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Original Article

Surface enhancement of Ti–6Al–4V fabricated by selective laser melting on bone-like apatite formation



A.N. Aufa ^a, Mohamad Zaki Hassan ^{a,*}, Zarini Ismail ^b,
 Norhaslinda Harun ^c, James Ren ^d, Mohd Faizal Sadali ^a

^a Razak Faculty of Technology and Informatics, Universiti Teknologi Malaysia, Jalan Sultan Yahya Petra, 54100 Kuala Lumpur

^b Faculty of Medicine and Health Sciences, Universiti Sains Islam Malaysia, Bandar Baru Nilai, 71800 Nilai, Negeri Sembilan

^c Diploma Programme Space, Universiti Teknologi Malaysia, Jalan Sultan Yahya Petra, 54100 Kuala Lumpur

^d School of Engineering, Liverpool John Moores University, James Parson Building, Byrom Street, Liverpool L3 3AF, United Kingdom

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ABSTRACT

Medical grade Ti–6Al–4V implants fabricated using selective laser melting (SLM) are recognized as a commercial biomaterial used for bone repairs and fracture fixation. However, there have been cases of failed bone remodeling and implant infections caused by deficient osteointegration. To improve osteogenesis, the Ti implants are treated by acid-etching. This study focused on the effect of surface treatment by using different percentages of sulfuric acid (H₂SO₄), hydrochloric acid (HCL), and hydrofluoric acid (HF) on the SLM Ti–6Al–4V. The microstructure and surface topography before and after treatment were evaluated. Then, the presence of a thicker anatase layer that was detectable on the surface was also observed by X-ray diffraction (XRD). The apatite-forming capabilities which indicated a sample bioactivity were assessed in simulated body fluid (SBF) for periods of 3 days and 7 days. The apatite formed on the surface of sample and XRD scanning revealed the deposition of Ca/P, suggesting successful bone-like apatite. The study discovered that these surface improvements were appropriate for the SLM Ti–6Al–4V prior to clinical applications and were likely to yield higher levels of osseointegration.

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* Corresponding author.

E-mail address: mzaki.kl@utm.my (M.Z. Hassan).

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1. Introduction

Titanium (Ti) alloy is one of the most commonly used orthopedic implant materials in clinical applications [1]. Among those, Ti–6Al4V is widely utilized in additive manufacturing (AM) as a bioinert substance that can improve osseointegration and enhance the bone-implant contact in open reduction internal fixation (ORIF). Additive manufacturing, specifically selective laser melting (SLM), is a powder bed process that enables irregular and complex-shaped fabrication with high accuracy [2,3]. Therefore, it offers the potential to produce materials that are more precisely suited to anatomical fracture sites than commercially available implants [4]. However, SLM showed surface roughness in the range of 5–20 μm , which may negatively influence tissue regeneration and osseointegration of medical implants that prefer a surface roughness of 3–5 μm for bone remodeling [4,5].

As a result of the poor surface roughness, complications arising from failed bone remodeling after fracture treatment may occur. Bacteria such as *Staphylococcus aureus* (*S. aureus*), *Staphylococcus epidermidis*, *Klebsiella pneumoniae*, and *Pseudomonas aeruginosa* colonize the surface of implant and form biofilms [6]. Whereas, for dental implants, *Prevotella intermedia*, *Porphyromonas gingivalis*, and *Fusobacterium nucleatum* are associated with infection [7]. The formation of biofilms further promotes the adherence of multiple species of bacteria on the surface, and thus can lead to chronic infection [8]. These infections are typically due to slow-growing microorganisms such as *Coagulase-negative staphylococci* (CoNS) that commonly live on a person's skin [9]. Bacteria that contribute to the spread and persistence of infections increase the osteoclastic and decrease the osteoblastic activity, causing bone tissue loss known as osteolysis [10]. Infections caused by *S. aureus* can lead to local inflammation implant-associated osteomyelitis such as erythema, swelling, or drainage of pus. Delaying treatment for these infections often results in pseudoarthrosis and soft tissue calcification due to insufficient bone remodeling [11].

Many attempts have been made to improve the surface of titanium alloys, which were hampered by poor cell adhesion and osseointegration [12–15]. Granato et al. [13] stated that the surface of Ti–6Al–4V treated with acid etching (AE) presented a higher mean bone-to-implant contact (BIC) and bone-area-fraction occupancy (BAFO) after 6 weeks in an *in vivo* animal model. This treatment procedure created micro pits on titanium surfaces, which improved osseointegration, and it was suggested that clinical trials be conducted to assess the long-term efficacy of various implant bulk materials. Liang et al. [16] also reported a similar finding, whereby a micro–nanocomposite structure on the titanium surface was obtained after the treatment. After 2 weeks of SBF immersion, apatite formed on the implant surface and thus confirming the bioactivity. Feng et al. [14] seeded rat bone marrow mesenchymal stem cells onto the etched Ti–6Al–4V surface and found that the sample promoted proliferation and differentiation due to its excellent wettability. The cell adhesion increased when the AE time was 5 min, and they noticed that the proliferation increased with increasing AE time. Furthermore, Kim et al. [15] demonstrated that the treated surface

SLM Ti–6Al–4V sample exhibited higher hydrophilicity than conventional titanium as a result of roughness enhancement. In addition, Doe et al. [17] inserted the acid-etched implants into rats' long bones and proved no significant toxicity or accumulation of metal ions. This treated Ti–6Al–4V biomaterial was suitable for implantation by offering superior stability and biosafety. Moreover, the substantial improvement of cell attachment on an implant was observed by Sola-Ruiz et al. [18] as a result of the increase of alkaline phosphatase (ALP) and osteocalcin-positive cells following the acid etching.

Surface roughness plays an important role in influencing the cell proliferation and differentiation of SLM Ti–6Al–4V implants. Frequently, modifying this implant could enhance the surface morphology and improve biocompatibility of implant prior to implantation. For example, Jamshidi et al. [19] etched an SLM Ti–6Al–4V sample in Hydrofluoric acid (HF) solution and found the improvement in the cellular affinity and tissue integration. Similarly, Gonzalez et al. [20] demonstrated that acid etching had a greater influence on the surface features of the samples where the osteoblast cells attached to and into the micropores, covering the implant. This osteogenic behavior significantly increases the alkaline phosphatase (ALP) compared to an untreated sample. These micropores can increase osteoblast proliferation, bone-implant contact (BIC) behavior, and bone remodeling, as reported by Xu et al. [21]. Zhang et al. [22] treated the surface of SLM Ti–6Al–4V with a combination of HF and hydrochloric acid (HCL) to evaluate the bioactive response. The treated sample exhibited a super-hydrophilic surface with apatite successfully formed, resulting in excellent biological activity. Tian et al. [23] investigated the effects of AE implant surfaces both *in vitro* and *in vivo*. The findings revealed that a surface with an appropriately sized microgroove/nanopore topography exhibited more satisfactory adhesive and osteogenic efficiency than the conventionally used implant surface in the market. The sample increased the protein expressions of Integrin alpha-2 precursor (ITGA2), phosphatidylinositol 3-kinase (PI3K), and phosphorylated serine/threonine kinase Akt (p-Akt) to enhance osseointegration. The adsorption of proteins onto SLM Ti–6Al–4V was directly related to surface wettability, which was greatly influenced by the surface roughness, as demonstrated by Chen et al. [24]. Soro et al. [25] stated that during SLM, defects were formed that might negatively affect the biological performance of an implant. Therefore, undergoing post-treatment is crucial. After AE, adequate penetration of etchant into the sample was found, thereby reducing the surface defects. Despite the many studies of improving the surface roughness via AE technique, only a limited number of studies have been reported to evaluate the effect of the combination of HF, HCL, and H_2SO_4 on the biological performance of SLM Ti–6Al–4V.

This study aimed to investigate the effect of surface treatment of SLM Ti–6Al–4V. Initially, the SLM Ti–6Al–4V alloy was subjected to acid etching (AE) to improve the surface morphology and roughness. The samples were then characterized via surface profiler, scanning electron microscopy (SEM), and X-Ray Diffraction (XRD). Then, the apatite-forming ability was carried out *in vitro* in SBF to demonstrate the influence of surface treatments on the bioactive performance prior to clinical application.

2. Materials and methods

2.1. Material

The Ti–6Al–4V powder was supplied by SLM Solution Group AG (ASTM F136), and the chemical composition is illustrated in Table 1. The particle size distribution is shown in Fig. 1 after analysis by using a low-vacuum scanning electron microscope LVSEM (M7800F, JOEL Ltd, Tokyo, Japan) at x200 magnification.

The HF and HCL used to treat the sample were supplied by Orioner Hightech Sdn. Bhd. (Setapak, Kuala Lumpur, Malaysia), while H₂SO₄ was purchased from BT Science Sdn. Bhd., (Cheras, Kuala Lumpur Malaysia).

2.2. Fabrication of Ti–6Al–4V by additive manufacturing

The Ti–6Al–4V samples were manufactured by using an SLM 125^{HL} (Estlandring, Lübeck, Germany) machine with a maximum power of 400W. The AutoFab software was used to convert the CAD file into STL format to fabricate the 3D shape of Ti–6Al–4V. The parameters of the SLM were based on the following equation (1):

$$E = \frac{P}{vht} \quad (1)$$

Where, laser power (P), scanning speed (v), hatching distance (h), layer thickness (t), and energy density (E) are all factors. The fabricated samples had dimensions of 2 mm × 2 mm × 2 mm and the setting parameters are tabulated in Table 2.

2.3. Acid etching of SLM-built Ti–6Al–4V

To produce 50 ml of 90% H₂SO₄ from 98% molarity, a 46 ml volume of H₂SO₄ was taken and 4 ml of distilled water was added slowly until the 50 ml mark on the measuring cylinder was reached. Steps were repeated for each of the concentrations as summarized in Table 3. The SLM Ti–6Al4V samples were first washed with distilled water, dried, and immersed in concentrations of the acid for a time of 30min.

2.4. Simulated body fluid (SBF)

The *in vitro* apatite-forming ability of the channel implants was examined by using a simulated body fluid (SBF) with pH value of 7.40 and an ion concentration, as illustrated in Table 4. The channel implants remained in SBF for 3 days and 7 days at 36.5 °C after which they were removed from the SBF, washed with distilled water, and dried. This SBF was prepared according to Kokubo et al. [26] by using 8.035g NaCl, 0.355g NaHCO₃, 0.225g KCl, 0.231g K₂HPO₄·3H₂O, 0.311g MgCl₂·6H₂O,

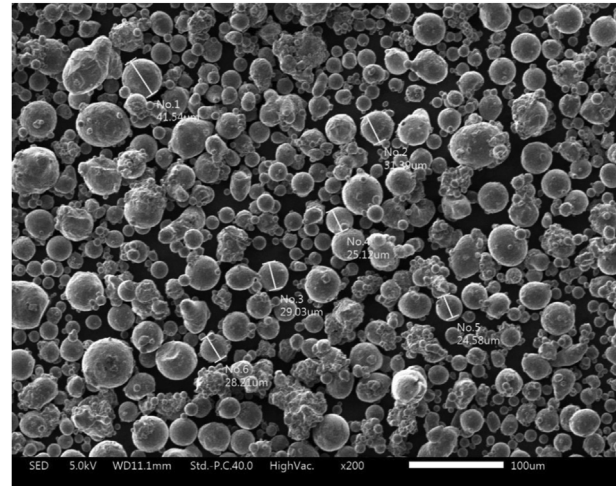


Fig. 1 – The particle size distribution of Ti–6Al–4V.

Table 2 – The input parameters for the fabrication of SLM Ti–6Al–4V.

| Parameter | Unit | Value |
|----------------------|---------|-------|
| Laser Power(P) | Watt, W | 175 |
| Hatching Distance(h) | mm | 0.12 |
| Thickness(t) | μm | 30 |
| Scanning Speed(v) | mm/s | 697.5 |

Table 3 – The AE performed in a mixture of H₂SO₄, HCL and HF solution for 30 min.

| Sample | H ₂ SO ₄ | HCL | HF |
|----------|--------------------------------|-----|----|
| Sample A | 50% | 30% | 2% |
| Sample B | 60% | 25% | 2% |
| Sample C | 70% | 20% | 2% |
| Sample D | 80% | 15% | 2% |
| Sample E | 90% | 10% | 2% |
| Sample F | Untreated | | |

39 ml 1M HCL, 0.292g CaCl₂, 0.072g Na₂SO₄ and 6.118g tris dissolved sequentially together in 1 L of solution based on ISO 23317.

2.5. Surface characterization of SLM fabricated Ti–6Al–4V

2.5.1. Density Measurement

The same samples were used for Archimedes. Measurements were performed at room temperature. The mass of the samples was measured before and after AE for each of the samples by using weighing scales (KERN & SOHN GmbH, Balingen, Germany).

Table 1 – Composition of Ti–6Al–4V.

| Titanium Alloy | Chemical Composition, % | | | | | | | |
|----------------|-------------------------|-----|-------|-------|---------|-------|-------|-------|
| | Al | V | Fe | O | H | C | N | Ti |
| Ti–6Al–4V | 5.5 | 3.5 | <0.30 | <0.20 | <0.0015 | <0.08 | <0.05 | Other |

Table 4 – Ion Concentration of Blood Plasma compared to SBF [26].

| Ions | Ion Concentration (mM) | |
|--------------------------------|------------------------|-------|
| | Blood Plasma | SBF |
| Na ⁺ | 142.0 | 142.0 |
| K ⁺ | 5.0 | 5.0 |
| Mg ²⁺ | 1.5 | 1.5 |
| Ca ²⁺ | 2.5 | 2.5 |
| Cl ⁻ | 103.0 | 103.0 |
| HCO ₃ ⁻ | 27.0 | 27.0 |
| HPO ₄ ²⁻ | 1.0 | 1.0 |
| SO ₄ ²⁻ | 0.5 | 0.5 |
| pH value | 7.2–7.4 | 7.40 |

2.5.2. SEM

The surface characterizations of the SLM Ti–6Al–4V after AE and SBF immersion had been performed involved field emission scanning (FESEM) (M7800F, JOEL Ltd, Tokyo, Japan).

2.5.3. XRD

The crystallographic structures of the sample were investigated by utilizing X-ray diffractometry (Siemens D5000 Aubrey, Texas, USA) by using Cu K α radiation ($\lambda = 0.1542$ nm) with 40 kV and 40 mA, a step size of 0.05° , a counting time

of 80s/step, and a diffraction angle 2θ between 20° and 90° .

2.5.4. Surface roughness

The surface roughness of the top view of the SLM samples was determined by using an Olympus LEXT 3D Measuring Laser Microscope (OLS4000, Shinjuku, Tokyo, Japan). The main source of light in the Olympus LEXT 3D (OLS4000) was a laser diode emitting a light beam from the visible spectrum with a wavelength, $\lambda = 405$ nm.

3. Results and discussion

3.1. Microstructure of untreated SLM Ti–6Al–4V alloy

Figure 2 shows the surface morphology of the untreated SLM Ti–6Al–4V fabricated at a layer thickness of 30 μ m and a hatching distance of 0.12 mm, with a laser scanning velocity of 697.5 mm/s. As can be seen in Fig. 2(a), fine needles existed on the uppermost surface attributed to the extremely high cooling rates of the SLM process. These are known as an α' -martensite microstructure, which are aligned at $\pm 45^\circ$ from the build-up direction and act to restrict the deformation. As a result, the SLM sample could offer significantly higher yield

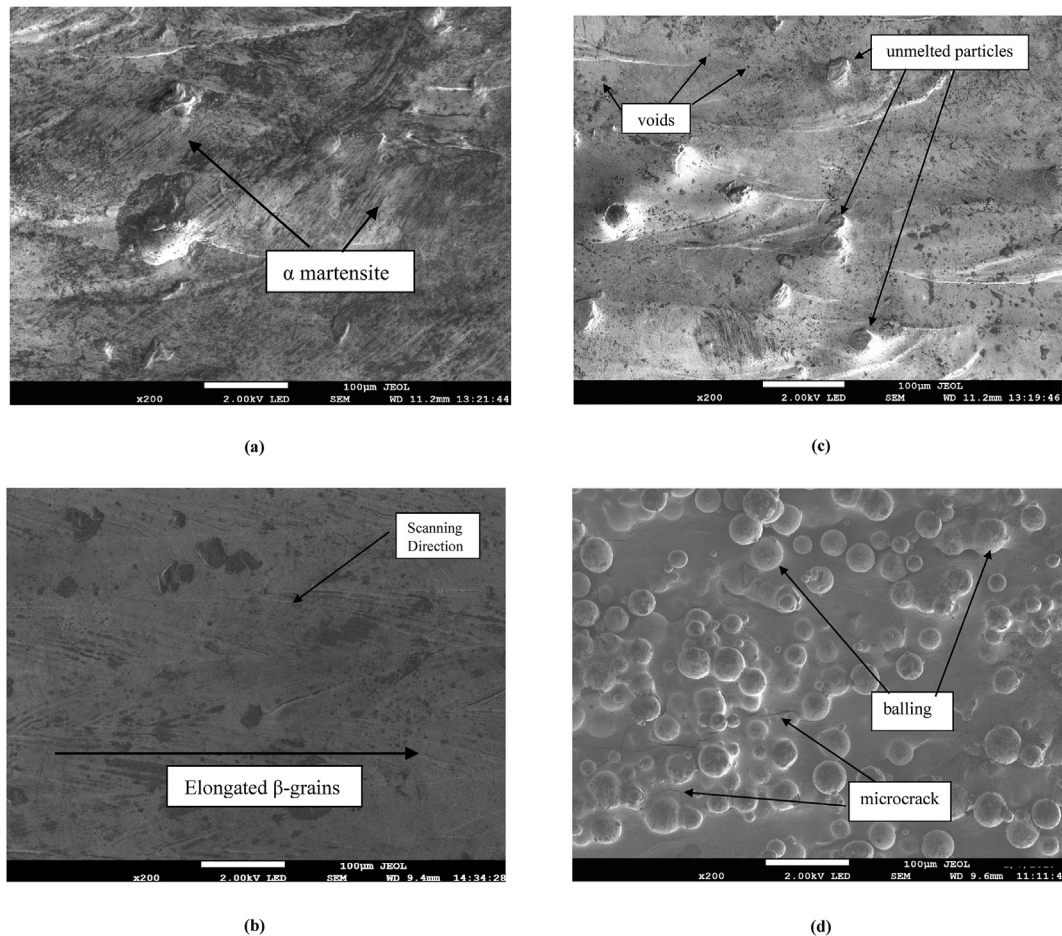


Fig. 2 – Typical micrograph of (a) uppermost surface, (b) β -grains, (c) un-melted particles, and (d) side of untreated SLM Ti–6Al–4V.

strength and ultimate stress. The precipitation of the α' may lead to the diffusion of the excess vanadium from the super-saturated α' [27,28], which α' colony size provided excellent mechanical characteristics and ductile fracture mode in SLM Ti–6Al–4V failure. However, the presence of undesired α martensite caused inadequate ductility in columnar prior- β grains because of their low capacity to tolerate crack initiation and propagation [29,30]. The crack propagations on the uppermost sample were clearly obtained on the SLM samples as shown in Fig. 2(a). These defects were commonly caused by the unstable melt-pools due to the non-optimum process parameters in SLM processing [31].

Figure 2(b) illustrates the untreated surface of SLM Ti–6Al–4V toward the scanning direction. This uppermost view reveals long, columnar grains which are oriented in the building direction. These were identified as prior- β grains which were up to several millimeters in length and grew epitaxially during the scanning process as a result of large temperature gradients in the region induced by repeated remelting and re-heating. The grains were very reliant on heat conduction, and they moved in the same direction as the scanning direction. Therefore, the formation of β -Ti could be caused by lower heat conduction during the melting of the powder playing a significant role in determining the orientation of the grains. These β -Ti precipitates arose from the decomposition of α' martensite during the annealing process. The scanning pattern, local heat transfer, and temperature gradient could all influence the preceding grain boundaries [32,33].

Figure 2(c) demonstrates an untreated SLM surface with a significant proportion of un-melted particles and voids. It happens caused by massive temperature differences in the heat transfer between the powder and solidification of melt pool during fabrication. The SLM Ti–6Al–4V process involves fine metal powders that are melted by a laser beam moving at a very high speed. Therefore, this melting process may create external defects, resulting in some of the powder being blown off. Due to this inherent issue, as-fabricated parts usually have partially melted particles on the surface when the given energy density is inadequate to totally melt the powder. The un-melted powder remained on the layer and caused porosity. This complete coverage of un-melted particles formed on the surface may be attributed to the higher surface roughness of the untreated SLM Ti–6Al–4V [34,35]. In addition, pores (Fig. 2(c)) occurred due to the high cooling rate during the solidification process, which dissolved gas was trapped in the molten pool. When high laser energy was applied to the molten pool, pores were created by gas bubbles confined between the layers [36]. According to Williams et al. [37], larger melt pools of SLM Ti–6Al–4V created greater voids, which might lead to fracture initiation on the surface and lower fatigue life.

Figure 2(d) represents the balling and micro-cracking that exist on the side surface of SLM Ti–6Al–4V. Balling is generally associated with the formation of several metallic agglomerates as spherical or ellipsoidal balls caused by the powder particles adhering to the SLM sample surface. During the layer-by-layer process, the balling phenomenon occurs due to the low level of liquid aggregation with the solid particle, which eventually retards the liquid flow and

particle rearrangement, leading to the formation of different-sized balls on the sample surface. The low energy input and low laser power followed by the high scanning speed cause the formation of balling due to insufficient melting and wetting with the surrounding parts [38]. These phenomena eventually lead to higher surface roughness of the SLM Ti–6Al–4V sample and greatly affect the fatigue performance of the SLM Ti–6Al–4V. The micro-cracks (Fig. 2(d)) initiated by the partially melted particles were caused by the shrinkage rate between the solidifying melt and the powder particles. As a result, micro-cracks occurred between the melt pool and the partially molten powder particles. The micro-cracks formed along the grain boundaries between the α' martensite in the microstructure of the SLM Ti–6Al–4V, which eventually reduced the ductility of an SLM-fabricated sample [39].

3.2. Acid etching of SLM Ti–6Al–4V

In general, AE treatment is used to modify the SLM Ti–6Al–4V surface, which may help lessen the effects of hydrogen generation on titanium embrittlement. Figure 3 summarizes the effect on sample density after AE treatment. As can be seen, after AE treatment for 30 min, the average mass loss ranged from 10% to 30%, and was associated with the removal of un-melted particles. The reduction in mass was more pronounced when a higher acid concentration was used. It was also related to immersion time and was attributed to factors such as surface passivation. The highest weight loss was observed for Sample E, which may be caused by the penetration of H_2SO_4 and HF into the samples due to improved permeability. At a high concentration, the hydrogen generation was negligible, so the structural wettability and absorption exerted the most influence. The decreased sample mass with increasing treatment time resulted in effectively removing the partially sintered powder particles attached to the surface. According to Bezuidenhout et al. [40], after 30 min of AE immersion, the sample mass decreased, and brown nitrogen dioxide (NO_2) gas was emitted. The combined reaction is proposed in Equation (2).

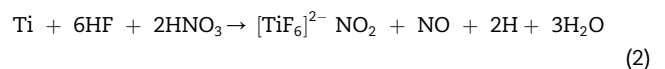


Figure 4 shows the SEM images of Sample A, Sample B, Sample C, Sample D, and Sample E following AE treatment. Overall, this process effectively removes the partly melted and retained metal particles from the surface of the samples. Figure 4 (a) illustrates a rougher layer, protuberances, and a submicron-nanoporous morphology texture, which could be related to the formation of an oxide film and irregular coarseness that form on the outer surface. This specific morphology positively affects the osseointegration of implantable devices to promote the adhesion of fibrin and osteogenic cells. In addition, a protuberance and spheroid-like shape were observed in Sample B (Fig. 4(b)). It was owing to the adoption of the balling effect that was introduced during the initial fabrication process. In contrast, the presence of micropores and protrusions were obtained in Sample C (Fig. 4(c)) and Sample D (Fig. 4(d)). These micropores were formed because of surface imperfections. A combination of pit and

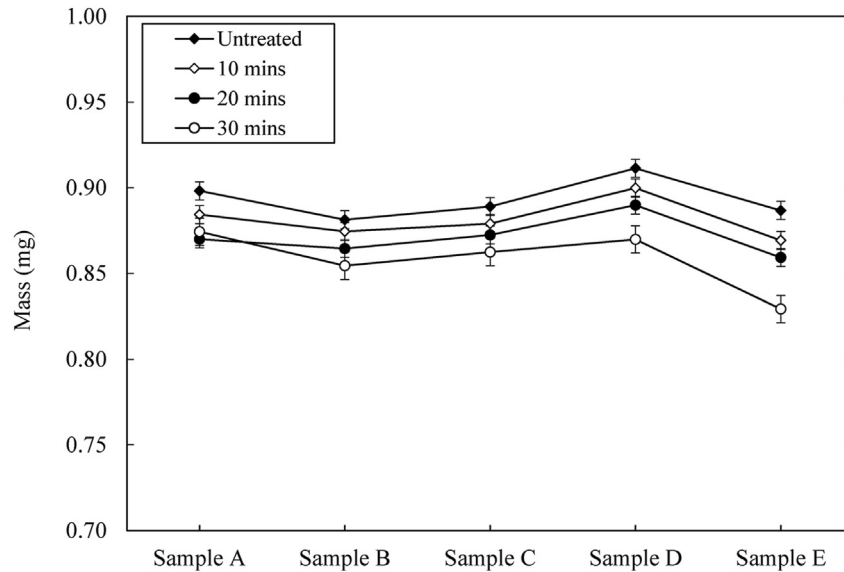


Fig. 3 – Typical mass of SLM Ti-6Al-4V measured following AE treatment.

valley surfaces significantly increases the proliferation and cell viability in biomedical implants. A rougher topographical feature allows protein attachment and improves the bone-implant contact after implantation [41]. Some microcracks were also observed on Sample D, due to corrosive behavior of H_2SO_4 . On the other hand, Sample D shows fewer protrusions with several spots of smooth surface as compared to Sample C. Nonetheless, Sample E (Fig. 4(e)) was significantly smoother after the acid etching treatment. It can be suggested that an increasing concentration of H_2SO_4 resulted in a finer sample surface and removed most of the residual. Some cracks were also noticed due to TiO_2 layer deterioration, mainly originating from the oxidization of the α -phase. Then, the SO_4^{2-} ions that exist on the surface, form a passive film and penetrate the crack. This α -structure might easily dissolve, creating pits and subsequently leading to the expansion of the corrosion area [42].

3.3. Roughness and morphology of acid etched samples

Figure 5 depicts a roughness profile for each of the samples after the AE process. In general, for all the treatments investigated in this study, specimens had a nearly identical surface roughened. In general, the average roughness (R_a) value for the five distinct Ti-6Al-4V specimens was between $1.56\ \mu\text{m}$ and $2.52\ \mu\text{m}$. However, the maximum R_z value ranged between $12.71\ \mu\text{m}$ and $19.30\ \mu\text{m}$. It was attributed to pit-valley creation and the formation of an oxide layer on the sample surfaces [43].

Besides the R_a , R_q , and R_z surface height parameters, it is possible to further characterize the surface roughness by performing a statistical analysis on the roughness data. The skewness (R_{sk}) and the kurtosis (R_{ku}) are relevant surface roughness parameters that allow a deeper understanding of surface morphology. Figure 6 shows the (R_{sk}) and the (R_{ku}) mean values for the roughness profiles of the different Ti-6Al-4V specimens. Based on Fig. 6, it was clear that Samples B, D and E showed R_{sk} to be negative, which

indicated that the AE surface led to a particular topography which deep valleys are predominant in the sample. The sharp peaks formed are effective for load-bearing applications which correlate with this work. The kurtosis (R_{ku}) parameter showed that all the tested specimens (Sample A to Sample E) presented a flat height distribution ($R_{ku} < 4$). These two roughness parameters allowed a more comprehensive understanding of the influence of the topography on the tribological behavior and osseointegration of these Ti-6Al-4V surfaces by indicating the low number of high peaks and predominance of deep valleys.

Nevertheless, Sample C yields the highest R_a , which indicates the beta phase ratio in the samples. It should also be noted that the $\beta > \alpha > \alpha'$ phase structures cause the resistance in the etched surface, with increases in the β phase causing decreased R_a values. Previous studies show that R_a values of $1\ \mu\text{m}$ – $5\ \mu\text{m}$ can promote cell proliferation and osseointegration for biomedical implants [44–46]. In this study, the maximum height of the R_z profile was obtained between 12.7 and $19.3\ \mu\text{m}$. This high surface roughness can be said to be due to the thickness of the oxide layer formed that is generated by the 20% HCl [47]. HCl is a nonoxidizing acid, titanium is only modestly resistive to ferric chloride solutions. However, the R_a value for Sample E increased significantly due to the larger volumes of debris and the number of fractures generated by the corrosive action of 90% H_2SO_4 . The R_a parameter of samples produced decreased after AE with HF as this treatment completely removed the oxides formed during raw powder processing. High roughness can limit cell growth space, forcing deposited cells into their sedimentation stage [48]. The poor surface finish obtained during SLM was caused by the presence of partially unmelted particles and the balling phenomenon, as illustrated in Fig. 2. This caused limited space for cells to adhere, leading to low initial attachment and subsequent proliferation which resulted in the poor biological behavior exhibited by the as-built sample [49].

The native SLM sample yielded an R_a of $5\ \mu\text{m}$ – $20\ \mu\text{m}$. This native SLM Ti-6Al-4V showed that the higher surface

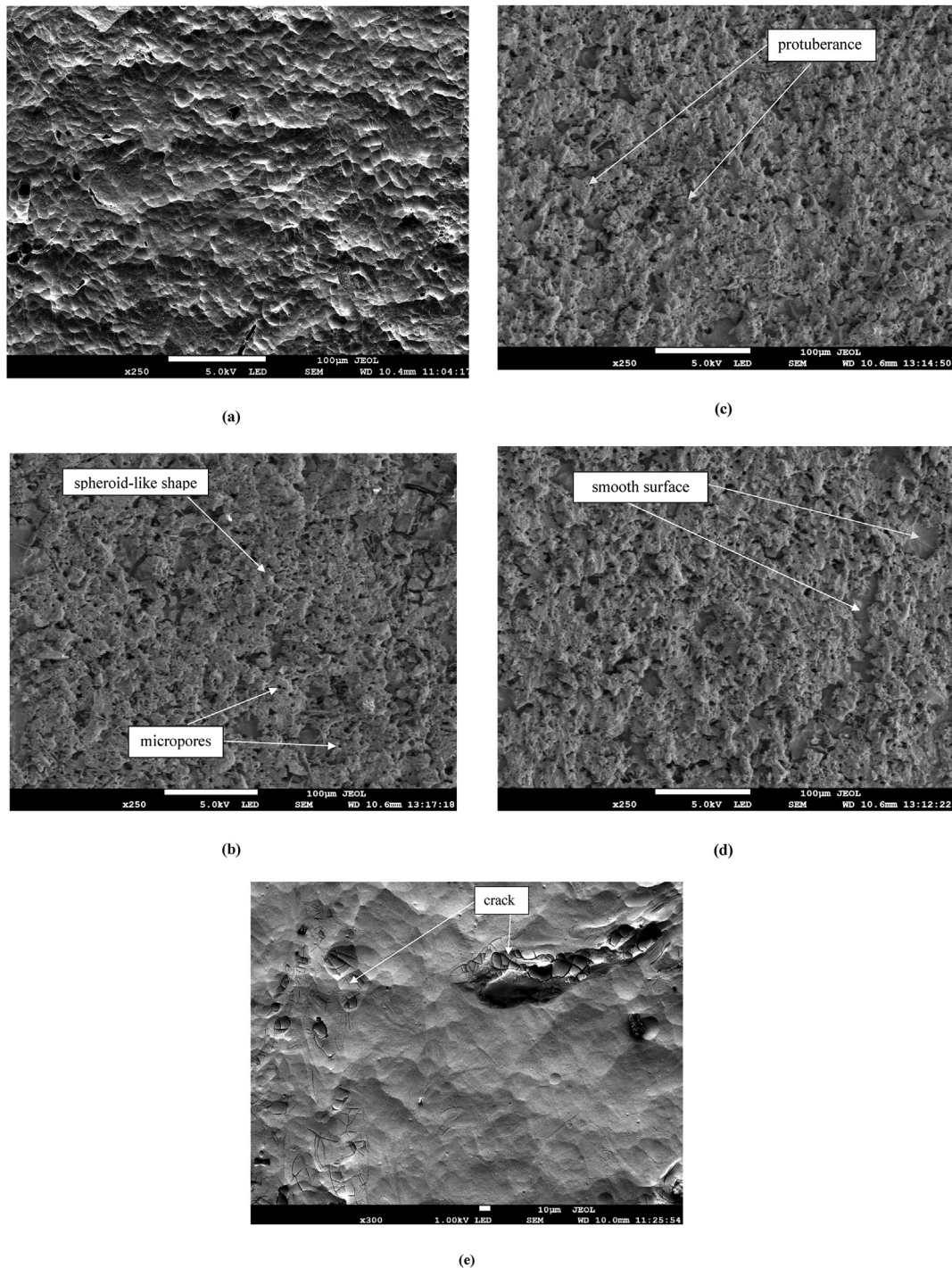


Fig. 4 – Typical SEM images of (a) Sample A, (b) Sample B, (c) Sample C, (d) Sample D and (e) Sample E SLM Ti–6Al–4V after AE treatment.

roughness caused by the processing parameter could lead to multiple stress concentrations, which eventually caused crack initiation on the SLM surface [50]. In addition, the corresponding surface roughness of the SLM sample showed Ra increased as the percentage of H_2SO_4 increased. The increase in roughness was mainly caused by mixtures with high percentages of H_2SO_4/HCl and the subsequent increase in the surface roughness of SLM-Ti by producing a micrometer-scale roughness condition. The acid etched samples caused

hydrogen desorption to occur, which consequently led to the presence of a titanium hydride (TiH_2) on the surface, leading to the increased roughness of SLM-produced samples [51].

3.4. Apatite-forming ability of SLM Ti–6Al–4V in SBF

Specifically, the bioactivity of fabricated SLM Ti–6Al–4V was investigated by determining the apatite layer on the surface of the samples. Samples were then submerged in the SBF

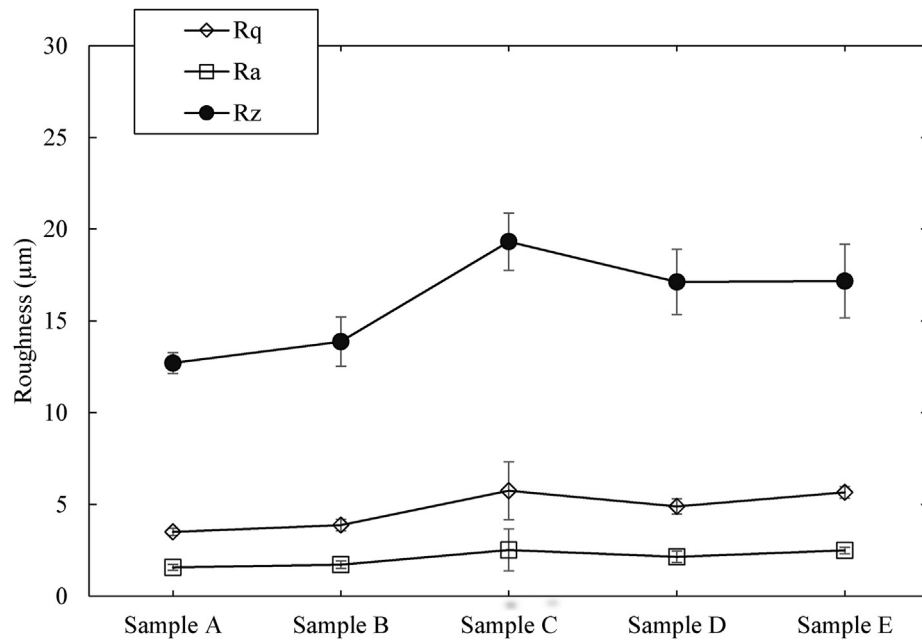


Fig. 5 – Ra, Rq, and Rz surface roughness parameters of SLM Ti–6Al–4V top surface.

solution after being pretreated with the AE procedure. Generally, the surface of all specimens was homogeneously covered by a layer of Ca/P after 3 days of immersion. This Ca/P layer consisted of apatite that formed as a byproduct of SBF immersion. When the treated Ti alloy was immersed in SBF, the Ca^{2+} ions released, and the pH value of the surrounding fluid increased. It is intended to enhance ionic activity and generate Ti–OH groups on SLM samples as shown in Fig. 7. As Ca^{2+} ions accumulate, the Ti surface becomes positive ions and interacts with negatively charged phosphate ions to generate amorphous calcium phosphate. This calcium phosphate is ultimately converted into bone-like crystalline apatite.

Figure 8 illustrates the morphological transformation for Sample A, Sample B, Sample C, Sample D, and Sample E of

SLM Ti–6Al–4V after 3 days of immersion in SBF. As seen in Sample A (Fig. 8(a)), globules apatite mostly covered the outer surfaces. In contrast, the area of Sample B (Fig. 8(b)) was dominated by crystals and precipitates. The precipitate that developed on both surfaces was Ca/P derived from calcium phosphate. Samples A and B clearly had smaller crystals, while Samples C (Fig. 8(c)), Sample D (Fig. 8(d)), and Sample E (Fig. 8(e)) had larger clusters and more precipitates. In addition, microcracks were seen on the Sample D surfaces. An obvious crack was also obtained from Sample E. It can be suggested that increasing H_2SO_4 concentration lengthens the crack on the Ti-alloy surface and tends to lead to corrosion. Nevertheless, this Ti-alloy also exhibited low corrosion resistance due to the presence of α' phases. Shi et al. [52] reported that the corrosion rate of an SLM Ti–6Al–4V submerged in H_2SO_4 was greater than that of other acid solutions. Also, Samples D and Sample E showed relatively small spheroidal structures in the form of elongated aggregates produced on the surface of the samples. This chemical treatment homogeneously improves the micro/nanostructures on the surface of an implant [53,54] and is beneficial for bone-like apatite formation [55].

The apatite-forming activity was further evaluated by the immersion in SBF for 7 days. The results showed a drastic improvement in apatite formation, as illustrated in Fig. 9(a). Figure 9(b) thereby confirmed that the bioactivity occurred which a compact and homogeneous apatite layer was observed as a layer of Ca/P deposit. The Ca/P formation for sample C in Fig. 9(c) was higher than after 3 days immersion in SBF. Whereby, XRD indicated that uniform elongated apatite crystals formed which suggested that the Ra of $2.50\ \mu\text{m}$ favored the formation of Ca/P. As for Fig. 9(d), there were uniform apatite formed on the surface. However, the presence of cracks, which may be attributed to the higher percentage of H_2SO_4 produced pits or microcracks. Nevertheless, similar

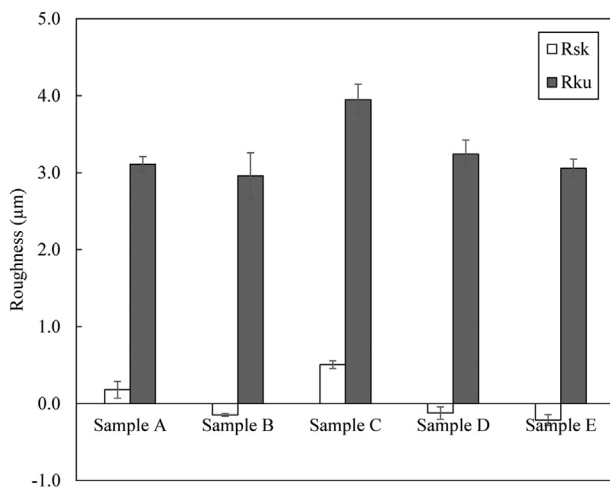


Fig. 6 – Surface roughness of Rsk and Rku of SLM Ti–6Al–4V.

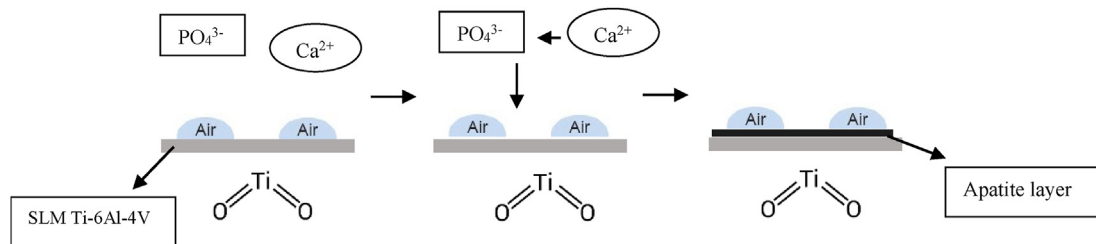


Fig. 7 – Schematic drawing of the mechanism of apatite formation on the SLM Ti–6Al–4V.

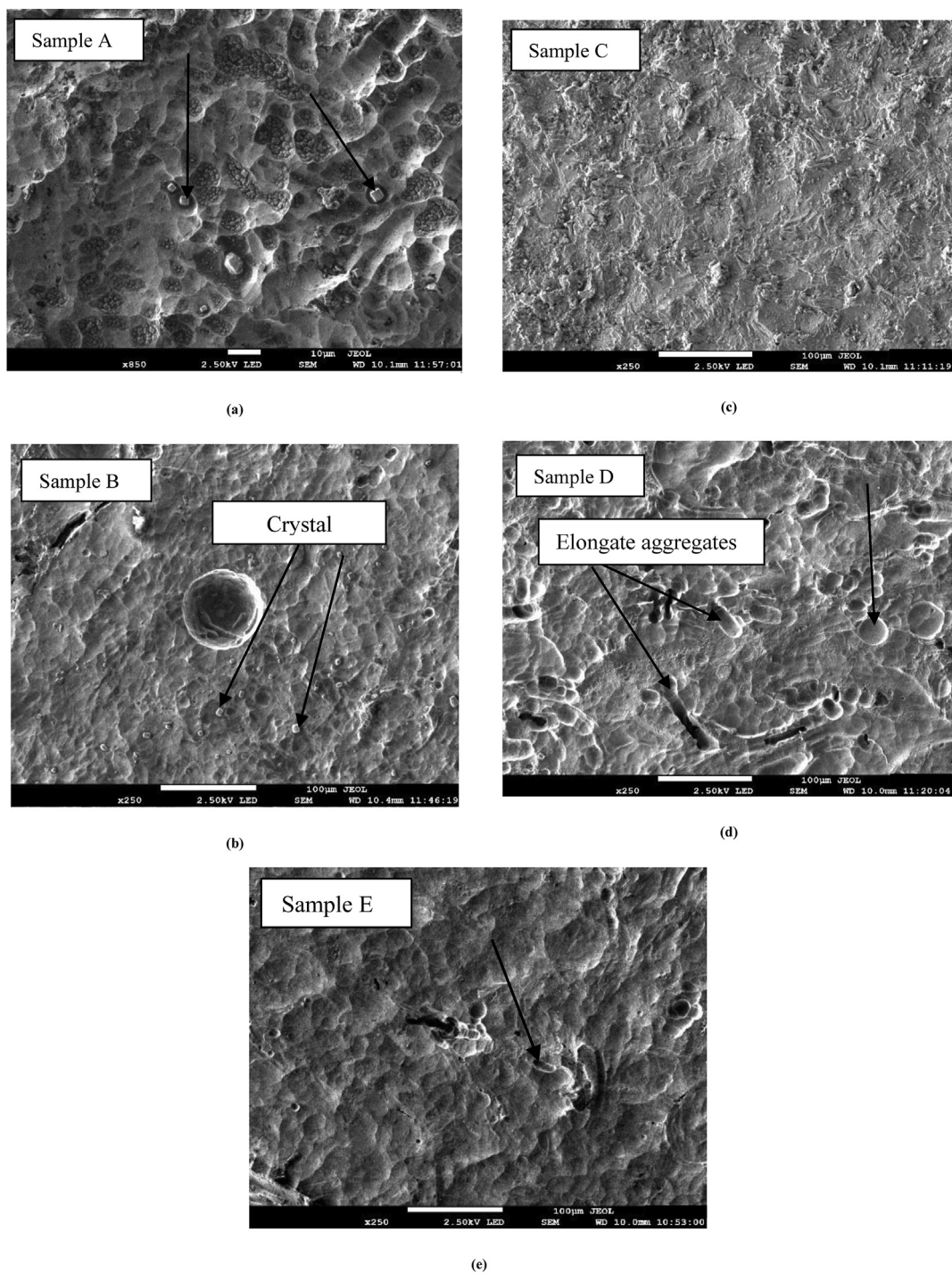


Fig. 8 – Morphological changes for (a) Sample A, (b) Sample B, (c) Sample C, (d) Sample D and (e) Sample E of SLM Ti–6Al–4V after 3 days immersed in SBF.

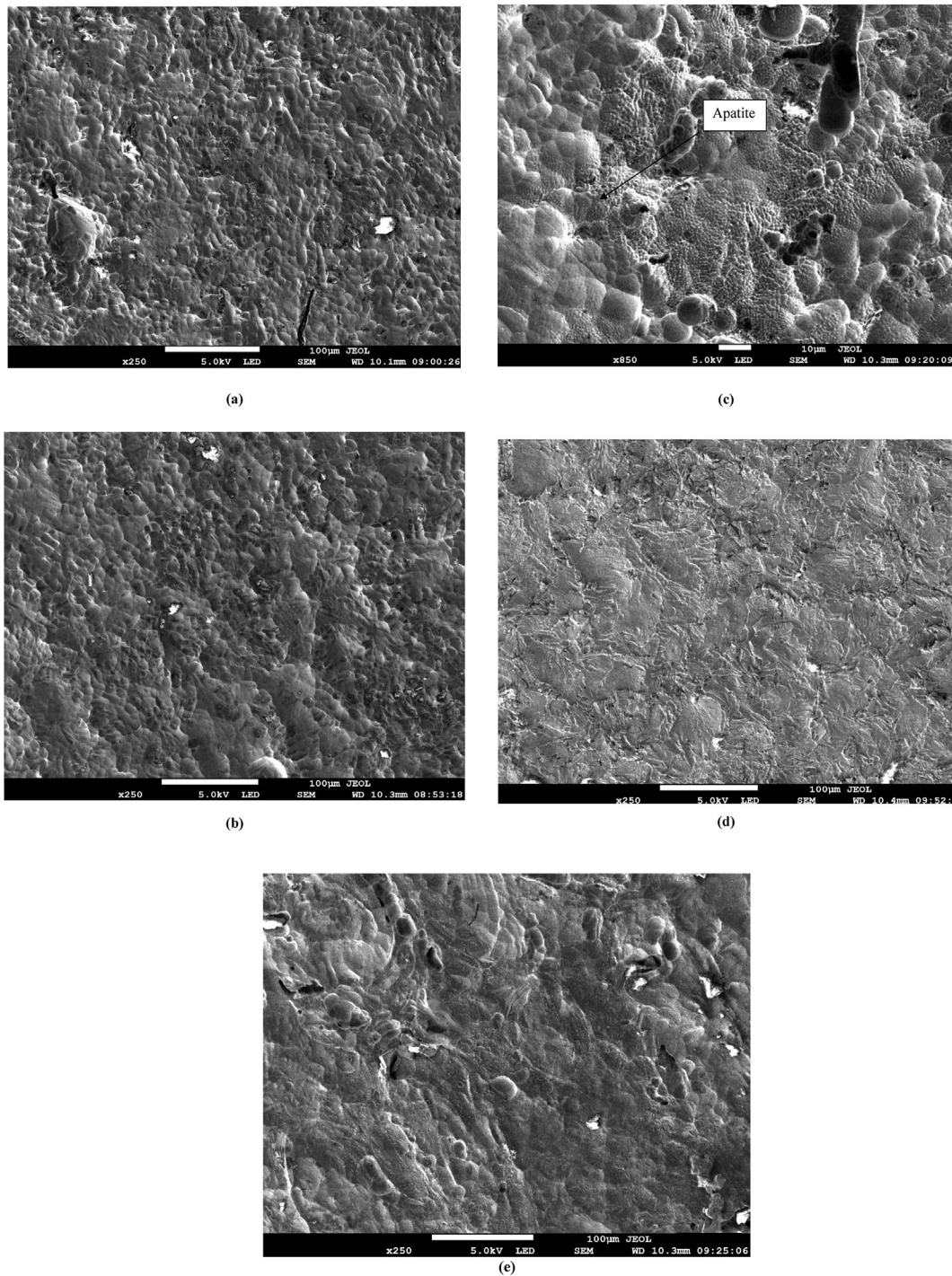


Fig. 9 – Morphological changes of (a) Sample A, (b) Sample B, (c) Sample C, (d) Sample D and (e) Sample E SLM Ti–6Al–4V after 7 days immersed in SBF.

cracks formed on Sample E in Fig. 9(e) after 7 days of SBF. The formation of cracks might be from the SBF that penetrated the porous layer and reacted with the inner compact layer from the corrosive behavior of strong acids from the AE treatment. The corrosive behavior of SBF and found that SLM Ti–6Al–4V showed the lowest corrosion potential as compared to other AM methods such as EBM [56]. This indicated that SLM is more favorable for biomedical applications as compared to other AM methods.

3.5. XRD Crystallinity

XRD analysis was performed on the SLM Ti–6Al–4V samples to assess the influence of treatment and SBF immersion on the crystalline structure of the samples. Figure 10 presents the XRD spectra of Sample B, Sample D and Sample E following SBF immersion for 7 days. From the XRD pattern, α and α' martensite is characterized by their similar structure. However, the presence of a rutile TiO₂ phase in the XRD diffraction peaks

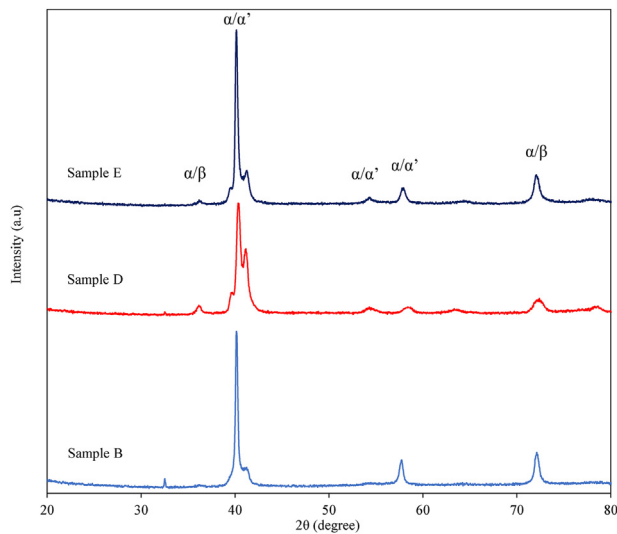
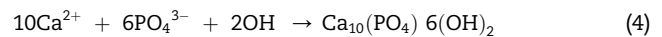


Fig. 10 – XRD diffraction pattern of α , α' , and α/β of SLM Ti–6Al–4V following immersion in SBF for 7 days.

(38° – 43°) for all the samples is attributed to higher heat during the SLM process [57]. Therefore, it was difficult to distinguish between the α and α' phases because they had the same crystal structure and similar lattice parameters. From the figure, XRD peaks were clearly visible in Sample A and Sample D, and the low intensity of these peaks in Sample E was related to the small thickness of the oxide layer. Sample D shows an intensity increase at the peaks, indicating the thickening of the apatite layers after soaking in SBF for 7 days. The low-intensity XRD peaks also observed in Sample D were assigned to anatase TiO_2 ($2\theta = 32.52^\circ$ and 85.40°). The anatase layer thickness was estimated to be about 25 nm–50 nm [58]. On the other hand, the TiO_2 stimulated the apatite structure after the SBF immersion in AM Ti–6Al–4V. The SLM process was characterized by a fast-cooling rate, which will influence the microstructure of Ti–6Al–4V by increasing the needle-shaped β phase which was significant in Sample B. The SLM fast cooling rate promoted a transformation from β phase to the martensitic α' phase. As for the β phase, it was more favorable than the α phase for assisting nucleation and growth of the apatite structure on the surface due to their different crystal structures. During the AE, the passive film coating of SLM Ti–6Al–4V was completely dissolved [as based on equation (3)], which suggested that the present oxide layer was thinner as compared to the untreated sample [59–61]. After soaking in SBF for 7 days, the ions from the titanate layer exchanged with the H_3O^{3+} leading to the formation of Ti–OH groups on the surface of the treated sample. This influenced the ability for apatite formation due to the presence of the rutile oxide phase (TiO_2) and increased the corrosion resistance of the SLM Ti–6Al–4V which prevents the release of ions into the body fluid [62]. The XRD pattern of the untreated sample did not reveal the formation of any oxide phase on its surface since no characteristic peaks of oxide phases were observed. The TiOH is incorporated with the Ca^{2+} , the byproduct of SBF, to provide the nuclei for apatite formation. These negatively charged PO_4^{3-} ions form a Ca–P layer which crystallized into bone-like apatite as suggested by the following equation (4).



4. Conclusion

In this study, the SLM Ti–6Al–4V was treated by using acid etchings at different percentage concentrations. The amount of apatite formed for each sample was then determined by immersing them in SBF. The main findings of this research are:

1. The as-built Ti–6Al–4V showed an out-of-equilibrium microstructure with very fine α' martensitic laths, unmelted particles, and crystallographic defects. After the AE process, there was improvement in the surface of the SLM Ti–6Al–4V with fewer crystallographic defects. Also, β grain boundaries were not observed.
2. A higher surface roughness resultant from balling, voids, staircase-chapping, and unmelted particles is mostly due to layer-by-layer fabrication of Ti–6Al–4V parts. The AE reduced the defects and then formed a uniform and relatively smooth titanium surface. Still, treating the samples with a high concentration of H_2SO_4 makes the samples corrode more quickly. This causes pits and cracks to form, which reduces the formation of bone-like apatite in SBF.
3. After the samples were treated by acid etching, the presence of a thicker anatase layer and the formation of a stable titanium oxide layer were detectable by X-ray diffraction, indicating good bioactivity.
4. After soaking in SBF, thick apatite layers consisting of Ca/P formed on the surface. The amount of apatite formed increased continuously under the SEM observation with the duration of time soaking

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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