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# Genetically Determined Serum 25-Hydroxyvitamin D Is Associated with Total, Trunk, and Arm Fat-Free Mass: A Mendelian Randomization Study

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#### **Abstract**

PURPOSE: Low serum vitamin D status has been associated with reduced muscle mass in observational studies although the relationship is controversial and a causal association cannot be determined from such observations. Two-sample Mendelian randomization (MR) was applied to assess the association between serum vitamin D (25(OH)D) and total, trunk, arm and leg fat-free mass (FFM).

METHODS: MR was implemented using summary-level data from the largest genome-wide association studies (GWAS) on vitamin D (n=73,699) and total, trunk, arm and leg FFM. Inverse variance weighted method (IVW) was used to estimate the causal estimates. Weighted median (WM)-based method, and MR-Egger, leave-one-out were applied as sensitivity analysis.

RESULTS: Genetically higher serum 25(OH)D levels had a positive effect on total (IVW = Beta: 0.042, p = 0.038), trunk (IVW = Beta: 0.045, p = 0.023) and arm (right arm IVW = Beta: 0.044, p = 0.002; left arm IVW = Beta: 0.05, p = 0.005) FFM. However, the association with leg FFM was not significant (right leg IVW = Beta: 0.03, p = 0.238; left leg IVW = Beta: 0.039, p = 0.100). The likelihood of heterogeneity and pleiotropy was determined to be low (statistically non-significant), and the observed associations were not driven by single SNPs. Furthermore, MR pleiotropy residual sum and outlier test did not highlight any outliers.

CONCLUSIONS: Our results illustrate the potentially causal, positive effect of serum 25(OH)D concentration on total, trunk and upper body appendicular fat-free mass.

Key words: Muscle mass, Mendelian randomization, 25-Hydroxyvitamin D, 25(OH)D, lean mass, fat-free mass.

## Introduction

itamin D is an essential nutrient for human health with roles in multiple biological pathways and low vitamin D status is associated with multiple chronic diseases (1) as well as being associated with musculoskeletal health (2, 3) highlighting this nutrient's significance in the global burden of disease. However, up to 40% of the European population may suffer from vitamin D insufficiency (serum 25-hydroxy vitamin D [25(OH)D] concentration <50 nmol/L) (4) and vitamin D deficiency (25(OH)D concentration <30 nmol/L)is widespread enough to be considered a global health issue (4-6).

Loss of muscle mass directly affects muscle strength and physical function and as such, sarcopenia, the progressive loss of muscle mass and strength in aging, and frailty (7, 8). Furthermore, muscle mass loss has been associated with a multitude of chronic conditions including cardiovascular disease (CVD) (9), type 2 diabetes mellitus (T2DM) (10), increased risk of falls and fractures (11), cognitive decline and depression (12, 13), and all-cause mortality (14). Older adults may spend more time indoors due to poor mobility/reduced muscle function which can further lead to an elevated risk of vitamin D inadequacy (15, 16), leading to a vicious cycle of vitamin D deficiency and loss of muscle mass.

Epidemiological studies suggest an association between low vitamin D status and reduced muscle mass (3, 17, 18) although some studies have found no such association (19, 20). However, such studies are limited as observational data cannot determine whether an association is causal. Mendelian randomization (MR) analysis uses functional polymorphisms (single nucleotide polymorphisms (SNPs)) associated with specific changes in exposures (in this case, serum 25(OH) D) as genetic instruments to determine whether the risk factor is a cause of the disease (21). A major advantage of MR analysis is that they are considerably less prone to confounding, residual bias, and reverse causation than conventional riskfactor epidemiology (22). MR analysis may also circumvent the financial, logistical and ethical limitations of randomised controlled trials (RCTs) and additionally, the data from such studies can inform the design of pilot RCTs and clinical trials by providing information for the potential magnitude of effect of nutrients on a given outcome in specific populations (23).

In the present study, we used MR analysis to determine whether a potential causal relationship exists between serum 25(OH)D concentration and total, trunk, arm and leg fat-free mass (FFM).

## **Methods**

# Study design

A two-sample MR study design was used. In a 2-sample MR, the ssummary statistics are provided from various studies for the association of the genetic instruments with the exposure and

| Table 1. Summary results of the 6 genetic loci associated with serum vitamin D |               |        |        |    |    |      |           |  |  |
|--|---------------|--------|--------|----|----|------|-----------|--|--|
| SNP  | Nearest gene  | GX     | GX SE  | EA | OA | EAF  | p-value   |  |  |
| rs3755967  | GC            | -0.089 | 0.0023 | T  | C  | 0.28 | 4.74E-343 |  |  |
| rs10741657   | CYP2R1        | 0.031  | 0.0022 | A  | G  | 0.4  | 2.05E-46  |  |  |
| rs12785878   | NADSYN1/DHCR7 | 0.036  | 0.0022 | T  | G  | 0.75 | 3.80E-62  |  |  |
| rs10745742   | AMDHD1        | 0.019  | 0.002  | T  | C  | 0.4  | 2.10E-20  |  |  |
| rs8018720  | SEC23A        | -0.019 | 0.0027 | C  | G  | 0.82 | 1.11E-11  |  |  |
| rs17216707   | CYP24A1       | 0.026  | 0.0027 | T  | C  | 0.79 | 8.14E-23  |  |  |

All serum vitamin D markers were associated at genome-wide significance (p < 5 x  $10^{-8}$ ); EA: effect allele; OA: other allele, EAF: effect allele frequency; GX: the per-allele effect on standard deviation units of the telomere length; GX SE: standard error of GX.

outcome. In our study, we obtained the summary statistics from the largest genome wide association studies (GWAS) on serum 25(OH)D (exposure (24)) and FFM (outcome). We applied methods to estimate the unbiased effect of serum 25(OH)D on FFM (total, trunk, arms and legs,).

# Genetic predictors of exposures

We used six SNPs identified to be associated with circulating 25(OH)D concentration by the SUNLIGHT meta-GWAS, which are samples of European ancestry (79,366 discovery samples and 42,757 replication samples) (Table 1). GWAS were performed within each cohort according to a uniform analysis plan. Additive genetic models using linear regression on natural-log-transformed 25(OH)D were fitted and a fixed-effects inverse variance weighted (IVW) meta-analysis across the contributing cohorts was performed (24).

# Association of genetic instruments with outcome

SNPs associated with bioelectrical-impedance-measured fat mass and total, trunk, arm and leg FFM were obtained from analyses by Neale Lab (http://www.nealelab.is). We retrieved the association of the six genetic instruments with SNPs associated with bioelectrical impedance measured FFM using data obtained from UK Biobank. Detailed descriptions of the methods used to measure body composition is available on the UK Biobank website (25). Briefly, whole body as well as site-specific (trunk, leg, arm) fat-free mass/fat mass were evaluated with bioelectrical-impedance analysis (Tanita BC418MA body composition analyser). Body composition of a subset of participants was also assessed using dual-energy X-ray absorptiometry (DXA) which showed high correlation with bio-impedance values (fat-free mass: r = 0.96) (25). The UK Biobank is a population-based cohort of approximately 500,000 individuals; 54% are female, the average age is 57 (range 37-73), while 94% report as being White British. Further details on the rationale, design and methodology for UK Biobank can be found elsewhere (26).

# Mendelian Randomisation analysis

We combined the effect of six instruments using inverse variance weighted (IVW) method as implemented in Two Sample MR package of the statistical software, R (R Core Team, Vienna, Austria. https://www.R-project.org/). We assessed the heterogeneity using Q value for IVW. To address the potential effect of pleiotropic variants on the final effect estimate, we conducted sensitivity analysis including weighted median (WM) and MR-Egger. Sensitivity analysis was conducted using the leave-one-out method. The weighted median (WM) estimate, as the weighted median of the SNPspecific estimates, provides correct estimates as long as SNPs accounting for ≥50% of the weight are valid instruments. WM MR allows some variants to be invalid instruments provided at least half are valid instruments. It uses inverse variance weights and bootstrapping to estimate confidence intervals (CIs) (27). MR-Egger has an ability to make estimates by assumption of all SNPs are invalid instruments as long as the assumption of instrument strength independent of direct effect (InSIDE) is satisfied (27). MR-Egger allows free estimation of the intercept, although further assumptions, such as the independence between instrument strength and direct effects, cannot be easily verified. Average directional pleiotropy across genetic variants was assessed from the p-value of the intercept term from MR-Egger (27). Causal estimates in MR Egger are less precise than those obtained by using IVW MR (28). Analysis using MR-Egger has a lower false positive rate but a higher false negative rate than IVW (29).

Further, to assess heterogeneity between individual genetic variant estimates, we used the Q' heterogeneity statistic [30] and the MR pleiotropy residual sum and outlier (MR-PRESSO) test (30). The Q' statistic uses modified 2nd order weights that are a derivation of a Taylor series expansion and take into account uncertainty in both numerator and denominator of the instrumental variable ratio (this eases the no-measurement-error [NOME] assumption) (30). The MR-PRESSO framework relies on the regression of variant-outcome associations on variantexposure associations and implements a global heterogeneity test by comparing the observed distance (residual sums of squares) of all variants to the regression line with the distance expected under the null hypothesis of no pleiotropy (31). In case of evidence of horizontal pleiotropy, the test compares individual variants expected and observed distributions to identify outlier variants. Further we applied on MR-Robust Adjusted Profile Score (RAPS) this method is able to correct for pleiotropy using robust adjusted profile scores. We consider as results, causal estimates that agreed in direction and magnitude across MR methods, pass nominal significance in

Table 2. Results of the Mendelian Randomization (MR) analysis for effects of serum vitamin D on total, trunk, arm and leg fat-free mass

| Exposure                  | Outcome                   | MR       |        |        | Heterogeneity |          |        | Pleiotropy |           |       |       |
|---------------------------|---------------------------|----------|--------|--------|---------------|----------|--------|------------|-----------|-------|-------|
|                           |                           | Method   | beta   | SE     | p             | Method   | Q      | P-value    | Intercept | SE    | p     |
| Vitamin D (Serum 25(OH)D) | Total fat-free mass       | MR Egger | 0.019  | 0.039  | 0.657         | MR-Egger | 11.018 | 0.026      | 0.001     | 0.002 | 0.503 |
|                           |                           | WM       | 0.031  | 0.015  | 0.029         |          |        |            |           |       |       |
|                           |                           | IVW      | 0.042  | 0.02   | 0.038         | IVW      | 12.506 | 0.029      |           |       |       |
|                           |                           | RAPS     | 0.036  | 0.016  | 0.03          |          |        |            |           |       |       |
|                           | Trunk fat-free mass       | MR Egger | 0.037  | 0.039  | 0.406         | MR-Egger | 11.479 | 0.022      | 0.0004    | 0.002 | 0.817 |
|                           |                           | WM       | 0.039  | 0.015  | 0.008         |          |        |            |           |       |       |
|                           |                           | IVW      | 0.045  | 0.02   | 0.023         | IVW      | 11.655 | 0.04       |           |       |       |
|                           |                           | RAPS     | 0.039  | 0.017  | 0.019         |          |        |            |           |       |       |
|                           | Arm fat-free mass (right) | MR Egger | 0.042  | 0.029  | 0.225         | MR-Egger | 6.415  | 0.17       | 0.0001    | 0.001 | 0.94  |
|                           |                           | WM       | 0.042  | 0.014  | 0.003         |          |        |            |           |       |       |
|                           |                           | IVW      | 0.044  | 0.015  | 0.002         | IVW      | 6.425  | 0.267      |           |       |       |
|                           |                           | RAPS     | 0.042  | 0.014  | 0.002         |          |        |            |           |       |       |
|                           | Arm fat-free mass (left)  | MR Egger | 0.033  | 0.0349 | 0.398         | MR-Egger | 8.746  | 0.068      | 0.001     | 0.002 | 0.589 |
|                           |                           | WM       | 0.041  | 0.015  | 0.007         |          |        |            |           |       |       |
|                           |                           | IVW      | 0.05   | 0.018  | 0.005         | IVW      | 9.499  | 0.091      |           |       |       |
|                           |                           | RAPS     | 0.046  | 0.016  | 0.005         |          |        |            |           |       |       |
|                           | Leg fat-free mass (right) | MR Egger | -0.025 | 0.0401 | 0.561         | MR-Egger | 10.775 | 0.029      | 0.003     | 0.002 | 0.171 |
|                           |                           | WM       | 0.015  | 0.0152 | 0.334         |          |        |            |           |       |       |
|                           |                           | IVW      | 0.03   | 0.0258 | 0.238         | IVW      | 18.269 | 0.003      |           |       |       |
|                           |                           | RAPS     | 0.022  | 0.021  | 0.281         |          |        |            |           |       |       |
|                           | Leg fat-free mass (left)  | MR Egger | -0.008 | 0.0382 | 0.838         | MR-Egger | 9.807  | 0.044      | 0.003     | 0.002 | 0.215 |
|                           |                           | WM       | 0.022  | 0.0148 | 0.143         |          |        |            |           |       |       |
|                           |                           | IVW      | 0.039  | 0.0234 | 0.1           | IVW      | 15.108 | 0.009      |           |       |       |
|                           |                           | RAPS     | 0.031  | 0.019  | 0.098         |          |        |            |           |       |       |

25(OH)D: 25-hydroxy vitamin D; WM: weighted median; IVW: inverse variance weighted; SE: standard error; beta: beta-coefficients; MR: Mendelian randomization; RAPS: robust adjusted profile score

IVW MR, and did not show evidence of bias from horizontal pleiotropy using heterogeneity tests. We used R version 3.4.2 (R Core Development Team 2017).

The MR studies assume that the SNPs (instrumental variables) are associated with the outcome only via the exposure (32), so we performed sensitivity analysis excluding SNPs with potentially pleiotropic effects. To assess the instrumental variable analysis "exclusion-restriction" assumption we used Ensembl release (http://useast.ensembl.org/index.html). Ensembl contains a base of SNP phenotypes.

### **Ethics**

This investigation uses published or publicly available summary data with no involvement of participants in the study. No original data were collected for this manuscript. Ethical approval for each of the studies included in the investigation can be found in the original publications (including informed consent from each subject).

#### **Results**

In total, 6 SNPs were identified as instrumental variables for serum 25(OH)D, none of which were significantly associated

with FFM. A list of all SNP associations is shown in Table 1. The results of MR analysis, displayed as beta-coefficient for interested outcomes per increase in serum 25(OH) D, demonstrate a positive and statistically significant effect on total FFM (MR Egger=  $\beta$ :0.019, p= 0.657 and IVW= $\beta$ : 0.042, p= 0.038; respectively, Table 2 and Fig. 1), trunk (MR Egger=  $\beta$ :0.037, p= 0.406 and IVW= $\beta$ : 0.045, p=0.023, respectively, Table 2 and Fig. 1) FFM. This data suggests that each 25 nmol/L increase in serum 25(OH)D is associated with an increase of 0.042 kg of total FFM. Serum 25(OH) D also demonstrated a positive and statistically significant effect on arm FFM (Right arm: MR Egger= β:0.043, p= 0.225 and IVW= $\beta$ : 0.044, p=0.002; Left arm: MR Egger=  $\beta$ :0.033, p= 0.398 and IVW= $\beta$ : 0.05, p=0.005, respectively, Table 2. However, results for leg FFM did not demonstrate a statistically significant effect (Right leg: MR Egger= β: -0.025, SE: 0.04, p= 0.561 and IVW=β: 0.03, SE: 0.026, p=0.238; Left leg: MR Egger=  $\beta$ : -0.008, SE: 0.038, p= 0.838 and IVW= $\beta$ : 0.039, SE: 0.023, p=0.1, respectively, Table 2).

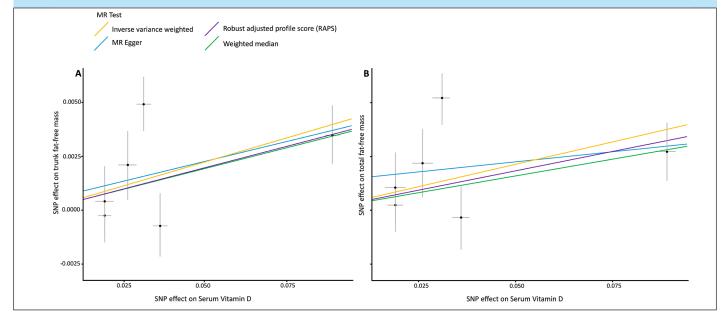


Figure 1. Scatter plots of the association of the effect of SNP-determined serum 25(OH)D on trunk (A) and total (B) fat-free mass

Each black point represents an SNP, plotted by the estimate of SNP on serum 25(OH)D level (x-axis, nmol/L) and the estimate of SNP on fat-free mass (y-axis, kg). The slopes of each line represent the potential causal associations for each method.

The horizontal pleiotropy test, with very negligible Egger regression intercept, also showed a low likelihood of pleiotropy for all our estimations (all p > 0.171, Table 2). Further the result of the MR-RAPS was identical with the IVW prediction, which again indicated a statistically low chance of pleiotropy. Heterogeneity tests highlighted no trace of heterogeneity (Table 2). Furthermore, MR-PRESSO analysis did not indicate any outliers for all estimates. Results of leave-one-out method demonstrated that the links are not driven by any single SNP.

# **Discussion**

Our results illustrate the potentially causal, positive effect of lifetime serum 25(OH)D concentration on total, trunk and arm FFM. These findings are in agreement with a number of cross-sectional, population-based studies, which have shown a positive relationship between serum 25(OH)D status and FFM in a wide range of age groups and clinical populations [3,17,18]. In a study of 100 adolescents (15.1  $\pm$  1.9 y), serum 25(OH)D was positively associated with lean body mass and inversely with fat mass (18). In a cross-sectional study of 127 pre-frail and frail elderly people (79.0  $\pm$  7.8 y) in The Netherlands, Tieland et al. (3) reported that low 25(OH)D status was associated with reduced muscle mass and poorer physical performance (3). Additionally, a meta-analysis of 12 studies with data from 22,590 individuals (mean range 50 – 88 yrs) reported that sarcopenic individuals had lower blood 25(OH)D concentrations than non-sarcopenic controls (17). Conversely, some studies have reported no such association between 25(OH)D and lean body mass (LBM) or FFM (19, 20).

Mechanistically, vitamin D is known to exert its effects of muscle tissue both by regulating expression of target genes via the vitamin D receptor (VDR) and by non-genomic regulation of skeletal muscle intracellular signaling pathways

(33). In animal models, vitamin D supplementation has been demonstrated to activate the mammalian target of rapamycin/ S6 kinase (mTOR/S6K) pathway, which leads to increased muscle protein synthesis (MPS) (34) essential for increases in muscle protein accrual and size (35). Cell culture models have also reported that vitamin D enhances the stimulating effect of leucine and insulin on muscle protein synthesis rates (36) and promotes myogenic differentiation and reduces the expression of myostatin, a known negative regulator of muscle size (37). Vitamin D has also been reported to stimulate the expression of genes involved in the control of cellular growth (33, 38). These varied mechanisms may partly explain the adverse effects of low vitamin D status on muscle mass and function.

The present study did not find a statistically significant relationship between genetically determined serum 25(OH)D concentration and leg FFM. This is not the first study to identify a discrepancy in the relationship between 25(OH)D status with upper and lower body appendicular lean mass. In a study of frail elderly Dutch people (n = 127; mean 79 y) 25(OH)D status was associated with appendicular lean mass (ALM) ( $\beta$ =0.012 [P=0.05]) but was not significantly associated with leg lean mass ( $\beta$ =0.008 [P=0.08]) (3). Furthermore, a cross-sectional study of the association of 25(OH)D status with muscle strength (n = 419; healthy men and women; 20-76 y) has also reported a stronger association between 25(OH)D and muscle strength in the arms compared to the legs (39). One potential explanation for this discrepancy is the reported greater distribution of VDR in type 2 muscle fibres (40) which make up a greater proportion of upper body skeletal muscle (41-44). Vitamin D affects both the diameter and the number of type 2 muscle fibres, which are important for not only young athletes but also the elderly, due to their capacity to reduce the risk of falls, for example (45, 46). Greater expression of VDR has been reported to stimulate muscle hypertrophy through a number of potential mechanisms including increased protein synthesis (47). Furthermore, the greater daily utilization of lower extremities, for example, due to locomotion and bearing the individuals body weight during movement, may provide a superior stimulus for muscle hypertrophy. Further research is clearly needed to elucidate the mechanisms by which vitamin D differentially affects lower and upper body appendicular muscle physiology.

This study highlights the importance of serum vitamin D concentrations in accruing and maintaining FFM, which itself is associated with lower risk of frailty and mortality (7, 11, 14). Addressing vitamin D insufficiency is challenging as the main source of vitamin D in humans is sun exposure (48) which is unlikely to become a widely accepted and implemented strategy. Furthermore, dietary intakes of vitamin D are typically low (49) due to low levels in common foodstuffs (50). Therefore, at a population level, food fortification with vitamin D, and at an individual level, supplementation may be the most effective methods to increase 25(OH)D status to sufficient levels (5).

A major strength of our study was the large sample population study with access to individual participant data of high validity and with the relevant SNPs available for both 25(OH)D serum concentration and FFM. Furthermore, the use of the Mendelian randomisation approach allowed us to examine the potential causal effects of serum 25(OH)D, largely without the disadvantages of confounding or reverse causation.

A potential limitation of this study is the use of segmental bioelectrical impedance analysis (BIA) as the method for determining FFM in the UK Biobank cohort. The accuracy of BIA measurement is known to be affected by hydration status; however the UK Biobank protocol did not specify any procedures to standardise some determinants of hydration before the assessment. This could potentially lead to inaccuracies in the values attained for FFM (51). Furthermore, evidence suggests that BIA is less accurate at high BMI levels (52) and considering the range of BMI included in the UK Biobank cohort, this should be taken into consideration with these results.

# **Conclusions**

Evidence for a potentially causal association of serum 25(OH)D with total, trunk and arm FFM was found. However, the relationship between serum 25(OH)D and leg FFM was not statistically significant. This finding highlights the importance of maintaining sufficient 25(OH)D status throughout the life course in order to maintain adequate lean mass, a factor associated with multiple chronic disease. Future research should address the causal role and potential mechanisms of serum 25(OH)D on FFM accrual and maintenance as well as the apparent lack of effect on leg FFM.

Availability of data and material: Not applicable.

Code availability: Not applicable.

Consent for publication: All the authors have read and approved the revised manuscript, and they are willing to publish it.

Author contributions: RK drafted the manuscript. MM conceived the study, acquired the data and performed the analyses. RK, MM, MI and IGD interpreted the findings. All authors contributed to critical reading and revision of the draft report. All authors approved the final version to be published.

Conflict of interest: RK has received a speaker honorarium for a symposium hosted by the British Association for Parenteral and Enteral Nutrition. RK has received payment from Myprotein UK for the production of educational media content. MM, MI and IGD declare no conflict of interest.

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