transmission of SARS-CoV-2 is likely, indicating that the virus can remain viable and infectious for hours in aerosols and up to days on surfaces. This is in agreement with a recent comparative analysis of 22 studies looking at the persistence of a broader panel of human coronaviruses on inanimate surfaces [17]. This study included prominent pathogenic coronavirus species such as Severe Acute Respiratory Syndrome (SARS), Middle East Respiratory Syndrome (MERS) and endemic human coronaviruses (HCoV) and concluded that: 1) viruses can remain infectious from 2 hours to 9 days; 2) incubation temperature is critical, as some viruses can remain viable at 4°C for up to 28 days whereas at 30–40°C viral viability is reduced.

3. Historical Milestones of Antimicrobial Light

The microbicidal effects of light have been widely known for more than a century. In 1885, Duclaux experimented with several microbial species and concluded that “sunlight is the best, cheapest, and most universally applicable microbicidal agent that we have” [18]. As early as 1877, Downes and Blunt observed that light could effectively kill a series of microorganisms and reported that this effect was dependent on light parameters such as intensity, duration (*i.e.*, light dose) and that the shortest wavelengths (*e.g.*, blue to ultraviolet light) were the most effective [19]. The first report on the virucidal effects of UV radiation dates back to 1928 when Rivers and Gates used UV light to inactivate viral particles in suspension and proved the efficacy of the method through subsequent subcutaneous inoculation of rabbits [20].

In 1903, Niels Finsen was awarded the Nobel Prize in Physiology or Medicine for his contribution to the treatment of infectious diseases, especially cutaneous tuberculosis, using visible light [21,22]. Virtually at the same time, Herman Von Tappeiner and Oscar Raab discovered by accident that the use of fluorescent dyes could enhance the microbial killing effect of visible light via photodynamic reactions [22]. By the 1930s, germicidal low-pressure

Mercury (Hg) discharge lamps emitting quasi-monochromatic UV-C light (peak emission at 254 nm) had been introduced into the market as highly efficient disinfection equipment [23]. Thus, since the pre-antibiotic era, light-based strategies have been extensively studied and used to treat and prevent infections [24]. However, each photoinactivation strategy has its pros and cons that must be carefully considered when designing a new microbial control strategy.

4. Natural Ultraviolet Light

Ultraviolet (UV) radiation is naturally and ubiquitously emitted by the sun, representing 10 % of its total light output. Only a small portion of the sunlight spectra has direct antimicrobial properties (UV-C). However, since most UV-C light is filtered by the atmospheric ozone layer, in practical terms, the antimicrobial activity associated with sunlight is mostly caused by photochemical reactions induced by UV-A and UV-B photons which are absorbed by endogenous chromophores such as amino acid residues, flavins, and porphyrin derivatives [25]. While UV-A alone does not seem to exert any significant virucidal effects, natural and artificial sunlight, as well as radiation in the UV-B spectrum, have been shown to inactivate bacteriophages and human viruses [26]. A model for the potential of solar UV radiation to inactivate viruses aerosolized in the atmosphere concluded that a full day of sun exposure would on average decrease the infectivity of UV-sensitive viruses by 3 log₁₀ [27].

Besides its virucidal potential, solar UV radiation can also play a protective role against infectious diseases via its modulating effect on vitamin D production [28]. Vitamin D is known to upregulate the production of human cathelicidin, LL-37. This antimicrobial peptide has both antimicrobial and antiendotoxin activities, and also attenuates the production of proinflammatory cytokines which typically accompany respiratory tract infections. Accordingly, it was recently suggested that vitamin D could reduce the incidence, severity, and risk of death due to respiratory tract infections, notably those caused by COVID-19 [29]. However, conclusive evidence for an association between vitamin D supplementation and decreased risk of respiratory tract infections is still lacking.

UV-C is directly absorbed by pyrimidine bases causing their dimerization, which leads to viral inactivation via DNA or RNA damage [30]. Thymine is the main chromophore in DNA while uracil is its counterpart in RNA. Upon UV-C exposure, thymine and uracil form cyclobutane-dimers and pyrimidine-protein cross-links [30]. The viral protein coat has been shown to

protect nucleic acids from UV-C radiation, by shielding the RNA, quenching the excited states of RNA, and/or by surrounding the bases with a hydrophobic environment and limiting the mobility of the individual bases. This results in a reduction of the overall rate of photoreactions, which allows the formation of non-cyclobutane-type dipyrimidines and uridine hydrates. Viral coating proteins themselves may suffer UV photodamage and become cross-linked to RNA.

The International Ultraviolet Association (IUVA) recently released a fact sheet detailing the efficacy of UV on SARS-CoV-2 [31] in which they reviewed all the appropriate requirements for the safety of UV-C disinfection devices and discussed the corresponding performance standards and validation protocols. Coronaviruses display a wide range of UV-C LD₉₀ (UV-C dose necessary to inactivate 90% of a microbial population) values, from 7 to 241 J/m² so one might assume that the UV-C susceptibility of the novel SARS-CoV-2 (COVID-19) virus probably lies within this range [32]. Therefore, based on previous studies with SARS-CoV-1 and other RNA-based coronaviruses, UV-C light can be used to effectively inactivate such pathogens present in the air, PPE, and on several types of surfaces [33,34].

5. Ultraviolet Germicidal Irradiation (UVGI)

UV-C lamps have long been used in hospital and industrial settings for decontamination purposes. In the context of a mitigation approach to infection spreading, UV-C can be particularly helpful in the inactivation of virus-containing aerosols and surfaces.

Air disinfection via upper-room germicidal UV-C light fixtures may be able to reduce viral transmission via the airborne route. Accordingly, an observational study during the 1957 influenza pandemic reported that patients housed in hospital wards with upper-room UV-C had an infection rate of 1.9 %, compared to an infection rate of 18.9 % among patients housed in wards without UV-C [35]. However, it is important to note that the germicidal effect of UV-C seems to be strongly dependent on the relative humidity of the air, with UV-C effectiveness against influenza virus decreasing with increasing relative humidity [36].

The potential of viral spreading via contaminated surfaces depends on the ability of the virus to maintain infectivity in the environment, which in turn is influenced by several biological, physical, and chemical factors, including the type of virus, temperature, relative humidity, and type of surface [37]. A study that assessed the influence of genetic content and relative

humidity on the virucidal effect of UV-C using viruses of different genetic composition concluded that single-stranded nucleic acid (ssRNA and ssDNA) viruses were more susceptible to UV inactivation than viruses with double-stranded nucleic acid (dsRNA and dsDNA) and that the UV dose necessary to achieve the same level of virus inactivation at 85 % relative humidity (RH) was higher than that at 55 % RH [37].

In an effort to safely expand the lifespan of the supply of medical equipment during the ongoing COVID-19 pandemic, it was estimated that 15-20 min exposure to 100 $\mu\text{W}/\text{cm}^2$ of UV-C from a standard biosafety cabinet could efficiently disinfect N95 masks. The authors noted that this was an empirical estimation, with the proposed dose corresponding to 30 times the one necessary to inactivate 90 % of respiratory viruses. The widespread implementation of such a procedure could potentially reduce the current PPE burden by 75 % [38]. Even though UV does not seem to affect the filtrating capacity of N95 respirators, it is important to note that high UV-C doses can lead to reduced tensile strength of its materials [39].

Ultraviolet blood irradiation (UBI) is an alternative therapeutic application of UV-C [40]. This technique involves the extraction of a small proportion of the body's total blood volume (typically 1-2 %), its irradiation with UV-C (254 nm) in a quartz tube followed by reinfusing it into the body. Rather than direct inactivation of pathogens, this procedure is thought to stimulate the host immune defense and has been used to treat both bacterial and viral infections, notably hepatitis [41].

The combination of multiple light wavelengths has been explored for cosmetic, environmental (water disinfection) and clinical (microbial catheter disinfection) applications. However, the precise photobiological mechanism of action and the experimental workflow to develop a marketable application is still missing [42,43].

It must be remarked that UV-C light at 254 nm can be harmful to the eyes and skin and, therefore, it is recommended to use it in setups that avoid direct human exposure. The use of far-UV-C (207-222 nm) has been proposed to minimize potential harm with human exposure [44]. This spectral range is strongly absorbed by amino acid residues and, therefore, is blocked by the acellular stratum corneum of the skin and the cornea of the eye, while still effective in the inactivation of viral particles and microbial cells present on surfaces and aerosols. However, this technology is still not broadly available in the market and the cost is far higher than

common Hg lamps. UV-C LED technology is still limited to very compact applications. The shortest wavelengths available are around 250 nm, with the price per Watt being up to 1,000 times higher than that Hg lamps, while displaying an energy efficiency (< 5 %) far lower than that of Hg lamps (25-40 %).

6. Photoantimicrobials and Photodynamic Therapy

Visible light can exert antiviral effects via photodynamic mechanisms that are initiated upon absorption of light by exogenous photosensitizer compounds, such as phenothiazinium salts, porphyrins, nanoparticles, and others [45-48]. The inactivation of microorganisms and viruses by visible light is based on the generation of lethal oxidant species via photosensitized oxidation reactions, which require three components: the chromophore, termed the photosensitizer (PS), light, and oxygen. After light absorption, excited oxygen states are quickly formed, initially in the singlet, and subsequently in the triplet states (*i.e.*, considering the photocycle of organic molecules). These species can release the excitation energy in the form of light (*e.g.*, fluorescence and phosphorescence) or heat (non-radiative decay) emission. Since excited states are intrinsically more reactive than ground states, energy and electron transfer reactions can occur. There are two main mechanisms of photosensitized oxidation: Type I reactions depend on the encounter of the excited species with biological substrates. These reactions usually occur through electron or hydrogen abstraction, leading to radical chain reactions; Type II reactions rely on energy transfer reaction from the PS triplet state to molecular oxygen, generating singlet oxygen ($^1\text{O}_2$) (Figure 1) [49].

[Figure 1]

Light energy is thus converted into oxidation potential that can damage biomolecules. Antimicrobial photodynamic therapy (aPDT) is based on this process and it has been used to treat localized microbial infections caused by viruses, bacteria, fungi, and parasites [50]. Among the many pathogens that can be targeted by aPDT, viruses are perhaps the most vulnerable, as they depend on entering a host cell for survival and replication and can be inactivated by damaging the capsid or envelope molecules (lipids, carbohydrates, proteins) or internal molecules (nucleic acids) (Figure 1). Thus, many viruses can be treated via aPDT, including papillomavirus (HPV), hepatitis A virus (HAV), and herpes simplex virus (HSV) [51-53]. Additionally, the disinfection of biological fluids (plasma and blood products) by

photoantimicrobials has been performed for decades and is a well-regarded technological application of these compounds. For instance, extracorporeal photoinactivation of coronaviruses and other clinically relevant pathogens using methylene blue (MB)-mediated aPDT has been reported [54-59].

It is possible that photosensitized oxidation can neutralize SARS-CoV-2 and, consequently, play a role in mitigating the ongoing pandemic; however, there is no data available on the photodynamic inactivation of this virus. Thus, here we sought to find and discuss scientific literature that could help predict whether COVID-19 is more or less susceptible to oxidant species generated during aPDT.

While all types of viruses can be neutralized by aPDT, the inactivation efficiency depends on both the PS and the virus [60,61]. As a rule of the thumb, RNA-type phages are more easily photoinactivated than their DNA-type counterparts, suggesting that SARS-CoV-2, which is an enveloped RNA-type virus, can be easily neutralized by aPDT [61,62]. Guanine bases are the major targets for oxidation by photosensitizing agents in both RNA and DNA [63]. The formation of RNA-protein crosslinks may also be an important lesion involved in virus inactivation via aPDT [64].

Enveloped viruses are more prone to aPDT neutralization than those without an envelope, due to the role of PS in damaging envelope components [65,66]. Initial studies on viral inactivation by aPDT demonstrated the importance of the PS reaching specific reaction sites, so-called “marked targets”, for efficient viral inactivation [67]. Other reports have confirmed the importance of PS binding on efficient virus inactivation via aPDT, and the PS membrane partition coefficients can be used as a predictor of its virus inactivation efficacy [68,69]. Transmission electron microscopy data has revealed that low PS concentrations degrade envelope surface glycoproteins blocking virus internalization, while higher PS concentrations can destroy lipid membranes [70]. These results can be interpreted in terms of the current mechanistic understanding of photosensitized oxidation, specifically the important role of direct-contact reactions. Irreversible membrane damage occurs with the abstraction of a hydrogen atom from an unsaturated fatty acid by direct reaction with the triplet excited state of the PS. Subsequent formation of peroxy and alkoxy radicals leads to the build-up of truncated lipid aldehydes, which are ultimately responsible for opening membrane pores [71]. The fact

that irreversible damage occurs due to contact-dependent reactions, indicates that the damage can be confined within the nanometer location site of the PS [72].

In terms of the application of aPDT to treat COVID-19 patients, it is encouraging to note that this technique is already used to treat several respiratory diseases [73]. PDT has been used for decades to treat lung cancers and its successful application in the treatment of laryngeal papillomas has also been reported [74]. Geralde *et al.* recently demonstrated that acute pneumonia caused by *Streptococcus pneumoniae* could be treated via inhalation of indocyanine green combined with extracorporeal administration of infrared light [75]. A prophylactic approach proposed by Soares *et al.* showed that aPDT can also be used to eliminate bacterial biofilms frequently associated with endotracheal tubes and that can lead to more severe stages of acute respiratory syndromes [76].

Considering that: 1) SARS-CoV-2 is an enveloped RNA virus, 2) aPDT is efficient at neutralizing such viruses, and 3) light is already used to treat lung and airway-related infections, we propose that aPDT is a good candidate for treating COVID-19 or as an adjunct to disinfect biological fluids. Alternatively, photosensitizers could also be used to decontaminate liquids and surfaces or be incorporated into polymeric matrices such as plastics, fabrics, paper, and paints to produce photoantimicrobial materials [50,55,77]. Allotropes of carbon such as fullerenes, carbon nanotubes, and graphene can also show light-activated antimicrobial effects, including the inactivation of viruses [66,78,79].

7. Antimicrobial Blue Light

Visible blue light exhibits microbicidal effects in the wavelength range of 405–470 nm [25,80–84]. Accordingly, antimicrobial blue light (aBL) has been explored in the treatment of infectious diseases and as a disinfection adjuvant in healthcare settings. Clinical trials have revealed the efficiency of aBL in the treatment of acne, *Helicobacter pylori* gastrointestinal infections, and dental infections [83,85–87]. aBL was recently shown to rescue mice from methicillin-resistant *Staphylococcus aureus* (MRSA) and *Pseudomonas aeruginosa* wound infections [88,89]. Oral anaerobic periodontopathogenic bacteria (*Porphyromonas gingivalis*, *Prevotella intermedia*, and *P. nigrescens*) were also inhibited or completely eradicated under blue light irradiation [90,91].

The exact mechanisms underlying the antimicrobial effects of blue light are not yet completely understood but appear to involve the formation of short-lived reactive oxygen species (ROS) [92]. The most widely accepted view of the process posits that the photochemical mechanisms of aBL are based on light energy excitation of endogenous microbial intracellular light receptors (chromophores), such as porphyrins and flavins. Once excited, these receptors undergo energy transfer processes that lead to the generation of cytotoxic ROS which react with intracellular components resulting in photodamage and cell death by oxidative stress [93]. Since endogenous photoreceptors appear to be absent in viruses, the mechanisms by which aBL affects these pathogens remains unclear. However, it is currently known that: 1) the use of exogenous photosensitizers improves the efficiency of inactivation by blue light, and 2) the inactivation is more pronounced when the virus is present in body fluids, *e.g.*, saliva, feces, and blood plasma [94,95].

High-intensity narrow-spectrum light at 405 nm has been used for continuous decontamination of inpatient and outpatient burn units and patient-occupied intensive care isolation rooms, as well as the treatment of surgical site infections in an orthopedic operating room [96-98]. Compared to UV-C light, aBL displays decreased deleterious effects on eukaryotic cells, reducing the possibility of human tissue injury. While one should avoid direct eye exposure to light in this spectral region because the eye lens absorbs visible radiation, aBL can be used in human-occupied environments. Additionally, aBL exposure of an elastomeric material did not degrade the material's properties and prevented bacterial adhesion to the material [99].

In a recent bioinformatics study, SARS-CoV-2 was reported to be dependent on porphyrins, which it captures from human hemoglobin, resulting in altered heme metabolism [100]. However, the *in silico* methods used to obtain such results have been questioned by a commentary publication, putting into doubt whether SARS-CoV-2 actually interacts with heme metabolism and accumulates porphyrins [101]. If this thesis is experimentally proven to be correct, aBL might be able to kill SARS-CoV-2 by photoexcitation of its acquired porphyrins. Thus, potential applications of aBL to prevent and control COVID-19 may include the disinfection aerosols, surfaces and health care PPE.

8. Photobiomodulation Therapy

Photobiomodulation (PBM) employs low levels of red or near-infrared (NIR) light to treat and heal wounds and injuries, reduce pain and inflammation, regenerate damaged tissue, and protect tissue at risk of dying [102]. Instead of directly targeting viruses, PBM mainly acts on the mitochondria, the cell's powerhouses, which absorb light in the red and near-infrared spectral region [103]. The photons are absorbed by chromophores present in the mitochondria. Cytochrome c oxidase (*i.e.*, unit IV in the respiratory chain) appears to play the main role in this process. Other molecular chromophores include light and heat-sensitive ion channels (transient receptor potential) that, upon light activation, lead to changes in calcium concentrations. Nanostructured water (interfacial water) is also likely to act as a chromophore. Upon irradiation, the mitochondrial membrane potential is raised and oxygen consumption and ATP generation are increased. Subsequent activation of signaling pathways and transcription factors leads to fairly long-lasting effects even after relatively brief exposure of the tissue to light.

In the early 1900s, Finsen reported that patients exposed to red light exhibited significantly better recovery from smallpox infections than unexposed counterparts [21]. Since then, PBM has been used in the treatment of acute lung injury, pulmonary inflammation, and models of acute respiratory distress syndrome (ARDS), due to its ability to substantially reduce systemic inflammation while preserving lung function. [104-106]. There are currently 90 published papers on PBM concerning “acute lung injury” [106] OR “pulmonary inflammation” [107] OR “lung inflammation” [105] OR “ARDS” [108] OR “pneumonia” [109] OR “lung oxidative stress” [110] OR “asthma” [111] many involving small animal models where it can be argued that light penetrates more easily than in humans. However, there is a significant systemic effect of PBM, where absorption of light by circulating blood leads to major biological effects in distant parts of the body [112]. Because COVID-19 involves a “cytokine storm”, PBM delivered to the torso (chest and back) might not only allow some light to reach the lungs but might also reduce the systemic inflammation responsible for COVID-19 sepsis-like syndrome [113] and disseminated intravascular coagulation [114] that can be deadly [115]. Moreover, PBM is more effective on hypoxic cells [116], suggesting it could be effective for COVID-19 infection, which seems to be characterized by severe hypoxia [117].

Hospitalized patients receiving mechanical ventilation or under high-oxygen continuous positive airway pressure (CPAP) treatment could be placed on an LED pad. These do not generate unacceptable levels of heat, so the high fever experienced by these patients should not

be a problem. LED-based PBM devices similar to these have been approved by the FDA for general health and wellness applications, and there are no reported adverse effects [118].

9. Ultrafast Laser Irradiation

Ultrashort pulse lasers (USPLs) emitting visible to near-infrared light have been used to inactivate a broad spectrum of viruses (human immunodeficiency virus, human papillomavirus, encephalomyocarditis virus, M13 bacteriophage, tobacco mosaic virus, and murine cytomegalovirus) with no damage to human or murine cells [119-124]. Regardless of wavelength, ultrafast laser irradiation does not promote ionization effects that could impair host cells. This irradiation does not appear to destroy either bovine serum albumin or single-stranded DNA, nor cause adverse effects like those produced by toxic or carcinogenic chemicals. Previous works suggest that the antimicrobial effect of USPLs is exerted via impulsive stimulated Raman scattering, whereby high-frequency resonance vibrations provoke vibrations of sufficient strength to destroy the capsid [122,123].

However, laser pulsing may not be necessary for its antimicrobial action. Recently, Kingsley *et al.* applied a tunable mode-locked Ti-Sapphire laser emitting femtosecond pulses at wavelengths of 400, 408, 425, 450, 465, and 510 nm to verify inactivation of murine norovirus (MNV) [92]. Continuous-wave (CW) lasers were also used. More than 99.9 % of virus inactivation was reported after irradiation with an average power of 150 mW at wavelengths of 408, 425, or 450 nm femtosecond-pulsed light for 3 hours, indicating that the inactivation mechanism is not wavelength-specific. Further, irradiation using a CW laser of similar power at 408 nm resulted in a considerable reduction in MNV numbers, suggesting that inactivation does not require pulsing.

Other applications for USPLs include their use for pathogen reduction in blood and whole inactivated virus vaccines [124,125]. Laser treatment resulted in 1-log, 2-log, and 3-log reductions in hepatitis A, human immunodeficiency, and murine cytomegalovirus, respectively, in human plasma with no changes in the structure of fibrinogen. [125]. Further, in mice USPL-induced inactivation of H1N1 influenza virus was more effective than formalin and did not cause damage to surface proteins or resulted in the production of carbonyl groups in proteins [126].

10. Concluding remarks

In summary, we have described how light-based strategies can be used to reduce SARS-CoV-2 transmission through air, water, and surfaces as well as potential therapeutic applications that can reduce its morbidity and mortality. From our perspective, light provides several practical answers to the new logistical and therapeutic challenges brought by COVID-19. Therefore, we suggest that the death toll and quarantine extent can be significantly mitigated if at least part of these strategies are encouraged and implemented by health systems. Given the urgent demand raised by the current uncontrolled pandemic we must be ready to use all the available armamentarium to fight COVID-19.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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References

1. N. Zhu, D. Zhang, W. Wang, X. Li, B. Yang, J. Song, X. Zhao, B. Huang, W. Shi, R. Lu, P. Niu, F. Zhan, X. Ma, D. Wang, W. Xu, G. Wu, G.F. Gao, W. Tan; China Novel Coronavirus Investigating and Research Team, A Novel Coronavirus from Patients with Pneumonia in China, 2019, *N. Engl. J. Med.* 382 (2020) 727-33. <http://dx.doi.org/10.1056/NEJMoa2001017>

2. C. Huang, Y. Wang, X. Li, L. Ren, J. Zhao, Y. Hu, L. Zhang, G. Fan, J. Xu, X. Gu, Z. Cheng, T. Yu, J. Xia, Y. Wei, W. Wu, X. Xie, W. Yin, H. Li, M. Liu, Y. Xiao, H. Gao, L. Guo, J. Xie, G. Wang, R. Jiang, Z. Gao, Q. Jin, J. Wang, B. Cao, Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China, *Lancet* 395 (2020) 497-506. [http://dx.doi.org/10.1016/S0140-6736\(20\)30183-5](http://dx.doi.org/10.1016/S0140-6736(20)30183-5)
3. J. Bedford, D. Enria, J. Giesecke, D.L. Heymann, C. Ihekweazu, G. Kobinger, H.C. Lane, Z. Memish, M.D. Oh, A.A. Sall, A. Schuchat, K. Ungchusak, L.H. Wieler; WHO Strategic and Technical Advisory Group for Infectious Hazards, COVID-19: towards controlling of a pandemic, *Lancet* 395 (2020) 1015-1018. [http://dx.doi.org/10.1016/S0140-6736\(20\)30673-5](http://dx.doi.org/10.1016/S0140-6736(20)30673-5)
4. C.R. Wells, P. Sah, S.M. Moghadas, A. Pandey, A. Shoukat, Y. Wang, Z. Wang, L.A. Meyers, B.H. Singer, A.P. Galvani, Impact of international travel and border control measures on the global spread of the novel 2019 coronavirus outbreak, *Proc. Natl. Acad. Sci. USA* 117 (2020) 7504-7509. <http://dx.doi.org/10.1073/pnas.2002616117>
5. B. Xu, M.U.G. Kraemer, Group OC-DC, Open access epidemiological data from the COVID-19 outbreak, *Lancet Infect Dis.* S1473-3099 (2020) 30119-5. [http://dx.doi.org/10.1016/S1473-3099\(20\)30119-5](http://dx.doi.org/10.1016/S1473-3099(20)30119-5)
6. C. Sohrabi, Z. Alsafi, N. O'Neill, M. Khan, A. Kerwan, A. Al-Jabir, C. Iosifidis, R. Agha, World Health Organization declares global emergency: A review of the 2019 novel coronavirus (COVID-19), *Int. J. Surg.* 76 (2020) 71-6. <http://dx.doi.org/10.1016/j.ijsu.2020.02.034>
7. M. Lipsitch, D.L. Swerdlow, L. Finelli L, Defining the Epidemiology of Covid-19 - Studies Needed, *N. Engl. J. Med.* 382 (2020) 1194-1196. <http://dx.doi.org/10.1056/NEJMp2002125>
8. F. Zhou, T. Yu, R. Du, G. Fan, Y. Liu, Z. Liu, J. Xiang, Y. Wang, B. Song, X. Gu, L. Guan, Y. Wei, H. Li, X. Wu, J. Xu, S. Tu, Y. Zhang, H. Chen, B. Cao, Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study, *Lancet* 395 (2020) 1054-1062. [http://dx.doi.org/10.1016/S0140-6736\(20\)30566-3](http://dx.doi.org/10.1016/S0140-6736(20)30566-3)
9. L. D'Antiga, Coronaviruses and immunosuppressed patients. The facts during the third epidemic, *Liver Transpl.* (2020). <http://dx.doi.org/10.1002/lt.25756>
10. Z. Wu, J.M. McGoogan, Characteristics of and Important Lessons From the Coronavirus Disease 2019 (COVID-19) Outbreak in China: Summary of a Report of 72 314

Cases From the Chinese Center for Disease Control and Prevention, JAMA (2020). <http://dx.doi.org/10.1001/jama.2020.2648>

11. C.N. Wu, L.Z. Xia, K.H. Li, W.H. Ma, D.N. Yu, B. Qu, B.X. Li, Y. Cao, High-flow nasal-oxygenation-assisted fiberoptic tracheal intubation in critically ill patients with COVID-19 pneumonia: a prospective randomised controlled trial, Br. J. Anaesth. S0007-0912 (2020) 30135-5. <http://dx.doi.org/10.1016/j.bja.2020.02.020>
12. T.P. Velavan, C.G. Meyer, The COVID-19 epidemic, Trop. Med. Int. Health 25 (2020) 278-80. <http://dx.doi.org/10.1111/tmi.13383>
13. G. Grasselli, A. Pesenti, M. Cecconi, Critical Care Utilization for the COVID-19 Outbreak in Lombardy, Italy: Early Experience and Forecast During an Emergency Response, JAMA (2020). <http://dx.doi.org/10.1001/jama.2020.4031>
14. The Lancet, COVID-19: protecting health-care workers, Lancet 395 (2020) 922. [http://dx.doi.org/10.1016/S0140-6736\(20\)30644-9](http://dx.doi.org/10.1016/S0140-6736(20)30644-9)
15. Y. Liu, R.M. Eggo, A.J. Kucharski, Secondary attack rate and superspreading events for SARS-CoV-2, Lancet 395 (2020) e47. [http://dx.doi.org/10.1016/S0140-6736\(20\)30462-1](http://dx.doi.org/10.1016/S0140-6736(20)30462-1)
16. N. van Doremalen, T. Bushmaker, D.H. Morris, M.G. Holbrook, A. Gamble, B.N. Williamson, A. Tamin, J.L. Harcourt, N.J. Thornburg, S.I. Gerber, J.O. Lloyd-Smith, E. de Wit, V.J. Munster, Aerosol and Surface Stability of SARS-CoV-2 as Compared with SARS-CoV-1, N. Engl. J. Med. 382 (2020) 1564-1567. <http://dx.doi.org/10.1056/NEJMc2004973>
17. G. Kampf, D. Todt, S. Pfaender, E. Steinmann, Persistence of coronaviruses on inanimate surfaces and their inactivation with biocidal agents, J. Hosp. Infect. 104 (2020) 246-51. <http://dx.doi.org/10.1016/j.jhin.2020.01.022>
18. E. Duclaux, Comptes rendus de la Société de Biologie; 1885; 1885. p. 395.
19. A. Downes, T. Blunt, The influence of light upon the development of bacteria, Nature 16 (1877) 218.
20. T. Rivers, F.L. Gates, Ultra-Violet Light and Vaccine Virus: II. The effect of monochromatic Ultra-Violet light upon vaccine virus, J. Exp. Med. 47 (1928) 45-9. <http://dx.doi.org/10.1084/jem.47.1.45>
21. N. Finsen, Phototherapy. London: Edward Arnold; 1901.
22. R. Ackroyd, C. Kelty, N. Brown, M. Reed, The history of photodetection and photodynamic therapy, Photochem. Photobiol. 74 (2001) 656-69. [http://dx.doi.org/10.1562/0031-8655\(2001\)0740656THOPAP2.0.CO2](http://dx.doi.org/10.1562/0031-8655(2001)0740656THOPAP2.0.CO2)

23. P. Brickner, R.L. Vincent, Ultraviolet germicidal irradiation safety concerns: a lesson from the Tuberculosis Ultraviolet Shelter Study: Murphy's Law affirmed, *Photochem. Photobiol.* 89 (2013) 819-21. <http://dx.doi.org/10.1111/php.12034>
24. T. St Denis, T. Dai, L. Izikson, C. Astrakas, R.R. Anderson, M.R. Hamblin, G.P. Tegos, All you need is light: antimicrobial photoinactivation as an evolving and emerging discovery strategy against infectious disease, *Virulence* 2 (2011) 509-20. <http://dx.doi.org/10.4161/viru.2.6.17889>
25. T. Dai, The antimicrobial effect of blue light: What are behind? *Virulence* 8 (2017) 649-52. <http://dx.doi.org/10.1080/21505594.2016.1276691>
26. Y. Qiu, Q. Li, B.E. Lee, N.J. Ruecker, N.F. Neumann, N.J. Ashbolt, X. Pang, UV inactivation of human infectious viruses at two full-scale wastewater treatment plants in Canada, *Water Res.* 147 (2018) 73-81. <http://dx.doi.org/10.1016/j.watres.2018.09.057>
27. A. Ben-David, J.L. Sagripanti, A model for inactivation of microbes suspended in the atmosphere by solar ultraviolet radiation, *Photochem. Photobiol.* 86 (2010) 895-908. <http://dx.doi.org/10.1111/j.1751-1097.2010.00738.x>
28. Abhimanyu, A.K. Coussens, The role of UV radiation and vitamin D in the seasonality and outcomes of infectious disease, *Photochem. Photobiol. Sci.* 16 (2017) 314-38. <http://dx.doi.org/10.1039/c6pp00355a>
29. W.B. Grant, H. Lahore, S.L. McDonnell, C.A. Baggerly, C.B. French, J.L. Aliano, H.P. Bhattoa, Evidence that Vitamin D Supplementation Could Reduce Risk of Influenza and COVID-19 Infections and Deaths, *Nutrients* 12 (2020) 988. <http://dx.doi.org/10.3390/nu12040988>
30. S.E. Beck, R.A. Rodriguez, M.A. Hawkins, T.M. Hargy, T.C. Larason, K.G. Linden, Comparison of UV-Induced Inactivation and RNA Damage in MS2 Phage across the Germicidal UV Spectrum, *Appl. Environ. Microbiol.* 82 (2015) 1468-74. <http://dx.doi.org/10.1128/AEM.02773-15>
31. IUVA. Fact Sheet on UV Disinfection for COVID-19. LEDs Magazine 2020.
32. W. Kowalski, T.J. Walsh, V. Petraitis. 2020 COVID-19 Coronavirus Ultraviolet Susceptibility. 2020.
33. M. Darnell, K. Subbarao, S.M. Feinstone, D.R. Taylor, Inactivation of the coronavirus that induces severe acute respiratory syndrome, SARS-CoV, *J. Virol. Methods* 121 (2004) 85-91. <http://dx.doi.org/10.1016/j.jviromet.2004.06.006>
34. C.M. Walker, G. Ko, Effect of ultraviolet germicidal irradiation on viral aerosols, *Environ. Sci. Technol.* 41 (2007) 5460-5. <http://dx.doi.org/10.1021/es070056u>

35. W.S. Jordan, The mechanism of spread of Asian influenza, *Am. Rev. Respir. Dis.* 83 (1961) 29-40. <http://dx.doi.org/10.1164/arrd.1961.83.2P2.29>
36. J. McDevitt, S.N. Rudnick, L.J. Radonovich, Aerosol susceptibility of influenza virus to UV-C light, *Appl. Environ. Microbiol.* 78 (2012) 1666-9. <http://dx.doi.org/10.1128/AEM.06960-11>
37. C. Tseng, C.S. Li, Inactivation of viruses on surfaces by ultraviolet germicidal irradiation, *J. Occup. Environ. Hyg.* 4 (2007) 400-95. <http://dx.doi.org/10.1080/15459620701329012>
38. K.J. Card, D. Crozier, A. Dhawan, M. Dinh, E. Dolson, N. Farrokhian, V. Gopalakrishnan, E. Ho, E.S. King, N. Krishnan, G. Kuzmin, J. Maltas, J. Pelesko, J.A. Scarborough, J.G. Scott, G. Sedor, D.T. Weaver, UV Sterilization of Personal Protective Equipment with Idle Laboratory Biosafety Cabinets During the Covid-19 Pandemic, *medRxiv* 2020. <http://dx.doi.org/10.1101/2020.03.25.20043489>
39. W. Lindsley, S.B. Jr. Martin, R.E. Thewlis, K. Sarkisian, J.O. Nwoko, K.R. Mead, J.D. Noti, Effects of Ultraviolet Germicidal Irradiation (UVGI) on N95 Respirator Filtration Performance and Structural Integrity, *J. Occup. Environ. Hyg.* 12 (2015) 509-17. <http://dx.doi.org/10.1080/15459624.2015.1018518>
40. M.R. Hamblin, Ultraviolet Irradiation of Blood: "The Cure That Time Forgot"? *Adv. Exp. Med. Biol.* 996 (2017) 295-309. http://dx.doi.org/10.1007/978-3-319-56017-5_25
41. J.T. Kuenstner, S. Mukherjee, S. Weg, T. Landry, T. Petrie, The treatment of infectious disease with a medical device: results of a clinical trial of ultraviolet blood irradiation (UVBI) in patients with hepatitis C infection, *Int. J. Infect. Dis.* 37 (2015) 58-63. <http://dx.doi.org/10.1016/j.ijid.2015.06.006>
42. K. Song, F. Taghipour, M. Mohseni, Microorganisms inactivation by wavelength combinations of ultraviolet light-emitting diodes (UV-LEDs), *Sci. Total Environ.* 665 (2019) 1103-10. <http://dx.doi.org/10.1016/j.scitotenv.2019.02.041>
43. S. Beck, H. Ryu, L.A. Boczek, J.L. Cashdollar, K.M. Jeanis, J.S. Rosenblum, O.R. Lawal, K.G. Linden, Evaluating UV-C LED disinfection performance and investigating potential dual-wavelength synergy, *Water Res.* 109 (2017) 207-16. <http://dx.doi.org/10.1016/j.watres.2016.11.024>
44. D. Welch, M. Buonanno, V. Grilj, I. Shuryak, C. Crickmore, A.W. Bigelow, G. Randers-Pehrson, G.W. Johnson, D.J. Brenner, Far-UVC light: A new tool to control the spread of airborne-mediated microbial diseases, *Sci. Rep.* 8 (2018) 2752. <http://dx.doi.org/10.1038/s41598-018-21058-w>

45. A. Teles, T.M.A. Oliveira, F.C. Bezerra, L. Alonso, A. Alonso, I.E. Borissevitch, P.J. Gonçalves, G.R.L. Souza, Photodynamic inactivation of Bovine herpesvirus type 1 (BoHV-1) by porphyrins, *J. Gen. Virol.* 99 (2018) 1301-6. <http://dx.doi.org/10.1099/jgv.0.001121>
46. M. Lim, Y.L. Lee, Y. Zhang, J.J, Photodynamic inactivation of viruses using upconversion nanoparticles, *Biomaterials* 33 (2012) 1912-20. <http://dx.doi.org/10.1016/j.biomaterials.2011.11.033>
47. A. Wiehe, J.M. O'Brien, M.O. Senge, Trends and targets in antiviral phototherapy, *Photochem. Photobiol. Sci.* 18 (2019) 2565-612. <http://dx.doi.org/10.1039/c9pp00211a>
48. L. Sobotta, P. Skupin-Mrugalska, J. Mielcarek, T. Goslinski, J. Balzarini, Photosensitizers mediated photodynamic inactivation against virus particles, *Mini. Rev. Med. Chem.* 15 (2015) 503-21. <http://dx.doi.org/10.2174/1389557515666150415151505>
49. M.S. Baptista, J. Cadet, P. Di Mascio, A.A. Ghogare, A. Greer, M.R. Hamblin, C. Lorente, S.C. Nunez, M.S. Ribeiro, A.H. Thomas, M. Vignoni, T.M. Yoshimura, Type I and Type II Photosensitized Oxidation Reactions: Guidelines and Mechanistic Pathways, *Photochem. Photobiol.* 93 (2017) 912-9. <http://dx.doi.org/10.1111/php.12716>
50. M. Wainwright, T. Maisch, S. Nonell, K. Plaetzer, A. Almeida, G.P. Tegos, M.R. Hamblin, Photoantimicrobials-are we afraid of the light? *Lancet Infect. Dis.* 17 (2017) e49-e55. [http://dx.doi.org/10.1016/S1473-3099\(16\)30268-7](http://dx.doi.org/10.1016/S1473-3099(16)30268-7)
51. J.P. Tardivo, M. Wainwright, M.S. Baptista, Local clinical phototreatment of herpes infection in Sao Paulo, *Photodiagnosis Photodyn. Ther.* 9 (2012) 118-21. <http://dx.doi.org/10.1016/j.pdpdt.2012.01.002>
52. M.J. Shikowitz, A.L. Abramson, K. Freeman, B.M. Steinberg, M. Nouri, Efficacy of DHE photodynamic therapy for respiratory papillomatosis: immediate and long-term results, *Laryngoscope* 108 (1998) 962-7. <http://dx.doi.org/10.1097/00005537-199807000-00002>
53. M.J. Casteel, K. Jayaraj, A. Gold, L.M. Ball, M.D. Sobsey, Photoinactivation of hepatitis A virus by synthetic porphyrins, *Photochem. Photobiol.* 80 (2004) 294-300. <http://dx.doi.org/10.1562/2004-04-05-RA-134>
54. E. Blázquez, C. Rodríguez, J. Ródenas, N. Navarro, R. Rosell, S. Pina-Pedrero, J.M. Campbell, M. Sibila, J. Segalés, J. Pujols, J. Polo, UV-C irradiation is able to inactivate pathogens found in commercially collected porcine plasma as demonstrated by swine bioassay, *Vet. Microbiol.* 239 (2019) 108450. <http://dx.doi.org/10.1016/j.vetmic.2019.108450>
55. M. Eickmann, U. Gravemann, W. Handke, F. Tolksdorf, S. Reichenberg, T.H. Müller, A. Seltsam, Inactivation of three emerging viruses - severe acute respiratory syndrome coronavirus, Crimean-Congo haemorrhagic fever virus and Nipah virus - in platelet

concentrates by ultraviolet C light and in plasma by methylene blue plus visible light, *Vox Sang.* 115 (2020) 146-151. <http://dx.doi.org/10.1111/vox.12888>

56. M. Wainwright, Pathogen inactivation in blood products, *Curr. Med. Chem.* 9 (2002) 127-43. <http://dx.doi.org/10.2174/0929867023371355>

57. L. Marciel, L. Teles, B. Moreira, M. Pacheco, L.M. Lourenço, M.G. Neves, J.P. Tomé, M.A. Faustino, A. Almeida, An effective and potentially safe blood disinfection protocol using tetrapyrrolic photosensitizers, *Future Med. Chem.* 9 (2017) 365-79. <http://dx.doi.org/10.4155/fmc-2016-0217>

58. S.J. Wagner, Virus inactivation in blood components by photoactive phenothiazine dyes, *Transfus. Med. Rev.* 16 (2002) 61-6. <http://dx.doi.org/10.1053/tmrv.2002.29405>

59. M.M. Judy, Photodynamic inactivation of enveloped viruses: potential application for blood banking, *J. Clin. Laser Med. Surg.* 8 (1990) 49-52. <http://dx.doi.org/10.1089/clm.1990.8.49>

60. L. Costa, M.A. Faustino, M.G. Neves, A. Cunha, A. Almeida, Photodynamic inactivation of mammalian viruses and bacteriophages, *Viruses* 4 (2012) 1034-74. <http://dx.doi.org/10.3390/v4071034>

61. R.A. Floyd, J.E. Schneider, D.P. Dittmer, Methylene blue photoinactivation of RNA viruses, *Antiviral Res.* 61 (2004) 141-51. <http://dx.doi.org/10.1016/j.antiviral.2003.11.004>

62. L. Costa, J.P. Tomé, M.G. Neves, A.C. Tomé, J.A. Cavaleiro, A. Cunha, M.A. Faustino, A. Almeida, Susceptibility of non-enveloped DNA- and RNA-type viruses to photodynamic inactivation, *Photochem. Photobiol. Sci.* 11 (2012) 1520-3. <http://dx.doi.org/10.1039/c2pp25156f>

63. H. Abe, S.J. Wagner, Analysis of viral DNA, protein and envelope damage after methylene blue, phthalocyanine derivative or merocyanine 540 photosensitization, *Photochem. Photobiol.* 61 (1995) 402-9. <http://dx.doi.org/10.1111/j.1751-1097.1995.tb08630.x>

64. J.E. Schneider Jr, T. Tabatabaie, L. Maidt, R.H. Smith, X. Nguyen, Q. Pye, R.A. Floyd, Potential mechanisms of photodynamic inactivation of virus by methylene blue. I. RNA-protein crosslinks and other oxidative lesions in Q beta bacteriophage, *Photochem. Photobiol.* 67 (1998) 350-7.

65. S. Rywkin, E. Ben-Hur, Z. Malik, A.M. Prince, Y.S. Li, M.E. Kenney, N.L. Oleinick, B. Horowitz, New phthalocyanines for photodynamic virus inactivation in red blood cell concentrates, *Photochem. Photobiol.* 60 (1994) 165-70. <http://dx.doi.org/10.1111/j.1751-1097.1994.tb05085.x>

66. F. Kasermann, C. Kempf, Photodynamic inactivation of enveloped viruses by buckminsterfullerene, *Antiviral Res.* 34 (1997) 65-70. [http://dx.doi.org/10.1016/s0166-3542\(96\)01207-7](http://dx.doi.org/10.1016/s0166-3542(96)01207-7)
67. E. Steinmann, U. Gravemann, M. Friesland, J. Doerrbecker, T.H. Müller, T. Pietschmann, A. Seltsam, Two pathogen reduction technologies--methylene blue plus light and shortwave ultraviolet light--effectively inactivate hepatitis C virus in blood products, *Transfusion* 53 (2013) 1010-8. <http://dx.doi.org/10.1111/j.1537-2995.2012.03858.x>
68. S. Jockusch, D. Lee, N.J. Turro, E.F. Leonar, Photo-induced inactivation of viruses: adsorption of methylene blue, thionine, and thiopyronine on Qbeta bacteriophage, *Proc. Natl. Acad. Sci. USA* 93 (1996) 7446-51. <http://dx.doi.org/10.1073/pnas.93.15.7446>
69. S.J. Wagner, A. Skripchenko, D. Robinette, J.W. Foley, L. Cincotta, Factors affecting virus photoinactivation by a series of phenothiazine dyes, *Photochem. Photobiol.* 67 (1998) 343-9.
70. D. Korneev, O. Kurskaya, K. Sharshov, J. Eastwood, M. Strakhovskaya, Ultrastructural Aspects of Photodynamic Inactivation of Highly Pathogenic Avian H5N8 Influenza Virus, *Viruses* 11 (2019) 10. <http://dx.doi.org/10.3390/v11100955>
71. I.O.L. Bacellar, M.C. Oliveira, L.S. Dantas, E.B. Costa, H.C. Junqueira, W.K. Martins, A.M. Durantini, G. Cosa, P. Di Mascio, M. Wainwright, R. Miotto, R.M. Cordeiro, S. Miyamoto, M.S. Baptista, Photosensitized Membrane Permeabilization Requires Contact-Dependent Reactions between Photosensitizer and Lipids, *J. Am. Chem. Soc.* 140 (2018) 9606-15. <http://dx.doi.org/10.1021/jacs.8b05014>
72. I.O.L. Bacellar, M.S. Baptista, Mechanisms of Photosensitized Lipid Oxidation and Membrane Permeabilization, *ACS Omega* 4 (2019) 21636-46. <http://dx.doi.org/10.1021/acsomega.9b03244>
73. J. Kurman, N. Pastis, S. Murgu, Photodynamic Therapy and Its Use in Lung Disease, *Curr. Pulmonol. Rep.* 8 (2019) 215-21. <http://dx.doi.org/10.1007/s13665-019-00241-y>
74. P. Agostinis, K. Berg, K.A. Cengel, T.H. Foster, A.W. Girotti, S.O. Gollnick, S.M. Hahn, M.R. Hamblin, A. Juzeniene, D. Kessel, M. Korblick, J. Moan, P. Mroz, D. Nowis, J. Piette, B.C. Wilson, J. Golab, Photodynamic therapy of cancer: an update, *CA Cancer J. Clin.* 61 (2011) 250-81. <http://dx.doi.org/10.3322/caac.20114>
75. M.C. Geralde, I.S. Leite, N.M. Inada, A.C. Salina, A.I. Medeiros, W.M. Kuebler, C. Kurachi, V.S. Bagnato, Pneumonia treatment by photodynamic therapy with extracorporeal illumination - an experimental model, *Physiol. Rep.* 5 (2017) 5. <http://dx.doi.org/10.14814/phy2.13190>

76. R.B. Soares, D.H. Costa, W. Miyakawa, M.G.T. Delgado, A.S. Garcez, T.M. Yoshimura, M.S. Ribeiro, S.C. Nunez, Photodynamic Activity on Biofilm in Endotracheal Tubes of Patients Admitted to an Intensive Care Unit, *Photochem. Photobiol.* (2020). <http://dx.doi.org/10.1111/php.13239>
77. N.E. Eleraky, A. Allam, S.B. Hassan, M.M. Omar, Nanomedicine Fight against Antibacterial Resistance: An Overview of the Recent Pharmaceutical Innovations, *Pharmaceutics* 12 (2020) 2. <http://dx.doi.org/10.3390/pharmaceutics12020142>
78. R. Yin, T. Agrawal, U. Khan, G.K. Gupta, V. Rai, Y.Y. Huang, M.R. Hamblin, Antimicrobial photodynamic inactivation in nanomedicine: small light strides against bad bugs, *Nanomedicine* 10 (2015) 2379-404. <http://dx.doi.org/10.2217/nmm.15.67>
79. E. Tegou, M. Magana, A.E. Katsogridaki, A. Ioannidis, V. Raptis, S. Jordan, S. Chatzipanagiotou, S. Chatzandroulis, C. Ornelas, G.P. Tegos, Terms of endearment: Bacteria meet graphene nanosurfaces, *Biomaterials* 89 (2016) 38-55. <http://dx.doi.org/10.1016/j.biomaterials.2016.02.030>
80. Y. Wang, X. Wu, J. Chen, R. Amin, M. Lu, B. Bhayana, J. Zhao, C.K. Murray, M.R. Hamblin, D.C. Hooper, T. Dai, Antimicrobial Blue Light Inactivation of Gram-Negative Pathogens in Biofilms: *In Vitro* and *In Vivo* Studies, *J. Infec.* 213 (2016) 1380-7. <http://dx.doi.org/10.1093/infdis/jiw070>
81. R.M. Amin, B. Bhayana, M.R. Hamblin, T. Dai, Antimicrobial blue light inactivation of *Pseudomonas aeruginosa* by photo-excitation of endogenous porphyrins: *In vitro* and *in vivo* studies, *Lasers Surg. Med.* 48 (2016) 562-8. <http://dx.doi.org/10.1002/lsm.22474>
82. F. Cieplik, A. Späth, C. Leibl, A. Gollmer, J. Regensburger, L. Tabenski, K.A. Hiller, T. Maisch, G. Schmalz, Blue light kills *Aggregatibacter actinomycetemcomitans* due to its endogenous photosensitizers, *Clin. Oral Investig.* 18 (2014) 1763-9. <http://dx.doi.org/10.1007/s00784-013-1151-8>
83. T. Dai, A. Gupta, C.K. Murray, M.S. Vrahas, G.P. Tegos, M.R. Hamblin, Blue light for infectious diseases: *Propionibacterium acnes*, *Helicobacter pylori*, and beyond? *Drug Resist. Updat.* 15 (2012) 223-36. <http://dx.doi.org/10.1016/j.drug.2012.07.001>
84. R.M. Tomb, T.A. White, J.E. Coia, J.G. Anderson, S.J. MacGregor, M. Maclean, Review of the Comparative Susceptibility of Microbial Species to Photoinactivation Using 380-480 nm Violet-Blue Light, *Photochem. Photobiol.* 94 (2018) 445-58. <http://dx.doi.org/10.1111/php.12883>

85. N.S. Soukos, J. Stultz, A.D. Abernethy, J.M. Goodson, Phototargeting human periodontal pathogens *in vivo*, *Lasers Med. Sci.* 30 (2015) 943-52. <http://dx.doi.org/10.1007/s10103-013-1497-9>
86. E. Genina, V. Titorenko, A. Belikov, A. Bashkatov, V. Tuchin, Adjunctive dental therapy via tooth plaque reduction and gingivitis treatment by blue light-emitting diodes tooth brushing, *J. Biomed. Opt.* 20 (2015) 128004. <http://dx.doi.org/10.1117/1.JBO.20.12.128004>
87. S. Ammad, M. Gonzales, C. Edwards, A.Y. Finlay, C. Mills, An assessment of the efficacy of blue light phototherapy in the treatment of acne vulgaris, *J. Cosmet. Dermatol.* 7 (2008) 180-8. <http://dx.doi.org/10.1111/j.1473-2165.2008.00386.x>
88. P. Yang, N. Wang, C. Wang, Y. Yao, X. Fu, W. Yu, R. Cai, M. Yao, 460nm visible light irradiation eradicates MRSA via inducing prophage activation, *J. Photochem. Photobiol. B* 166 (2017) 311-22. <http://dx.doi.org/10.1016/j.jphotobiol.2016.12.001>
89. T. Dai, A. Gupta, Y.Y. Huang, R. Yin, C.K. Murray, M.S. Vrahas, M.E. Sherwood, G.P. Tegos, M.R. Hamblin, Blue light rescues mice from potentially fatal *Pseudomonas aeruginosa* burn infection: efficacy, safety, and mechanism of action, *Antimicrob. Agents Chemother.* 57 (2013) 1238-45. <http://dx.doi.org/10.1128/AAC.01652-12>
90. M. Fukui, M. Yoshioka, K. Satomura, H. Nakanishi, M. Nagayama, Specific-wavelength visible light irradiation inhibits bacterial growth of *Porphyromonas gingivalis*, *J. Periodontal Res.* 43 (2008) 174-8. <http://dx.doi.org/10.1111/j.1600-0765.2007.01009.x>
91. N.S. Soukos, S. Som, A.D. Abernethy, K. Ruggiero, J. Dunham, C. Lee, A.G. Doukas, J.M. Goodson, Phototargeting oral black-pigmented bacteria, *Antimicrob. Agents Chemother.* 49 (2005) 1391-6. <http://dx.doi.org/10.1128/AAC.49.4.1391-1396.2005>
92. D. Kingsley, R. Kuis, R. Perez, I. Basaldua, P. Burkins, A. Marcano, A. Johnson, Oxygen-dependent laser inactivation of murine norovirus using visible light lasers, *Virol. J.* 15 (2018) 117. <http://dx.doi.org/10.1186/s12985-018-1019-2>
93. Y. Wang, C.K. Murray, M.R. Hamblin, D.C. Hooper, T. Dai, Antimicrobial blue light inactivation of pathogenic microbes: State of the art, *Drug Resist. Updat.* 33-35 (2017) 1-22. <http://dx.doi.org/10.1016/j.drug.2017.10.002>
94. R.M. Tomb, M. Maclean, J.E. Coia, E. Graham, M. McDonald, C.D. Atreya, S.J. MacGregor, J.G. Anderson, New Proof-of-Concept in Viral Inactivation: Virucidal Efficacy of 405 nm Light Against Feline Calicivirus as a Model for Norovirus Decontamination, *Food Environ. Virol.* 9 (2017) 159-67. <http://dx.doi.org/10.1007/s12560-016-9275-z>

95. R.M. Tomb, M. Maclean, P.R. Herron, P.A. Hoskisson, S.J. MacGregor, J.G. Anderson, Inactivation of Streptomyces phage C31 by 405 nm light: Requirement for exogenous photosensitizers? Bacteriophage 4 (2014) e32129. <http://dx.doi.org/10.4161/bact.32129>
96. S.E. Bache, M. Maclean, S.J. MacGregor, J.G. Anderson, G. Gettinby, J.E. Coia, I. Taggart, Clinical studies of the High-Intensity Narrow-Spectrum light Environmental Decontamination System (HINS-light EDS), for continuous disinfection in the burn unit inpatient and outpatient settings, Burns 38 (2012) 69-76. <http://dx.doi.org/10.1016/j.burns.2011.03.008>
97. M. Maclean, M.G. Booth, J.G. Anderson, S.J. MacGregor, G.A. Woolsey, J.E. Coia, K. Hamilton, G. Gettinby, Continuous Decontamination of an Intensive Care Isolation Room during Patient Occupancy Using 405 Nm Light Technology, J. Infect. Prev. 14 (2013) 176-81. <http://dx.doi.org/10.1177/1757177413483646>
98. L.J. Murrell, E.K. Hamilton, H.B. Johnson, M. Spencer, Influence of a visible-light continuous environmental disinfection system on microbial contamination and surgical site infections in an orthopedic operating room, Am. J. Infect. Control 47 (2019) 804-10. <http://dx.doi.org/10.1016/j.ajic.2018.12.002>
99. D. Irving, D.A. Lamprou, M. Maclean, S.J. MacGregor, J.G. Anderson, M.H. Grant, A comparison study of the degradative effects and safety implications of UVC and 405 nm germicidal light sources for endoscope storage, Polym. Degrad. Stabil. 133 (2016) 249-54. <http://dx.doi.org/10.1016/j.polymdegradstab.2016.09.006>
100. W. Liu, H. Li, COVID-19: Attacks the 1-Beta Chain of Hemoglobin and Captures the Porphyrin to Inhibit Human Heme Metabolism, ChemRxiv (2020). <http://dx.doi.org/10.26434/chemrxiv.11938173>
101. R. Read, Flawed methods in “COVID-19: Attacks the 1-Beta Chain of Hemoglobin and Captures the Porphyrin to Inhibit Human Heme Metabolism”, ChemRxiv (2020). <http://dx.doi.org/10.26434/chemrxiv.12120912.v2>
102. M.R. Hamblin, C. Ferraresi, Y.Y. Huang, L. Freitas, J.D. Carroll, Low-level Light Therapy: Photobiomodulation. Bellingham, WA: SPIE Press; 2017.
103. L.F. De Freitas, M.R. Hamblin, Proposed Mechanisms of Photobiomodulation or Low-Level Light Therapy, IEEE J. Sel. Top. Quantum Electron. 22 (2016) 7000417. <http://dx.doi.org/10.1109/JSTQE.2016.2561201>
104. L. Sergio, A.M.C. Thome, L. Trajano, A.L. Mencalha, A.S. da Fonseca, F. de Paoli, Photobiomodulation prevents DNA fragmentation of alveolar epithelial cells and alters the

mRNA levels of caspase 3 and Bcl-2 genes in acute lung injury, *Photochem. Photobiol. Sci.* 17 (2018) 975-83. <http://dx.doi.org/10.1039/c8pp00109j>

105. A.A. de Brito, E.C. da Silveira, N.C. Rigonato-Oliveira, S.S. Soares, M.A.R. Brandao-Rangel, C.R. Soares, T.G. Santos, C.E. Alves, K.Z. Herculano, R.P. Vieira, A. Lino-Dos-Santos-Franco, R. Albertini, F. Aimbire, A.P. de Oliveira, Low-level laser therapy attenuates lung inflammation and airway remodeling in a murine model of idiopathic pulmonary fibrosis: Relevance to cytokines secretion from lung structural cells, *J. Photochem. Photobiol. B* 203 (2020) 111731. <http://dx.doi.org/10.1016/j.jphotobiol.2019.111731>

106. M. da-Palma-Cruz, R.F. da Silva, D. Monteiro, H.M.M.A. Rehim, C.C. Grabulosa, A.P.L. de Oliveira, A. Lino-Dos-Santos-Franco, Photobiomodulation modulates the resolution of inflammation during acute lung injury induced by sepsis, *Lasers Med. Sci.* 34 (2019) 191-9. <http://dx.doi.org/10.1007/s10103-018-2688-1>

107. F.M. de Lima, A.B. Villaverde, R. Albertini, J.C. Corrêa, R.L. Carvalho, E. Munin, T. Araújo, J.A. Silva, F. Aimbire, Dual Effect of low-level laser therapy (LLLT) on the acute lung inflammation induced by intestinal ischemia and reperfusion: Action on anti- and pro-inflammatory cytokines, *Lasers Surg Med* 43 (2011) 410-20. <http://dx.doi.org/10.1002/lsm.21053>

108. M.C. Oliveira Jr, F.R. Greiffo, N.C. Rigonato-Oliveira, R.W. Custódio, V.R. Silva, N.R. Damaceno-Rodrigues, F.M. Almeida, R. Albertini, R.Á. Lopes-Martins, L.V. de Oliveira, Pde. T. de Carvalho, A.P. Ligeiro de Oliveira, E.C. Leal Jr, R.P. Vieira, Low level laser therapy reduces acute lung inflammation in a model of pulmonary and extrapulmonary LPS-induced ARDS, *J. Photochem. Photobiol. B* 134 (2014) 57-63. <http://dx.doi.org/10.1016/j.jphotobiol.2014.03.021>

109. N.M. Burduli, N.G. Piliieva, [Changes in plasma hemostatic parameters under intravascular laser irradiation of blood in patients with community-acquired pneumonia], *Ter. Arkh.* 82 (2010) 36-8.

110. J.L. Costa Carvalho, A.A. de Brito, A.P. de Oliveira, H.C. de Castro Faria Neto, T.M. Pereira, R.A. de Carvalho, E. Anatriello, F. Aimbire, The chemokines secretion and the oxidative stress are targets of low-level laser therapy in allergic lung inflammation, *J Biophotonics* 9 (2016) 1208-21. <http://dx.doi.org/10.1002/jbio.201600061>

111. N.C. Rigonato-Oliveira, A.A. de Brito, L.B. Vitoretti, G. de Cunha Moraes, T. Gonçalves, K.Z. Herculano, C.E. Alves, A. Lino-Dos-Santos-Franco, F. Aimbire, R.P. Vieira, A.P. Ligeiro de Oliveira, Effect of Low-Level Laser Therapy (LLLT) in Pulmonary

Inflammation in Asthma Induced by House Dust Mite (HDM): Dosimetry Study, *Int. J. Inflam.* (2019) 3945496. <http://dx.doi.org/10.1155/2019/3945496>

112. A.D. Liebert, B.T. Bicknell, R.D. Adams, Protein conformational modulation by photons: a mechanism for laser treatment effects, *Med. Hypotheses* 82 (2014) 275-81. <http://dx.doi.org/10.1016/j.mehy.2013.12.009>

113. W. Yu, L.H. Chi, J.O. Naim, R.J. Lanzafame. Improvement of host response to sepsis by photobiomodulation, *Lasers Surg Med* 21 (1997) 262-8. [http://dx.doi.org/10.1002/\(sici\)1096-9101\(1997\)21:3<262::aid-lsm6>3.0.co;2-o](http://dx.doi.org/10.1002/(sici)1096-9101(1997)21:3<262::aid-lsm6>3.0.co;2-o)

114. P. Rola, A. Doroszko, E. Szahidewicz-Krupska, P. Rola, P. Dobrowolski, R. Skomro, A. Szymczyszyn, G. Mazur, A. Derkacz, Low-Level Laser Irradiation Exerts Antiaggregative Effect on Human Platelets Independently on the Nitric Oxide Metabolism and Release of Platelet Activation Markers, *Oxid. Med. Cell. Longev.* (2017) 6201797. <http://dx.doi.org/10.1155/2017/6201797>

115. F.A. Klok, M.J.H.A. Kruip, N.J.M. van der Meer, M.S. Arbous, D.A.M.P.J. Gommers, K.M. Kant, F.H.J. Kaptein, J. van Paassen, M.A.M. Stals, M.V. Huisman, H. Endeman, Incidence of thrombotic complications in critically ill ICU patients with COVID-19, *Thromb. Res.* S0049-3848 (2020) 30120-1. <http://dx.doi.org/10.1016/j.thromres.2020.04.013>

116. P.R. Sekhejane, N.N. Houreld, H. Abrahamse, Irradiation at 636 nm positively affects diabetic wounded and hypoxic cells *in vitro*, *Photomed. Laser Surg.* 29 (2011) 521-30. <http://dx.doi.org/10.1089/pho.2010.2877>

117. N.D. Caputo, R.J. Strayer, R. Levitan, Early Self-Prone in Awake, Non-intubated Patients in the Emergency Department: A Single ED's Experience during the COVID-19 Pandemic, *Acad. Emerg. Med.* (2020). <http://dx.doi.org/10.1111/acem.13994>

118. P. Cassano, M.A. Caldieraro, R. Norton, D. Mischoulon, N.H. Trinh, M. Nyer, C. Dording, M.R. Hamblin, B. Campbell, D.V. Iosifescu, Reported Side Effects, Weight and Blood Pressure, After Repeated Sessions of Transcranial Photobiomodulation, *Photomed. Laser Surg.* 37 (2019) 651-6. <http://dx.doi.org/10.1089/photob.2019.4678>

119. K. Tsen, S.W.D. Tsen, C.L. Chang, C.F. Hung, T.C. Wu, J.G. Kiang, Inactivation of viruses by coherent excitations with a low power visible femtosecond laser, *Viol. J.* (2007) 50. <http://dx.doi.org/10.1186/1743-422X-4-50>

120. K.T. Tsen, S.W. Tsen, Q. Fu, S.M. Lindsay, K. Kibler, B. Jacobs, T.C. Wu, B. Karanam, S. Jagu, R.B. Roden, C.F. Hung, O.F. Sankey, B. Ramakrishna, J.G. Kiang, Photonic approach to the selective inactivation of viruses with a near-infrared subpicosecond fiber laser, *J. Biomed. Opt.* 14 (2009) 064042. <http://dx.doi.org/10.1117/1.3275477>

121. K.T. Tsen, S.W. Tsen, Q. Fu, S.M. Lindsay, Z. Li, S. Cope, S. Vaiana, J.G. Kiang, Studies of inactivation of encephalomyocarditis virus, M13 bacteriophage, and *Salmonella typhimurium* by using a visible femtosecond laser: insight into the possible inactivation mechanisms, J. Biomed. Opt. 16 (2011) 078003. <http://dx.doi.org/10.1117/1.3600771>
122. S.W.D. Tsen, T. Chapa, W. Beatty, K.T. Tsen, D. Yu, S. Achilefu, Inactivation of enveloped virus by laser-driven protein aggregation, J. Biomed. Opt. 17 (2020) 128002. <http://dx.doi.org/10.1117/1.JBO.17.12.128002>
123. S.W. Tsen, D.H. Kingsley, C. Poweleit, S. Achilefu, D.S. Soroka, T.C. Wu, K.T. Tsen, Studies of inactivation mechanism of non-enveloped icosahedral virus by a visible ultrashort pulsed laser, Virol. J. 11 (2014) 20. <http://dx.doi.org/10.1186/1743-422X-11-20>
124. S.W. Tsen, T. Chapa, W. Beatty, B. Xu, K.T. Tsen, S. Achilefu, Ultrashort pulsed laser treatment inactivates viruses by inhibiting viral replication and transcription in the host nucleus, Antiviral Res. 110 (2014) 70-6. <http://dx.doi.org/10.1016/j.antiviral.2014.07.012>
125. S.W. Tsen, D.H. Kingsley, K. Kibler, B. Jacobs, S. Sizemore, S.M. Vaiana, J. Anderson, K.T. Tsen, S. Achilefu, Pathogen reduction in human plasma using an ultrashort pulsed laser, PLoS One 9 (2014) e111673. <http://dx.doi.org/10.1371/journal.pone.0111673>
126. S.W. Tsen, N. Donthi, V. La, W.H. Hsieh, Y.D. Li, J. Knoff, A. Chen, T.C. Wu, C.F. Hung, S. Achilefu, K.T. Tsen, Chemical-free inactivated whole influenza virus vaccine prepared by ultrashort pulsed laser treatment, J. Biomed. Opt. 20 (2015) 051008. <http://dx.doi.org/10.1117/1.JBO.20.5.051008>

Figure legends

Figure 1. Mechanisms of photosensitized oxidation reactions. The photosensitizer (PS) is a molecule capable of absorbing light depending on its specific absorption spectra. Once excited, the PS is converted from the ground state ^1PS to its singlet excited $^1\text{PS}^*$ and triplet excited $^3\text{PS}^*$ states. Via Type I (contact-dependent) reactions both $^1\text{PS}^*$ and $^3\text{PS}^*$ can react directly with O_2 or biomolecules, like carbohydrates, lipids, proteins, or nucleic acids, resulting in the formation of radicals capable of initiating redox chain reactions. Otherwise, $^3\text{PS}^*$ can react with molecular oxygen ($^3\text{O}_2$), via the Type II (energy transfer) reaction, generating the reactive state of singlet oxygen ($^1\text{O}_2$).

Figure 1.

