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Metabolic syndrome: an overview on its genetic associations and gene-diet interactions

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Abstract

Metabolic syndrome (MetS) is a cluster of cardio-metabolic risk factors that includes central obesity, hyperglycemia, hypertension, and dyslipidemias and whose inter-related occurrence may increase the odds of developing type 2 diabetes and cardiovascular diseases. MetS has become one of the most studied condition, nevertheless due to its complex etiology this has not been fully elucidated. Recent evidence describes that both genetic and environmental factors play an important role on its development. With the advent of genomic-wide association studies (GWAS), single nucleotide polymorphisms (SNPs) have gained special importance. In this review, we present an update of the genetics surrounding MetS as a single entity as well as its corresponding risk factors, considering SNPs and gene-diet interactions related to cardio-metabolic markers. Here, we focus on the conceptual aspects, diagnostic criteria as well as the role of genetics, particularly on SNPs and polygenic risk scores (PRS) for inter-individual analysis. In

addition, this review highlights future perspectives of personalized nutrition with regards to the approach of MetS and how individualized multi-omics approaches could improve the current outlook.

Introduction

In recent decades, chronic diseases, such as metabolic and cardiovascular disorders, have become particularly important worldwide. For almost a century now, a set of risk factors has been described to increase the risk of developing type 2 diabetes (T2D) and cardiovascular disease (CVD),¹ two of the top 10 worldwide diseases that cause mortality.² It is forecasted that by 2040, the metabolic risk factors of high blood pressure, high body mass index (BMI), and high fasting blood glucose will be among the five leading global risk factors for years of life lost (YLLs), and the differences between riskattributable YLLs in the better/worse health scenarios will be at least 2.6 times.³ The term "metabolic syndrome" (MetS) began to be used until earlies 1980s to identify cardiometabolic abnormalities and recognizing their deleterious synergistic role for health.^{1,4} These factors include central obesity, insulin resistance, dyslipidemia and elevated blood glucose, characterized by an increase in triglyceride (TG) and a decrease of high-density lipoprotein cholesterol (HDL-C) serum levels, as well as high blood pressure. Later, a model with insulin resistance was proposed as the central axis of this condition, although the obesity factor was omitted, being called as "syndrome x".⁵ Since its conceptualization, various definitions have been suggested for the diagnosis of metabolic syndrome. These include common clinical markers, although other biochemical markers associated with inflammation, cardiovascular risk and energy metabolism have been also described. 6,7

Among the most common definitions (Table I), are those proposed by the world health organization (WHO) in 1989,⁸ which highlighted the presence of insulin resistance (or its substitutes) as essential components. Subsequently, in 2001, the US National

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Cholesterol Education Program: Adult Treatment Panel III (NCEP-ATPIII) published its diagnostic criteria, focusing on the detection of people with higher cardiovascular risk by giving an equal weighting to the presence of all markers.⁹ In 2004, the International Diabetes Federation (IDF) proposed a definition that sought to be applicable for the detection of the risk of CVD and T2D worldwide in a simple manner.¹⁰ Central obesity was included as an essential marker, establishing different cut-off points for waist circumference, specific to each ethnicity, in conjunction with at least two additional factors for diagnosis. Interestingly, the IDF mentioned additional criteria for future MetS research, such as tomographic evaluation of visceral and hepatic adiposity, adipose tissue biomarkers, apolipoprotein B (Apo B), low-density lipoprotein cholesterol (LDL-C) particle size, formal measurements of insulin resistance and glucose tolerance test, as well as inflammatory and thrombotic markers.¹⁰

There are other proposals for definitions such as the European Group for the Study of Insulin Resistance (EGIR),¹¹ the American Association of Clinical Endocrinologists (AACE),¹² the American Heart Association/National Heart, Lung, and Blood Institute (AHA/NHLBI).¹³ However, the efforts to harmonize definitions resulted in a consensus to use the IDF definition by Alberti *et al* in 2009.¹⁴ Although these different diagnostic criteria for MetS vary in terms of the factor considered "central", all definitions include common clinical markers and have undeniably contributed to the detection of the increasing prevalence of MetS worldwide.¹⁵

Lately, the approach of concepts such as metabolic health and subphenotyping of metabolic risk has attracted attention to the scientific community. There is evidence which suggests that cardiometabolic risk stratification is not superior to established risk

prediction models. However, the addition of risk factors and the clustering approaches to identify subphenotypes might be informative to improve the prediction of cardiometabolic risk in subgroups of individuals with particular characteristics such as those in different BMI categories or with diabetes diagnosis. In addition, the communication of cardiometabolic risk to patients is easiest through the concept of metabolic health. Evidence is still missing with regards whether the allocation of individuals to a specific pathophysiological risk group could be helpful for prevention and treatment of cardiometabolic diseases.¹⁶

Due to the complex etiology behind the presence of each of the individual factors that comprise the MetS, its study as an interrelated entity becomes even more intricate. Its analysis across the '*omics*' sciences offers a picture of greater potential, by improving the ability of novel biomarkers to refine risk assessment for the disease.¹⁷ Genetics and nutrigenetics stand out, explaining part of the interindividual variability in the presence of MetS and its relationship with dietary aspects. Although non-genetic factors such as diet and lifestyle remain the main trigger for the development of components of MetS linked to obesity, there is a growing evidence explaining how genetic variants, and their interaction with other environmental factors, modulate the risk of developing MetS.¹⁸

The aim of this review is to explore the most relevant and recent evidence on how genetic factors, particularly single nucleotide polymorphisms (SNPs) and their interaction with diet can determine an individual's risk for developing MetS or maintaining optimal cardiometabolic health. In turn, the pathway and challenges towards personalized nutrition in cardiometabolic health are discussed.

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1. How do genetics affect cardiometabolic risk?

The risk components for MetS are complex entities, influenced by environmental factors such as diet, physical exercise, and lifestyle.¹⁹ The so-called "obesogenic environment" that promotes an unhealthy diet, sedentary lifestyle, along with aspects of urbanization and difficult access to adequate nutrition, also affect the other markers of cardiometabolic risk. ²⁰

However, genetic variation remains an important force of phenotypic variation in MetS components.²¹ In polygenic studies of disease, single nucleotide polymorphisms (SNPs) have been widely associated with an individual's predisposition to develop cardiometabolic abnormalities, including components of MetS.^{22,23} The estimate of heritability of MetS as an entity varies between 10-30%.²⁴⁻²⁶ On the other hand, as individual components, HDL-C is the trait with the highest heritability (30-80%), followed by waist circumference (30-70%), systolic blood pressure (20-71%), TG levels (30-72%), diastolic blood pressure (10-50%), and insulin levels (20-55%).^{24,26}

Genome wide association studies (GWAS) and candidate gene studies have identified numerous SNPs significantly associated with the presence of MetS and its individual cardiometabolic components (Table II).

One of the most recent GWAS using data from the UK Biobank,²⁷ found 93 independent *loci* with a p-value <5x10⁻⁸, associated with the MetS as a binary trait. Eighty out of these 93 variants had not been previously identified by their association with the trait; however, several variants had been associated with some of the individual components of the MetS. In a Korean population, 43 significantly associated *loci* were identified, in which 17

were novel.²⁸ Moreover, the rs662799 variant in the *APOA5* gene was associated with the presence of MetS, as previously reported in the Taiwanese population.²⁹ The authors also found significant association of the rs16944558 SNP in the *COLEC12* gene with the presence of MetS and an interaction with rs662799 in *APOA5* that promotes high levels of TG and low HDL-C.²⁹ Another study in India, comprising 10,093 individuals, reported two variants near the *CETP* gene, nominally associated with MetS, in addition to other modest signals reported for the first time.³⁰ On the other hand, in the Han Chinese population, two *loci* were identified in the *APOA5* and *ALDH2* genes, rs651821 and rs671, respectively, associated with the presence of MetS. In combined analyses they reported ORs and 95% CI of 1.28 (1.20, 1.36) and 0.71 (0.67, 0.76) for MetS risk, with the presence of C and A alleles in rs651821 and rs671 SNPs, respectively.³¹

There is evidence of the widely studied cluster region *APOA1/C3/A4/A5*, through the rs964184 SNP, associated with the presence of MetS and several lipid phenotypes in Finnish cohorts.³² Also, in a population of European ancestry, 29 common variants associated with MetS (or at least with a couple of individual traits) were identified.³³ Some of the genes that showed the greatest significance were *LPL*, *CETP*, *APOA5*, *GCKR*, *LIPC*, *TRIB1*, among others.

Notably, most of evidence indicates that the top genetic signals are located mostly in genes related to lipid metabolism and obesity pathways.³⁴ Through metabolic and transcriptomic studies, it has been shown that most of the SNPs of greater association with MetS have shown significant association with various lipid metabolites such as very low-density lipoprotein cholesterol (VLDL-C),³³ intermediate-density lipoprotein cholesterol (IDL-C) and Apo B and none with glucose or glycoproteins,³² which may

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indicate that the genetic contribution to the development of MetS could be rather directed by the lipid component of this entity.

Regarding the approach of complex diseases such as MetS, it has been suggested to analyze their genetic etiology through polygenic risk scores (PRS), which include a series of risk alleles previously associated with the presence of MetS,³⁵ either as an entity or their individual traits, which added and weighted (beta or OR values) result in a score that can help to identify individuals with greater genetic susceptibility to develop the disease.

In the past, there were studies including PRS of tens of *loci* that conferred a higher risk of developing T2D compared to that of individual SNPs.³⁶ However, these marginally improved the predictive power of previously known clinical risk factors.³⁷ With the appearance of GWAS in larger cohort populations, the number of variants associated with MetS has increased, improving the predictive power of derived PRS.^{38,39}

The aim of identifying and classifying risk groups at the extremes of the population distribution (i.e., those with highest percentiles) is becoming more popular due to the progress made in including a greater number of SNPs in the calculation of MetS PRS, as well as larger sample sizes.^{40,41} However, the fact that greater inclusion of ethnically diverse populations, which are currently underrepresented in the GWAS is required, should not be overlooked.⁴² This will allow in the future to improve the generalization and application of the indisputable benefits that PRS could bring to the clinical practice.

2. Genetic variants (SNPs) associated to cardiometabolic clinical markers

2.1 Obesity

Depending on the selected criteria, obesity can be one of the central components of MetS. However, due to its complexity, much of the etiology behind its origin is still unknown. Higher genetic susceptibility can increase the risk of obesity, although the impact of genes is greater when combined with the increasingly worsening obesogenic environment.⁴³ Recent evidence suggests that despite midlife obesity being independently associated with CVD, obesity influenced by genetic predisposition (i.e. individuals with genetically predicted high BMI) is less harmful than obesity influenced by environmental factors (i.e. individuals with obesity despite a genetically predicted low BMI). However, additional influencing factors such as other genetic variants could still affect these associations.⁴⁴

Obesity is defined by an excessive accumulation of fat mass, however, there is large variability in the risk associated to metabolic disease according to different subphenotypes. Evidence suggests this could be partly due to the variability in fat distribution patterns.⁴⁵ The accumulation of visceral fat and an impaired ability to expand subcutaneous fat in the lower part of the body contributes to the increased incidence of cardiometabolic diseases.⁴⁶ In addition, genetic factors are a key determinant of fat distribution⁴⁷ with an heritability of 22-61%.⁴⁸ Within the last decade, hundreds of genetic variants associated to measures of fat distribution have been identified in GWAS,⁴⁹ mostly in European population. This is important since fat distribution patterns also differ among populations. For example, Asians have lower BMIs but higher total body fat and visceral fat accumulation than Europeans.⁴⁸

The ratio of visceral adipose tissue (VAT) to subcutaneous adipose tissue (SAT) and the non-alcoholic fatty liver disease (NAFLD) lately named "metabolic dysfunction-associated

fatty liver disease" (MAFLD), play important roles in the increase of associated cardiometabolic diseases. VAT drains directly to the liver through portal circulation and contains a larger number of inflammatory and immune cells and a greater percentage of large and more metabolically active adipocytes, more sensitive to lipolysis and more insulin-resistant than SAT adipocytes.⁵⁰ Since most of the mechanisms of lipid metabolism are mediated by hormonal pathways,⁵¹ and the adipose tissue is an endocrine organ itself, there is evidence that supports the association of adipokines to the development of metabolic diseases and T2D trough VAT augmentation. The chronic inflammation, neurohormonal activation and insulin resistance are among the proposed mechanisms involved in the progression of MetS and its comorbidities (Figure 1).52 Notably, MAFLD is also an important cause of insulin resistance, its close relationship with visceral obesity conceals the role of fatty liver from VAT as the main pathomechanism of this relationship.⁵³ To this matter, the determination of major hepatokines and adipokines has been proposed to cluster insulin resistance in MAFLD and VAT in a pathomecanism-based way, i.e. attempt to differentiate the drivers of insulin resistance in metabolic dysregulation of white adipose tissue, skeletal muscle, or liver dysfunction.

Non-syndromic obesity can be classified as monogenic and polygenic, according to the participation of genes in its etiology. With regards the monogenic form, severe obesity (mainly early onset) is principally characterized by relative hyperphagia. Evidence suggests that around 5% of severe obesity cases in children can be attributed to this origin.⁵⁴ Four of the genes of the greatest association and study include the leptin (*LEP*), leptin receptor (*LEPR*), pro-opiomelanocortin (*POMC*) and melanocortin receptor-4 (*MC4R*).⁵⁵ These genes belong to the leptin/melanocortin pathway and play an important

role in the regulation of food intake since they encode proteins that act in the process at the hypothalamic level. These are centrally or peripherally produced molecules that influence appetite regulation.⁵⁶ Genetic variants in those genes have shown to be associated with variability on BMI and weight. For example, the variant Tyr35Ter (rs13447324) in *MC4R* has been associated with an excess of 7 kg of greater body weight in carriers (approximately 1 in 5,000 people).⁵⁷ However, despite being a mutation that results in the loss of total function (LoF) of *MC4R*,⁵⁸ it has been found to present incomplete penetrance, partly because normal-weight carriers of monogenic variants possess additional common variants that predispose them to a lower weight,⁵⁹ i.e., its low polygenic risk for obesity could compensate, at least in part, the risk caused by the mutation in *MC4R*.

In counterpart, obesity of polygenic origin, also called 'common obesity', represents the highest proportion of cases in the world, besides to being complex and exacerbated by the environment. The genetic component of common obesity is mostly given by the cumulative presence of multiple common genetic variants with little individual etiological contribution although, when added together, it can explain a greater proportion of the variability of body weight. The most studied variants for their abundance in the human genome are also SNPs. To date, more than 900 nearly independent SNPs associated with BMI,⁶⁰ and around 346 SNPs with body fat distribution.⁴⁹ To quantify the genetic predisposition of an individual, several PRS models have been proposed, including 12,⁶¹ 20,⁶² 32,⁶³ 56,⁶⁴ 97,⁶⁵ and 941 SNPs,⁶⁰ managing to explain a maximum proportion of the phenotypic variation of BMI of around 6%.⁶⁰ There are also several studies where these PRS have been replicated.⁴³ Furthermore, some genome-wide polygenic scores (GPS)

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included 2.1 million genetic variants for the quantification of genetic predisposition to obesity over the life course of more than 300,000 people of European ancestry. ⁶⁶ Longitudinally, a significant weight gradient was found between GPS deciles, which began in early childhood and reached a maximum difference of 12 kg at age 18. Notably, GPS allowed to identify 1.6% of the population with an increase in BMI like those with monogenic mutations. The model explained about 9% of the BMI variation, similarly to those results obtained in a Norwegian population.⁶⁷

Despite the modest, but significant advances in the study of polygenic obesity, there is still a gap between BMI heritability that can be explained by SNPs (h^2_{SNPs}) according to the literature (21-30%).^{65,68} However, the great potential of increasing the sample size of several populations in GWAS to detect a greater amount of common and rare associated *loci* and thus be able to explain more proportion of the total h^2_{SNP} is recognized.

Among the genes associated with obesity, the most widely studied is the fat mass and obesity-associated (*FTO*) gene.⁶⁹ In 2007, its polymorphisms were the first to be reproducibly associated with BMI.⁷⁰ Since then, the association of being a carrier of risk alleles in SNPs in the *FTO* gene has been replicated by several authors.⁷¹ However, the contribution of these SNPs to BMI variability remains modest, with carriers of the rs9939609 risk allele estimated to weigh about 3 kgs more and being 1.67 times more likely to develop obesity compared to non-carriers.⁷⁰ Other genes identified by their association with obesity have been studied. Interestingly, many of them have been found to be involved in neurogenesis, central nervous system (CNS) development, and in signaling pathways related to appetite and dietary intake regulation.⁶⁰ A compendium of human genes that regulate eating behavior and body weight was published, including 578

genes which were ordered according to their biological role in the regulation of body weight and classified by their expression patterns or functional characteristics.⁷²

Among the main genes whose *loci* have been identified and replicated by various studies for their association to body weight and BMI are *FTO*,^{70,73} *MC4R*,^{74,75} *POMC*,^{63,76} *BDNF*, ^{77,78} *TCF7L2*,^{49,65,79} *LINC01875*, *TMEM18*,⁸⁰⁻⁸² *ADCY3*,^{49,76,83} among many others.

Beyond genetic factors, some of the latest advances in obesity study have been directed to the identification of multiomic signatures of BMI. Despite the use of a single targeted metric (for example, BMI) or a single specific biomarker that provides useful information to quantify health and disease states, multiomic blood profiling which includes human genomes and longitudinal measurements of metabolomics, proteomics, clinical laboratory tests, gut microbiomes, physical activity, and health/lifestyle data, could help close the knowledge gaps between BMI and heterogeneous physiological states in a multifaceted manner. With regards to lifestyle interventions, the multiomic signatures can predict responses in a heterogeneous way; omics-inferred BMI behaves different than the actual BMI measurement in response to treatment. This highlights the fact that multiomic profiling could be a resource to quantify the changes in obesity status and metabolic health for predictive and preventive medicine.⁸⁴

2.2. Insulin resistance and type 2 diabetes

Insulin resistance, defined as the inability of insulin to stimulate the use of glucose in the body that can eventually lead to T2D, is the central component of most pathophysiological models of MetS. For several authors, this represents the underlying factor for the development of this pathological entity. Insulin sensitivity/resistance is closely related to

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coronary artery disease (CAD),⁸⁵ one of the main outcomes of MetS. Its presence has increased in recent years, with a current prevalence of 15.5-40% worldwide.⁸⁶

Depending on the diagnostic criteria, insulin resistance may be represented by different biochemical concepts and parameters. The inclusion of easily accessible biochemical parameters associated with hyperglycemia and insulin resistance, such as plasma glucose measurements, arose from the need to have criteria that would allow a lower cost and affordable diagnosis, particularly in studies including large study populations.⁸⁷ According to the American Diabetes Association (ADA), diabetes could be diagnosed by a fasting plasma glucose \geq 126 mg/dl (7.0 mmol/L) or a 2-h plasma glucose \geq 200 mg/dl (11.1 mmol/L) during OGTT or A1C \geq 6.5% (48 mmol/mol) or by classic symptoms of hyperglycemia or hyperglycemic crisis, a random plasma glucose \geq 200 mg/dL (11.1 mmol/L).⁸⁸

Insulin resistance has been studied through GWAS and association studies where common genetic variants involved in the presence of T2D as a phenotypic trait have been evaluated. To date, more than 240 *loci* associated with T2D have been found, corresponding to more than 400 genetic variants.⁴¹ The vast majority of *loci* related to this pathology are those associated with insulin secretion and beta cell function in the pancreas and to a lesser extent with insulin resistance.⁸⁹ This has a possible explanation in the complex etiology and pathophysiology of T2D, as well as its close relationship with obesity and plasma lipid levels. Some of the genes that have shown association with IR and T2D are *PPARG*,⁹⁰ *IRS1*,⁹¹ *ADAMTS9*,⁹² *KLF14*,⁹³ *ARL15*,⁹⁴ *FTO*,⁹⁵ among others.

One of the genes with the greatest contribution to T2D susceptibility is *TCF7L2*,⁹⁶ in which the risk allele has been related to insulin secretion.⁹⁷ These effects have been

replicated in various populations by GWAS,^{83,98} with the SNP rs7903146 being one of the most relevant SNPs. This SNP, which is located in an intronic region, has similar allelic frequencies worldwide, with the exception of East Asia where the risk allele remains relatively rare.⁹⁹ The presence of the T allele in rs7903146 has been shown to strongly predict the development of T2D and it is associated with increased expression of *TCF7L2* in human islets, as well as altered insulin secretion both *in vitro* and *in vivo* studies.¹⁰⁰

Another gene of great importance for its association with diabetes mellitus, insulin resistance and MetS is *HMGA1* which has among its functions the regulation of insulin receptor (*INSR*) gene expression. HMGA1 is a master regulatory factor for gluconeogenesis and glycogenolysis, as well as a positive regulator of insulin expression; ¹⁰¹ consequently, being associated with the presence of T2D in different populations.¹⁰² One of the studies of greater collaborative effort aimed to elucidate the genetic architecture of T2D hypothesized that the heritability of T2D, not yet fully explained by common variants, could be given by low-frequency variants.¹⁰³ However, the associated variants that they found (approximately 126 variants) were mostly common and identified by previous studies.

2.3. Serum triglyceride and HDL cholesterol levels

The presence of MetS increases the risk of cardiovascular disease and atherosclerosis, however, alterations in lipid metabolism are an independent risk factor for CAD.¹⁰⁴ The lipid markers considered for MetS are TG and HDL-C levels. The presence of hypertriglyceridemia (HTG) alone, defined as an elevation of circulating TG levels, usually >150-175 mg/dL (1.71-2 mmol/L), or in conjunction with other abnormalities such as

decreased levels of HDL-C tend to cluster in families.¹⁰⁵ Genetic factors influencing plasma TG levels explain about 40% of interindividual variations, while 49% is explained on the variation of the TG/HDL-C index.¹⁰⁶

In the postgenomic era, HTG, formerly classified as primary or secondary, is accepted as a complex ethology phenotype, except for the very rare familial chylomicronemia syndrome (FCS), with an autosomal recessive Mendelian inheritance. In most cases, their predisposition is given by the presence of common genetic variants of small effects that interact with rare heterozygous variants of great effect in genes that regulate the synthesis or catabolism of triglyceride-rich lipoproteins or with non-genetic factors, this can lead to the expression of more severe HTG phenotypes.¹⁰⁷

GWAS studies have identified SNPs in at least 45 *loci* associated with plasma TG levels alone or in combination with other lipoproteins.^{108,109} For example, the *locus* 1q21-23 has been associated with different lipid traits such as HTG, where the group of genes *APOA1/C3/A4/A5* is related to the presence of FCS.¹¹⁰ Similarly, the *USF1* gene is related to various target genes related to glucose and lipid metabolism.¹¹¹

The polygenic risk for HTG can be quantified by using PRS.^{112,113} A recent study found that ~2% of patients with severe HTG had a high PRS (16 *loci*) compared to 9.5% of normolipemic controls.¹⁰⁷ However, the genotype-phenotype relationship is probabilistic and non-deterministic. Some of the *loci* belonging to this PRS are located in the *DOCK7*, *KLHL8*, *GALNT2*, *MLXIPL*, *LPL*, *FADS*, *APOA*, *CETP*, *SUGP1*, *PLTP* genes, among others. The results found by recent studies suggest that non-genetic factors may lead to the presence of HTG in people with genetic predisposition, either through a rare variant or a high polygenic risk, or both.

Low HDL-C levels are a widely discussed risk marker for CAD and its functionality has been reappraised in recent years. Its role is mainly affected by both the heterogeneity of this lipoprotein class and the extensive remodeling of the HDL size and lipid and protein content during their physiological maturation.¹¹⁴ However, as a risk marker, represents the most common lipid abnormality in patients with CAD.¹¹⁵ HDL particles have multiple antiatherogenic effects, mainly through the removal of cholesterol from peripheral tissues to the liver, this reverse transport of cholesterol prevents macrophages from arterial walls from turning into foamy cells, which are progenitors of atherosclerotic plaque.¹¹⁶ Additionally, HDL has antioxidant, antithrombotic, and anti-inflammatory properties.¹¹⁷

As for the genetic factors associated with decreased HDL-C levels, there are monogenic forms that lead to extreme phenotypes; however, these causes are rare and explain a minimal portion (~1%) of low HDL-C cases. For example, *APOA1* deficiency causes HDL-C levels <5 mg/dl (0.13 mmol/L), normal LDL-C and TG levels .^{118,119} Likewise, defects in genes such as *ABCA1*,^{118,120} *LCAT*,¹²¹ and *LPL*,¹²² among others, cause deficiencies and familial syndromes causing extremely low HDL-C levels.

The most common genetic disorder causing low HDL-C levels is familial hypoalphalipoproteinemia (FHA)¹²³ but even in this case, studies suggest that most hereditary patterns for low HDL-C are polygenic.¹²⁴ Several studies have constructed PRS for the risk of low HDL-C in conjunction (or not) with other lipid traits and risk of CVD. ^{107,125}

Regarding the relationship of HDL-C levels and MetS, a 5-year longitudinal follow-up study showed that the incidence of MetS is higher in individuals with decreased HDL-C

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levels.¹²⁶ The authors mention that this important factor appears to be a pre-existing phase of MetS and could be a crucial state for prevention.

2.4 High blood pressure

Hypertension (HTN) is characterized by high systolic and/or diastolic blood pressure, and is a major risk factor for heart failure, stroke, kidney disease, and cardiovascular death.

Several studies, mostly GWAS have set out to reveal the genetic architecture of HTN, uncovering hundreds of novel common and rare variants in trans-ethnic study populations of hundreds of thousands individuals.^{128,129} Additionally, most blood pressure-associated SNPs are non-coding and found in regulatory elements of the genome.¹³⁰

However, being an extremely complex entity, the environment and gene-environment interactions have a contribution of great weight that is not taken into account in most current GWAS studies.¹³¹ Within these elements of interaction, prenatal environmental factors such as intrauterine and parental¹³² and other postnatal factors such as the living environment, lifestyle, age, sex, socioeconomic status, ethnicity, among others, are also considered.¹³³

Environmental risk factors traditionally considered such as an excess sodium in the diet were adaptive traits for the hot, humid, salt-free environment of ancient Africa. From an evolutionary point of view, there is a discrepancy between the current lifestyle and that of ancestors, which results in a poor adaptation that can lead to an increased risk of developing HTN.¹³⁴

In Figure 2, we present a summary of some of the genes most frequently mentioned in GWAS and candidate genes studies.

3. Gene-diet interactions associated to cardiometabolic markers.

The study of the genetic component, responsible for the variation in phenotypes associated with MetS, has managed to explain only a proportion of this variability. Part of this "lost heritability" could be given by the interactions between genes and the environment (GxE) of individuals.¹³⁵

One of the most studied GxE with regards to the components of MetS are those of the *FTO* gene and its interaction with diet. For example, in a recent study in a Middle Eastern population, dietary fiber consumption was found to modulate the association of a PRS of 6 SNPs in relation to obesity, where people with a higher PRS but a consumption \geq 14 g/day were less likely to develop obesity, compared to those with lower PRS but low fiber intake.¹³⁶ Previous studies also found similar interactions.^{137,138}

Other interactions of the *FTO* gene with dietary patterns such as adherence to a Mediterranean diet have been studied, for example, modulating the risk of diabetes. The increased risk effect of diabetes in individuals with the common variant rs9939609 is neutralized with high adherence to the Mediterranean diet.¹³⁹

Similarly, a study that included a PRS with 16 genetic variants previously associated with obesity found significant interaction between higher animal protein intake and higher body fat mass in people within the higher genetic risk group, and a protective effect of higher plant protein intake for people belonging to the lower genetic risk group.¹⁴⁰

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The consumption of fried food has also shown interaction in a PRS with 32 variants associated with BMI, where the greatest effect of genetic predisposition was observed in individuals with a high consumption (≥4 times per week) of fried foods, compared to those who consumed them less than 1 time per week, regardless of their high PRS.¹⁴¹

Sodium intake is a significant environmental factor in modifying blood pressure values, and unexplained blood pressure variability in GWAS may result from gene-environment interactions (GxE).^{142,143} For example, sodium consumption has shown interaction with various *loci* in genes such as *CLGN*, *MKNK1*, *EPHA6* and *CASP4*,¹⁴³ among others; however, the mechanisms through which these regulate blood pressure are still unknown. Nevertheless, interesting interaction trends have been observed, where people with higher PRS are less sensitive to modifications in dietary sodium intake, denoting that apparently for them the greatest influence comes from genes.¹⁴⁴

On the other hand, it has been postulated that dietary fatty acid consumption might not uniformly influence individuals' blood lipid levels. A recent publication showed evidence of interactions of variants such as rs5882 in the *CETP* gene, where a higher consumption of monounsaturated fatty acids in carriers of the main allele presented lower serum TG levels, as well as the rs13708 variant of the *LPL* gene whose interaction was observed in higher HDL-C levels when consuming a diet with higher lipid content.¹⁴⁵

These findings may explain the variation in the effectiveness of certain dietary interventions for the prevention or treatment of obesity and other traits of MetS, highlighting the importance of a healthy diet and lifestyle, especially in people who might have a higher genetic susceptibility to develop MetS.

4. What is the future of personalized nutrition for the improvement of cardiometabolic health?

Personalized nutrition has its focus on the design of "tailored" nutritional recommendations and treatments, based on the individual genetic arrangement, to treat or prevent various pathologies and metabolic disorders.¹⁴⁶ Personalized nutrition is a strategy that can help individuals to adopt lasting changes in dietary behavior, which are beneficial to their health.¹⁴⁷

As previously mentioned, several studies focused on gene-environment interaction have provided relevant results in the study of MetS and its components. These could be a key piece in the implementation of personalized interventions based on genetic arrangement. Additionally, the fact that nutrigenetics has improved from analyzing individual genetic variants to integrating PRS and looking for complex interactions with pathological states such as obesity, denotes the important and necessary evolution in this field.¹⁴⁸ However, what evidence do we have so far of its effectiveness and implementation in the clinical practice? Through the generation of PRS_{MetS} and their validation on big datasets it has been possible to test early prevention strategies in individuals with high genetic risk, demonstrating benefits, for example, in reducing cardiometabolic events such as atherosclerotic cardiovascular disease (Fig. 3).¹⁴⁹

Furthermore, there are some studies that have evaluated the effectiveness of interventions based on personalized nutrition.¹⁵⁰⁻¹⁵² These results suggest that the use of this strategy offers advantages over generalized "one-size-fits-all" diets, improving clinical outcomes and achieving greater reductions in discretionary food intake, as well as, facilitating the change in people's dietary behavior and improving acceptance and

adherence to nutritional treatment. However, multiple questions remain, and it is not entirely clear which aspects of personalization offer the greatest advantage over generalized strategies.

Regarding cardiometabolic health, studies have revealed that different dietary patterns and macronutrient intake have weight in modifying and reversing the presence of MetS components.^{153,154} For example, there have been revealed genetic variants that could be considered when referring to certain dietary treatments, such as the low-carbohydrate diet, whose results could be affected by the presence of SNPs such as rs694066 (*GAL*)¹⁵⁵ and rs5950584 (*AGTR2*),¹⁵⁶ where people carrying different alleles tend to lose more or less weight and body fat when following a ketogenic diet.

There are recent GWAS that have been shown evidence about the potential clinical utility in the assessment and treatment of MetS and other related cardiometabolic traits (Table III).^{149,157}

However, it is necessary to consider other factors, not only genetic, but others such as epigenetics, metabolomics, microbiome, to name a few, in the study of such complex entities as MetS.¹⁵⁸ This leads to the advancement from personalized nutrition to precision nutrition, whose main goal is to efficiently anticipate people's response to dietary recommendations, by considering the greatest number of biological-environmental aspects associated with health and disease states.^{159,160}

Conclusions

There is evidence of the important role of genetics on the development of MetS and its individual components, however, there is still an important gap in the clinical implementation of personalized nutrition, successfully achieving prevention and decreasing of the prevalence of this pathological entity around the world. The most current evidence mentions that the study of the interaction of genes with the growing obesogenic environment is possibly a key piece to unite this gap. The advance towards precision nutrition in cardiometabolic health issues is still in its infancy, however, its progress has been increased in the last decade with the growth of omics sciences.

Authors' contributions

DPO wrote the initial draft and ID and FFGG reviewed and contributed extensively to the manuscript writing.

Author Disclosure Statement

The authors declare that no conflict of interest exist.

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References

1. Moller DE, Kaufman KD. Metabolic syndrome: a clinical and molecular perspective. Annual review of medicine 2005;56(45-62, doi:10.1146/annurev.med.56.082103.104751

2. Organization WH. The top 10 causes of death: major causes of death. World Health Organization Media Center Fact Sheets Available from: <u>http://www</u> who int/mediacentre/factsheets/fs310/en/index2 html [Last downloaded on 2013 Aug 30] 2016;

3. Foreman KJ, Marquez N, Dolgert A, et al. Forecasting life expectancy, years of life lost, and allcause and cause-specific mortality for 250 causes of death: reference and alternative scenarios for 201640 for 195 countries and territories. Lancet (London, England) 2018;392(10159):2052-2090, doi:10.1016/s0140-6736(18)31694-5

4. Hanefeld M. Das metabolische syndrom. Dt Gesundh Wesen 1981;36(545-551

5. Reaven GM. Banting lecture 1988. Role of insulin resistance in human disease. Diabetes 1988;37(12):1595-607, doi:10.2337/diab.37.12.1595

6. Van Gaal LF, Mertens IL, De Block CE. Mechanisms linking obesity with cardiovascular disease. Nature 2006;444(7121):875-80, doi:10.1038/nature05487

7. Taglieri N, Koenig W, Kaski JC. Cystatin C and cardiovascular risk. Clinical chemistry 2009;55(11):1932-43, doi:10.1373/clinchem.2009.128397

8. Alberti KG, Zimmet PZ. Definition, diagnosis and classification of diabetes mellitus and its complications. Part 1: diagnosis and classification of diabetes mellitus provisional report of a WHO consultation. Diabetic medicine : a journal of the British Diabetic Association 1998;15(7):539-53, doi:10.1002/(sici)1096-9136(199807)15:7<539::aid-dia668>3.0.co;2-s

9. Third Report of the National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (Adult Treatment Panel III) final report. Circulation 2002;106(25):3143-421

10. Alberti KG, Zimmet P, Shaw J. Metabolic syndrome--a new world-wide definition. A Consensus Statement from the International Diabetes Federation. Diabetic medicine : a journal of the British Diabetic Association 2006;23(5):469-80, doi:10.1111/j.1464-5491.2006.01858.x

11. Balkau B, Charles MA. Comment on the provisional report from the WHO consultation. European Group for the Study of Insulin Resistance (EGIR). Diabetic medicine : a journal of the British Diabetic Association 1999;16(5):442-3, doi:10.1046/j.1464-5491.1999.00059.x

12. Einhorn D, Reaven GM, Cobin RH, et al. American College of Endocrinology position statement on the insulin resistance syndrome. Endocrine practice : official journal of the American College of Endocrinology and the American Association of Clinical Endocrinologists 2003;9(3):237-52

13. Grundy SM, Cleeman JI, Daniels SR, et al. Diagnosis and management of the metabolic syndrome: an American Heart Association/National Heart, Lung, and Blood Institute Scientific Statement. Circulation 2005;112(17):2735-52, doi:10.1161/circulationaha.105.169404

14. Alberti KG, Eckel RH, Grundy SM, et al. Harmonizing the metabolic syndrome: a joint interim statement of the International Diabetes Federation Task Force on Epidemiology and Prevention; National Heart, Lung, and Blood Institute; American Heart Association; World Heart Federation; International Atherosclerosis Society; and International Association for the Study of Obesity. Circulation 2009;120(16):1640-5, doi:10.1161/circulationaha.109.192644

15. Saklayen MG. The Global Epidemic of the Metabolic Syndrome. Current hypertension reports 2018;20(2):12, doi:10.1007/s11906-018-0812-z

16. Stefan N, Schulze MB. Metabolic health and cardiometabolic risk clusters: implications for prediction, prevention, and treatment. The lancet Diabetes & endocrinology 2023;11(6):426-440, doi:10.1016/s2213-8587(23)00086-4

17. Tahir UA, Gerszten RE. Omics and Cardiometabolic Disease Risk Prediction. Annual review of medicine 2020;71(163-175, doi:10.1146/annurev-med-042418-010924

18. Stančáková A, Laakso M. Genetics of metabolic syndrome. Reviews in endocrine & metabolic disorders 2014;15(4):243-52, doi:10.1007/s11154-014-9293-9

19. Wang Q, Chair SY, Wong EM. The effects of a lifestyle intervention program on physical outcomes, depression, and quality of life in adults with metabolic syndrome: A randomized clinical trial. International journal of cardiology 2017;230(461-467, doi:10.1016/j.ijcard.2016.12.084

20. Guo F, Bostean G, Berardi V, et al. Obesogenic environments and cardiovascular disease: a path analysis using US nationally representative data. BMC public health 2022;22(1):703, doi:10.1186/s12889-022-13100-4

21. Elder SJ, Lichtenstein AH, Pittas AG, et al. Genetic and environmental influences on factors associated with cardiovascular disease and the metabolic syndrome. Journal of lipid research 2009;50(9):1917-26, doi:10.1194/jlr.P900033-JLR200

22. Tekola-Ayele F, Doumatey AP, Shriner D, et al. Genome-wide association study identifies African-ancestry specific variants for metabolic syndrome. Molecular Genetics and Metabolism 2015;116(4):305-313, doi:https://doi.org/10.1016/j.ymgme.2015.10.008

23. Wu Y, Yu Y, Zhao T, et al. Interactions of Environmental Factors and APOA1-APOC3-APOA4-APOA5 Gene Cluster Gene Polymorphisms with Metabolic Syndrome. PloS one 2016;11(1):e0147946, doi:10.1371/journal.pone.0147946

24. Bellia A, Giardina E, Lauro D, et al. "The Linosa Study": epidemiological and heritability data of the metabolic syndrome in a Caucasian genetic isolate. Nutrition, metabolism, and cardiovascular diseases : NMCD 2009;19(7):455-61, doi:10.1016/j.numecd.2008.11.002

25. Musani SK, Martin LJ, Woo JG, et al. Heritability of the Severity of the Metabolic Syndrome in Whites and Blacks in 3 Large Cohorts. Circulation Cardiovascular genetics 2017;10(2), doi:10.1161/circgenetics.116.001621

26. Henneman P, Aulchenko YS, Frants RR, et al. Prevalence and heritability of the metabolic syndrome and its individual components in a Dutch isolate: the Erasmus Rucphen Family study. Journal of medical genetics 2008;45(9):572-7, doi:10.1136/jmg.2008.058388

27. Lind L. Genome-Wide Association Study of the Metabolic Syndrome in UK Biobank. Metabolic syndrome and related disorders 2019;17(10):505-511, doi:10.1089/met.2019.0070

28. Oh SW, Lee JE, Shin E, et al. Genome-wide association study of metabolic syndrome in Korean populations. PloS one 2020;15(1):e0227357, doi:10.1371/journal.pone.0227357

29. Lin E, Kuo PH, Liu YL, et al. Detection of susceptibility loci on APOA5 and COLEC12 associated with metabolic syndrome using a genome-wide association study in a Taiwanese population. Oncotarget 2017;8(55):93349-93359, doi:10.18632/oncotarget.20967

30. Prasad G, Bandesh K, Giri AK, et al. Genome-Wide Association Study of Metabolic Syndrome Reveals Primary Genetic Variants at CETP Locus in Indians. Biomolecules 2019;9(8), doi:10.3390/biom9080321

31. Zhu Y, Zhang D, Zhou D, et al. Susceptibility loci for metabolic syndrome and metabolic components identified in Han Chinese: a multi-stage genome-wide association study. Journal of cellular and molecular medicine 2017;21(6):1106-1116, doi:10.1111/jcmm.13042

32. Kristiansson K, Perola M, Tikkanen E, et al. Genome-wide screen for metabolic syndrome susceptibility Loci reveals strong lipid gene contribution but no evidence for common genetic basis for clustering of metabolic syndrome traits. Circulation Cardiovascular genetics 2012;5(2):242-9, doi:10.1161/circgenetics.111.961482

33. Kraja AT, Vaidya D, Pankow JS, et al. A bivariate genome-wide approach to metabolic syndrome: STAMPEED consortium. Diabetes 2011;60(4):1329-39, doi:10.2337/db10-1011

34. Nagrani R, Foraita R, Gianfagna F, et al. Common genetic variation in obesity, lipid transfer genes and risk of Metabolic Syndrome: Results from IDEFICS/I.Family study and meta-analysis. Scientific reports 2020;10(1):7189, doi:10.1038/s41598-020-64031-2

35. Chatterjee N, Shi J, García-Closas M. Developing and evaluating polygenic risk prediction models for stratified disease prevention. Nature reviews Genetics 2016;17(7):392-406, doi:10.1038/nrg.2016.27
36. Lango H, Palmer CN, Morris AD, et al. Assessing the combined impact of 18 common genetic variants of modest effect sizes on type 2 diabetes risk. Diabetes 2008;57(11):3129-35, doi:10.2337/db08-0504

37. Ripatti S, Tikkanen E, Orho-Melander M, et al. A multilocus genetic risk score for coronary heart disease: case-control and prospective cohort analyses. Lancet (London, England) 2010;376(9750):1393-400, doi:10.1016/s0140-6736(10)61267-6

38. Khera AV, Emdin CA, Drake I, et al. Genetic Risk, Adherence to a Healthy Lifestyle, and Coronary Disease. The New England journal of medicine 2016;375(24):2349-2358, doi:10.1056/NEJMoa1605086

39. Inouye M, Abraham G, Nelson CP, et al. Genomic Risk Prediction of Coronary Artery Disease in 480,000 Adults: Implications for Primary Prevention. Journal of the American College of Cardiology 2018;72(16):1883-1893, doi:10.1016/j.jacc.2018.07.079

40. Abraham G, Havulinna AS, Bhalala OG, et al. Genomic prediction of coronary heart disease. European heart journal 2016;37(43):3267-3278, doi:10.1093/eurheartj/ehw450

41. Mahajan A, Taliun D, Thurner M, et al. Fine-mapping type 2 diabetes loci to single-variant resolution using high-density imputation and islet-specific epigenome maps. Nature Genetics 2018;50(11):1505-1513, doi:10.1038/s41588-018-0241-6

42. Sirugo G, Williams SM, Tishkoff SA. The Missing Diversity in Human Genetic Studies. Cell 2019;177(1):26-31, doi:10.1016/j.cell.2019.02.048

43. Brandkvist M, Bjørngaard JH, Ødegård RA, et al. Quantifying the impact of genes on body mass index during the obesity epidemic: longitudinal findings from the HUNT Study. BMJ (Clinical research ed) 2019;366(I4067, doi:10.1136/bmj.I4067

44. Ojalehto E, Zhan Y, Jylhävä J, et al. Genetically and environmentally predicted obesity in relation to cardiovascular disease: a nationwide cohort study. EClinicalMedicine 2023;58(101943, doi:10.1016/j.eclinm.2023.101943

45. Shungin D, Winkler TW, Croteau-Chonka DC, et al. New genetic loci link adipose and insulin biology to body fat distribution. Nature 2015;518(7538):187-196, doi:10.1038/nature14132

46. Kaess BM, Pedley A, Massaro JM, et al. The ratio of visceral to subcutaneous fat, a metric of body fat distribution, is a unique correlate of cardiometabolic risk. Diabetologia 2012;55(10):2622-2630, doi:10.1007/s00125-012-2639-5

47. Stefan N. Causes, consequences, and treatment of metabolically unhealthy fat distribution. The lancet Diabetes & endocrinology 2020;8(7):616-627, doi:10.1016/s2213-8587(20)30110-8

48. Schleinitz D, Böttcher Y, Blüher M, et al. The genetics of fat distribution. Diabetologia 2014;57(7):1276-86, doi:10.1007/s00125-014-3214-z

49. Pulit SL, Stoneman C, Morris AP, et al. Meta-analysis of genome-wide association studies for body fat distribution in 694 649 individuals of European ancestry. Human molecular genetics 2019;28(1):166-174, doi:10.1093/hmg/ddy327

50. Ibrahim MM. Subcutaneous and visceral adipose tissue: structural and functional differences. Obesity reviews : an official journal of the International Association for the Study of Obesity 2010;11(1):11-8, doi:10.1111/j.1467-789X.2009.00623.x

51. Nielsen TS, Jessen N, Jørgensen JO, et al. Dissecting adipose tissue lipolysis: molecular regulation and implications for metabolic disease. Journal of molecular endocrinology 2014;52(3):R199-222, doi:10.1530/jme-13-0277

52. Fahed G, Aoun L, Bou Zerdan M, et al. Metabolic Syndrome: Updates on Pathophysiology and Management in 2021. International journal of molecular sciences 2022;23(2), doi:10.3390/ijms23020786

53. Stefan N, Schick F, Birkenfeld AL, et al. The role of hepatokines in NAFLD. Cell metabolism 2023;35(2):236-252, doi:10.1016/j.cmet.2023.01.006

54. Farooqi IS, Keogh JM, Yeo GS, et al. Clinical spectrum of obesity and mutations in the melanocortin 4 receptor gene. The New England journal of medicine 2003;348(12):1085-95, doi:10.1056/NEJMoa022050

55. Fairbrother U, Kidd E, Malagamuwa T, et al. Genetics of Severe Obesity. Current diabetes reports 2018;18(10):85, doi:10.1007/s11892-018-1053-x

56. Broberger C. Brain regulation of food intake and appetite: molecules and networks. Journal of internal medicine 2005;258(4):301-27, doi:10.1111/j.1365-2796.2005.01553.x

57. Turcot V, Lu Y, Highland HM, et al. Protein-altering variants associated with body mass index implicate pathways that control energy intake and expenditure in obesity. Nat Genet 2018;50(1):26-41, doi:10.1038/s41588-017-0011-x

58. Brumm H, Mühlhaus J, Bolze F, et al. Rescue of melanocortin 4 receptor (MC4R) nonsense mutations by aminoglycoside-mediated read-through. Obesity (Silver Spring, Md) 2012;20(5):1074-81, doi:10.1038/oby.2011.202

59. Chami N, Preuss M, Walker RW, et al. The role of polygenic susceptibility to obesity among carriers of pathogenic mutations in MC4R in the UK Biobank population. PLoS medicine 2020;17(7):e1003196, doi:10.1371/journal.pmed.1003196

60. Yengo L, Sidorenko J, Kemper KE, et al. Meta-analysis of genome-wide association studies for height and body mass index in ~700000 individuals of European ancestry. Human molecular genetics 2018;27(20):3641-3649, doi:10.1093/hmg/ddy271

61. Li S, Zhao JH, Luan J, et al. Physical activity attenuates the genetic predisposition to obesity in 20,000 men and women from EPIC-Norfolk prospective population study. PLoS medicine 2010;7(8), doi:10.1371/journal.pmed.1000332

62. Sandholt CH, Sparsø T, Grarup N, et al. Combined analyses of 20 common obesity susceptibility variants. Diabetes 2010;59(7):1667-73, doi:10.2337/db09-1042

63. Speliotes EK, Willer CJ, Berndt SI, et al. Association analyses of 249,796 individuals reveal 18 new loci associated with body mass index. Nat Genet 2010;42(11):937-48, doi:10.1038/ng.686

64. Peterson RE, Maes HH, Holmans P, et al. Genetic risk sum score comprised of common polygenic variation is associated with body mass index. Human genetics 2011;129(2):221-30, doi:10.1007/s00439-010-0917-1

65. Locke AE, Kahali B, Berndt SI, et al. Genetic studies of body mass index yield new insights for obesity biology. Nature 2015;518(7538):197-206, doi:10.1038/nature14177

66. Khera AV, Chaffin M, Wade KH, et al. Polygenic Prediction of Weight and Obesity Trajectories from Birth to Adulthood. Cell 2019;177(3):587-596.e9, doi:10.1016/j.cell.2019.03.028

67. Brandkvist M, Bjørngaard JH, Ødegård RA, et al. Genetic associations with temporal shifts in obesity and severe obesity during the obesity epidemic in Norway: A longitudinal population-based cohort (the HUNT Study). PLoS medicine 2020;17(12):e1003452, doi:10.1371/journal.pmed.1003452

68. Yang J, Bakshi A, Zhu Z, et al. Genetic variance estimation with imputed variants finds negligible missing heritability for human height and body mass index. Nat Genet 2015;47(10):1114-20, doi:10.1038/ng.3390

69. Fawcett KA, Barroso I. The genetics of obesity: FTO leads the way. Trends in genetics : TIG 2010;26(6):266-74, doi:10.1016/j.tig.2010.02.006

70. Frayling TM, Timpson NJ, Weedon MN, et al. A common variant in the FTO gene is associated with body mass index and predisposes to childhood and adult obesity. Science (New York, NY) 2007;316(5826):889-94, doi:10.1126/science.1141634

71. Ran S, Jiang Z-X, He X, et al. Replication of FTO Gene associated with lean mass in a Meta-Analysis of Genome-Wide Association Studies. Scientific reports 2020;10(1):5057, doi:10.1038/s41598-020-61406-3

72. Ignatieva EV, Afonnikov DA, Saik OV, et al. A compendium of human genes regulating feeding behavior and body weight, its functional characterization and identification of GWAS genes involved in brain-specific PPI network. BMC genetics 2016;17(Suppl 3):158, doi:10.1186/s12863-016-0466-2

73. Meyre D, Delplanque J, Chèvre J-C, et al. Genome-wide association study for early-onset and morbid adult obesity identifies three new risk loci in European populations. Nature Genetics 2009;41(2):157-159, doi:10.1038/ng.301

74. Loos RJF, Lindgren CM, Li S, et al. Common variants near MC4R are associated with fat mass, weight and risk of obesity. Nature Genetics 2008;40(6):768-775, doi:10.1038/ng.140

75. Wade KH, Lam BYH, Melvin A, et al. Loss-of-function mutations in the melanocortin 4 receptor in a UK birth cohort. Nature medicine 2021;27(6):1088-1096, doi:10.1038/s41591-021-01349-y

76. Wen W, Cho Y-S, Zheng W, et al. Meta-analysis identifies common variants associated with body mass index in east Asians. Nature Genetics 2012;44(3):307-311, doi:10.1038/ng.1087

77. Tachmazidou I, Süveges D, Min JL, et al. Whole-Genome Sequencing Coupled to Imputation Discovers Genetic Signals for Anthropometric Traits. American journal of human genetics 2017;100(6):865-884, doi:10.1016/j.ajhg.2017.04.014

78. Berndt SI, Gustafsson S, Mägi R, et al. Genome-wide meta-analysis identifies 11 new loci for anthropometric traits and provides insights into genetic architecture. Nat Genet 2013;45(5):501-12, doi:10.1038/ng.2606

Huang LO, Rauch A, Mazzaferro E, et al. Genome-wide discovery of genetic loci that uncouple excess adiposity from its comorbidities. Nat Metab 2021;3(2):228-243, doi:10.1038/s42255-021-00346-2
Willer CJ, Speliotes EK, Loos RJ, et al. Six new loci associated with body mass index highlight a

neuronal influence on body weight regulation. Nat Genet 2009;41(1):25-34, doi:10.1038/ng.287
81. Scherag A, Dina C, Hinney A, et al. Two New Loci for Body-Weight Regulation Identified in a Joint Analysis of Genome-Wide Association Studies for Early-Onset Extreme Obesity in French and German

Study Groups. PLOS Genetics 2010;6(4):e1000916, doi:10.1371/journal.pgen.1000916

82. Graff M, Scott RA, Justice AE, et al. Genome-wide physical activity interactions in adiposity - A meta-analysis of 200,452 adults. PLoS Genet 2017;13(4):e1006528, doi:10.1371/journal.pgen.1006528

83. Wojcik GL, Graff M, Nishimura KK, et al. Genetic analyses of diverse populations improves discovery for complex traits. Nature 2019;570(7762):514-518, doi:10.1038/s41586-019-1310-4

84. Watanabe K, Wilmanski T, Diener C, et al. Multiomic signatures of body mass index identify heterogeneous health phenotypes and responses to a lifestyle intervention. Nature medicine 2023;29(4):996-1008, doi:10.1038/s41591-023-02248-0

85. Rewers M, Zaccaro D, D'Agostino R, et al. Insulin sensitivity, insulinemia, and coronary artery disease: the Insulin Resistance Atherosclerosis Study. Diabetes care 2004;27(3):781-7, doi:10.2337/diacare.27.3.781

86. Do HD, Lohsoonthorn V, Jiamjarasrangsi W, et al. Prevalence of insulin resistance and its relationship with cardiovascular disease risk factors among Thai adults over 35 years old. Diabetes research and clinical practice 2010;89(3):303-8, doi:10.1016/j.diabres.2010.04.013

87. Pacini G, Mari A. Methods for clinical assessment of insulin sensitivity and beta-cell function. Best practice & research Clinical endocrinology & metabolism 2003;17(3):305-22, doi:10.1016/s1521-690x(03)00042-3

88. ElSayed NA, Aleppo G, Aroda VR, et al. 2. Classification and Diagnosis of Diabetes: Standards of Care in Diabetes-2023. Diabetes care 2023;46(Suppl 1):S19-s40, doi:10.2337/dc23-S002

89. Brown AE, Walker M. Genetics of Insulin Resistance and the Metabolic Syndrome. Curr Cardiol Rep 2016;18(8):75-75, doi:10.1007/s11886-016-0755-4

90. Voight BF, Scott LJ, Steinthorsdottir V, et al. Twelve type 2 diabetes susceptibility loci identified through large-scale association analysis. Nature genetics 2010;42(7):579-589, doi:10.1038/ng.609

91. Rung J, Cauchi S, Albrechtsen A, et al. Genetic variant near IRS1 is associated with type 2 diabetes, insulin resistance and hyperinsulinemia. Nat Genet 2009;41(10):1110-5, doi:10.1038/ng.443

92. Boesgaard TW, Gjesing AP, Grarup N, et al. Variant near ADAMTS9 known to associate with type 2 diabetes is related to insulin resistance in offspring of type 2 diabetes patients--EUGENE2 study. PloS one 2009;4(9):e7236, doi:10.1371/journal.pone.0007236

93. Vujkovic M, Keaton JM, Lynch JA, et al. Discovery of 318 new risk loci for type 2 diabetes and related vascular outcomes among 1.4 million participants in a multi-ancestry meta-analysis. Nat Genet 2020;52(7):680-691, doi:10.1038/s41588-020-0637-y

94. Mahajan A, Go MJ, Zhang W, et al. Genome-wide trans-ancestry meta-analysis provides insight into the genetic architecture of type 2 diabetes susceptibility. Nat Genet 2014;46(3):234-44, doi:10.1038/ng.2897

95. Do R, Bailey SD, Desbiens K, et al. Genetic variants of FTO influence adiposity, insulin sensitivity, leptin levels, and resting metabolic rate in the Quebec Family Study. Diabetes 2008;57(4):1147-50, doi:10.2337/db07-1267

96. Grant SF, Thorleifsson G, Reynisdottir I, et al. Variant of transcription factor 7-like 2 (TCF7L2) gene confers risk of type 2 diabetes. Nat Genet 2006;38(3):320-3, doi:10.1038/ng1732

97. Weedon MN. The importance of TCF7L2. Diabetic medicine : a journal of the British Diabetic Association 2007;24(10):1062-6, doi:10.1111/j.1464-5491.2007.02258.x

98. Masotti M, Guo B, Wu B. Pleiotropy informed adaptive association test of multiple traits using genome-wide association study summary data. Biometrics 2019;75(4):1076-1085, doi:10.1111/biom.13076

99. Grant SFA. The TCF7L2 Locus: A Genetic Window Into the Pathogenesis of Type 1 and Type 2 Diabetes. Diabetes care 2019;42(9):1624-1629, doi:10.2337/dci19-0001

100. Lyssenko V, Lupi R, Marchetti P, et al. Mechanisms by which common variants in the TCF7L2 gene increase risk of type 2 diabetes. J Clin Invest 2007;117(8):2155-2163, doi:10.1172/JCI30706

101. Chiefari E, Foti DP, Sgarra R, et al. Transcriptional Regulation of Glucose Metabolism: The Emerging Role of the HMGA1 Chromatin Factor. Frontiers in endocrinology 2018;9(357, doi:10.3389/fendo.2018.00357

102. Bianco A, Chiefari E, Nobile CG, et al. The Association between HMGA1 rs146052672 Variant and Type 2 Diabetes: A Transethnic Meta-Analysis. PloS one 2015;10(8):e0136077,

doi:10.1371/journal.pone.0136077

103. Fuchsberger C, Flannick J, Teslovich TM, et al. The genetic architecture of type 2 diabetes. Nature 2016;536(7614):41-47

104. Baroni MG, Berni A, Romeo S, et al. Genetic study of common variants at the Apo E, Apo AI, Apo CIII, Apo B, lipoprotein lipase (LPL) and hepatic lipase (LIPC) genes and coronary artery disease (CAD): variation in LIPC gene associates with clinical outcomes in patients with established CAD. BMC medical genetics 2003;4(8, doi:10.1186/1471-2350-4-8

105. Hegele RA, Ginsberg HN, Chapman MJ, et al. The polygenic nature of hypertriglyceridaemia: implications for definition, diagnosis, and management. The lancet Diabetes & endocrinology 2014;2(8):655-66, doi:10.1016/s2213-8587(13)70191-8

106. Shearman AM, Ordovas JM, Cupples LA, et al. Evidence for a gene influencing the TG/HDL-C ratio on chromosome 7q32.3-qter: a genome-wide scan in the Framingham study. Human molecular genetics 2000;9(9):1315-20, doi:10.1093/hmg/9.9.1315

107. Dron JS, Wang J, Cao H, et al. Severe hypertriglyceridemia is primarily polygenic. Journal of clinical lipidology 2019;13(1):80-88, doi:10.1016/j.jacl.2018.10.006

108. Ripatti P, Rämö JT, Söderlund S, et al. The Contribution of GWAS Loci in Familial Dyslipidemias. PLoS Genet 2016;12(5):e1006078, doi:10.1371/journal.pgen.1006078

109. Willer CJ, Schmidt EM, Sengupta S, et al. Discovery and refinement of loci associated with lipid levels. Nat Genet 2013;45(11):1274-1283, doi:10.1038/ng.2797

110. Mar R, Pajukanta P, Allayee H, et al. Association of the APOLIPOPROTEIN A1/C3/A4/A5 gene cluster with triglyceride levels and LDL particle size in familial combined hyperlipidemia. Circulation research 2004;94(7):993-9, doi:10.1161/01.res.0000124922.61830.f0

111. Suviolahti E, Lilja HE, Pajukanta P. Unraveling the complex genetics of familial combined hyperlipidemia. Annals of medicine 2006;38(5):337-51, doi:10.1080/07853890600865759

112. Buscot MJ, Magnussen CG, Juonala M, et al. The Combined Effect of Common Genetic Risk Variants on Circulating Lipoproteins Is Evident in Childhood: A Longitudinal Analysis of the

Cardiovascular Risk in Young Finns Study. PloS one 2016;11(1):e0146081, doi:10.1371/journal.pone.0146081

113. Ripatti P, Rämö JT, Mars NJ, et al. Polygenic Hyperlipidemias and Coronary Artery Disease Risk. Circulation: Genomic and Precision Medicine 2020;13(2):e002725,

doi:doi:10.1161/CIRCGEN.119.002725

114. Casula M, Colpani O, Xie S, et al. HDL in Atherosclerotic Cardiovascular Disease: In Search of a Role. Cells 2021;10(8), doi:10.3390/cells10081869

115. Di Angelantonio E, Sarwar N, Perry P, et al. Major lipids, apolipoproteins, and risk of vascular disease. Jama 2009;302(18):1993-2000, doi:10.1001/jama.2009.1619

116. Rye KA, Bursill CA, Lambert G, et al. The metabolism and anti-atherogenic properties of HDL. Journal of lipid research 2009;50 Suppl(Suppl):S195-200, doi:10.1194/jlr.R800034-JLR200

117. Jia C, Anderson JLC, Gruppen EG, et al. High-Density Lipoprotein Anti-Inflammatory Capacity and Incident Cardiovascular Events. Circulation 2021;143(20):1935-1945,

doi:10.1161/circulationaha.120.050808

118. Santos RD, Asztalos BF, Martinez LR, et al. Clinical presentation, laboratory values, and coronary heart disease risk in marked high-density lipoprotein-deficiency states. Journal of clinical lipidology 2008;2(4):237-47, doi:10.1016/j.jacl.2008.06.002

119. Rhee EJ, Byrne CD, Sung KC. The HDL cholesterol/apolipoprotein A-I ratio: an indicator of cardiovascular disease. Current opinion in endocrinology, diabetes, and obesity 2017;24(2):148-153, doi:10.1097/med.00000000000315

120. Huang L, Fan B, Ma A, et al. Inhibition of ABCA1 protein degradation promotes HDL cholesterol efflux capacity and RCT and reduces atherosclerosis in mice. Journal of lipid research 2015;56(5):986-97, doi:10.1194/jlr.M054742

121. van den Bogaard B, Holleboom AG, Duivenvoorden R, et al. Patients with low HDL-cholesterol caused by mutations in LCAT have increased arterial stiffness. Atherosclerosis 2012;225(2):481-5, doi:10.1016/j.atherosclerosis.2012.09.022

122. Rahalkar AR, Giffen F, Har B, et al. Novel LPL mutations associated with lipoprotein lipase deficiency: two case reports and a literature review. Canadian journal of physiology and pharmacology 2009;87(3):151-60, doi:10.1139/y09-005

123. Klos KL, Kullo IJ. Genetic determinants of HDL: monogenic disorders and contributions to variation. Current opinion in cardiology 2007;22(4):344-51, doi:10.1097/HCO.0b013e3281a8acad
124. Kral BG, Becker DM. Familial occurrence of abnormalities of high-density lipoprotein cholesterol.

Journal of clinical lipidology 2007;1(1):31-40, doi:10.1016/j.jacl.2007.01.006

125. Sun L, Pennells L, Kaptoge S, et al. Polygenic risk scores in cardiovascular risk prediction: A cohort study and modelling analyses. PLoS medicine 2021;18(1):e1003498,

doi:10.1371/journal.pmed.1003498

126. Wang C, Li H, Chen K, et al. Association of polymorphisms rs1800012 in COL1A1 with sportsrelated tendon and ligament injuries: A meta-analysis. 2015.

127. James PA, Oparil S, Carter BL, et al. 2014 evidence-based guideline for the management of high blood pressure in adults: report from the panel members appointed to the Eighth Joint National Committee (JNC 8). Jama 2014;311(5):507-20, doi:10.1001/jama.2013.284427

128. Giri A, Hellwege JN, Keaton JM, et al. Trans-ethnic association study of blood pressure determinants in over 750,000 individuals. Nat Genet 2019;51(1):51-62, doi:10.1038/s41588-018-0303-9 129. Evangelou E, Warren HR, Mosen-Ansorena D, et al. Genetic analysis of over 1 million people identifies 535 new loci associated with blood pressure traits. Nat Genet 2018;50(10):1412-1425, doi:10.1038/s41588-018-0205-x

130. Warren HR, Evangelou E, Cabrera CP, et al. Genome-wide association analysis identifies novel blood pressure loci and offers biological insights into cardiovascular risk. Nat Genet 2017;49(3):403-415, doi:10.1038/ng.3768

131. Cooper R. Hypertension, Genes, and Environment: Challenges for Prevention and Risk Prediction. Circulation 2018;137(7):662-664, doi:10.1161/circulationaha.117.032196

132. Li J, Tsuprykov O, Yang X, et al. Paternal programming of offspring cardiometabolic diseases in later life. Journal of hypertension 2016;34(11):2111-26, doi:10.1097/hjh.000000000001051

133. Ji LD, Tang NLS, Xu ZF, et al. Genes Regulate Blood Pressure, but "Environments" Cause Hypertension. Frontiers in genetics 2020;11(580443, doi:10.3389/fgene.2020.580443

134. Ji LD, Tang NL, Xu J. AGTR1 has undergone natural selection in Euro-Asian populations in relation to ambient temperature that predisposes Chinese populations to essential hypertension. International journal of cardiology 2016;209(278-80, doi:10.1016/j.ijcard.2016.02.031

135. Gauderman WJ, Mukherjee B, Aschard H, et al. Update on the State of the Science for Analytical Methods for Gene-Environment Interactions. American journal of epidemiology 2017;186(7):762-770, doi:10.1093/aje/kwx228

136. Hosseini-Esfahani F, Koochakpoor G, Daneshpour MS, et al. The interaction of fat mass and obesity associated gene polymorphisms and dietary fiber intake in relation to obesity phenotypes. Scientific reports 2017;7(1):18057, doi:10.1038/s41598-017-18386-8

137. Villegas R, Goodloe RJ, McClellan BE, Jr., et al. Gene-carbohydrate and gene-fiber interactions and type 2 diabetes in diverse populations from the National Health and Nutrition Examination Surveys (NHANES) as part of the Epidemiologic Architecture for Genes Linked to Environment (EAGLE) study. BMC genetics 2014;15(69, doi:10.1186/1471-2156-15-69

138. Rukh G, Sonestedt E, Melander O, et al. Genetic susceptibility to obesity and diet intakes: association and interaction analyses in the Malmö Diet and Cancer Study. Genes & nutrition 2013;8(6):535-47, doi:10.1007/s12263-013-0352-8

139. Ortega-Azorín C, Sorlí JV, Asensio EM, et al. Associations of the FTO rs9939609 and the MC4R rs17782313 polymorphisms with type 2 diabetes are modulated by diet, being higher when adherence to the Mediterranean diet pattern is low. Cardiovascular diabetology 2012;11(137, doi:10.1186/1475-2840-11-137

140. Goni L, Cuervo M, Milagro FI, et al. A genetic risk tool for obesity predisposition assessment and personalized nutrition implementation based on macronutrient intake. Genes & nutrition 2015;10(1):445, doi:10.1007/s12263-014-0445-z

141. Qi Q, Chu AY, Kang JH, et al. Fried food consumption, genetic risk, and body mass index: genediet interaction analysis in three US cohort studies. BMJ (Clinical research ed) 2014;348(g1610, doi:10.1136/bmj.g1610

142. He J, Kelly TN, Zhao Q, et al. Genome-wide association study identifies 8 novel loci associated with blood pressure responses to interventions in Han Chinese. Circulation Cardiovascular genetics 2013;6(6):598-607, doi:10.1161/circgenetics.113.000307

143. Li C, He J, Chen J, et al. Genome-Wide Gene-Sodium Interaction Analyses on Blood Pressure: The Genetic Epidemiology Network of Salt-Sensitivity Study. Hypertension (Dallas, Tex : 1979) 2016;68(2):348-55, doi:10.1161/hypertensionaha.115.06765

144. Nierenberg JL, Li C, He J, et al. Blood Pressure Genetic Risk Score Predicts Blood Pressure Responses to Dietary Sodium and Potassium. Hypertension (Dallas, Tex : 1979) 2017;70(6):1106-1112, doi:doi:10.1161/HYPERTENSIONAHA.117.10108

145. Hannon BA, Edwards CG, Thompson SV, et al. Genetic Variants in Lipid Metabolism Pathways Interact with Diet to Influence Blood Lipid Concentrations in Adults with Overweight and Obesity. Lifestyle genomics 2020;13(6):155-163, doi:10.1159/000507021 146. Ordovas JM, Ferguson LR, Tai ES, et al. Personalised nutrition and health. BMJ (Clinical research ed) 2018;361(bmj.k2173, doi:10.1136/bmj.k2173

147. Kraemer K, Cordaro J, Fanzo J, et al. Personalized Nutrition: Paving the way to better population health. In: Good Nutrition: Perspectives for the 21st Century. Karger Publishers: 2016; pp. 235-248.
148. Frazier-Wood AC. Dietary Patterns, Genes, and Health: Challenges and Obstacles to be

148. Frazier-Wood AC. Dietary Patterns, Genes, and Health: Challenges and Obstacles to be Overcome. Current nutrition reports 2015;4(82-87, doi:10.1007/s13668-014-0110-6

149. Song H, Koh Y, Rhee TM, et al. Prediction of incident atherosclerotic cardiovascular disease with polygenic risk of metabolic disease: Analysis of 3 prospective cohort studies in Korea. Atherosclerosis 2022;348(16-24, doi:10.1016/j.atherosclerosis.2022.03.021

150. Kan J, Ni J, Xue K, et al. Personalized Nutrition Intervention Improves Health Status in Overweight/Obese Chinese Adults: A Randomized Controlled Trial. Frontiers in nutrition 2022;9(919882, doi:10.3389/fnut.2022.919882

151. Celis-Morales C, Livingstone KM, Marsaux CF, et al. Effect of personalized nutrition on healthrelated behaviour change: evidence from the Food4Me European randomized controlled trial. International journal of epidemiology 2017;46(2):578-588, doi:10.1093/ije/dyw186

152. Livingstone KM, Celis-Morales C, Navas-Carretero S, et al. Personalised nutrition advice reduces intake of discretionary foods and beverages: findings from the Food4Me randomised controlled trial. The international journal of behavioral nutrition and physical activity 2021;18(1):70, doi:10.1186/s12966-021-01136-5

153. Hyde PN, Sapper TN, Crabtree CD, et al. Dietary carbohydrate restriction improves metabolic syndrome independent of weight loss. JCI insight 2019;4(12), doi:10.1172/jci.insight.128308

154. Lutsey PL, Steffen LM, Stevens J. Dietary intake and the development of the metabolic syndrome: the Atherosclerosis Risk in Communities study. Circulation 2008;117(6):754-61, doi:10.1161/circulationaha.107.716159

155. Ruaño G, Windemuth A, Kocherla M, et al. Physiogenomic analysis of weight loss induced by dietary carbohydrate restriction. Nutrition & metabolism 2006;3(20, doi:10.1186/1743-7075-3-20
156. Seip RL, Volek JS, Windemuth A, et al. Physiogenomic comparison of human fat loss in response to diets restrictive of carbohydrate or fat. Nutrition & metabolism 2008;5(4, doi:10.1186/1743-7075-5-4
157. van Walree ES, Jansen IE, Bell NY, et al. Disentangling Genetic Risks for Metabolic Syndrome. Diabetes 2022;71(11):2447-2457, doi:10.2337/db22-0478

158. Corella D, Coltell O, Mattingley G, et al. Utilizing nutritional genomics to tailor diets for the prevention of cardiovascular disease: a guide for upcoming studies and implementations. Expert review of molecular diagnostics 2017;17(5):495-513, doi:10.1080/14737159.2017.1311208

159. Allison DB, Bassaganya-Riera J, Burlingame B, et al. Goals in Nutrition Science 2015-2020. Frontiers in nutrition 2015;2(26, doi:10.3389/fnut.2015.00026

160. Ferguson LR, De Caterina R, Görman U, et al. Guide and Position of the International Society of Nutrigenetics/Nutrigenomics on Personalised Nutrition: Part 1 - Fields of Precision Nutrition. Journal of nutrigenetics and nutrigenomics 2016;9(1):12-27, doi:10.1159/000445350

161. Wan JY, Goodman DL, Willems EL, et al. Genome-wide association analysis of metabolic syndrome quantitative traits in the GENNID multiethnic family study. Diabetology & metabolic syndrome 2021;13(1):59, doi:10.1186/s13098-021-00670-3

162. Kong S, Cho YS. Identification of female-specific genetic variants for metabolic syndrome and its component traits to improve the prediction of metabolic syndrome in females. BMC medical genetics 2019;20(1):99, doi:10.1186/s12881-019-0830-y

163. Moon S, Lee Y, Won S, et al. Multiple genotype–phenotype association study reveals intronic variant pair on SIDT2 associated with metabolic syndrome in a Korean population. Human genomics 2018;12(1):48, doi:10.1186/s40246-018-0180-4

164. Lee HS, Kim Y, Park T. New Common and Rare Variants Influencing Metabolic Syndrome and Its Individual Components in a Korean Population. Scientific reports 2018;8(1):5701, doi:10.1038/s41598-018-23074-2

165. Shim U, Kim HN, Sung YA, et al. Pathway Analysis of Metabolic Syndrome Using a Genome-Wide Association Study of Korea Associated Resource (KARE) Cohorts. Genomics & informatics 2014;12(4):195-202, doi:10.5808/gi.2014.12.4.195

166. Zabaneh D, Balding DJ. A genome-wide association study of the metabolic syndrome in Indian Asian men. PloS one 2010;5(8):e11961, doi:10.1371/journal.pone.0011961

Risk factors	ATP III [®]	WHO ⁸	IDF ¹⁴	AHA/NHLBI ¹³
Blood pressure	≥130/≥85 mmHg	≥160/≥90 mmHg	≥130/≥85 mmHg	≥130/≥85 mmHg or drug treatment
Glucose levels	Fasting: ≥110 mg/dL (6.1 mmol/L) or T2D	IGT or IFG orT2D and/or IR	≥100 mg/dL (5.6 mmol/L) or drug treatment or T2D	≥100 mg/dL (5.6 mmol/L) or T2D or drug treatment
Triglyceride levels	≥150 mg/dL (1.7 mmol/L)	≥150 mg/dL (1.7 mmol/L)	≥150 mg/dL (1.7 mmol/L) or drug treatment	≥150 mg/dL (1.7 mmol/L) or drug treatment
HDL-C	<40 mg/dL (1 mmol/L) M <50 mg/dL (1.7 mmol/L) W	<35 mg/dL (0.9 mmol/L) M <39 mg/dL (1 mmol/L) W	<40 mg/dL (1 mmol/L) M <50 mg/dL (1.7 mmol/L) W or drug treatment	<40 mg/dL (1 mmol/L) M <50 mg/dL (1.7 mmol/L) W or drug treatment
Abdominal obesity	WC >102 cm (40 in) M >88 cm (35 in) W	WHR >0.90 M >0.85 W and/or BMI >30 kg/m ²	Population- and country-specific definitions	WC ≥102 cm (≥40 in) M ≥88 cm (≥35 in) W
Microalbuminuria	-	Urinary excretion rate ≥20 µg min ⁻¹ or albumin:creatinine ratio ≥20 mg g ⁻¹	-	-
Diagnosis of MetS:	≥ 3 risk factors	IFG, IGT or T2D and/or IR together with ≥2 additional risk factors	Abdominal obesity and ≥2 risk factors	≥ 3 risk factors

Table I. Diagnostic criteria for MetS, according to ATP III, WHO, IDF and AHA/NHLBI.

AHA/NHLBI, American Heart Association/National Heart, Lung, and Blood Institute; ATP III, the Panel for the Treatment of Adults III; BMI, Body Mass Index; F, female; HDL-C, high-density lipoprotein cholesterol; IDF, International Diabetes Federation; IFG, impaired fasting glucose, defined by WHO as fasting <110 mg/dL (<6.1 mmol/L) and 2-h post glucose load \geq 110 mg/dL (\geq 6.7 mmol/L) <180 mg/dL (<10 mmol/L); IGT, impaired glucose tolerance, defined by WHO as fasting \geq 100 mg/dL (\geq 5.6 mmol/L) <110mg/dL (<6.1 mmol/L) and 2-h <120 mg/dL (<6.7 mmol/L); IR, insulin resistance; M, male; MetS, metabolic syndrome; T2D, type 2 diabetes; WC, waist circumference; WHO, World Health Organization; WHR, waist-hip ratio.

Publication year	Title	Discovery sample and ancestry	Reference
2021	Genome-wide association analysis of metabolic syndrome quantitative traits in the GENNID multiethnic family study	1520 Multiethnic	161
2020	Genome-wide association study of metabolic syndrome in Korean populations.	7423 East Asian	28
2019	Genome-Wide Association Study of the Metabolic Syndrome in UK Biobank.	291107 European	27
2019	Genome-Wide Association Study of Metabolic Syndrome Reveals Primary Genetic Variants at CETP Locus in Indians	2158 South Asian	30
2019	Identification of female-specific genetic variants for metabolic syndrome and its component traits to improve the prediction of metabolic syndrome in females.	4659 East Asian	162
2018	Multiple genotype-phenotype association study reveals intronic variant pair on SIDT2 associated with metabolic syndrome in a Korean population	7198 East Asian	163
2018	New Common and Rare Variants Influencing Metabolic Syndrome and Its Individual Components in a Korean Population	8373 East Asian	164
2017	Detection of susceptibility loci on APOA5 and COLEC12 associated with metabolic syndrome using a genome-wide association study in a Taiwanese population.	10300 East Asian	29
2017	Susceptibility loci for metabolic syndrome and metabolic components identified in Han Chinese: a multi-stage genome-wide association study.	1742 East Asian	31
2014	Pathway Analysis of Metabolic Syndrome Using a Genome-Wide Association Study of Korea Associated Resource (KARE) Cohorts.	8842 East Asian	165
2012	Genome-wide screen for metabolic syndrome susceptibility Loci reveals strong lipid gene contribution but no evidence for common genetic basis for clustering of metabolic syndrome traits	10564 European	32
2011	A bivariate genome-wide approach to metabolic syndrome: STAMPEED consortium.	22161 European	33
2010	A genome-wide association study of the metabolic syndrome in Indian Asian men.	2554 South Asian	166

Table II. Principal GWAS published with metabolic syndrome as associated trait.

Table III. Evidence from recent large GWAS and PRS studies and the	eir potential clinical
utility in the assessment and treatment of MetS and other related cardi	ometabolic traits.

Author, year	Populatio n studied	PRS model	Results	Conclusions	Potential Clinical utility
Song H et <i>al.</i> , 2022	3 Korean cohorts of 28,445, 8840 and 4333 individual s.	PRS _{Mets} - ASCVD: construction of the most optimal combination of PRS's for prediction of ASCVD.	6.7% of the population was at high genetic risk with 3.3-fold (95% C.I. 1.7- 6.1, p<0.001) higher risk for incident ASCVD.	The polygenic risk of metabolic disease independently predicts those at an increased risk of ASCVD, identifying those at a genetically high risk of incident ASCVD.	The combination of PRS ASCVD and conventional risk factors (such as age, sex, BMI, smoking, hypertension, diabetes and hyperlipidemia) could provide a better performance for predicting ASCVD, especially in younger individuals.
Eva S. van Walree <i>et al.</i> , 2022	3 cohorts of European and multi- ancestry represent ation	Polygenic risk score drafted from the MetS factor GWAS.	PRS predicts 5.9% of the variance in MetS. Of the 235 loci identified in the GWAS, 53 (22.5%) overlap with loci identified for two or more MetS components.	Genetic correlations are best captured by a genetic one factor model. The MetS components genetic overlapping indicates that this entity is a complex, heterogeneous disorder.	These results provide mechanistic insights into the genetics of MetS and suggestions for drug targets, especially fenofibrate, which has the promise of tackling multiple MetS components.
Hardy, D.S. <i>et</i> <i>al</i> ., 2021	10,681 European American s and African American s.	PRS and its interaction with dietary patterns to increase MetS risk.	Among each racial group within PRS tertiles, the Western dietary pattern was associated with development and cycling of MetS status between visits, and the high-fat dairy pattern with being free from MetS (p < 0.017).	The influence of dietary patterns or MetS risk appears to differ by genetic predisposition and racial ancestry.	f Dietary patterns assessment becomes more important, especially for individuals with higher genetic risk for MetS. Routinely clinical- nutritional consultations could provide a clearer image of the overall increased risk for MetS in some individuals.

PRS, polygenic risk score; ASCVD, atherosclerotic cardiovascular disease; PRS_{MetS}-ASCVD, metabolic PRS to identify atherosclerotic cardiovascular disease incidence; BMI, body mass index; MetS, Metabolic syndrome; GWAS, Genome-wide association study.

Figure legends

Figure 1. Main pathomecanisms associated to the development of MetS.

Ang II, angiotensin II; CRP, C-reactive protein; DAMPs, damage-associated molecular patterns; FFAs, free fatty acids; GLUT4, glucose transporter type 4; IL-6, interleukin 6; IR, insulin resistance; IRS-1/PI3K, insulin receptor substrate-associated phosphoinositide 3-kinase activity; LDL, low density lipoprotein; LOX-1 lipoprotein receptor-1; LPS, lipopolysaccharides; MetS, metabolic syndrome; NF-kB, nuclear factor kappa-light-chain enhancer of activated B cells; NO, nitric oxide; PAMPs, pathogen-associated molecular patterns; RAS, renin–angiotensinogen system; ROS, reactive oxygen species; TLRs, Toll-like receptors; TNF α , tumor necrosis factor α ; VAT, visceral adipose tissue.

Figure 2. Main genes associated to the presence of MetS components.

Figure 3. Generation, validation, and application of $\mathsf{PRS}_{\mathsf{MetS}}$ and its components.



Main pathomecanisms associated to the development of MetS.

Ang II, angiotensin II; CRP, C-reactive protein; DAMPs, damage-associated molecular patterns; FFAs, free fatty acids; GLUT4, glucose transporter type 4; IL-6, interleukin 6; IR, insulin resistance; IRS-1/PI3K, insulin receptor substrate-associated phosphoinositide 3-kinase activity; LDL, low density lipoprotein; LOX-1 lipoprotein receptor-1; LPS, lipopolysaccharides; MetS, metabolic syndrome; NF-kB, nuclear factor kappalight-chain enhancer of activated B cells; NO, nitric oxide; PAMPs, pathogen-associated molecular patterns; RAS, renin-angiotensinogen system; ROS, reactive oxygen species; TLRs, Toll-like receptors; TNF a, tumor necrosis factor a; VAT, visceral adipose tissue.

566x347mm (72 x 72 DPI)



Main genes associated to the presence of MetS components.

382x247mm (72 x 72 DPI)



Generation, validation, and application of PRSMetS and its components.

365x218mm (72 x 72 DPI)