

## LJMU Research Online

Murtagh, CF, Hall, EC, Brownlee, TE, Drust, B, Williams, AG and Erskine, RM

The genetic association with athlete status, physical performance and injury risk in soccer.

http://researchonline.ljmu.ac.uk/id/eprint/19704/

Article

**Citation** (please note it is advisable to refer to the publisher's version if you intend to cite from this work)

Murtagh, CF, Hall, EC, Brownlee, TE, Drust, B, Williams, AG and Erskine, RM (2023) The genetic association with athlete status, physical performance and injury risk in soccer. International Journal of Sports Medicine. ISSN 0172-4622

LJMU has developed LJMU Research Online for users to access the research output of the University more effectively. Copyright © and Moral Rights for the papers on this site are retained by the individual authors and/or other copyright owners. Users may download and/or print one copy of any article(s) in LJMU Research Online to facilitate their private study or for non-commercial research. You may not engage in further distribution of the material or use it for any profit-making activities or any commercial gain.

The version presented here may differ from the published version or from the version of the record. Please see the repository URL above for details on accessing the published version and note that access may require a subscription.

For more information please contact <a href="mailto:researchonline@ljmu.ac.uk">researchonline@ljmu.ac.uk</a>

http://researchonline.ljmu.ac.uk/

### **Accepted Manuscript**

# **International Journal of Sports Medicine**

# The genetic association with athlete status, physical performance and injury risk in soccer

Conall F Murtagh, Elliott C Hall, Thomas E Brownlee, Barry Drust, Alun G Williams, Robert M Erskine.

Affiliations below.

DOI: 10.1055/a-2103-0165

Please cite this article as: Murtagh C F, Hall E C, Brownlee T E et al. The genetic association with athlete status, physical performance and injury risk in soccer. International Journal of Sports Medicine 2023. doi: 10.1055/a-2103-0165

Conflict of Interest: The authors declare that they have no conflict of interest.

#### Abstract:

The aim of this review was to critically appraise the literature concerning the genetic association with player status, physical performance and injury risk in soccer. The objectives were to provide guidance on which genetic markers could potentially be used as part of future practice in soccer; and to provide direction for future research in this area. The most compelling evidence identified six genetic polymorphisms to be associated with soccer athlete status (ACE I/D; ACTN3 rs1815739; AGT rs699; MCT1 rs1049434; NOS3 rs2070744; PPARA rs4253778), six with physical performance (ACTN3 rs1815739; AMPD1 rs17602729; BDNF rs6265; COL2A1 rs2070739; COL5A1 rs12722; NOS3 rs2070744), and seven with injury risk (ACTN3 rs1815739; CCL2 rs2857656; COL1A1 rs1800012; COL5A1 rs12722; EMILIN1 rs2289360; IL6 rs1800795; MMP3 rs679620). As well as replication by independent groups, large-scale genome-wide association studies are required to identify new genetic markers. Future research should also investigate the physiological mechanisms associating these polymorphisms with specific phenotypes. Further, researchers should investigate the above associations in female and non-Caucasian soccer players, as almost all published studies have recruited male participants of European ancestry. Only after robust, independently replicated genetic data have been generated, can genetic testing be considered an additional tool to potentially inform future practice in soccer.

#### **Corresponding Author:**

Dr. Robert M Erskine, Liverpool John Moores University, School of Sport and Exercise Sciences, Liverpool, United Kingdom of Great Britain and Northern Ireland, R.M.Erskine@ljmu.ac.uk

#### Affiliations:

Conall F Murtagh, Liverpool John Moores University, School of Sport and Exercise Sciences, Liverpool, United Kingdom of Great Britain and Northern Ireland

Conall F Murtagh, Liverpool Football Club and Athletic Grounds Ltd, Sports Science Department, Liverpool, United Kingdom of Great Britain and Northern Ireland

Elliott C Hall, Liverpool John Moores University, School of Sport and Exercise Sciences, Liverpool, United Kingdom of Great Britain and Northern Ireland

#### [...]

Robert M Erskine, University College London, Institute of Sport, Exercise and Health, London, United Kingdom of Great Britain and Northern Ireland

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



The genetic association with athlete status, physical performance and injury risk in soccer

Conall F. Murtagh<sup>1,2</sup>, Elliott C. R. Hall<sup>1,3</sup>, Thomas E. Brownlee<sup>4</sup>, Barry Drust<sup>4</sup>, Alun G. Williams<sup>5,6</sup> and Robert M. Erskine<sup>1,6</sup>

<sup>1</sup>School of Sport and Exercise Sciences, Liverpool John Moores University, Liverpool, UK; <sup>2</sup>Liverpool Football Club, Liverpool, UK; <sup>3</sup>Faculty of Health Sciences and Sport, University of Stirling, Stirling, UK; <sup>4</sup>School of Sport, Exercise and Rehabilitation Sciences, University of Birmingham, Birmingham, UK; <sup>5</sup>Department of Sport and Exercise Science, Manchester Metropolitan University, Manchester, UK; <sup>6</sup>Institute of Sport, Exercise and Health, University College London, London, UK.

#### Address for correspondence:

Dr Rob Erskine, School of Sport and Exercise Sciences, Liverpool John Moores University, Liverpool, L3 3AF, United Kingdom.

#### Email: <u>R.M.Erskine@ljmu.ac.uk;</u>

 C. F. Murtagh ORCID:
 0000-0002-3924-7452

 E. C. R. Hall ORCID:
 0000-0003-0540-0735

 T. E. Brownlee ORCID:
 0000-0002-3355-1867

 B. Drust ORCID:
 0000-0003-2092-6962

 A. G. Williams ORCID:
 0000-0002-8052-8184

 R. M. Erskine ORCID:
 0000-0002-5705-0207

Running title: Genetic variation and football

#### 1 Abstract

The aim of this review was to critically appraise the literature concerning the genetic 2 association with player status, physical performance and injury risk in soccer. The objectives 3 were to provide guidance on which genetic markers could potentially be used as part of future 4 practice in soccer; and to provide direction for future research in this area. The most 5 compelling evidence identified six genetic polymorphisms to be associated with soccer 6 athlete status (ACE I/D; ACTN3 rs1815739; AGT rs699; MCT1 rs1049434; NOS3 rs2070744; 7 PPARA rs4253778), six with physical performance (ACTN3 rs1815739; AMPD1 rs17602729; 8 BDNF rs6265; COL2A1 rs2070739; COL5A1 rs12722; NOS3 rs2070744), and seven with 9 injury risk (ACTN3 rs1815739; CCL2 rs2857656; COL1A1 rs1800012; COL5A1 rs12722; 10 EMILIN1 rs2289360; IL6 rs1800795; MMP3 rs679620). As well as replication by 11 independent groups, large-scale genome-wide association studies are required to identify new 12 genetic markers. Future research should also investigate the physiological mechanisms 13 associating these polymorphisms with specific phenotypes. Further, researchers should 14 investigate the above associations in female and non-Caucasian soccer players, as almost all 15 published studies have recruited male participants of European ancestry. Only after robust, 16 independently replicated genetic data have been generated, can genetic testing be considered 17 an additional tool to potentially inform future practice in soccer. 18

19

Key words: physical performance, SNP, polymorphism, DNA, injuries, football

20

#### 21 Introduction

Human physical performance and injury risk are influenced by various environmental and 22 genetic factors [1, 2]. The heritability of being a high-level athlete across a range of sports is 23 estimated to be 66% [2], which is itself dependent on the heritability of the other phenotypes, 24 including risk of soft-tissue injury [3, 4] and the physiological and anatomical characteristics 25 known to contribute to competitive sport performance [5-8]. Although such characteristics, 26 e.g. strength, power, speed, agility and aerobic capacity [9-17], and injury risk factors, e.g. 27 previous injury, maturity status, playing position, match intensity, etc. [18-24], have been 28 extensively investigated in soccer players, the genetic contribution to high-level soccer 29 success remains unclear. Variation in the sequence of nucleotides within or adjacent to 30 certain genes has the ability to alter the type/abundance/function of the proteins encoded for 31 by those specific genes [25], thus having the potential to affect tissue characteristics [26, 27] 32 and ultimately physical performance [28] or injury risk [29]. Numerous case-control studies 33 have identified genetic variants (e.g. single nucleotide polymorphisms, or SNPs) that 34 distinguish talented soccer players from non-athletes [28, 30-34], while some studies have 35 investigated the association of specific SNPs with physical performance [28, 35-37] and 36 injury risk [22, 38-47] in soccer players. However, some study designs are limited by small 37 sample sizes and/or cohorts of mixed sports (where soccer is but one), which restrict the 38 ability to detect a genetic association due to a lack of statistical power, or may reveal false 39 positives. Such examples can confuse the perceived importance of genetic variation in high-40 level soccer. The purpose of this narrative review was therefore to critically appraise the 41 scientific literature concerning the genetic association with high-level athlete status, physical 42 performance and injury risk in soccer. Having identified robust evidence, the main objectives 43 were to provide guidance for researchers and practitioners on which genetic markers could 44 potentially be used to identify talent and/or manage the training/recovery/injury prevention of 45

3

talented players as part of future practice; and to provide direction for future research in thisarea.

48

#### 49 Literature Search

A literature search was conducted for empirical research studies and review articles using 50 PubMed and SPORTDiscus databases from inception to March 2023. Key terms were 51 searched for within the article title, abstract, and keywords using conjunctions "OR" and 52 "AND" with truncation "\*." Combinations of the following Boolean's phrases comprised the 53 search terms: 'Soccer'; 'Football'; 'Genetic'; 'Genotype'; 'Polymorphism'; 'Power'; 'Speed'; 54 'Sprint'; 'Acceleration'; 'Agility'; 'Strength'; 'Force'; 'Aerobic'; 'Fitness'; 'Endurance'; 55 'Injury'. The included publications met the following criteria: (1) contained relevant data 56 concerning genomics in soccer; (2) included soccer players; and (3) were written in English. 57 Studies were excluded on the basis that they: (1) did not contain any relevant genomics data; 58 and (2) were conference abstracts. Studies were not included/excluded on the basis of soccer 59 athlete status (i.e. standard of competitive soccer player) or the biological sex or geographic 60 ancestry of participants. 61

62

#### 63 The genetic association with soccer athlete status

Numerous case-control studies have associated specific gene variants with soccer athlete status [28, 30-34]. Logically, and all else being equal, a genotype or allele that is more prevalent in high-level soccer players than non-athletes may have enhanced soccer performance or reduced susceptibility to injury, thereby increasing the likelihood of that soccer player becoming successful. "Success" and "high-level athlete status" are defined here as being selected to represent a professional soccer club (including their academy for youth

Perhaps the most investigated gene variant regarding high-level athlete status is the 73 ACTN3 rs1815739 SNP. ACTN3 XX homozygotes cannot produce alpha-actinin-3 [48], a 74 structural sarcomeric protein found in type II skeletal muscle fibres that inhibits the transition 75 of larger, stronger, more powerful type II fibres towards a smaller, weaker more oxidative 76 phenotype [26]. This probably explains the association between this SNP and skeletal muscle 77 fibre type composition, with XX homozygotes presenting with a greater proportion of slow 78 twitch muscle fibres [49, 50]. Type I fibres are smaller and contract slower than type II fibres. 79 and therefore produce less force and power than type II fibres [51, 52]. Therefore, it follows 80 that muscle volume (the main determinant of power [53]), maximum strength and power are 81 all lower in XX homozygotes compared to R-allele carriers, which appears to be the case in 82 young, healthy men [27]. Regarding high-level soccer, evidence suggests that professional 83 male players (n = 60) have a higher frequency of the ACTN3 R allele compared to 123 non-84 athletes [34]. Hence, the ACTN3 rs1815739 SNP could be a determinant of high-level soccer 85 playing status, as it appears to predispose players to greater muscular power performance, 86 although it should be noted that the sample size of this relatively early study was relatively 87 small, thus larger-scale studies are required to confirm this finding. 88

More recently, it has been reported that the frequency distribution of the *ACTN3* rs1815739 R allele (67.5 vs. 60.4%), the angiotensin-I converting enzyme insertion/deletion polymorphism (*ACE* I/D) D allele (65.0 vs. 48.9%), peroxisome proliferator-activated receptor  $\alpha$  (*PPARA*) rs4253778 C allele (24.3 vs. 17.3%) and uncoupling protein 2 (*UCP2*) rs66033955 T allele (44.4 vs. 35.8%) were higher in 246 male soccer players representing clubs in the Russian Premier (*n* = 51), National and Second Division leagues (*n* = 81) and academies (114 youth players aged 10.6  $\pm$  0.1 years), compared to 872 non-athletes [30] (Table 1). As the *ACTN3* R, *ACE* D and *PPARA* C alleles have previously been associated with strength/power phenotypes in over 20 case-control studies, and *UCP2* rs66033955 has been associated with endurance performance [54], these results provide evidence to support the hypothesis that, whilst muscular endurance is important to meet the aerobic demands of high-level soccer, strength, power and speed capabilities may play a more important role in determining match and individual player success [10, 13, 55, 56].

In accordance with such findings in Russian soccer players [30], Juffer and colleagues 102 [32] reported that the ACE I/D II genotype was less common in 54 professional male soccer 103 players compared to 52 endurance athletes [32], thus suggesting that endurance is not as 104 important as strength and power in professional soccer players. However, similar to the study 105 by Santiago and colleagues [34], the sample size of this relatively early study is modest. 106 Interestingly, these findings are somewhat in contrast to the study by Eynon et al. [57], who 107 reported a higher frequency of the *NOS3* rs2070744 C allele in a similar sample (n = 60) of 108 professional male players compared to 100 non-athletes, while the opposing T allele has 109 previously been found to be more common in high-level power-oriented athletes than non-110 athletes [58, 59]. Contrary to the study by Eynon et al. [57], the T allele was found to be more 111 common in 536 high-level academy players than non-athletes [28] (Table 1). The reason for 112 this discrepancy between studies is not clear, especially considering the studies by Eynon et 113 al. [57], Santiago et al. [34] and Juffer et al. [32] all appear to have been performed in the 114 same population (i.e. La Liga soccer players). In further support of the association between 115 strength and power genetic profiles and soccer playing status, academy male soccer players 116 from Italian clubs were associated with a higher frequency of the VDR rs10735810 FF 117 genotype [33], which has previously been associated with greater maximum isometric 118 quadriceps strength in patients with chronic obstructive pulmonary disease and age-matched 119

120 controls [60]. However, a study investigating the role of the *ACE* I/D polymorphism in 199 121 professional male soccer players representing Lithuanian clubs found a lower frequency of 122 *ACE* DD genotype compared to non-athletes, thus suggesting that endurance capacity may be 123 more important for professional soccer performance in the Lithuanian National League [31]. 124 Such research implies that specific genetic profiles may be advantageous for high-level 125 soccer performance in different countries, perhaps due to between-country differences in 126 playing demands [61] and/or differences in player anthropometry [62].

Indeed, research involving 535 high-level academy male soccer players from multiple clubs across two continents (an English Premier League academy and academies of the highest national category in Uruguay) found that academy players had a higher frequency of the PPARA rs4253778 C allele, AGT rs699 G allele and NOS3 rs2070744 T allele than agematched non-athletes [28] (Table 1). Interestingly, all three of these alleles have previously been associated with elite power athlete status [63-66] and superior power performance [35, 63-65, 67]. However, the physical determinants of high-level youth soccer playing status are known to depend on maturity status [28], so any investigation regarding genetic profiling in youth athletes should account for the confounding effect of maturation. Indeed, when segregated according to peak height velocity (PHV, i.e. the peak rate of somatic growth that is associated with puberty), academy male soccer players have distinct genetic profiles, with 137 post-PHV players demonstrating a greater frequency of the PPARA rs4253778 C allele 138 (25.8% vs. 19.0% vs. 18.5%) and the AGT rs699 G allele (53.5% vs. 44.8% vs. 42.7%) 139 compared to pre-PHV players and non-athletes, respectively [28]. Moreover, the ACTN3 140 rs1815739 XX genotype was more common in pre-PHV players than non-athletes (29.0% vs. 141 15.9%, respectively). Thus, the differences in PPARA rs4253778, AGT rs699 and ACTN3 142 rs1815739 allele/genotype frequency distributions between academy players and non-athletes 143 suggest that the polygenic profile of pre-PHV academy players favours fatigue resistance, 144

rather than power/speed, and that post-PHV academy players would demonstrate a polygenicprofile more advantageous for power/speed performance [28] (Table 1).

Recent research has also documented that the G alleles of two SNPs in ADRB2 147 (rs1042713 and rs1042714), which have previously been associated with power athlete status 148 in a Polish population [68], were over-represented in a small group (n=48, age 12-18 years) 149 of academy male soccer players compared to non-athletes [36]. However, this study had a 150 limited small sample size and did not account for the confounding factor of maturity status, 151 thus, results should be interpreted with caution. Moreover, this study only included academy 152 soccer players from a single club, which means the findings are not necessarily relevant in 153 other academies, either within the same national league, or between nations. 154

In another study, Massidda and colleagues [69] recruited 694 professional male soccer 155 players from Italy, Poland, Lithuania and Ukraine, and investigated the association between 156 the *MCT1* (monocarboxylate transporter-1) rs1049434 SNP and playing position (by dividing 157 players into four sub-groups: defenders, midfielders, forwards, and goalkeepers). In this 158 study, the AA genotype was more frequent in forwards (n = 148) than non-athletes (n = 781). 159 As this SNP is thought to be associated with a higher lactate clearance [70-72], it was 160 postulated to contribute to the previously documented better repeated sprint ability (RSA) in 161 forwards [73], who complete more sprints during a match than midfielders and defenders 162 [74]. The results from this study [69], and analyses documenting the unique demands of each 163 position in high-level soccer match-play [75], suggest that future case control genetic studies 164 in soccer should consider playing position. However, research measuring the activity profile 165 166 (and fitness requirements) of each position recommends that, due to the contrasting matchplay demands [75], there should also be a separation between central and lateral midfielders 167 and defenders. It is therefore recommended that future genetics studies recruit a large enough 168 sample to gain sufficient statistical power to perform playing position analyses by stratifying 169

A further two studies have investigated the genetic association with playing position 172 in professional men's soccer [76, 77]. However, regarding the above points and the limited 173 number of soccer players (n=43) of mixed geographic ancestry (and the lack of control 174 group) in the study by Clos et al [76], it is unlikely that this study had sufficient statistical 175 power to detect an association of genotype with playing position. Although a recent study by 176 Petr et al. [77] did include a control group (n=107) and a larger cohort of soccer players 177 (n=99), the latter sample size was still relatively modest, considering this needed to be 178 segregated into much smaller groups for playing position and genetic analyses. 179

In summary, there is convincing evidence that genetic variation is associated with 180 high-level soccer playing status, and that this association favours strength and power. 181 However, due to physical and technical requirements varying between national leagues [61], 182 and to be representative of high-level soccer on a global scale, case-control genetic 183 association studies should include large sample sizes from multiple clubs in different 184 countries and continents. Further, the fact that physical performance [10] and physiological 185 factors associated with muscular power [78] differ according maturity status, not accounting 186 for maturation is likely to confound any genetic association with playing status in academy 187 soccer players. Studies that have segregated such players according to maturity status have 188 shown that pre-PHV players have a genetic profile that favours endurance performance, 189 while the genetic profile of post-PHV (physically mature) players is more aligned with 190 191 strength and power (in accordance with that of senior professional players). Finally, when investigating the genetic association with soccer playing position, studies should include a 192 sample size large enough to account for the stratification into multiple positional groups and 193 subsequent genotype groups. 194

#### - - /

#### 196 The genetic association with muscular power in soccer players

While it is important to establish if a specific combination of gene variants may characterise high-level soccer in different countries, case-control studies alone do not reveal the relationship between specific genetic polymorphisms and physical performance capabilities. Cross-sectional studies, investigating associations between gene variants and specific soccer performance predictors, may provide more diagnostic information for applied soccer practitioners.

Previous research has found that acceleration (10 m speed) and sprint (20 m speed) 203 capabilities are important determinants of high-level male soccer player status in academy 204 players at all stages of maturity [10, 79], whereas muscular power (horizontal and vertical 205 jump performance) was only a determinant of playing status in post-PHV, not pre-PHV 206 players [10]. The genetic association with explosive performance capacities could be due to 207 certain gene variants influencing the protein abundance/expression within key tissues (e.g. 208 skeletal muscle, tendon), which, in turn, affect the mechanical properties of those tissues and, 209 thus, the players' force/power/speed-generating capacity. Of these specific traits, 210 acceleration, sprint and agility performance have been shown to be independent capabilities 211 [80] underpinned by a different combination of physiological factors. While early 212 acceleration is associated with longer ground contact times (0.12-0.20 s) and relies on 213 contractile force capabilities [81], sprinting is associated with shorter ground contact times 214 (0.09-0.12 s) and therefore relies more on the ability of the muscle-tendon unit to perform 215 fast stretch-shortening cycle actions [82]. Agility is associated with unilateral reactive 216 217 strength capabilities [83] and the main physiological underpinning factors appear to be a combination of neural (inter-muscular co-ordination, neural firing frequency and motor unit 218 recruitment and synchronization) and muscle-tendon unit mechanical properties. Similarly, 219

vertical and horizontal-forward countermovement jump (CMJ) capabilities are independent qualities [13], and are controlled by different co-ordination strategies [84], with horizontalforward CMJs requiring greater biceps femoris activation than vertical CMJs [85]. As power assessed in the vertical and horizontal planes are underpinned by independent physiological factors in high-level soccer players, we will discuss the current body of literature documenting the genetic association with each performance capacity separately.

226

#### 227 Vertical jump performance

One of the main physiological determinants of vertical jump performance in soccer players is 228 quadriceps femoris muscle volume [11]. Indeed, genetic variants associated with muscle size 229 and hypertrophy have also been associated with favourable vertical jump capabilities [35, 230 37]. The ACTN3 rs1815739 R allele has previously been associated with increased serum 231 testosterone [86], greater muscle volume [27] and a greater composition of type II skeletal 232 muscle fibres [49], which are larger (therefore produce more force) and able to shorten 233 quicker, thus able to generate more power, than type I fibres [51, 52]. Pimenta and colleagues 234 [37] documented that the ACTN3 rs1815739 R allele was associated with greater bilateral 235 vertical CMJ performance in 200 professional male soccer players from the Brazilian premier 236 division (Table 1). Furthermore, in 220 Brazilian youth players aged 14-20 years, ACTN3 237 rs1815739 R-allele carriers and ACE DD homozygotes jumped higher than their XX and I-238 allele carrying counterparts [87], although it should be noted that these authors did not 239 account for maturity status in these participants, which may have affected the strength of their 240 associations. These results are partly supported by Ginevičienė et al. [35], who showed that 241 ACTN3 RR genotype was associated with greater bilateral vertical CMJ performance in 152 242 male regional or national level athletes, although only 32 were soccer players. The same 243 study also found that PPARA rs4253778 (another SNP associated with muscle hypertrophy 244

and type II skeletal muscle fibre composition [88]) CC homozygotes also had greater 245 estimated total body muscle mass and outperformed G-allele carriers in the vertical jump 246 247 assessment. However, the results of this study should be treated with caution due to the nonelite standard of athletes, the small sample size of Caucasian only athletes, and the mix of 248 sports the athletes competed in, of which only a small proportion were soccer players. 249 Meanwhile, Massidda and colleagues [89] recruited 90 high-level, professional, male Italian 250 251 soccer players and performed a total genotype score regression analysis, demonstrating that the combination of the ACTN3 rs1815739, ACE I/D and bradykinin receptor B2 (BDKRB2) 252 rs1799722 polymorphisms explained 18–24% of the variance in vertical jump performance. 253 However, there is limited research on the association between polymorphisms of the 254 BDKRB2 gene and power performance, and the ACTN3 rs1815739 [28, 90], ACE I/D [91] 255 and PPARA rs4253778 [28] polymorphisms have not always shown consistent associations 256 with vertical CMJ performance in high-level soccer players, so more research is required to 257 confirm such findings. 258

While greater quadriceps femoris muscle volume is thought to allow the knee 259 extensors to produce more force when propelling the body in the vertical direction [11], 260 research has also found an association between greater compliance of the vastus lateralis 261 tendon-aponeurosis complex and higher vertical CMJ performance [92]. COL5A1 rs12722 262 CC genotype has previously been associated with a more extensible patellar tendon [93], 263 thus, it is possible that the tendon of CC homozygotes can store and release more energy, 264 enabling power to be amplified during vertical jumps. More recent research showed that, in 265 535 academy male soccer players (age 8-23 years), COL5A1 rs12722 CC homozygotes 266 achieved greater bilateral vertical CMJ height than TT homozygotes [28] (Table 1). 267 Assuming that COL5A1 rs12722 CC homozygotes have more elastic tendons than TT 268 homozygotes [93], the findings by Murtagh et al. [28] are somewhat in agreement with Kubo 269

and colleagues [92], who reported that greater compliance of the VL tendon-aponeurosis 270 complex facilitates improved CMJ performance. Interestingly, the association reported by 271 272 Murtagh et al. [28] was only significant in the academy soccer players (and not in nonathletes), possibly as the soccer players could co-ordinate the vertical CMJ actions better, 273 thus gaining greater utilisation of the stretch shortening cycle and elastic tendon recoil 274 properties. Nevertheless, this has only been investigated in one cohort and the association of 275 COL5A1 rs12722, and other SNPs related to tendon elasticity, needs to be further 276 investigated. Overall, it appears that studies targeting genetic polymorphisms that are thought 277 to influence the physiological determinants of vertical jump performance could help elucidate 278 if genetic variation can influence such a task. 279 280

#### 281 Horizontal jump performance

Horizontal jump performance is a key determinant of soccer playing status in physically 282 mature academy soccer players [10, 13]. Interestingly, this performance measure was also 283 shown to be a key physical factor at a young age, influencing future contract status and 284 playing minutes after reaching professional status [94]. In a cohort of 535 academy male 285 soccer players and 151 age- and sex-matched non-athletes, BDNF rs6265 CC homozygotes 286 demonstrated greater horizontal CMJ distance [28]. Neural function is largely determined by 287 neurotrophins, of which BDNF is one of the most active [95], regulating neuronal survival, 288 growth, maintenance, neurogenesis and synaptic plasticity [96]. As the rs6265 C allele is 289 associated with a greater abundance of exercise-induced serum BDNF concentration [97], the 290 findings by Murtagh and colleagues [28] suggest that individuals with potentially enhanced 291 292 neuromuscular characteristics perform better in the performance tasks that require the body to be propelled in the horizontal-forward direction. Such findings are in agreement with 293 previous research documenting that high-level soccer players produce greater hamstring 294

activation during horizontal compared to vertical jumping [13], and the horizontal jump is
believed to be a more complex action, requiring more inter-muscular co-ordination [84, 85].

Overall, there is limited research investigating the relationship between genetic variants and horizontal jump performance in soccer players but the study by Murtagh and colleagues [28] suggests the *BDNF* rs6265 SNP plays a role. Considering the importance of horizontal power for determining high-level soccer playing status and future contract status, more research investigating the association between genetic variation and such a key performance variable is of paramount importance if we are to fully understand the genetic contribution to success in soccer.

#### **The genetic association with maximum strength in soccer players**

As power is underpinned by strength, it follows that strength is likely an important performance measure in soccer players. Indeed, previous work has shown that high-level academy soccer players are stronger (using the isometric mid-thigh pull strength test) than age-matched non-athletes [9]. Although numerous studies have found genetic associations 309 with strength in different populations [98-102], to our knowledge, there are no published 310 studies that have investigated a genetic association with strength in soccer players. However, 311 work from a doctoral thesis did explore the association of four SNPs (PPARA rs4253778, 312 NOS3 rs2070744, COLIA1 rs2249492 and VDR rs2228570) with isometric mid-thigh pull 313 maximum voluntary force (MVF) in 148 high-level male academy soccer players and 93 age-314 and sex-matched non-athletes [103]. Although there were no associations with MVF 315 regarding any of these four SNPs, subsequent genotyping for a further six SNPs (ACTN3 316 rs1815739; AGT rs699; AMPD1 rs17602729; BDNF rs6265; COL2A1 rs2070739; COL5A1 317 rs12722) revealed that COL2A1 rs2070739 CC homozygotes were stronger than T allele 318

carriers, regardless of maturity and athlete status (Fig. 1). This association may be due to the 319 SNP influencing RNA stability, potentially altering the structure of the collagen network in 320 321 and around the muscle (as proposed by Baumert et al. [104]). Increasing the number of lateral attachments within the muscle has the potential to increase the force per muscle cross-322 sectional area, thus augmenting MVF [53, 105], although this hypothesis remains speculative. 323 It should be noted that the isometric mid-thigh pull test used in the aforementioned study is 324 325 an applied measure of strength commonly used in the field. However, it is not specific to a particular muscle group and may therefore lack the sensitivity required to detect more than 326 327 this one genetic association with strength in soccer players. Thus, future studies may wish to incorporate strength tests that isolate key muscle groups, such as the knee extensors, knee 328 flexors or plantar flexors. 329

Although strength was not measured, a separate study demonstrated that fat-free mass 330 was relatively greater in T-allele carriers of the MCT1 A1470T SNP, compared to AA 331 homozygotes in physically mature (~16-18 years of age) academy male soccer players [106]. 332 Due to the association between muscle size and strength [53, 107], it is feasible that this SNP 333 may also be associated with MVF in physically mature soccer players, but this has not yet 334 been investigated. In summary, there is very limited research investigating the genetic 335 association with strength in soccer players and, given the importance of strength in soccer [9], 336 further work is required to elucidate more genetic markers associated with strength in soccer 337 players. 338

Insert Figure 1 near here.

- 339
- 340
- 341

342 The genetic association with acceleration and sprint performance

Accelerating and sprinting require soccer players to optimise the production of peak power to 343 propel the body forward in a cyclical motion. In academy soccer players aged ~18 years, it 344 has been shown that peak power during vertical CMJ is correlated with quadriceps femoris 345 muscle volume [11], and running speed has recently been associated independently with 346 greater muscle volume [108]. The ACE D [109, 110], NOS3 rs2070744 T [58] and AGT 347 (rs699) G [63, 65] alleles have all been associated with elite power athlete status and are 348 thought to exert their favourable effect on power performance by promoting skeletal muscle 349 hypertrophy [63, 65, 111]. Indeed, in a cohort of 212 high-level Brazilian professional male 350 351 players, ACE DD homozygotes demonstrated faster 20 m sprint speeds compared to players of II genotype but there was no difference in muscular power (vertical CMJ) performance 352 [91]. In a separate study from the same population (200 high-level Brazilian professional 353 male players), ACTN3 rs1815739 RR players sprinted faster over 10, 20 and 30 m and 354 jumped higher than XX homozygotes [37]. Furthermore, in 220 Brazilian youth players aged 355 14-20 years, ACTN3 rs1815739 R-allele carriers and ACE DD homozygotes sprinted faster 356 than their XX and I-allele carrying counterparts [87], although it should be noted that this 357 study did not account for the participants' maturity status, which may have affected the 358 strength of the associations. Finally, in 535 high-level male academy soccer players, NOS3 359 TT genotype was not associated with muscular power (neither horizontal nor vertical jump 360 performance), but TT homozygotes did exhibit greater acceleration/sprint performance than 361 their C-allele carrier counterparts (regardless of maturity status) [28] (Table 1). It could 362 therefore be postulated that the ACE DD, ACTN3 RR and NOS3 TT genotypes may stimulate 363 an increase in the number of sarcomeres arranged in series, thus increasing muscle fibre 364 length, which is the main determinant of maximal muscle fibre contraction velocity [53]. 365 Alternatively, simply increasing strength (e.g. through muscle hypertrophy) is known to 366 increase sprint performance [112], thus by influencing muscle size, these polymorphisms 367

may indirectly affect sprint performance. Furthermore, in a pilot study, comprising 26 male 368 players from the under-19 youth team of a Serie A club in Italy, MCT1 rs1049434 A-allele 369 370 carriers performed better than players of TT genotype in the final two sprints of an RSA assessment ( $6 \times 30$  m sprints with an active recovery between sprints). This finding may be 371 linked to the contribution of monocarboxylate transporter-1 (MCT1) to increased metabolic 372 use of lactate after repeated sprinting activity [113]. However, given the limited sample size, 373 374 results should be interpreted with caution. Finally, the AGT G allele has been associated with greater 5 and 20 m sprint performance in a small cohort (n = 48) of academy male soccer 375 376 players aged 12-18 years [36]. Thus, given the age range of players, it is highly likely that there was variation in maturity status, which has been shown to influence the genetic 377 association with athlete status and sprint performance in high-level academy soccer players 378 [28]. Interestingly, no association was found between this SNP and acceleration/sprint 379 performance in a much larger cohort of 535 high-level male academy players, where maturity 380 status was accounted for [28]. Consequently, more large-scale research (including larger 381 sample sizes) is required to confirm the associations between acceleration/sprint speed and 382 the NOS3 rs2070744 and AGT rs699 SNPs, and between RSA and the MCT1 rs1049434 383 SNP. 384

While muscle fibre length (or specifically, the number of serial-aligned sarcomeres) is 385 the main determinant of maximal muscle contraction velocity [53], the extensibility of the 386 tendon is related to speed and horizontal-forward power capabilities in sprinters [114] and 387 post-PHV academy soccer players [12]. Recent research shows that, at all stages of maturity, 388 male academy players and age-matched non-athletes of COL5A1 rs12722 CC genotype 389 achieved quicker acceleration (10 m) and sprint (20 m) times than CT heterozygotes [28] 390 (Table 1). As the CC genotype has previously been associated with a more extensible knee 391 extensor tendon [93], it is feasible that the tendons of CC homozygotes might store and 392

release more energy, thus amplifying power during sprint and acceleration. As well as the 393 aforementioned findings in COL5A1 CC homozygotes, Murtagh et al. [28] showed that 394 COL2A1 rs2070739 CC homozygotes accelerated and sprinted quicker than their T-allele 395 carrier counterparts, possibly due to having a greater capacity to store and release mechanical 396 energy in the tendon. However, this association was only significant in non-athletes and not 397 academy soccer players. It is possible that the specific training environment and physical 398 399 interventions in elite soccer clubs, which are often tailored to improve individual athletic weaknesses, could have confounded the association between the COL2A1 rs2070739 SNP 400 401 and speed performance in academy soccer players. Overall, the limited research investigating the association between genetic variation (particularly SNPs potentially associated with 402 tendon properties) and speed performance in soccer players is positive but future research in 403 independent groups is required to confirm such findings. 404

In addition to having a more compliant patellar tendon [114], research showing that 405 sprinters producing more horizontal force during acceleration are able to highly activate their 406 hamstring muscles just before ground contact [115] suggests that neuromuscular activation is 407 also a major factor in determining sprint performance. In the study by Murtagh et al. [28], 408 BDNF rs6265 CC homozygotes demonstrated greater horizontal power, acceleration and 409 sprint performance. Therefore, as the C allele is associated with a greater abundance of 410 exercise-induced serum BDNF concentration [97], these findings suggest that individuals 411 with potentially enhanced neuromuscular characteristics perform better in the performance 412 tasks that require the body to be propelled in the horizontal-forward direction. Research in 413 just 48 academy soccer players aligns with these findings, showing that three SNPs thought 414 to be related with neural function were associated with 5 m (acceleration) and 20 m (sprint) 415 speed performance [36]. The G alleles of the CPNE5 rs3213537 and CBLN2 rs8093502 SNPs 416 (both genes are thought to play a role in synaptic function [116, 117]), and the A allele of 417

CNTN4 rs62247016 SNP (the CNTN4 gene encodes contactin 4, which is thought to play a 418 role in the formation of axon connections in the developing nervous system [118]), were all 419 420 associated with greater sprint performance [36]. However, the findings of this study should be interpreted with caution, as not only was the sample size limited, but it comprised players 421 from just one academy, and maturity status was not accounted for. Overall, it seems that 422 SNPs associated with neural function/adaptation have a positive impact on sprint 423 performance; however, the research is limited and more evidence is required from large-scale 424 independent studies. 425

Since it is clear that neuromuscular function (e.g. inter- and intra-muscular co-426 ordination [83]) primarily underpins speed performance prior to maturation, i.e. in pre-PHV 427 athletes [78], it seems logical that certain genetic variants may have a greater influence on 428 speed/power performance in pre-PHV soccer players. While no neural related genetic variant 429 has been shown to exert this association specifically in this population to date, AMPD1 430 rs17602729 GG homozygotes did achieve quicker acceleration and sprint times in pre-PHV 431 academy players only [28]. It is possible that pre-PHV AMPD1 GG homozygotes can repeat 432 high intensity acceleration and sprint efforts more frequently during soccer match-play, 433 which may lead to preferential adaptations in acceleration and sprint performance in GG 434 homozygotes compared to their A allele-carrying counterparts. The AMPD1 G>A SNP is the 435 only genetic variant to date that appears to influence acceleration and speed in pre-PHV 436 academy players on an individual SNP basis [28]. Such findings support the argument that 437 genetic research in academy soccer players should be specific to maturity status. 438

439

440 The genetic association with agility performance

Only one study to date has investigated the relationship between agility and genetic profile in 441 soccer players [28]. The main physiological factors underpinning agility capacity appear to 442 be a combination of neural (inter-muscular co-ordination, neural firing frequency and motor 443 unit synchronization) and muscle-tendon unit mechanical properties. It is therefore logical 444 that agility performance was associated with a combination of SNPs, rather than one 445 individual genetic variant. Indeed, a total genotype score model derived from four SNPs 446 (AMPD1 G>A rs17602729, BDNF C>T rs6265, COL5A1 C>T rs12722, COL2A1 C>T 447 rs2070739) that were individually associated with various power/sprint performance 448 449 variables was shown to be positively correlated with agility performance in pre-PHV but not post-PHV academy male soccer players. These significant correlations in pre-PHV players 450 could be due to the academy soccer environment increasingly impacting on agility 451 performance (and overall physical performance) as the player matures. It has previously been 452 demonstrated that training exposure in academy soccer increases (and includes more 453 personalized athletic development and resistance training as well as soccer activities) as the 454 players progress to older age group teams [9], thus suggesting environmental influences 455 could increasingly confound any genetic effect. Nevertheless, such a novel polygenic analysis 456 shows that, while one SNP may not have a significant impact on explosive performance, a 457 combination of SNPs could have a favourable effect, especially in pre-PHV individuals. 458

459

#### 460 The genetic association with aerobic capacity in soccer players

Due to the aerobic underpinnings of soccer [119], potential genetic associations with endurance performance could also influence high-level soccer player status and/or endurance capacity in soccer players. Perhaps the most well-known genetic variant associated with prolonged endurance performance is the *ACE* I/D polymorphism, i.e. the II genotype is commonly associated with greater endurance capacity/performance [120, 121], although not

always [122]. However, only two studies have investigated the association of the ACE I/D 466 polymorphism with soccer athlete status [31, 32]. The study by Juffer and colleagues [32] 467 found that the II genotype frequency was lower in professional male soccer players than 468 endurance runners, supporting the hypothesis that high-level soccer players tend to have a 469 power/strength oriented genetic profile, as suggested by others [28, 32]. However, the study 470 by Ginevičienė and colleagues [31] found a lower frequency of DD genotype in 199 sub-elite 471 472 professional male soccer players representing Lithuanian clubs compared to non-athletes. The contrasting results in these two studies [31, 32] suggest the influence of ACE genotype on 473 474 soccer athlete status may depend on the standard of player (e.g. elite [32] vs. sub-elite [31]) and the associated differences in training/match demands. 475

A different study in 200 professional male soccer players incorporated an applied 476 measure of aerobic capacity and found that ACTN3 XX homozygotes had a greater Yoyo test 477 478 score than RR and RX genotypes [37], i.e. those players with an 'endurance genotype' exhibited a greater aerobic capacity. In a different study of 212 participants from the same 479 population, higher frequencies of the ACE II genotype (compared to I/D or DD genotypes) 480 were observed in "excellent and good performance groups" regarding the YoYo intermittent 481 recovery test [91]. In a separate study in 220 Brazilian youth soccer players, ACE II 482 homozygotes were found to perform better in the YoYo test than D-allele carriers, although 483 this was only in the under-16 year age group and thus a limited sample size of 41 [87]. 484 Interestingly, a study in high-level academy male soccer players found that maturity status 485 was associated with allele and genotype frequency distribution, with pre-PHV players having 486 a greater frequency of ACTN3 XX genotype [28], and post-PHV players demonstrating a 487 greater frequency of the PPARA rs4253778 C allele and the AGT rs699 G allele compared to 488 pre-PHV players and non-athletes. This suggests that pre-PHV academy players are recruited 489 in part due to their high aerobic capacity. In English Category 1 soccer academies, male 490

players do not compete 11 vs. 11 on a full-sized pitch until they reach ~13 years of age 491 (approximately when PHV occurs in boys [123]). While 11 vs. 11 match-play requires a 492 493 greater percentage of time spent at low speeds and more frequent explosive sprint actions, match-play on smaller pitches with fewer players requires a greater distance per minute, more 494 time running at moderate speeds, and a higher work load per minute [124], thus placing a 495 greater physiological demand on aerobic rather than anaerobic capacity. The playing 496 demands of high-level soccer therefore differ according to maturity status and, at pre-PHV 497 level, aerobic capacity appears to be more important than power/ speed, which would explain 498 the apparent endurance-favourable genetic profile of pre-PHV academy soccer players. 499

Another study combined the genotype score of five other SNPs (VEGFA rs2010963, 500 ADRB2 rs1042713 and rs1042714, CRP rs1205 and PPARGC1A rs8192678), and found that 501 those male academy players with a favourable combination responded better to eight weeks' 502 aerobic training when measured with the Yo-Yo intermittent recovery test level 1 [125]. 503 Unfortunately, the authors of this study did not specify which alleles/genotypes were classed 504 as "favourable", thus preventing replication by other groups, whether in research or applied 505 practice. Furthermore, it should be noted that this study only included 42 participants, who 506 were not associated with a high-level soccer academy. A combination of small sample size, 507 low sensitivity of field-based performance measure, and small effect size may have limited 508 statistical power in this study, thus these results should be interpreted with caution. In 509 summary, very few studies have investigated the genetic association with endurance capacity 510 in soccer players, and more rigorous research is required to provide more confidence that 511 genetic variation influences these important phenotypes in soccer players. 512

513

514

Insert Table 1 near here.

#### 516 The genetic association with injury risk in soccer players

517 Player availability is negatively affected by injury [126]. Consequently, team success is associated with low injury rates [127], having more players available for matches and training 518 [128], and losing fewer days to injury during a season [18]. Despite scientific advances 519 identifying a number of risk factors, including player age, previous injury and joint range of 520 motion [1], as well as maturity status [24], playing position [23] and biological sex [129], 521 inter-individual variation in the frequency and severity of soccer injury suggests not all 522 factors are fully understood. With injury prevention representing a key objective for players 523 and coaches, it is logical that athletes and practitioners believe there may be value in genetic 524 data concerning injury susceptibility [130], not least because previous injury increases future 525 injury risk [1]. 526

The majority of soccer injuries occur in the lower extremities without external contact 527 [20]. Muscle strains commonly occur in the quadriceps, hamstrings and calves [21], whilst 528 the knee and ankle are the most common sites for ligament sprains [21]. Extensive 529 mechanical stress during contractile activity, such as acceleration, deceleration and kicking, 530 are implicated in muscle strain, whilst ligament sprains occur when collagenous fibres are 531 excessively stretched as a joint is forced to go beyond its normative range. Common SNPs 532 have the potential to influence the structure, function or expression of proteins within tissues 533 and affect their phenotypes [25], and the heritability of soft-tissue injury is estimated to be as 534 high as 40% [3]. Therefore, genetic differences between players might alter the mechanical 535 properties of musculoskeletal soft-tissues, such as muscle, ligament and tendon, which are 536 537 commonly injured in soccer [21, 131]. The complexity of soccer injuries means that any genetic influence is likely to be polygenic, rather than being due to a single SNP. The high 538 cost and complexity of studying multiple SNPs and their interaction means that, to date, most 539

genetic associations with soccer injuries are from candidate gene studies. This approach 540 involves selecting specific genes or variants known or suspected to contribute to a particular 541 542 phenotype. For example, certain SNPs can influence the structure and/or function of collagenous tissues [132], and several SNPs in collagen genes have been associated with 543 ligament and tendon injuries in athletes from other sports [133-135]. Whilst there are a 544 growing number of SNPs related to injury in various cohorts [136], this section summarises 545 those SNPs associated with soccer injuries, offering potential mechanisms where possible. It 546 should be noted that a considerable number of SNPs have been associated with injury in 547 cohorts across multiple sports [136], and that the aetiology of soccer injuries is often similar 548 to injuries occurring in those sports. Consequently, it is possible that SNPs associated with 549 injury in other sports, of which some are not discussed in this section, may also be associated 550 with soccer injury. 551

552

#### 553 ACTN3 rs1815739

As previously mentioned, carriers of the ACTN3 rs1815739 R allele have larger and stronger 554 muscles than XX homozygotes [27, 137], which is important because low muscle strength is 555 a risk factor for soft-tissue injury [138]. Research suggests that professional male soccer 556 players of ACTN3 XX genotype exhibit greater exercise-induced muscle damage than RR 557 homozygotes [139], which may help explain why this genotype has been associated with a 558 higher rate of non-contact musculoskeletal soft-tissue injuries in professional male soccer 559 players [140], although this study included just 43 participants (Table 2). Another study of 560 169 professional male players reported a higher rate of non-contact muscle injuries in XX 561 562 homozygotes than R-allele carriers, with XX homozygotes most likely to suffer severe injury [44] (Table 2). A separate study in professional players found X-allele carriers were most 563 likely to be injured, with XX homozygotes having the longest absence after injury [47]. 564

However, this study comprised just 46 participants, of which 22 were male and 24 female. In addition to the well-documented physiological and biomechanical differences between men and women, soccer match demands [141] and injury risk [129, 142] also differ between men's and women's soccer, thus pooling male and female players in the same study is problematic when deciphering the genetic association with soccer injuries. However, it is worth noting that this is one of only two studies to include female participants in a study investigating the genetic association with injury in soccer players.

Elsewhere, no association was reported between the ACTN3 rs1815739 SNP and 572 hamstring injuries in 107 male professional players [40], nor with injury prevalence in 402 573 male academy players [29]. Similarly, a recent study investigating the ACTN3 rs1815739 574 SNP and injury in 191 female soccer players found no difference in non-contact injury 575 incidence between genotype groups [143]. Nevertheless, the study by Hall and colleagues 576 [29] demonstrated that high-level academy players with at least one X allele missed more 577 days following ankle injuries than RR homozygotes (Table 2), suggesting greater muscle size 578 and strength associated with the R allele [26, 27] may enhance limb stability and favour 579 return to play. It appears that the ACTN3 rs1815739 SNP may influence soccer injuries, 580 where potential mechanisms include muscle strength enhancement and/or the provision of 581 structural stability to the sarcomere during contraction [144]. 582

583

#### 584 COL1A1 rs1800012

The rs1800012 SNP is a C>A substitution in the *COL1A1* gene [145], encoding the type I collagen  $\alpha$ 1 chain. This is important when considering injury risk, as type I collagen is the major fibrillar collagen providing structural stability to ligaments and tendons [146]. The rare AA genotype appears protective against ACL [133] and Achilles tendon [147] injuries in

athletes, while professional male soccer players with ACL ruptures are also less likely to be 589 AA homozygotes [39] (Table 2), although others found no associations between this SNP and 590 injury [29, 40, 46]. Differing conclusions in these studies may relate to the specific injury 591 types investigated, as it appears COL1A1 variants are most likely to influence injuries to 592 ligament and/or tendon. Nevertheless, the rs1800012 A allele has been associated (as part of a 593 haplotype) with significantly higher COL1A1 gene transcription [148]. If higher COL1A1 594 transcriptional activity increases the structural stability of ligaments and tendons, this might 595 explain the apparent protective effect of the rare rs1800012 genotype against ACL and 596 597 Achilles tendon injuries.

598

#### 599 COL5A1 rs12722 and rs13946

The rs12722 SNP is a C>T substitution within the *COL5A1* gene [149], encoding the type V 600 collagen  $\alpha$ 1 chain, a vital structural component of tendons and ligaments during collagen 601 fibrillogenesis [150]. Two studies with relatively small sample sizes (both n < 80) reported 602 603 severe hamstring injuries in TC heterozygous professional male soccer players [45, 46], with no association reported elsewhere [40, 42]. More severe musculoskeletal injuries appear to be 604 experienced by TT homozygous players than C-allele carriers [42], whilst a case-control 605 study comparing players with and without ACL rupture found that carrying C alleles in two 606 different COL5A1 SNPs (rs12722 and rs13946) was apparently protective [41] (Table 2). In 607 contrast, a higher prevalence of soft-tissue and ligament injuries has been reported in pre-608 PHV academy male players carrying the C allele and CC genotype, respectively [29] (Table 609 2). Therefore, more research is required to determine which *COL5A1* rs12722 allele is indeed 610 611 protective and why, although from this limited evidence, it is possible that it is linked to the maturity status of the player. 612

#### 614 EMILIN1 rs2289360

615 The rs2289360 SNP is a T>C substitution within the *EMILIN1* gene [151], which encodes the elastin microfibril interface-1 (EMILIN-1) protein. This protein assists the fusion of elastin 616 fibres during elastogenesis [152] and provides elasticity to ligaments and tendons [153]. In 617 professional male soccer players, the TT genotype has been associated with more severe 618 MCL injuries, but not with the incidence or severity of muscle or tendon injuries [38, 46] 619 (Table 2). Another study reported no association with hamstring injuries in 107 professional 620 male players [40], whilst in 402 academy male players, CC homozygotes had the highest 621 overall injury prevalence and the 'protective' TT genotype was more frequent in post- than 622 pre-PHV players [29] (Table 2). These findings suggest that fewer 'genetically pre-disposed' 623 (i.e. CC homozygotes) players progress from pre- to post- PHV in a soccer academy. 624 However, in the same study, academy players carrying the T allele experienced the most days 625 missed following ankle injuries [29]. Together with the association of the TT genotype with 626 MCL injury in professional players [38, 46], this suggests that this EMILIN1 SNP might 627 influence soccer-related ligament injury predisposition and recovery. However, contrasting 628 results exist between and within studies regarding which rs2289360 genotypes and/or alleles 629 are protective, requiring further investigation to understand a potential role in soccer injuries. 630 Hall et al. [29] hypothesised that the C allele might augment collagen tissue stiffness, causing 631 stiffer tendons in CC homozygotes and increasing injury risk as a result of greater strain on 632 muscle fascicles during eccentric activity. In contrast, the T allele might protect against injury 633 634 in general (but particularly muscle injury) as a product of increased tendon compliance. However, such increased compliance may be insufficient to withstand more excessive 635 external loads, leading to greater tendon damage (and even rupture) requiring a longer period 636 of healing and rehabilitation. 637

ch in nd in e str onfir onfir c-C stitial with o

#### 639 MMP3 rs679620 and rs3025058

640 The rs679620 SNP is a T>C substitution within the *MMP3* gene [154]. The MMPs, a family of collagen-degrading enzymes, are physiological regulators of extracellular matrix (ECM) 641 remodelling [155] and are important for muscle regeneration [156]. The T allele was 642 associated with hamstring injury in 107 professional male soccer players [40], and with 643 longer absence following knee injuries in academy soccer players [29] (Table 2). As the T 644 allele sits in linkage disequilibrium with the MMP3 rs3025058 5A allele, which increases 645 MMP3 expression [157], the rs679620 T allele might increase RNA expression and intensify 646 the degradation of proteins, such as collagen and elastin, thereby weakening the structural 647 integrity of ligament and ECM. However, further research is required to confirm this 648 hypothesis. 649

650

#### 651 CCL2 rs2857656

The rs2857656 SNP is a G>C substitution in the CCL2 gene affecting plasma C-C motif 652 chemokine ligand-2 (CCL2) levels [158]. CCL2 is expressed within the interstitial space 653 654 between myofibres after damaging activity [159] and mediates systemic changes with chronic exercise [160]. In professional male soccer players (n = 73-74), those with the GG genotype 655 656 suffered more severe muscle injuries than C-allele carriers [45, 46], although others report no association with hamstring injuries in 107 professional male players [40], nor with injury 657 prevalence or recovery time in male academy players [29] (Table 2). Plasma CCL2 is lower 658 in GG homozygotes [158], who exhibit lower muscle strength than C-allele carriers [161]. 659 Considering the prospective role of CCL2 in satellite cell proliferation [162], higher CCL2 660

during recovery from muscle injury may be advantageous to players carrying at least oners2857656 C allele.

663

#### 664 IL6 rs1800795

The rs1800795 SNP is a C>G substitution within the *IL6* gene [163]. The myokine IL-6 is 665 produced in skeletal muscle after exercise and is functionally related to growth, atrophy and 666 collagen synthesis [164, 165]. Interestingly, the GG genotype was associated with greater 667 hamstring injury risk in 107 professional male players [40] (Table 2). Contrastingly, CC 668 homozygotes had higher prevalence of any injury and muscle injury in 402 academy male 669 players [29] (Table 2). Having both pro- and anti-inflammatory capacities [166] makes it 670 plausible that IL-6 could increase or reduce injury risk. Increased plasma IL-6 associated with 671 the G allele [163] may heighten soft-tissue injury risk. Specifically, because eccentric 672 contractions induce pro-inflammatory IL-6 expression [167], and because cytokines trigger 673 tenocyte apoptosis and ECM degradation [168], GG homozygotes may experience these 674 675 unfavourable changes via increased IL-6, thus heightening injury risk. This could explain the association of the GG genotype with hamstring injury risk [40] if eccentric contractions 676 initiate a process of elevated ECM degradation above that which is required for repair and 677 recovery. In contrast, increased injury risk in CC homozygotes reported in academy players 678 [29] could relate to an increased inflammatory response following eccentric activity 679 associated with the C allele [169], where recovery between intense training and/or 680 competition may be impaired by sustained inflammation. 681

682

683

Insert Table 2 near here.

684

#### 685 Future directions for genetics research in soccer

In addition to the SNPs highlighted above, several other SNPs have been identified as 686 potentially important but require further investigation, e.g. due to low sample size or lack of 687 replication by independent research groups. For example, there are currently no published 688 studies that have investigated the genetic association with strength in soccer players, although 689 previously unpublished work from our group suggests the COL2A1 rs2070739 SNP is 690 associated with maximum strength in 148 high-level male academy soccer players and 93 691 age- and sex-matched non-athletes, regardless of maturity status and athlete status (Fig. 1). 692 Further, in a study with a limited sample size, the combined genotype score of five SNPs 693 (VEGF rs2010963, ADRB2 rs1042713 and rs1042714, CRP rs1205 and PPARGC1A 694 rs8192678) was associated with a greater adaptation to eight weeks' aerobic training in 695 academy players [125]. Thus, replication by independent groups is necessary to provide more 696 confidence in the results from these studies. 697

698 Regarding the genetic association with injury risk, numerous studies have found individual SNP associations that require replication by independent groups. For example, 699 IGF2 rs3213221 GC genotype, GEFT rs11613457 GG genotype, HGF rs5745678 T allele 700 and *LIF* rs9290271 T allele were suggested to be protective against severe non-contact 701 injuries, whilst *HGF* rs5745697 CC and rs1011694 AA homozygotes suffered fewer injuries, 702 albeit in relatively small sample sizes [45, 46]. Male academy players with at least one MYLK 703 rs28497577 T allele required longer recovery time following knee injury [29], whilst 704 professional male players with the SOX15 GG genotype were injured more often than T-705 706 allele carriers [45], although not in another study by the same group [46]. Further, hamstring injury risk was associated with the TNC rs2104772 A allele, and with the HIF1A CC and 707 NOS3 rs1799983 GG genotypes, in 107 professional male players [40]. However, others have 708 reported no association between the TNC rs2104772 SNP and injury [46], which could be due 709

to the latter study having a smaller sample size (*n* = 73, potentially reducing statistical power to detect an association). Recently, the *ACE* I/D D allele was associated with reduced muscle injury risk in 710 professional male players from Italy and Japan [43], and in 402 male academy players, those homozygous for the *VEGFA* rs2010963 C allele had a higher prevalence of ligament and tendon injuries [29], while the *MCT1* rs1049434 AA genotype has been associated with higher muscle injury incidence in 173 high-level soccer players [170]. It is pertinent to mention additional SNPs, which were not associated with soccer injury. These include no association between muscle injury and three *VDR* SNPs [171] or *COL5A1* rs12722 [42], between *COL12A1* rs970547 and ACL injury [39], between *TTN* rs2742327 and non-contact musculoskeletal injuries [46], or between hamstring injuries and 32 of 37 investigated SNPs [40].

As well as replication of robust studies by independent research groups, further investigation is required to conduct large-scale genome-wide association studies to identify new genetic markers associated with athlete status, physical performance and injury risk in soccer. To identify a substantial proportion of the gene variants with small effect sizes that collectively contribute to the genetic component of variability in a complex trait, such as athlete status, physical performance and injury, a sample of at least 2,000 is probably 726 required [172]. Furthermore, only a handful of genetic markers have been investigated to the 727 extent whereby their function is known to impact physiology/biomechanics, which is crucial 728 if we are to truly understand the potential impact of genetic testing in soccer. Moreover, to 729 date, all genetic studies in soccer players have recruited almost exclusively male participants. 730 Given the differences in match demands between women's and men's soccer [141], as well 731 as the physiological and biomechanical sex differences, and variability in injury risk between 732 male and female soccer players [129, 142], we cannot assume that the genetic associations 733 found with athlete status, physical performance and injury risk in male soccer players will be 734

the same for female players. Therefore, future research should investigate the genetic 735 association with athlete status, physical performance characteristics and injury risk in 736 737 women's soccer. Finally, almost all studies published to date have recruited participants of European ancestry due to the evidence that genetic variation differs between geographic 738 ancestries [173]. To provide a more comprehensive understanding of the genetic contribution 739 to success as a soccer player, however, future studies should investigate the genetic 740 association with athlete status, physical performance characteristics and injury risk in soccer 741 players from different geographic ancestries. 742

743

#### 744 Summary

Throughout this review, we have critically appraised the literature pertaining to the genetic 745 association with physical performance and injury risk in soccer players. Using the most 746 robust evidence, we have suggested directions for future research in this area, and provided 747 guidance on the most compelling evidence to help practitioners understand the importance of 748 genetic variation and its influence on soccer athlete status, physical performance (Table 1) 749 and injury risk (Table 2). Gaining a more detailed insight into the genetic contribution to 750 soccer-related physical performance and injury risk could potentially help *future* talent 751 selection and physical development procedures in elite soccer clubs. However, it should be 752 stressed that genetics research in soccer is still in its infancy, and future studies should 753 attempt to replicate those more robust studies highlighted in this review by recruiting large 754 cohorts from various leagues around the world, and distinguishing between maturity status 755 when recruiting academy players. Further, very few studies have investigated genetic 756 757 associations with athlete status, physical performance or injury in female players, or players of non-European geographic ancestry, so future studies should include these populations. 758 Regarding the genetic association with soccer playing position, studies should include a 759

sample size large enough to account for the stratification into six positional groups (i.e. 760 goalkeepers, wide defenders, central defenders, wide midfielders, central midfielders, and 761 forwards) and subsequent genotype groups. Furthermore, the candidate gene approach, 762 although practical, relies on a degree of prior knowledge and is restricted to the specific SNPs 763 selected for analysis. However, reduced costs and improved accessibility to more powerful 764 approaches, such as genome-wide association studies, may aid the discovery of new SNPs to 765 be investigated in larger cohorts. Studies with broader scope in terms of sample size, injury 766 frequency and the number SNPs investigated are required to advance the existing evidence 767 768 for a genetic contribution to soccer injuries. If large-scale studies following our recommended criteria could be initiated through collaborative analyses (and replicated by 769 independent groups), genetics research could have a positive impact in elite soccer clubs by 770 improving the level and detail of individual physical programming aimed to overcome and/or 771 develop potential genetic deficiencies and/or strengths, respectively. 772

#### 773 **References**

1. Arnason A, Sigurdsson SB, Gudmundsson A et al. Risk factors for injuries in football.

Am J Sports Med 2004; 32: 5S-16S. doi:10.1177/0363546503258912

- De Moor MH, Spector TD, Cherkas LF et al. Genome-wide linkage scan for athlete
  status in 700 British female DZ twin pairs. Twin Res Hum Genet 2007; 10: 812-820.
  doi:10.1375/twin.10.6.812
- Hakim AJ, Cherkas LF, Spector TD et al. Genetic associations between frozen
  shoulder and tennis elbow: a female twin study. Rheumatology (Oxford) 2003; 42:
  739-742. doi:10.1093/rheumatology/keg159
- Magnusson K, Turkiewicz A, Hughes V et al. High genetic contribution to anterior 782 4. ~69. ligament rupture: Heritability Br J Sports Med 783 cruciate 2020. doi:10.1136/bjsports-2020-102392. doi:10.1136/bjsports-2020-102392 784
- Bouchard C, An P, Rice T et al. Familial aggregation of VO(2max) response to
  exercise training: results from the HERITAGE Family Study. J Appl Physiol (1985)
  1999; 87: 1003-1008. doi:10.1152/jappl.1999.87.3.1003
- 788 6. Peeters MW, Thomis MA, Loos RJ et al. Heritability of somatotype components: a
  789 multivariate analysis. Int J Obes (Lond) 2007; 31: 1295-1301.
  790 doi:10.1038/sj.ijo.0803575
- 791 7. Simoneau JA, Bouchard C. Genetic determinism of fiber type proportion in human
  792 skeletal muscle. FASEB J 1995; 9: 1091-1095. doi:10.1096/fasebj.9.11.7649409
- 793 8. Zhai G, Ding C, Stankovich J et al. The genetic contribution to longitudinal changes
  794 in knee structure and muscle strength: a sibpair study. Arthritis Rheum 2005; 52:
  795 2830-2834. doi:10.1002/art.21267
- 9. Brownlee TE, Murtagh CF, Naughton RJ et al. Isometric maximal voluntary forceevaluated using an isometric mid-thigh pull differentiates English Premier League

youth soccer players from a maturity-matched control group. Sci Med Football 2018;

799 2: 209-215. doi:10.1080/24733938.2018.1432886

- Murtagh CF, Brownlee TE, O'Boyle A et al. Importance of Speed and Power in Elite
  Youth Soccer Depends on Maturation Status. J Strength Cond Res 2018; 32: 297-303.
  doi:10.1519/jsc.0000000002367
- Murtagh CF, Nulty C, Vanrenterghem J et al. The Neuromuscular Determinants of
  Unilateral Jump Performance in Soccer Players Are Direction-Specific. Int J Sports
  Physiol Perform 2018; 13: 604-611. doi:10.1123/ijspp.2017-0589
- Murtagh CF, Stubbs M, Vanrenterghem J et al. Patellar tendon properties distinguish
  elite from non-elite soccer players and are related to peak horizontal but not vertical
  power. Eur J Appl Physiol 2018; 118: 1737-1749. doi:10.1007/s00421-018-3905-0
- Murtagh CF, Vanrenterghem J, O'Boyle A et al. Unilateral jumps in different
  directions: a novel assessment of soccer-associated power? J Sci Med Sport 2017; 20:
  1018-1023. doi:10.1016/j.jsams.2017.03.016
- Abbott W, Clifford T. The influence of muscle strength and aerobic fitness on
  functional recovery in professional soccer. J Sports Med Phys Fitness 2021.
  doi:10.23736/s0022-4707.21.13401-2. doi:10.23736/s0022-4707.21.13401-2
- Buchheit M, Simpson BM, Hader K et al. Occurrences of near-to-maximal speedrunning bouts in elite soccer: insights for training prescription and injury mitigation.
  Sci Med Footb 2021; 5: 105-110. doi:10.1080/24733938.2020.1802058

81816.Dos'Santos T, Cowling I, Challoner M et al. What are the significant turning demands

- of match play of an English Premier League soccer team? J Sports Sci 2022; 40:
- 820 1750-1759. doi:10.1080/02640414.2022.2109355
- Manzi V, Annino G, Savoia C et al. Relationship between aerobic fitness and
  metabolic power metrics in elite male soccer players. Biol Sport 2022; 39: 599-606

- 18. Arnason A, Sigurdsson SB, Gudmundsson A et al. Physical fitness, injuries, and team 823 824 performance in soccer. Med Sci **Sports** Exerc 2004: 36: 278-285. 825 doi:10.1249/01.Mss.0000113478.92945.Ca
- 826 19. Carling C, Orhant E, LeGall F. Match injuries in professional soccer: inter-seasonal
  827 variation and effects of competition type, match congestion and positional role. Int J
  828 Sports Med 2010; 31: 271-276. doi:10.1055/s-0029-1243646
- 829 20. Ekstrand J, Hägglund M, Waldén M. Injury incidence and injury patterns in
  830 professional football: the UEFA injury study. Br J Sports Med 2011; 45: 553-558.
  831 doi:10.1136/bjsm.2009.060582
- Hawkins RD, Hulse MA, Wilkinson C et al. The association football medical research
  programme: an audit of injuries in professional football. Br J Sports Med 2001; 35:
  43-47
- Hall ECR, Larruskain J, Gil SM et al. An injury audit in high-level male youth soccer
  players from English, Spanish, Uruguayan and Brazilian academies. Phys Ther Sport
  2020; 44: 53-60. doi:10.1016/j.ptsp.2020.04.033
- Hall ECR, Larruskain J, Gil SM et al. Playing Position and the Injury Incidence Rate
  in Male Academy Soccer Players. J Athl Train 2022; 57: 696-703. doi:10.4085/10626050-0346.21
- Hall ECR, Larruskain J, Gil SM et al. Injury risk is greater in physically mature
  versus biologically younger male soccer players from academies in different
  countries. Phys Ther Sport 2022; 55: 111-118. doi:10.1016/j.ptsp.2022.03.006
- Tabor HK, Risch NJ, Myers RM. Candidate-gene approaches for studying complex
  genetic traits: practical considerations. Nat Rev Genet 2002; 3: 391-397.
  doi:10.1038/nrg796

- 847 26. Seto JT, Quinlan KG, Lek M et al. ACTN3 genotype influences muscle performance
  848 through the regulation of calcineurin signaling. J Clin Invest 2013; 123: 4255-4263
- 27. Erskine RM, Williams AG, Jones DA et al. The individual and combined influence of
- ACE and ACTN3 genotypes on muscle phenotypes before and after strength training.
  Scand J Med Sci Sports 2014; 24: 642-648. doi:10.1111/sms.12055
- 852 28. Murtagh CF, Brownlee TE, Rienzi E et al. The genetic profile of elite youth soccer
  853 players and its association with power and speed depends on maturity status. PLoS
  854 One 2020; 15: e0234458. doi:10.1371/journal.pone.0234458
- Hall ECR, Baumert P, Larruskain J et al. The genetic association with injury risk in
  male academy soccer players depends on maturity status. Scand J Med Sci Sports
  2022; 32: 338-350. doi:10.1111/sms.14077
- 858 30. Egorova ES, Borisova AV, Mustafina LJ et al. The polygenic profile of Russian
  859 football players. J Sports Sci 2014; 32: 1286-1293.
  860 doi:10.1080/02640414.2014.898853
- 31. Gineviciene V, Jakaitiene A, Tubelis L et al. Variation in the ACE, PPARGC1A and
  PPARA genes in Lithuanian football players. Eur J Sport Sci 2014; 14 Suppl 1: S289295. doi:10.1080/17461391.2012.691117
- 32. Juffer P, Furrer R, González-Freire M et al. Genotype distributions in top-level soccer
  players: a role for ACE? Int J Sports Med 2009; 30: 387-392. doi:10.1055/s-00281105931
- Micheli ML, Gulisano M, Morucci G et al. Angiotensin-converting enzyme/vitamin D
  receptor gene polymorphisms and bioelectrical impedance analysis in predicting
  athletic performances of Italian young soccer players. J Strength Cond Res 2011; 25:
  2084-2091. doi:10.1519/JSC.0b013e31820238aa

- 871 34. Santiago C, González-Freire M, Serratosa L et al. ACTN3 genotype in professional soccer players. Br J Sports Med 2008; 42: 71-73. doi:10.1136/bjsm.2007.039172 872
- Ginevičienė V, Pranckevičienė E, Milašius K et al. Relating fitness phenotypes to genotypes in Lithuanian elite athletes. . Acta Medica Lituanica 2010; 17: 1-10. 874 doi:10.2478/v10140-010-0001-0 875

873

35.

- Pickering C, Suraci B, Semenova EA et al. A Genome-Wide Association Study of 876 36. Sprint Performance in Elite Youth Football Players. J Strength Cond Res 2019; 33: 877 2344-2351. doi:10.1519/jsc.000000000003259 878
- Pimenta EM, Coelho DB, Veneroso CE et al. Effect of ACTN3 gene on strength and 879 37. endurance in soccer players. J Strength Cond Res 2013; 27: 3286-3292. 880 doi:10.1519/JSC.0b013e3182915e66 881
- 38. Artells R, Pruna R, Dellal A et al. Elastin: a possible genetic biomarker for more 882 severe ligament injuries in elite soccer. A pilot study. Muscles Ligaments Tendons J 883 2016; 6: 188-192 884
- 39. Ficek K, Cieszczyk P, Kaczmarczyk M et al. Gene variants within the COL1A1 gene 885 are associated with reduced anterior cruciate ligament injury in professional soccer 886 players. J Sci Med Sport 2013; 16: 396-400. doi:10.1016/j.jsams.2012.10.004 887
- Larruskain J, Celorrio D, Barrio I et al. Genetic Variants and Hamstring Injury in 40. 888 Soccer: An Association and Validation Study. Med Sci Sports Exerc 2018; 50: 361-889 368. doi:10.1249/mss.000000000001434 890
- Lulińska-Kuklik E, Rahim M, Domańska-Senderowska D et al. Interactions between 891 41. COL5A1 Gene and Risk of the Anterior Cruciate Ligament Rupture. J Hum Kinet 892 2018; 62: 65-71 893

olymoi ssional 0000000 n-conta 2017; associ rs: inf ; 14: 22 peficien ndings

- Massidda M, Bachis V, Corrias L et al. Influence of the COL5A1 rs12722 on
  musculoskeletal injuries in professional soccer players. J Sports Med Phys Fitness
  2015; 55: 1348-1353
- Massidda M, Miyamoto-Mikami E, Kumagai H et al. Association between the ACE
  I/D polymorphism and muscle injuries in Italian and Japanese elite football players. J
  Sports Sci 2020; 38: 2423-2429. doi:10.1080/02640414.2020.1787683
- Massidda M, Voisin S, Culigioni C et al. ACTN3 R577X Polymorphism Is
  Associated With the Incidence and Severity of Injuries in Professional Football
  Players. Clin J Sport Med 2019; 29: 57-61. doi:10.1097/jsm.0000000000487
- Pruna R, Artells R, Lundblad M et al. Genetic biomarkers in non-contact muscle
  injuries in elite soccer players. Knee Surg Sports Traumatol Arthrosc 2017; 25: 33113318. doi:10.1007/s00167-016-4081-6
- 906 46. Pruna R, Artells R, Ribas J et al. Single nucleotide polymorphisms associated with
  907 non-contact soft tissue injuries in elite professional soccer players: influence on
  908 degree of injury and recovery time. BMC Musculoskelet Disord 2013; 14: 221
- 909 47. Rodas G, Moreno-Pérez V, Del Coso J et al. Alpha-Actinin-3 Deficiency Might
  910 Affect Recovery from Non-Contact Muscle Injuries: Preliminary Findings in a Top911 Level Soccer Team. Genes (Basel) 2021; 12
- 912 48. Beggs AH, Byers TJ, Knoll JH et al. Cloning and characterization of two human
  913 skeletal muscle alpha-actinin genes located on chromosomes 1 and 11. J Biol Chem
  914 1992; 267: 9281-9288
- Ahmetov II, Druzhevskaya AM, Lyubaeva EV et al. The dependence of preferred 49. 915 competitive racing distance on muscle fibre type composition and ACTN3 genotype 916 skaters. Physiol 2011; 96: 1302-1310. 917 in speed Exp doi:10.1113/expphysiol.2011.060293 918

- 919 50. Vincent B, De Bock K, Ramaekers M et al. ACTN3 (R577X) genotype is associated
  920 with fiber type distribution. Physiol Genomics 2007; 32: 58-63.
  921 doi:10.1152/physiolgenomics.00173.2007
- Bottinelli R, Canepari M, Pellegrino MA et al. Force-velocity properties of human
  skeletal muscle fibres: myosin heavy chain isoform and temperature dependence. J
  Physiol 1996; 495 ( Pt 2): 573-586
- 925 52. Gilliver SF, Degens H, Rittweger J et al. Variation in the determinants of power of
  926 chemically skinned human muscle fibres. Exp Physiol 2009; 94: 1070-1078.
  927 doi:10.1113/expphysiol.2009.048314
- 53. Jones DA, Rutherford OM, Parker DF. Physiological changes in skeletal muscle as a 928 strength training. J Physiol 74: result of Q Exp 1989; 233-256. 929 doi:10.1113/expphysiol.1989.sp003268 930
- 931 54. Ahmetov II, Fedotovskaya ON. Current Progress in Sports Genomics. Adv Clin Chem
  932 2015; 70: 247-314. doi:10.1016/bs.acc.2015.03.003
- Faude O, Koch T, Meyer T. Straight sprinting is the most frequent action in goal
  situations in professional football. J Sports Sci 2012; 30: 625-631.
  doi:10.1080/02640414.2012.665940
- 936 56. Murtagh CF, Naughton RJ, McRobert AP et al. A Coding System to Quantify
  937 Powerful Actions in Soccer Match Play: A Pilot Study. Res Q Exerc Sport 2019; 90:
  938 234-243. doi:10.1080/02701367.2019.1576838

57. Eynon N, Ruiz JR, Yvert T et al. The C allele in NOS3 -786 T/C polymorphism is
associated with elite soccer player's status. Int J Sports Med 2012; 33: 521-524.
doi:10.1055/s-0032-1306337

- 942 58. Gómez-Gallego F, Ruiz JR, Buxens A et al. The -786 T/C polymorphism of the
  943 NOS3 gene is associated with elite performance in power sports. Eur J Appl Physiol
  944 2009; 107: 565-569. doi:10.1007/s00421-009-1166-7
- Sessa F, Chetta M, Petito A et al. Gene polymorphisms and sport attitude in Italian
  athletes. Genet Test Mol Biomarkers 2011; 15: 285-290. doi:10.1089/gtmb.2010.0179
- 947 60. Hopkinson NS, Li KW, Kehoe A et al. Vitamin D receptor genotypes influence
  948 quadriceps strength in chronic obstructive pulmonary disease. Am J Clin Nutr 2008;
  949 87: 385-390. doi:10.1093/ajcn/87.2.385
- 950 61. Dellal A, Chamari K, Wong dP et al. Comparison of physical and technical
  951 performance in European soccer match-play: FA Premier League and La Liga. . Eur J
  952 Sport Sci 2011; 11: 51-59
- 953 62. Bloomfield J, Polman R, Butterly R et al. Analysis of age, stature, body mass, BMI
  954 and quality of elite soccer players from 4 European Leagues. J Sports Med Phys
  955 Fitness 2005; 45: 58-67
- Gomez-Gallego F, Santiago C, González-Freire M et al. The C allele of the AGT
  Met235Thr polymorphism is associated with power sports performance. Appl Physiol
  Nutr Metab 2009; 34: 1108-1111. doi:10.1139/h09-108
- 959 64. Ruiz JR, Arteta D, Buxens A et al. Can we identify a power-oriented polygenic
  960 profile? J Appl Physiol (1985) 2010; 108: 561-566.
  961 doi:10.1152/japplphysiol.01242.2009
- 962 65. Zarębska A, Sawczyn S, Kaczmarczyk M et al. Association of rs699 (M235T)
  963 polymorphism in the AGT gene with power but not endurance athlete status. J
- 964 Strength Cond Res 2013; 27: 2898-2903. doi:10.1519/JSC.0b013e31828155b5

- 965 66. Petr M, Maciejewska-Skrendo A, Zajac A et al. Association of Elite Sports Status
  966 with Gene Variants of Peroxisome Proliferator Activated Receptors and Their
  967 Transcriptional Coactivator. Int J Mol Sci 2019; 21
- 968 67. Petr M, Stastny P, Pecha O et al. PPARA intron polymorphism associated with power
  969 performance in 30-s anaerobic Wingate Test. PLoS One 2014; 9: e107171
- 970 68. Sawczuk M, Maciejewska-Karlowska A, Cieszczyk P et al. Association of the
  971 ADRB2 Gly16Arg and Glu27Gln polymorphisms with athlete status. J Sports Sci
  972 2013; 31: 1535-1544. doi:10.1080/02640414.2013.786184
- 973 69. Massidda M, Mendez-Villanueva A, Ginevičienė V et al. Association of
  974 Monocarboxylate Transporter-1 (MCT1) A1470T Polymorphism (rs1049434) with
  975 Forward Football Player Status. Int J Sports Med 2018; 39: 1028-1034.
  976 doi:10.1055/a-0634-6387
- 977 70. Merezhinskaya N, Fishbein WN, Davis JI et al. Mutations in MCT1 cDNA in patients
  978 with symptomatic deficiency in lactate transport. Muscle Nerve 2000; 23: 90-97.
  979 doi:10.1002/(sici)1097-4598(200001)23:1<90::aid-mus12>3.0.co;2-m
- 71. Cupeiro R, Benito PJ, Maffulli N et al. MCT1 genetic polymorphism influence in
  high intensity circuit training: a pilot study. J Sci Med Sport 2010; 13: 526-530.
  doi:10.1016/j.jsams.2009.07.004
- 72. Cupeiro R, Pérez-Prieto R, Amigo T et al. Role of the monocarboxylate transporter
  MCT1 in the uptake of lactate during active recovery. Eur J Appl Physiol 2016; 116:
  1005-1010. doi:10.1007/s00421-016-3365-3
- 73. Aziz AR, Mukherjee S, Chia MY et al. Validity of the running repeated sprint ability
  test among playing positions and level of competitiveness in trained soccer players.
  Int J Sports Med 2008; 29: 833-838. doi:10.1055/s-2008-1038410

- P89 74. Di Salvo V, Baron R, Tschan H et al. Performance characteristics according to
  playing position in elite soccer. Int J Sports Med 2007; 28: 222-227. doi:10.1055/s2006-924294
- 75. Carling C, Bloomfield J, Nelsen L et al. The role of motion analysis in elite soccer:
  contemporary performance measurement techniques and work rate data. Sports Med
  2008; 38: 839-862. doi:10.2165/00007256-200838100-00004
- 995 76. Clos E, Pruna R, Lundblad M et al. ACTN3's R577X Single Nucleotide
  996 Polymorphism Allele Distribution Differs Significantly in Professional Football
  997 Players according to Their Field Position. Med Princ Pract 2021; 30: 92-97
- Petr M, Thiel D, Kateřina K et al. Speed and power-related gene polymorphisms
  associated with playing position in elite soccer players. Biol Sport 2022; 39: 355-366
- 1000 78. Viru A, Loko J, Harro M et al. Critical periods in the development of performance
  1001 capacity during childhood and adolescence. Eur J Phys Edu 1999; 4: 75–119
- Devismes M, Aeles J, Philips J et al. Sprint force-velocity profiles in soccer players:
  impact of sex and playing level. Sports Biomech 2021; 20: 947-957.
  doi:10.1080/14763141.2019.1618900
- Little T, Williams AG. Specificity of acceleration, maximum speed, and agility in 1005 80. professional players. J Strength Cond Res 2005; 19: 76-78. soccer 1006 doi:10.1519/14253.1 1007
- 1008 81. Mero A. Force-Time Characteristics and Running Velocity of Male Sprinters during
  1009 the Acceleration Phase of Sprinting. Res Q Exerc Sport 1988; 59: 94-98.
  1010 doi:10.1080/02701367.1988.10605484
- 1011 82. Waldron M, Murphy A. A comparison of physical abilities and match performance
  1012 characteristics among elite and subelite under-14 soccer players. Pediatr Exerc Sci
  1013 2013; 25: 423-434. doi:10.1123/pes.25.3.423

43

- 101483.Venturelli M, Bishop D, Pettene L. Sprint training in preadolescent soccer players. Int1015J Sports Physiol Perform 2008; 3: 558-562. doi:10.1123/ijspp.3.4.558
- 1016 84. Nagano A, Komura T, Fukashiro S. Optimal coordination of maximal-effort
  1017 horizontal and vertical jump motions--a computer simulation study. Biomed Eng
  1018 Online 2007; 6: 20
- Fukashiro S, Besier TF, Barrett R et al. Direction control in standing horizontal and
  vertical jumps. . Int J Sport Health Sci 2005; 3: 272–279
- 102186.Ahmetov II, Donnikov AE, Trofimov DY. Actn3 genotype is associated with1022testosteronelevelsofathletes.BiolSport2014;31:105-108.1023doi:10.5604/20831862.1096046
- 1024 87. Dionísio TJ, Thiengo CR, Brozoski DT et al. The influence of genetic polymorphisms
  1025 on performance and cardiac and hemodynamic parameters among Brazilian soccer
  1026 players. Appl Physiol Nutr Metab 2017; 42: 596-604. doi:10.1139/apnm-2016-0608
  1027 88. Ahmetov, II, Williams AG, Popov DV et al. The combined impact of metabolic gene
- polymorphisms on elite endurance athlete status and related phenotypes. Hum Genet
  2009; 126: 751-761. doi:10.1007/s00439-009-0728-4
- 89. Massidda M, Scorcu M, Calò CM. New genetic model for predicting phenotype traits 1030 in sports. Int J Sports Physiol Perform 2014; 9: 554-560. doi:10.1123/ijspp.2012-0339 1031 90. Coelho DB, Pimenta E, Rosse IC et al. The alpha-actinin-3 r577x polymorphism and 1032 physical performance in soccer players. J Sports Med Phys Fitness 2016; 56: 241-248 1033 91. Coelho DB, Pimenta EM, Rosse IC et al. Polymorphism of the angiotensin converting 1034 enzyme gene (ACE-I/D) differentiates the aerobic and speed performance of football 1035 players. J Sports Med Phys Fitness 2022; 62: 192-198. doi:10.23736/s0022-1036 4707.21.12060-2 1037

- 1038 92. Kubo K, Kawakami Y, Fukunaga T. Influence of elastic properties of tendon
  1039 structures on jump performance in humans. J Appl Physiol (1985) 1999; 87: 20901040 2096. doi:10.1152/jappl.1999.87.6.2090
- 1041 93. Kubo K, Yata H, Tsunoda N. Effect of gene polymorphisms on the mechanical
  1042 properties of human tendon structures. Springerplus 2013; 2: 343
- Deprez DN, Fransen J, Lenoir M et al. A retrospective study on anthropometrical, 1043 94. physical fitness, and motor coordination characteristics that influence dropout, 1044 contract status, and first-team playing time in high-level soccer players aged eight to 1045 1046 eighteen years. J Strength Cond Res 2015; 29: 1692-1704. doi:10.1519/jsc.0000000000000806 1047
- 1048 95. Zigova T, Pencea V, Wiegand SJ et al. Intraventricular administration of BDNF
  1049 increases the number of newly generated neurons in the adult olfactory bulb. Mol Cell
  1050 Neurosci 1998; 11: 234-245. doi:10.1006/mcne.1998.0684
- McAllister AK, Katz LC, Lo DC. Neurotrophins and synaptic plasticity. Annu Rev
  Neurosci 1999; 22: 295-318. doi:10.1146/annurev.neuro.22.1.295
- 1053 97. Egan MF, Kojima M, Callicott JH et al. The BDNF val66met polymorphism affects
  1054 activity-dependent secretion of BDNF and human memory and hippocampal function.
  1055 Cell 2003; 112: 257-269. doi:10.1016/s0092-8674(03)00035-7
- 1056 98. Charlier R, Caspers M, Knaeps S et al. Limited potential of genetic predisposition
  1057 scores to predict muscle mass and strength performance in Flemish Caucasians
  1058 between 19 and 73 years of age. Physiol Genomics 2017; 49: 160-166.
  1059 doi:10.1152/physiolgenomics.00085.2016
- 1060 99. Dong B, Li Q, Zhang T et al. Population Genetic Polymorphism of Skeletal Muscle
  1061 Strength Related Genes in Five Ethnic Minorities in North China. Front Genet 2021;
  1062 12: 756802

- 1063 100. He L, Khanal P, Morse CI et al. Associations of combined genetic and epigenetic
  1064 scores with muscle size and muscle strength: a pilot study in older women. J Cachexia
  1065 Sarcopenia Muscle 2020; 11: 1548-1561
- 1066 101. He L, Van Roie E, Bogaerts A et al. Genetic predisposition score predicts the
  1067 increases of knee strength and muscle mass after one-year exercise in healthy elderly.
  1068 Exp Gerontol 2018; 111: 17-26. doi:10.1016/j.exger.2018.06.030
- 1069 102. Maciejewska-Skrendo A, Leźnicka K, Leońska-Duniec A et al. Genetics of Muscle
  1070 Stiffness, Muscle Elasticity and Explosive Strength. J Hum Kinet 2020; 74: 143-159
- 1071 103. Brownlee TE. An investigation into the capabilities and affecting factors of isometric
  1072 mid-thigh pull force production in elite youth soccer players. [Doctoral Thesis]:
  1073 Liverpool John Moores University; 2016. doi:10.24377/LJMU.t.00006072
- 1074 104. Baumert P, Stewart CE, Lake MJ et al. Variations of collagen-encoding genes are
  1075 associated with exercise-induced muscle damage. Physiol Genomics 2018; 50: 6911076 693. doi:10.1152/physiolgenomics.00145.2017
- 1077 105. Erskine RM, Williams AG, Jones DA et al. Do PTK2 gene polymorphisms contribute
  1078 to the interindividual variability in muscle strength and the response to resistance
  1079 training? A preliminary report. J Appl Physiol (1985) 2012; 112: 1329-1334.
  1080 doi:10.1152/japplphysiol.01137.2011
- 106. Massidda M, Eynon N, Bachis V et al. Association Between MCT1 A1470T
  Polymorphism and Fat-Free Mass in Well-Trained Young Soccer Players. J Strength
  Cond Res 2016; 30: 1171-1176. doi:10.1519/jsc.00000000001176
- 107. Erskine RM, Fletcher G, Folland JP. The contribution of muscle hypertrophy to
  strength changes following resistance training. Eur J Appl Physiol 2014; 114: 12391249. doi:10.1007/s00421-014-2855-4

46

- 1087 108. Nuell S, Illera-Domínguez V, Carmona G et al. Hamstring Muscle Volume as an
  1088 Indicator of Sprint Performance. J Strength Cond Res 2021; 35: 902-909.
  1089 doi:10.1519/jsc.00000000003976
- 1090 109. Boraita A, de la Rosa A, Heras ME et al. Cardiovascular adaptation, functional
  1091 capacity and Angiotensin-converting enzyme I/D polymorphism in elite athletes. Rev
  1092 Esp Cardiol 2010; 63: 810-819. doi:10.1016/s1885-5857(10)70166-3
- 1093 110. Papadimitriou ID, Papadopoulos C, Kouvatsi A et al. The ACTN3 gene in elite Greek
  1094 track and field athletes. Int J Sports Med 2008; 29: 352-355. doi:10.1055/s-20071095 965339
- 1096 111. Pescatello LS, Kostek MA, Gordish-Dressman H et al. ACE ID genotype and the
  1097 muscle strength and size response to unilateral resistance training. Med Sci Sports
  1098 Exerc 2006; 38: 1074-1081. doi:10.1249/01.mss.0000222835.28273.80
- 1099 112. Delecluse C. Influence of strength training on sprint running performance. Current
  1100 findings and implications for training. Sports Med 1997; 24: 147-156.
  1101 doi:10.2165/00007256-199724030-00001
- 1102 113. Dubouchaud H, Butterfield GE, Wolfel EE et al. Endurance training, expression, and
  physiology of LDH, MCT1, and MCT4 in human skeletal muscle. Am J Physiol
  Endocrinol Metab 2000; 278: E571-579. doi:10.1152/ajpendo.2000.278.4.E571
- 1105 114. Stafilidis S, Arampatzis A. Muscle tendon unit mechanical and morphological
  1106 properties and sprint performance. J Sports Sci 2007; 25: 1035-1046.
  1107 doi:10.1080/02640410600951589
- 1108 115. Morin JB, Gimenez P, Edouard P et al. Sprint Acceleration Mechanics: The Major
  1109 Role of Hamstrings in Horizontal Force Production. Front Physiol 2015; 6: 404

- 1110 116. Ding X, Jin Y, Wu Y et al. Localization and cellular distribution of CPNE5 in
  1111 embryonic mouse brain. Brain Res 2008; 1224: 20-28.
  1112 doi:10.1016/j.brainres.2008.05.051
- 1113 117. Reiner A, Yang M, Cagle MC et al. Localization of cerebellin-2 in late embryonic
  1114 chicken brain: implications for a role in synapse formation and for brain evolution. J
  1115 Comp Neurol 2011; 519: 2225-2251
- 1116 118. Fernandez T, Morgan T, Davis N et al. Disruption of contactin 4 (CNTN4) results in
  1117 developmental delay and other features of 3p deletion syndrome. Am J Hum Genet
  1118 2004; 74: 1286-1293
- 1119 119. Stølen T, Chamari K, Castagna C et al. Physiology of soccer: an update. Sports Med
  2005; 35: 501-536. doi:10.2165/00007256-200535060-00004
- 1121 120. Bueno S, Pasqua LA, de Araújo G et al. The Association of ACE Genotypes on
  1122 Cardiorespiratory Variables Related to Physical Fitness in Healthy Men. PLoS One
  1123 2016; 11: e0165310
- 1124 121. Konopka MJ, van den Bunder J, Rietjens G et al. Genetics of long-distance runners
  and road cyclists-A systematic review with meta-analysis. Scand J Med Sci Sports
  2022; 32: 1414-1429
- 1127 122. Papadimitriou ID, Lockey SJ, Voisin S et al. No association between ACTN3 R577X
  1128 and ACE I/D polymorphisms and endurance running times in 698 Caucasian athletes.
  1129 BMC Genomics 2018; 19: 13. doi:10.1186/s12864-017-4412-0
- 1130 123. Mirwald RL, Baxter-Jones AD, Bailey DA et al. An assessment of maturity from
  1131 anthropometric measurements. Med Sci Sports Exerc 2002; 34: 689-694.
  1132 doi:10.1097/00005768-200204000-00020

124. Casamichana D, Castellano J, Castagna C. Comparing the physical demands of
friendly matches and small-sided games in semiprofessional soccer players. J Strength
Cond Res 2012; 26: 837-843. doi:10.1519/JSC.0b013e31822a61cf

- 1136 125. Pickering C, Kiely J, Suraci B et al. The magnitude of Yo-Yo test improvements
  1137 following an aerobic training intervention are associated with total genotype score.
  1138 PLoS One 2018; 13: e0207597
- 126. Parry L, Drust B. Is injury the major cause of elite soccer players being unavailable to
  train and play during the competitive season? Phys Ther Sport 2006; 7: 58-64
- 1141 127. Eirale C, Tol JL, Farooq A et al. Low injury rate strongly correlates with team success
  in Qatari professional football. Br J Sports Med 2013; 47: 807-808
- 1143 128. Carling C, Le Gall F, McCall A et al. Squad management, injury and match 1144 performance in a professional soccer team over a championship-winning season. Eur J 1145 Sport Sci 2015; 15: 573-582. doi:10.1080/17461391.2014.955885
- 129. Larruskain J, Lekue JA, Diaz N et al. A comparison of injuries in elite male and
  female football players: A five-season prospective study. Scand J Med Sci Sports
  2018; 28: 237-245. doi:10.1111/sms.12860
- 130. Varley I, Patel S, Williams AG et al. The current use, and opinions of elite athletes
  and support staff in relation to genetic testing in elite sport within the UK. Biol Sport
  2018; 35: 13-19
- 1152 131. Tucker AM. Common soccer injuries. Diagnosis, treatment and rehabilitation. Sports
  1153 Med 1997; 23: 21-32. doi:10.2165/00007256-199723010-00003
- 132. Laguette MJ, Abrahams Y, Prince S et al. Sequence variants within the 3'-UTR of the
  COL5A1 gene alters mRNA stability: implications for musculoskeletal soft tissue
  injuries. Matrix Biol 2011; 30: 338-345. doi:10.1016/j.matbio.2011.05.001

- 133. Posthumus M, September AV, Keegan M et al. Genetic risk factors for anterior 1157 cruciate ligament ruptures: COL1A1 gene variant. Br J Sports Med 2009; 43: 352-1158 356. doi:10.1136/bjsm.2008.056150 1159
- Posthumus M, September AV, O'Cuinneagain D et al. The COL5A1 gene is 134. 1160 associated with increased risk of anterior cruciate ligament ruptures in female 1161 participants. Am J Sports Med 2009; 37: 2234-2240. doi:10.1177/0363546509338266

1162

- 135. September AV, Cook J, Handley CJ et al. Variants within the COL5A1 gene are 1163 associated with Achilles tendinopathy in two populations. Br J Sports Med 2009; 43: 1164 357-365. doi:10.1136/bjsm.2008.048793 1165
- Ahmetov II, Hall ECR, Semenova EA et al. Advances in sports genomics. Adv Clin 136. 1166 Chem 2022; 107: 215-263. doi:10.1016/bs.acc.2021.07.004 1167
- 137. Broos S, Malisoux L, Theisen D et al. Evidence for ACTN3 as a Speed Gene in 1168 Isolated Human Muscle Fibers. PLoS One 2016; 11: e0150594 1169
- Timmins RG, Bourne MN, Shield AJ et al. Short biceps femoris fascicles and 1170 138. eccentric knee flexor weakness increase the risk of hamstring injury in elite football 1171 (soccer): a prospective cohort study. Br J Sports Med 2016; 50: 1524-1535. 1172 doi:10.1136/bjsports-2015-095362 1173
- Pimenta EM, Coelho DB, Cruz IR et al. The ACTN3 genotype in soccer players in 139. 1174 response to acute eccentric training. Eur J Appl Physiol 2012; 112: 1495-1503. 1175 doi:10.1007/s00421-011-2109-7 1176
- Clos E, Pruna R, Lundblad M et al. ACTN3 single nucleotide polymorphism is 140. 1177 associated with non-contact musculoskeletal soft-tissue injury incidence in elite 1178 professional football players. Knee Surg Sports Traumatol Arthrosc 2019; 27: 4055-1179 4061. doi:10.1007/s00167-019-05381-x 1180

- 141. Bradley PS, Dellal A, Mohr M et al. Gender differences in match performance
  characteristics of soccer players competing in the UEFA Champions League. Hum
  Mov Sci 2014; 33: 159-171. doi:10.1016/j.humov.2013.07.024
- Waldén M, Hägglund M, Werner J et al. The epidemiology of anterior cruciate 142. 1184 ligament injury in football (soccer): a review of the literature from a gender-related 1185 perspective. Knee Surg Sports Traumatol Arthrosc 2011; 19: 3-10. 1186 doi:10.1007/s00167-010-1172-7 1187
- 1188 143. Del Coso J, Rodas G, Buil M et al. Association of the ACTN3 rs1815739
  1189 Polymorphism with Physical Performance and Injury Incidence in Professional
  1190 Women Football Players. Genes (Basel) 2022; 13
- 144. Baumert P, Lake MJ, Stewart CE et al. Genetic variation and exercise-induced muscle
  damage: implications for athletic performance, injury and ageing. Eur J Appl Physiol
  2016; 116: 1595-1625
- 1194 145. Mann V, Hobson EE, Li B et al. A COL1A1 Sp1 binding site polymorphism
  predisposes to osteoporotic fracture by affecting bone density and quality. J Clin
  Invest 2001; 107: 899-907
- 1197 146. Boot-Handford RP, Tuckwell DS. Fibrillar collagen: the key to vertebrate evolution?
  1198 A tale of molecular incest. Bioessays 2003; 25: 142-151. doi:10.1002/bies.10230
- 147. Posthumus M, September AV, Schwellnus MP et al. Investigation of the Sp1-binding 1199 site polymorphism within the COL1A1 gene in participants with Achilles tendon 1200 injuries controls. J Sci 2009; 12: 1201 and Med Sport 184-189. doi:10.1016/j.jsams.2007.12.006 1202
- 148. Jin H, van't Hof RJ, Albagha OM et al. Promoter and intron 1 polymorphisms of
  COL1A1 interact to regulate transcription and susceptibility to osteoporosis. Hum
  Mol Genet 2009; 18: 2729-2738. doi:10.1093/hmg/ddp205

 1212
 151.
 Zacch

 1213
 pressu

 1214
 152.
 Rande

 1215
 transfe

 1216
 Am Se

 1217
 153.
 Kanne

 1218
 10: 31

 1219
 154.
 Posthe

 1220
 chrom

 1221
 155.
 Birkee

1206 149. Collins M, Posthumus M. Type V collagen genotype and exercise-related phenotype
1207 relationships: a novel hypothesis. Exerc Sport Sci Rev 2011; 39: 191-198.
1208 doi:10.1097/JES.0b013e318224e853

1209 150. Wenstrup RJ, Florer JB, Brunskill EW et al. Type V collagen controls the initiation of
1210 collagen fibril assembly. J Biol Chem 2004; 279: 53331-53337.
1211 doi:10.1074/jbc.M409622200

- 1212 151. Zacchigna L, Vecchione C, Notte A et al. Emilin1 links TGF-beta maturation to blood
  1213 pressure homeostasis. Cell 2006; 124: 929-942. doi:10.1016/j.cell.2005.12.035
- 214 152. Randell A, Daneshtalab N. Elastin microfibril interface-located protein 1,
  215 transforming growth factor beta, and implications on cardiovascular complications. J
  216 Am Soc Hypertens 2017; 11: 437-448. doi:10.1016/j.jash.2017.04.010
- 217 153. Kannus P. Structure of the tendon connective tissue. Scand J Med Sci Sports 2000;
  218 10: 312-320. doi:10.1034/j.1600-0838.2000.010006312.x
- Posthumus M, Collins M, van der Merwe L et al. Matrix metalloproteinase genes on
  chromosome 11q22 and the risk of anterior cruciate ligament (ACL) rupture. Scand J
  Med Sci Sports 2012; 22: 523-533. doi:10.1111/j.1600-0838.2010.01270.x
- 1222 155. Birkedal-Hansen H, Moore WG, Bodden MK et al. Matrix metalloproteinases: a
  1223 review. Crit Rev Oral Biol Med 1993; 4: 197-250.
  1224 doi:10.1177/10454411930040020401

1225 156. Chen X, Li Y. Role of matrix metalloproteinases in skeletal muscle: migration,
1226 differentiation, regeneration and fibrosis. Cell Adh Migr 2009; 3: 337-341

1227 157. Medley TL, Kingwell BA, Gatzka CD et al. Matrix metalloproteinase-3 genotype
1228 contributes to age-related aortic stiffening through modulation of gene and protein
1229 expression. Circ Res 2003; 92: 1254-1261. doi:10.1161/01.Res.0000076891.24317.Ca

- 1230 158. Guo YQ, Zheng LN, Wei JF et al. Expression of CCL2 and CCR2 in the hippocampus
  and the interventional roles of propofol in rat cerebral ischemia/reperfusion. Exp Ther
  Med 2014; 8: 657-661
- 1233 159. Hubal MJ, Chen TC, Thompson PD et al. Inflammatory gene changes associated with
  the repeated-bout effect. Am J Physiol Regul Integr Comp Physiol 2008; 294: R16281637. doi:10.1152/ajpregu.00853.2007
- 1236 160. Catoire M, Kersten S. The search for exercise factors in humans. FASEB J 2015; 29:
  1237 1615-1628. doi:10.1096/fj.14-263699
- 1238 161. Hubal MJ, Devaney JM, Hoffman EP et al. CCL2 and CCR2 polymorphisms are
  1239 associated with markers of exercise-induced skeletal muscle damage. J Appl Physiol
  1240 (1985) 2010; 108: 1651-1658. doi:10.1152/japplphysiol.00361.2009
- 1241 162. Yahiaoui L, Gvozdic D, Danialou G et al. CC family chemokines directly regulate
  1242 myoblast responses to skeletal muscle injury. J Physiol 2008; 586: 3991-4004
- 1243 163. Fishman D, Faulds G, Jeffery R et al. The effect of novel polymorphisms in the 1244 interleukin-6 (IL-6) gene on IL-6 transcription and plasma IL-6 levels, and an 1245 association with systemic-onset juvenile chronic arthritis. J Clin Invest 1998; 102: 1246 1369-1376
- 1247 164. Andersen MB, Pingel J, Kjær M et al. Interleukin-6: a growth factor stimulating
  1248 collagen synthesis in human tendon. J Appl Physiol (1985) 2011; 110: 1549-1554.
  1249 doi:10.1152/japplphysiol.00037.2010
- 165. Muñoz-Cánoves P, Scheele C, Pedersen BK et al. Interleukin-6 myokine signaling in
  skeletal muscle: a double-edged sword? FEBS J 2013; 280: 4131-4148
- 1252 166. Pedersen BK, Febbraio MA. Muscle as an endocrine organ: focus on muscle-derived
  1253 interleukin-6. Physiol Rev 2008; 88: 1379-1406. doi:10.1152/physrev.90100.2007

- 1254 167. Nieman DC, Nehlsen-Cannarella SL, Fagoaga OR et al. Influence of mode and
  1255 carbohydrate on the cytokine response to heavy exertion. Med Sci Sports Exerc 1998;
  1256 30: 671-678. doi:10.1097/00005768-199805000-00005
- 1257 168. Millar NL, Wei AQ, Molloy TJ et al. Cytokines and apoptosis in supraspinatus
  1258 tendinopathy. J Bone Joint Surg Br 2009; 91: 417-424. doi:10.1302/03011259 620x.91b3.21652
- 1260 169. Yamin C, Duarte JA, Oliveira JM et al. IL6 (-174) and TNFA (-308) promoter 1261 polymorphisms are associated with systemic creatine kinase response to eccentric 1262 exercise. Eur J Appl Physiol 2008; 104: 579-586. doi:10.1007/s00421-008-0728-4
- 1263 170. Massidda M, Eynon N, Bachis V et al. Influence of the MCT1 rs1049434 on Indirect
  1264 Muscle Disorders/Injuries in Elite Football Players. Sports Med Open 2015; 1: 33
- 1265 171. Massidda M, Corrias L, Bachis V et al. Vitamin D receptor gene polymorphisms and
  1266 musculoskeletal injuries in professional football players. Exp Ther Med 2015; 9:
  1267 1974-1978
- 1268 172. Bouchard C. Overcoming barriers to progress in exercise genomics. Exerc Sport Sci
  1269 Rev 2011; 39: 212-217
- Mills M, Yang N, Weinberger R et al. Differential expression of the actin-binding 1270 173. proteins, alpha-actinin-2 and -3, in different species: implications for the evolution of 1271 functional redundancy. Mol 2001; 10: 1335-1346. Hum Genet 1272 doi:10.1093/hmg/10.13.1335 1273
- 1274 174. Reneland R, Lithell H. Angiotensin-converting enzyme in human skeletal muscle. A
  1275 simple in vitro assay of activity in needle biopsy specimens. Scand J Clin Lab Invest
  1276 1994; 54: 105-111. doi:10.3109/00365519409086516

- 1277 175. Gordon SE, Davis BS, Carlson CJ et al. ANG II is required for optimal overload1278 induced skeletal muscle hypertrophy. Am J Physiol Endocrinol Metab 2001; 280:
  1279 E150-159. doi:10.1152/ajpendo.2001.280.1.E150
- 176. Rigat B, Hubert C, Alhenc-Gelas F et al. An insertion/deletion polymorphism in the
  angiotensin I-converting enzyme gene accounting for half the variance of serum
  enzyme levels. J Clin Invest 1990; 86: 1343-1346
- 1283 177. Charbonneau DE, Hanson ED, Ludlow AT et al. ACE genotype and the muscle
  hypertrophic and strength responses to strength training. Med Sci Sports Exerc 2008;
  40: 677-683
- 178. North KN, Beggs AH. Deficiency of a skeletal muscle isoform of alpha-actinin
  (alpha-actinin-3) in merosin-positive congenital muscular dystrophy. Neuromuscul
  Disord 1996; 6: 229-235. doi:10.1016/0960-8966(96)00361-6
- 1289 179. Sethi AA, Nordestgaard BG, Tybjaerg-Hansen A. Angiotensinogen gene
  polymorphism, plasma angiotensinogen, and risk of hypertension and ischemic heart
  disease: a meta-analysis. Arterioscler Thromb Vasc Biol 2003; 23: 1269-1275.
  doi:10.1161/01.Atv.0000079007.40884.5c
- 1293 180. Morisaki T, Gross M, Morisaki H et al. Molecular basis of AMP deaminase
  1294 deficiency in skeletal muscle. Proc Natl Acad Sci U S A 1992; 89: 6457-6461
- 1295 181. Kuntz LA, Rossetti L, Kunold E et al. Biomarkers for tissue engineering of the
  1296 tendon-bone interface. PLoS One 2018; 13: e0189668
- 1297 182. Birk DE, Fitch JM, Babiarz JP et al. Collagen fibrillogenesis in vitro: interaction of
  1298 types I and V collagen regulates fibril diameter. J Cell Sci 1990; 95 (Pt 4): 649-657.
- doi:10.1242/jcs.95.4.649

- 1300 183. Halestrap AP, Meredith D. The SLC16 gene family-from monocarboxylate
  1301 transporters (MCTs) to aromatic amino acid transporters and beyond. Pflugers Arch
  1302 2004; 447: 619-628. doi:10.1007/s00424-003-1067-2
- 1303 184. Massidda M, Flore L, Kikuchi N et al. Influence of the MCT1-T1470A polymorphism
  1304 (rs1049434) on repeated sprint ability and blood lactate accumulation in elite football
  1305 players: a pilot study. Eur J Appl Physiol 2021; 121: 3399-3408. doi:10.1007/s004211306 021-04797-z
- 1307 185. Nakayama M, Yasue H, Yoshimura M et al. T(-786)--> C mutation in the 5'-flanking
  1308 region of the endothelial nitric oxide synthase gene is associated with myocardial
  1309 infarction, especially without coronary organic stenosis. Am J Cardiol 2000; 86: 6281310 634. doi:10.1016/s0002-9149(00)01041-9
- 186. Kersten S. Integrated physiology and systems biology of PPARα. Mol Metab 2014; 3:
  1312 354-371
- 1313 187. Ahmetov, II, Mozhayskaya IA, Flavell DM et al. PPARalpha gene variation and 1314 physical performance in Russian athletes. Eur J Appl Physiol 2006; 97: 103-108. 1315 doi:10.1007/s00421-006-0154-4
- 1316 188. Auton A, Brooks LD, Durbin RM et al. A global reference for human genetic
  1317 variation. Nature 2015; 526: 68-74
- 1318 189. McKee TJ, Perlman G, Morris M et al. Extracellular matrix composition of
  1319 connective tissues: a systematic review and meta-analysis. Sci Rep 2019; 9: 10542
- 1320 190. Oh VM, Chua BM, Heng CK et al. Association of intronic single-nucleotide
  polymorphisms in the EMILIN1 gene with essential hypertension in a Chinese
  population. J Hum Hypertens 2012; 26: 553-561. doi:10.1038/jhh.2011.68
- 1323 191. Nie G, Wen X, Liang X et al. Additional evidence supports association of common1324 genetic variants in MMP3 and TIMP2 with increased risk of chronic Achilles





#### 1329 Figure legend

**Figure 1.** Association between the *COL2A1* rs2070739 SNP and isometric mid-thigh pull maximum voluntary force (MVF) normalised to body mass (BM) in 148 high-level male academy soccer players (ASP) and 93 age- and sex-matched non-athletes (NA). \* Main effect for genotype between CC (black bars) and CT/TT (chequered bars) genotypes (P = 0.033), ^ main effect for cohort and maturity status (P < 0.001).

1335

#### 1336 Table headings

Table 1. Key polymorphisms associated with athlete status and physical performance
phenotypes in soccer players. Alleles for each polymorphism are reported according to the
forward DNA strand.

Table 2. Key polymorphisms associated with injury risk in soccer players. Alleles foreach polymorphism are reported according to the forward DNA strand.

1342

#### Table 1. Key polymorphisms associated with athlete status and physical performance phenotypes in soccer players. Alleles for each polymorphism are reported

according to the forward DNA strand.

Gene	Encoded protein	Protein function	Alleles and rs number	Polymorphism function	Associations with phenotypes of interest in soccer
ACE	Angiotensin-I converting enzyme	ACE converts angiotensin-I (Ang I) to Ang II and is expressed in SkM [174], where Ang II modulates SkM hypertrophy after mechanical loading [175].	Insertion (I)/ Deletion (D) of 287 base pairs rs1799752 / rs4340 / rs13447447 / rs4646994	D-allele carriers express higher ACE activity than II homozygotes [176].	<ul> <li><i>Phenotype: Athlete status</i></li> <li>D allele more frequent in pro players vs. NA [30, 32].</li> <li>Possibly due to D allele influencing circulating ACE and Ang II in SkM, potentially leading to larger [177], more powerful muscles.</li> <li><i>Phenotype: Sprint Performance</i></li> <li>DD genotype associated with faster 20 m sprint speed compared to II genotype but no difference in vertical CMJ performance in 212 high-level pro players [91].</li> </ul>
ACTN3	α-actinin-3	Binds actin to the Z- line in type II SkM fibres [48]; blocks calcineurin, inhibiting fast-to-slow myofibre transition [26].	C(R)>T(X) rs1815739	X allele leads to a stop codon at amino acid 577, preventing protein production [178]. Consequently, the R allele is associated with greater type II SkM fibre composition [49].	<ul> <li>Phenotype: Athlete status</li> <li>X allele (generally associated with endurance phenotypes) more frequent in pre-PHV academy players vs. non-athletes (NA) [28];</li> <li>R allele (generally associated with strength/power phenotypes) more frequent in pro players vs. NA [30, 34];</li> <li>Above discrepancy probably due to variability in match demands between pre-(endurance-based) and post-PHV/adult (power/speed-based) players [28];</li> <li>Combination of 'favourable' alleles from PPARA rs4253778 (C), NOS3 rs2070744 (T), ACTN3 rs1815739 (R), and AGT rs699 (G) SNPs correlated with agility performance in pre-PHV academy players [28].</li> <li>Phenotype: CMJ (Vertical Power) Performance</li> <li>R allele associated with greater bilateral vertical CMJ height in pro players [37].</li> <li>Phenotype: Aerobic Capacity</li> <li>XX genotype associated with greater YoYo score in pro players [37].</li> </ul>
AGT	Angiotensinogen	Precursor of Ang II.	A>G rs699	G allele associated with higher blood AGT • concentration and higher •	<i>Phenotype: Athlete status</i> D allele more frequent in pro players vs. NA [30, 32]. G allele more frequent in academy players vs. NA [28];

			• blood pressure [179].	Combination of 'favourable' alleles from <i>PPARA</i> rs4253778 (C), <i>NOS3</i> rs2070744 (T), <i>ACTN3</i> rs1815739 (R), and <i>AGT</i> rs699 (G) SNPs correlated with agility performance in pre-PHV academy players [28].
AMPD1	Adenosine Monophosphate Deaminase 1	Enzyme catalysing G>A the deamination of rs17602 AMP to IMP in SkM	A allele results in a premature stop codon and non-functional enzyme, causing impaired AMP metabolism, which can lead to muscle fatigue, weakness and cramping [180].	<ul> <li>Phenotype: Horizontal Acceleration/ Sprint/ Agility Performance</li> <li>GG genotype associated with faster 10 m acceleration and 20 m sprint times than pre-PHV A-allele carriers in pre-PHV academy players [28].</li> <li>Phenotype: Agility (Change of Direction) Performance</li> <li>Combination of 'favourable' alleles from AMPD1 rs17602729 (G), BDNF rs6265 (C), COL5A1 rs12722 (C), COL2A1 rs2070739 (C) SNPs correlated with agility performance in pre-PHV academy players [28].</li> </ul>
BDNF	Brain-derived neurotrophic factor	Promotes neurone C>T rs6. growth, differentiation, and maintenance [95].	265 C allele results in valine rather than methionine (T allele) at amino acid 66, leading to greater abundance of exercise- induced serum BDNF concentration [97].	<ul> <li><i>Phenotype: Horizontal Acceleration/ Sprint/ Agility Performance</i></li> <li>CC genotype associated with greater 10 m acceleration and 20 m sprint performance in academy players [28], possibly linked to C allele's relation with higher exercise-induced serum BDNF concentration [97], potentially enhancing neuromuscular activation/control during acceleration/sprint performance.</li> <li><i>Phenotype: Agility (Change of Direction) Performance</i></li> <li>Combination of 'favourable' alleles from <i>AMPD1</i> rs17602729 (G), <i>BDNF</i> rs6265 (C), <i>COL5A1</i> rs12722 (C), <i>COL2A1</i> rs2070739 (C) SNPs correlated with agility performance in pre-PHV academy players [28].</li> </ul>
COL2A1	Pro-α1 (II) chain	Major component of C>T type II collagen, rs20707 providing structure and strength to connective tissue, e.g. at the enthesis [181].	Not yet known but 3'-UTR SNPs have the potential to alter the level, location, or timing of gene expression [25].	<ul> <li>Phenotype: Horizontal Acceleration/ Sprint/ Agility Performance</li> <li>CC genotype associated with faster 10 m acceleration times than T allele carriers in academy players [28].</li> <li>Phenotype: Agility (Change of Direction) Performance</li> <li>Combination of 'favourable' alleles from AMPD1 rs17602729 (G), BDNF</li> <li>rs6265 (C), COL5A1 rs12722 (C), COL2A1 rs2070739 (C) SNPs correlated with agility performance in pre-PHV academy players [28].</li> </ul>
COL5A1	Pro-α1 (V) chain	Major component of C>T type V collagen, rs1272 which regulates the diameter of collagen fibrils [182].	Not yet known but 3'-UTR 2 SNPs have the potential to alter the level, location, or timing of gene expression [25].	<b>Phenotype: CMJ (Vertical Power) Performance</b> CC genotype associated with greater bilateral vertical CMJ height in academy players [28], possibly linked to more extensible knee extensor tendon in CC homozygotes [93], enabling greater storage and release of energy, thus amplifying power during vertical CMJ.

genotype ag greater releration,
b), <i>BDNF</i> correlated
781 NA
mulation 71].
C), NOS3 correlated

						<ul> <li><i>Phenotype: Horizontal Acceleration/ Sprint/ Agility Performance</i></li> <li>CC genotype associated with quicker acceleration, sprint and horizontal CMJ performance in academy players [28], possibly linked to CC genotype associated with more extensible knee extensor tendon [93], enabling greater storage and release of energy, thus amplifying power during acceleration, sprint and horizontal CMJ.</li> <li><i>Phenotype: Agility (Change of Direction) Performance</i></li> <li>Combination of 'favourable' alleles from <i>AMPD1</i> rs17602729 (G), <i>BDNF</i></li> </ul>
						rs6265 (C), <i>COL5A1</i> rs12722 (C), <i>COL2A1</i> rs2070739 (C) SNPs correlated with agility performance in pre-PHV academy players [28].
MCT1	Monocarboxylate transporter-1	Transports lactate and H <sup>+</sup> across SkM sarcolemma, found predominantly in oxidative fibres [183].	A>T rs1049434	Not yet known but intronic SNPs have the potential to influence gene expression and mRNA stability [25]. T allele results in aspartic acid replacing glutamic acid at position 490, potentially causing a defect in MCT1's ability to transport lactate.		<ul> <li><i>Phenotype: Athlete status</i></li> <li>A allele and AA genotype more frequent in 148 pro forwards than 781 NA [69].</li> <li><i>Phenotype: Horizontal Acceleration/ Sprint/ Agility Performance</i></li> <li>A-allele associated with faster time in final two sprints of RSA assessment compared to TT homozygotes in 26 academy players [184].</li> <li>The above associations may be linked to increased blood lactate accumulation after high-intensity exercise in T-allele carriers vs. AA homozygotes [71].</li> </ul>
NOS3	Nitric oxide synthase-3	Potentially stimulating muscle hypertrophy through NO-mediated vasodilatation [58].	C>T rs2070744	T allele increases gene promoter activity, thus increasing eNOS and NO synthesis [185].		<ul> <li>Phenotype: Athlete status</li> <li>T allele greater in academy players vs. NA [28].</li> <li>Combination of 'favourable' alleles from PPARA rs4253778 (C), NOS3 rs2070744 (T), ACTN3 rs1815739 (R), and AGT rs699 (G) SNPs correlated with agility performance in pre-PHV academy players [28].</li> <li>Phenotype: Horizontal Acceleration/ Sprint/ Agility Performance</li> <li>TT genotype associated with greater acceleration and sprint performance in academy players [28], possibly linked to greater number of sarcomeres in series, thus increasing SkM contraction velocity [53].</li> </ul>
PPARA	Peroxisome proliferator- activated receptor-α	Present in SkM and promotes uptake, utilization, and catabolism of fatty	G>C rs4253778	Not yet known but intronic SNPs have the potential to influence gene expression	•	<b>Phenotype:</b> Athlete status C allele more frequently distributed in post-PHV academy [28] and pro players [30] vs. NA, possibly due to C allele's association with greater SkM type II fibre composition [187].

acids (FAs) by and mRNA stability [25]. • ( upregulation of genes involved in FA transport, FA binding activation, and peroxisomal and mito chondrial FA β- oxidation [186].	Combination of 'favourable' alleles from <i>PPARA</i> rs4253778 (C), <i>NOS3</i> rs2070744 (T), <i>ACTN3</i> rs1815739 (R), and <i>AGT</i> rs699 (G) SNPs correlated with agility performance in pre-PHV academy players [28].
---	--

SNP, single nucleotide polymorphism; MAF, minor allele frequency (according to European ancestry [188]); SkM, skeletal muscle; NA, non-athletes; PHV, peak height

Downloaded by: Liverpool John Moores University. Copyrighted material

velocity; *RSA*, repeated sprint ability.

Table 2. Key polymorphisms associated with injury risk in soccer players. Alleles for each polymorphism are reported according to the forward DNA strand.

Gene	Encoded protein	Protein function	Alleles and rs number	Polymorphism function	Associations with injury/ poor recovery
ACTN3	α-actinin-3	Binds actin to the Z- line in type II SkM fibres [48]; blocks calcineurin, inhibiting fast-to-slow myofibre transition [26].	C(R)>T(X) rs1815739	X allele leads to a stop codon at amino acid 577, preventing protein production [178]. Consequently, the R allele is associated with greater type II SkM fibre composition [49].	<ul> <li>XX genotype associated with greater indices of exercise-induced muscle damage than RR genotype in pro players [139], possibly due to less structural stability of the sarcomere during contraction [144] and/or having smaller, weaker and less powerful muscles [27], thus potentially experiencing relatively greater ground reaction forces.</li> <li>XX genotype associated with higher rate of non-contact musculoskeletal soft-tissue injuries in 42 pro players [140].</li> <li>XX genotype associated with higher rate of non-contact muscle injuries in XX homozygotes than R-allele carriers in pro players, with XX homozygotes most likely to suffer severe injury [44].</li> <li>X-allele carriers most likely to be injured in pro players, with XX homozygotes having the longest absence after injury [47].</li> <li>X-allele carriers missed more days following ankle injuries than RR homozygotes in academy players [29], suggesting greater muscle size and strength associated with the R allele [27] may enhance limb stability and favour return to play.</li> <li>No association with hamstring injuries in pro players [40].</li> </ul>
CCL2	C-C motif chemokine ligand-2 (CCL2)	CCL2 is expressed within the interstitial space between myofibres after damaging activity [159] and mediates systemic changes with chronic exercise [160].	G>C rs2857656	Affects plasma CCL2 levels [158]. Plasma CCL2 is lower in GG homozygotes [158], who exhibit lower muscle strength than C-allele • carriers [161].	GG genotype associated with more severe muscle injuries than C-allele carriers in pro players [45, 46]. Not associated with hamstring injuries in pro players [40]. Not associated with injury prevalence or recovery time in academy players [29]. Considering the prospective role of CCL2 in satellite cell proliferation [162], higher CCL2 during recovery from muscle injury may be advantageous to soccer players carrying at least one C allele.
COL1A1	Pro-α1 (I) chain	Major component of type I collagen, a structural protein found in most connective tissues,	C>A rs1800012	Affects binding site for the $OL1A1$ Sp1 transcription factor. A allele associated with greater $\alpha$ 1 (I) chain	Rare AA genotype appears protective against ACL [133] and Achilles tendon [147] injuries in athletes. Pro soccer players with ACL ruptures are less likely to be AA homozygotes [39].

_		including ligament and tendon [189].		mRNA and protein [145].	•	However, others found no associations between this SNP and injury [29, 40, 46].
COL5A1	Pro-α1 (V) chain	Major component of type V collagen, which regulates the diameter of collagen fibrils [182].	C>T rs12722	Not yet known but 3'-UTR SNPs have the potential to alter the level, location, or timing of gene expression [25].	•	TC genotype associated with severe hamstring injuries in pro players [45, 46] but sample sizes were low in these studies (n=74). More severe musculoskeletal injuries are exhibited by TT homozygous players than C-allele carriers [42]. C-allele carriers in two different <i>COL5A1</i> SNPs (rs12722 and rs13946) was protective against ACL rupture in pro players [41]. Higher prevalence of soft-tissue and ligament injuries in pre-PHV academy players with the C allele and CC genotype, respectively [29]. Considering rs12722 may influence mRNA stability and affect the tensile strength and stiffness of collagen fibrils [149], the <i>COL5A1</i> rs12722 SNP has a plausible association with musculoskeletal injuries in soccer.
EMILIN1	Elastin microfibril interfacer-1	ECM glycoprotein that assists the fusion of elastin fibres during elastogenesis [152] and provides elasticity to ligaments and tendons [153].	C>T rs2289360	Currently unknown. CC genotype associated with hypertension, possibly by reducing arterial compliance [190].	•	CC genotype associated with the highest overall injury prevalence, while T- allele carriers missed the most days following ankle injuries in academy players [29]. TT genotype associated with more severe MCL injuries, but not with the incidence or severity of muscle or tendon injuries in pro players [38, 45]. Not associated with hamstring injuries in pro players [40].
IL6	Interleukin-6 (IL- 6)	IL-6 is a myokine produced in SkM after exercise and is functionally related growth, atrophy and collagen synthesis [164, 165]. Potential role as signaling molecule associated with post-exercise satellite cell proliferation.	C>G rs1800795	Possibly affects glucocorticoid receptor and transcription. G allele associated with increased plasma IL6 [163].	•	GG genotype associated with greater hamstring injury risk in pro players [40]. Contrastingly, CC homozygotes associated with higher prevalence of all injury types and specifically muscle injury in post-PHV academy players [29]. Having both pro- and anti- [166] inflammatory capacities, it is plausible that IL-6 could increase or reduce injury risk. Increased plasma IL-6 associated with the G allele [163] may heighten soft- tissue injury risk. Because eccentric contractions induce pro-inflammatory IL-6 expression [167], and because cytokines trigger tenocyte apoptosis and ECM degradation [168], GG homozygotes may experience these unfavourable changes via increased IL-6, heightening injury risk.
MMP3	Matrix metalloproteinase	One of >20 MMPs that can catalytically	C>T	Currently unknown. C>T substitution replaces	•	rs679620 T allele associated with hamstring injury in pro players [40]. rs679620 T allele associated with longer absence following knee injuries in

- 3	degrade collagens and ECM substrates [155]. Also activates other MMPs and important for SkM	rs679620	glutamate residue with lysin residue but does not affect MMP3 protein function [191].	academy soccer players [29]. rs679620 T allele is in linkage disequilibrium with <i>MMP3</i> rs3025058 5A allele, which increases MMP3 expression [157]. rs679620 T allele might increase RNA expression and intensify the degradation of proteins such as collagen and elastin weakening the structural
	regeneration [156].			degradation of proteins, such as collagen and elastin, weakening the structural integrity of ligament and ECM. However, further research is required.

*SNP*, single nucleotide polymorphism; *MAF*, minor allele frequency (according to European ancestry [188]); *SkM*, skeletal muscle; *PHV*, peak height velocity.



