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Environmental DNA metabarcoding reveals the effects of seafloor litter and trawling on marine biodiversity



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ABSTRACT

Environmental DNA (eDNA) techniques are emerging as promising tools for monitoring marine communities. However, they have not been applied to study the integrated effects of anthropogenic pressures on marine biodiversity. This study examined the relationships between demersal community species composition, key environmental features, and anthropogenic impacts such as fishing effort and seafloor litter using eDNA data in the central Tyrrhenian Sea. The results indicated that both fishing effort and seafloor litter influenced species composition and diversity. The adaptive traits of marine species played a critical role in their response to debris accumulation and fishing. Mobile species appeared to use relocation strategies, while sessile species showed flexibility in the face of disturbance. Epibiotic species relied on passive transport. The use of eDNA-based methods is a valuable resource for monitoring anthropogenic impacts during scientific surveys, enhancing our ability to monitor marine ecosystems and more effectively assess the effects of pollution.

1. Introduction

Marine biodiversity plays a key role in maintaining the health and balance of marine ecosystems (Goulletquer et al., 2014). The seafloor hosts many marine species, encompassing a wide range of organisms that collectively contribute to the complex structural arrangement of life in the depths of the sea. However, this priceless biodiversity faces significant challenges from direct and/or indirect anthropogenic impacts, including fishing, and pollutants (Halpern et al., 2008).

Commercial trawling is recognized as a major anthropogenic disturbance to seafloor ecosystems, leading to significant changes in benthic community composition, in terms of abundance and spatial distribution (Beauchard et al., 2021; Eigaard et al., 2017; Oberle et al., 2016 Clark et al., 2016; Pusceddu et al., 2014). The effects of trawling on benthic communities vary depending on the sensitivity of the animals and the resilience of the population (Hiddink et al., 2017; Kaiser et al., 2006; Tiano et al., 2020). Fishing activity has direct and indirect impacts on the environment. Direct effects include substrate scraping, sediment resuspension, and benthos destruction. Indirect effects include

post-fishing mortality and long-term changes in the community. Additionally, trawling can alter the availability of food and habitats, affecting species differently, depending on their feeding strategy (Johnson et al., 2015; Mangano et al., 2017). Indeed, it could increase food availability for opportunistic species (Hiddink et al., 2008) or remove certain prey for more selective organisms (Hiddink et al., 2017). Previous studies (Bozzano and Sardaà, 2002; Groenewold et al., 2000; Kaiser and Spencer, 1994; Lejeune et al., 2023) showed that scavengers may benefit from the discards produced by trawling, while more vulnerable species, such as sessile filter feeders, were removed.

Nowadays, marine litter represents one of the most predominant and persistent pollutants (Deudero and Alomar, 2015). Litter waste mainly originates from the mainland and is transported to the sea by surface runoff and rivers (Boucher et al., 2020). Once enters the ocean, litter can be trapped on the seafloor and persist for extended periods ranging from decades to even centuries (Canals et al., 2021). Seafloor litter pollution has emerged as a significant environmental concern, posing potential threats to marine biodiversity and ecosystem functioning (Kühn et al., 2015). The seafloor is a key habitat for numerous marine species, and

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the accumulation of litter disrupts the fundamental equilibrium and functioning of many ecological systems (Angiolillo and Fortibuoni, 2020; de Carvalho-Souza et al., 2018; Galgani et al., 2018). Indeed, persistent pollution not only inflicts physical harm upon marine organisms through entanglement and ingestion but also disrupts seafloor ecosystems by altering nutrient availability and by modifying the physical and chemical properties of the environment (Angiolillo and Fortibuoni, 2020; Sbrana et al., 2022). Finally, a relevant consequence of litter accumulation consists of the phenomenon known as biofouling: various benthic organisms colonize and establish communities on submerged surfaces, including litter (Mancini et al., 2021; Pinochet et al., 2020; Póvoa et al., 2021; Rech et al., 2018). Biofilm formation on plastic, the so-called Plastisphere (Zettler et al., 2013), could alter the physical, chemical, and biological characteristics of plastic in aquatic environments, causing an increase in plastic density and sinking. Moreover, benthic plastic debris substrates may have the potential to change the biodiversity and structure of benthic communities (Subías-Baratau et al., 2022).

In the last decade, various monitoring programs and modelling approaches have been applied to assess the accumulation of litter at sea and the importance of its drivers. Recently, (Cau et al., 2024) used a machine learning approach (i.e., Random Forests) to model and investigate pathways from a large available dataset of marine litter distribution in the western and central Mediterranean Sea, producing suitable results.

Understanding the risk posed by anthropogenic pressures on the seafloor, such as trawling activities and marine litter, is crucial for developing effective conservation and management strategies. This could benefit greatly from a multidisciplinary framework that includes sources, distribution, and environmental impacts. (Hardesty et al., 2019). The assessment of seafloor impacts is particularly urgent in the Mediterranean Sea, which is known to be a biodiversity hotspot at high risk of degradation, due to its semi-enclosed and highly anthropised basin (Boucher and Billard, 2020; Coll et al., 2012; Cózar et al., 2015; Deudero and Alomar, 2015). Nevertheless, knowledge about the impacts on the seafloors remains limited, probably due to the difficulties involved in sampling the diversity and understanding ecosystem processes and ecological interactions in these environments (Sandra et al., 2023). Many studies focused on the impact of marine litter and trawling on indicative species of Vulnerable Marine Ecosystems (Consoli et al., 2020; Downie et al., 2021; Horton and Barnes, 2020; Mačić et al., 2022) but there is a lack of studies that address the problem from a broader ecological perspective (i.e., at the scale of populations or communities) (Hardesty et al., 2019).

Ecological investigations involve the examination of species occurrence patterns as a fundamental task. Traditionally, assessing biodiversity in marine ecosystems has relied on scientific surveys, traps, or fishery-dependent data (Angiolillo et al., 2023; Kipson et al., 2011; Mallet and Pelletier, 2014; Pennino et al., 2016). However, these methods have various limitations, such as their high costs, logistical challenges, and potential disturbance to the fragile ecosystems under investigation (Brandt et al., 2016). As a result, there is a need to explore alternative approaches able to overcome these limitations while providing valuable insights into the effects of anthropogenic pressures on marine communities. Environmental DNA (eDNA) techniques have gained increasing recognition and usage in biodiversity studies, offering promising opportunities for monitoring the impacts of different human-induced stressors (Yoccoz, 2018). By analysing genetic material shed by organisms into the surrounding environment, eDNA sequencing allows for a non-invasive and efficient assessment of species presence and diversity (Bohmann et al., 2014; Thomsen and Willerslev, 2015). Recently, it has also been shown that eDNA can be collected efficiently and inexpensively during normal fishing activities of the commercial fleet (Maiello et al., 2022, 2023). eDNA has been successfully applied in various contexts, including the detection of rare or elusive species, monitoring invasive species, and assessing community dynamics (Cicala

et al., 2024; Albonetti et al., 2023; Fonseca et al., 2023; Juhel et al., 2022). However, the potential application of eDNA metabarcoding techniques to investigate the impact of seafloor litter pollution on biodiversity has not yet been implemented.

The aim of this study is therefore to examine the impact of seafloor litter pollution and trawling activities on marine communities. The study uses opportunistically collected eDNA data to identify species occurrence and relate them to the various levels of fishing effort and litter pollution. The metabarcoding approach overcomes the limitations of traditional methods in assessing marine communities, promoting the study of the effects of anthropogenic pressures on organisms under investigation. The association of species with marine debris and fishing activities could provide insight into possible biological interactions and the broader consequences of these pressures on marine biodiversity.

This research would represent a significant step towards understanding the complex relationship between anthropogenic pressures and marine communities. In particular, it could help expand knowledge on marine litter, with potential implications for the development of targeted mitigation and management strategies.

2. Material and methods

2.1. Area of study

Environmental DNA samples were gathered during summer 2020 from 24 sampling locations aboard a commercial bottom-trawler (Fig. 1). The study area is located in the central Tyrrhenian Sea (FAO Geographical Sub Area 9 – Western Mediterranean Sea) and covers a bathymetric range spanning from the continental shelf edge (average depth \sim 70 m) to the deep slope (\sim 820 m), with a distance from the coast going from 4 to 24 km.

This area has a very narrow continental shelf, characterised by fine sands and muddy bottoms (Ardizzone et al., 2018). The coastal area is heavily populated with large urban centres, industrial settlements, and important ports, being close to the city of Rome. The Tiber River run-off significantly affects the chemical-physical traits of the area, bringing contaminants such as marine debris (Cesarini et al., 2023; Inghilesi et al., 2008). During winter, the river Tiber discharge is carried along the coast and dispersed offshore by the strong Tyrrhenian Sea currents from the northwest. In contrast, during summer, weak sea breezes cause downwelling conditions that limit the river's fresh waters near the mouth (Inghilesi et al., 2008).

2.2. Species distribution from eDNA samples

The analysis performed in this study integrated and developed the ones in Maiello et al. (2023), and as a result, the initial stage of the operational procedure, which includes the data acquisition from the eDNA sample, is the same in both studies. The data acquisition process from eDNA will be briefly described.

Two approaches were adopted to sample eDNA aboard the commercial trawler: *slush* water and the *metaprobe* rolls of gauze. The *metaprobe* is a hollow-perforated reusable plastic ball (the 3D project is freely available at: https://github.com/GiuliaMaiello/Metaprobe-2.0), into which three rolls of sterile gauze were placed to passively absorb DNA from the surrounding environment. To account for possible contamination linked with the fishing boat background, field blanks were collected along with environmental samples.

Samples were processed in hight containment rooms to avoid crosscontamination. After eDNA extraction, two taxonomically informative mitochondrial regions were PCR amplified from eDNA samples: a ~167 bp fragment of the 12 S gene and a ~313 bp fragment of the COI gene. The first fragment was amplified using the fish-specific Tele02 primers, while the second fragment was amplified using highly degenerated universal metazoan primers. A positive PCR control (*Sebastes mentella*, a subarctic species absent in the Mediterranean Sea) and a negative



Fig. 1. A: Map of the 24 eDNA sampling sites (white circles) and outgroup hauls (blue circles); B: Map of fishing effort (hours of bottom trawl fishing activity, log-transformed); C: Map of seafloor litter (number of objects • km⁻², log-transformed) in the central Tyrrhenian Sea (FAO Geographical Sub Area, GSA 9–western Mediterranean Sea).

control were included, to monitor the degree of contamination associated with laboratory procedures.

Bioinformatic analysis followed the OBITOOLS pipeline (Boyer et al., 2016). Sequences were clustered into Molecular Operational Taxonomic Units (MOTU) using SWARM (Mahé et al., 2015), setting a clustering threshold at d = 3 for Tele02 12 S and d = 13 for the universal COI (Kemp et al., 2019; Siegenthaler et al., 2019; Wangensteen et al., 2018). The taxonomic assignment was then performed with ECOTAG, followed by the examination of ambiguous and poorly resolved MOTUs/taxa (e. g., non-Mediterranean taxa) using BLASTn search in the NCBI system. Finally, the data were filtered to retain only sequences showing identity matches >98% and removing potential contamination noise using blanks and negative controls with the decontam package in R (Davis et al., 2018), using the prevalence method with a threshold of 0.5.

2.3. Anthropogenic pressures and environmental features

In this study, we aimed to investigate the structure of demersal

communities, with a focus on the primary edaphic factors and key sources of anthropogenic disturbance such as pollution and fishing effort, using information obtained through eDNA metabarcoding. To achieve this goal, we quantified various descriptors at each sampling site, integrating them into the statistical analyses described in the next sections.

- Seafloor litter data were extrapolated from a Random Forests (RF) model on the whole western Mediterranean Sea (https://doi.org/10 .17632/r2b6svy7h7.1; Cau et al., 2024). Input data used for the RF model were collected by the MEDiterranean International bottom Trawl Survey (MEDITS) from 2013 to 2020, according to the standardized official protocol (MEDITS working group, 2012). A long series of datasets has provided a more accurate representation of litter presence on the seabed and its potential impact on species with multi-year life cycles. The MEDITS protocol for monitoring marine litter (in agreement with the requirements of the Marine Strategy Directive Framework (Directive, 2008/56/EC)) is based on a stratified random sampling design, providing reliable estimates for the area between 0 and 1000 m in depth, and the total number of objects collected is then standardized according to the swept area, expressed in number of objects km⁻².
- The Mean Fishing effort (bottom trawling) was calculated as the mean annual hours of trawling, over the last 5 years before the sampling (2016–2020), in the cell of a 1 km² grid defined for the area of study, using the Vessel Monitoring System data provided by the Italian Ministry of Agriculture, Food Sovereignty and Forests and applying the procedures described in (Russo et al., 2014, 2016). Gridded fishing effort data were used to assign an effort value to each sampling site by matching the sites with their respective grid cells.
- Cumulative human impacts: we integrated data from various sources to identify anthropogenic descriptors that affect marine communities: Mean Distance from Shipping Lanes (https://knb.ecoin formatics.org/view/doi:10.5063/F1S180FS); Impact of Port Areas (https://msi.nga.mil/Publications/WPI); Impact of Urban Areas (https://www.efrainmaps.es/english-version/free-downloads/eur ope/).
- The distance (in Km) from the Tiber River mouth, assuming that rivers are the main carriers of nutrients and pollutants for the marine communities.
- The Sea bottom Depth (m) estimated from the NOAA ETOPO1 Global Relief Model using the R package *marmap* (Pante and Bouhet, 2015), assuming that depth is a major forcer of demersal community structure (Pérès and Picard, 1964).
- The Distance from the coast (Km), computed using the *dist2Line* function of the R package geosphere (Hijmans, 2019).

2.4. Data analysis

The final dataset comprised, for each of the 24 sampling locations: species presence as detected by eDNA metabarcoding, number of seafloor litter, fishing effort, environmental features, and anthropogenic pressures.

Taxonomic composition of marine species, resulting from eDNA data analysis, was examined in presence-absence format. Only species directly related to seafloor litter were maintained in the analysis (i.e., individuals of benthic and necto-benthic species, identified from the FishBase database for fishes (http://www.fishbase.org) or SeaLifeBase (http://www.sealifebase.org) for invertebrates), thus excluding those embracing strictly pelagic behaviours. Furthermore, we excluded rare species from the analysis, defined as those found in less than 25% of the sampling sites.

The total amount of marine litter (number of objects \bullet km⁻²) and the Mean Fishing effort (total annual hours of bottom trawling) were used to perform, in parallel, a hierarchical cluster analysis (using a Manhattan distance and the "ward.D2" agglomeration method as provided by the

hclust function of R). We used an additional set of five hauls, corresponding to areas with low fishing effort and low litter levels (outgroups), to provide a comparative framework for evaluating the groups resulting from the cluster analysis. Then, we categorized the sampling sites based on a combined scale of litter (low/high) and fishing pressure (low/high).

We performed a redundancy analysis (RDA) using the *vegan* package in R (Oksanen, 2022) to analyse the community structure, under the influence of environmental features and anthropogenic pressures. RDA is a multivariate technique used to explore the relationship between a set of response variables and a set of explanatory variables (Israels, 1984). RDA maximize the amount of the dependent variables' variance that can be explained by a linear combination of the explanatory factors. In this study, the presence/absence of the different species (response variable) was modelled with respect to the environmental and anthropogenic features (explanatory variables).

Marine community composition, extrapolated from eDNA data, has been set on the family level, so to gain into the ecological roles and patterns within broader taxonomic groups.

The association between biodiversity patterns and combinations of

Table 1

List of taxa selected for this study, identified through eDNA metabarcoding (COI and 12 S) detection, to investigate relationship within the species community in the central Tyrrhenian Sea (Lazio region).). Code numbers refer to the family level.

Class	Family	Species	Code	Class	Family	Species	Code
Anthozoa	Alcyoniidae	Alcyonium acaule	1	Malacostraca	Sergestidae	Robustosergia robusta	
	Epizoanthidae	Epizoanthus arenaceus	2	Polychaeta	Acoetidae	Panthalis oerstedi	63
	Funiculinidae	Funiculina quadrangularis	3		Onuphidae	Paradiopatra calliopae	64
	Sagartiidae	Sagartiogeton spp.	4		Spionidae	Laonice cirrata	65
Ascidiacea	Ascidiidae	Ascidia spp.	5	Teleostei	Argentinidae	Argentina sphyraena	66
Asteroidea	Astronectinidae	Astronecten irregularis	6	releoster	ingentindue	Glossanodon leioglossus	00
risteroideu	Luidiidaa	Luidia sarsi	7		Bothidae	Armoglossus laterna	68
Directorie	Torodinidoo	Dailatanada masatana	,		Doulidae	Amoodoosus tuteritu	00
bivalvia	Teredinidae		8			Arnogiossus rueppeui	=0
Cephalopoda	Eledonidae	Eledone cirrhosa	9		Callionymidae	Synchiropus phaeton	70
	Enoploteuthidae	Abralia veranyi	10		Caproidae	Capros aper	71
	Histioteuthidae	Histioteuthis bonnellii	11		Centrolophidae	Centrolophus niger	72
		Histioteuthis reversa			Cepolidae	Cepola macrophthalma	73
	Loliginidae	Alloteuthis media	13		Chlorophthalmidae	Chlorophthalmus agassizi	74
		Alloteuthis subulata			Citharidae	Citharus linguatula	75
	Octopodidae	Octopus vulgaris	15		Congridae	Conger conger	76
	-	Pteroctopus tetracirrhus			-	Gnathophis mystax	
		Scaeurgus unicirrhus			Epigonidae	Epigonus denticulatus	78
	Ommastrephidae	Illex coindetii	18			Enigonus telesconus	
	••••••	Todarodes sagittatus			Gadidae	Gadiculus argenteus	80
		Todaropsis eblanae			Guadate	Micromesistius poutassou	00
	Onwebsteuthides	Omushatauthia hambaii				Trisontomic conclonus	
	Constitute		00		0.1.1.1.	Trisopierus capetantis	00
	Seplidae	Sepia elegans	22		Gobiidae	Lesueurigodius friesu	83
	Sepiolidae	Heteroteuthis dispar	23		Gonostomatidae	Cyclothone braueri	84
		Neorossia caroli				Gonostoma denudatum	
		Rondeletiola minor			Lophiidae	Lophius budegassa	86
		Rossia macrosoma				Lophius piscatorius	
		Sepietta oweniana			Lotidae	Molva molva	88
		Stoloteuthis leucoptera			Macrouridae	Coelorinchus caelorhincus	89
Crinoidea	Antedonidae	Antedon mediterranea	29			Hymenocephalus italicus	
		Leptometra phalangium				Nezumia sclerorhynchus	
Echinoidea	Brissidae	Brissopsis lvrifera	31		Merlucciidae	Merluccius merluccius	92
	Echinidae	Echinus acutus	32		Moridae	Mora moro	93
Elasmobranchii	Dalatiidae	Dalatias licha	33		Mullidae	Mullus barbatus	94
Liubinobruneim	Etmonteridae	Etmonterus spinar	34		Myctophidae	Ceratos copelus maderensis	95
	Hevanchidae	Havanchus arisaus	35		wyctopindae	Dianhus holti))
	Doptopolidao	Calaus malastomus	35			Diaphus notin	
	Pendancinuae	Distance consistents	30			Electron a micro	
	кајшае	Dipurus oxyrinchus	3/			Electrona risso	
		Leucoraja circularis				Hygophum benoiti	
		Raja clavata				Lampanyctus crocodilus	
	Scyliorhinidae	Scyliorhinus canicula	40			Lobianchia dofleini	
Gastropoda	Cocculinidae	Coccopigya spp.	41			Myctophum punctatum	
Holocephali	Chimaeridae	Chimaera monstrosa	42			Notoscopelus elongatus	
Hydrozoa	Aglaopheniidae	Lytocarpia myriophyllum	43		Nemichthyidae	Nemichthys scolopaceus	104
	Bougainvilliidae	Bougainvillia muscus	44		Nettastomatidae	Facciolella oxyrhyncha	105
	Campanulariidae	Clytia hemisphaerica	45			Nettastoma melanurum	
	Plumulariidae	Nemertesia ramosa	46		Notacanthidae	Notacanthus bonaparte	107
Malacostraca	Aristeidae	Aristeus antennatus	47		Peristediidae	Peristedion cataphractum	108
	Cirolanidae	Natatolana borealis	49		Phycidae	Phycis blennoides	109
	Gonenlacidae	Gonenlay rhomboides	50		Scophthalmidae	Lepidorhombus boscii	110
	Nenbronidae	Nenbrons norvegicus	51		beophinimude	Lepidorhombus whiffiagonis	110
	Dandalidae	Designika acanthonotus	52		Sternontychidae	Armronalacus hamimmus	112
	1 anuanuac	Decionika adwardsii	52		Stemoptychiuae	Maurolicus musllari	114
		Plesionika katana ammua			Ctomiidoo	Chaulia dua alagni	114
		Plesionika neterocarpus			Stollindae	Chaming has	114
	n · 1 · ·	Plesionika martia	-		m 1.1.1.1	Stomias Doa	
	Pasiphaeidae	Pasiphaea multidentata	56		Trachichthyidae	Hoplostethus mediterraneus	116
		Pasiphaea sivado			Trachyrincidae	Trachyrincus scabrus	117
	Penaeidae	Parapenaeus longirostris	58		Trichiuridae	Lepidopus caudatus	118
	Polychelidae	Polycheles typhlops	59		Triglidae	Chelidonichthys cuculus	119
	Sergestidae	Eusergestes arcticus	60			Lepidotrigla cavillone	
		Paraservestes vivilar				Triola lyra	

site groups was studied using a multi-level pattern analysis (*indicspecies* package in R; De Cáceres et al., 2010). This function generates various combinations of the input clusters and evaluates each combination against the species in the input matrix. It then selects the combination with the highest association value for each species and assesses the statistical significance of this relationship through a permutation test.

3. Results

A total of 305 species (193 families) were identified through eDNA metabarcoding (Supplementary Table 1), and among them, 60 pelagic families were excluded from the analysis. Finally, after removing species present in less than 25% of the sampling sites, a total of 122 species (79 families) were retained for the study (Table 1).

The random forest model yielded a total estimation of 46.37 litter objects per square kilometre, with a mean value of 1.93 n. objects·km⁻² (\pm 0.25 n. objects·km⁻²) in each sampling site. Total fishing effort was 54.47 h, with a mean of 2.27 (\pm 0.28) hours. The cluster analysis separated sampling locations into Low litter (N = 17), High litter (N = 7), Low effort (N = 7) and High effort (N = 17) subgroups for seafloor litter and fishing effort respectively (Fig. 2A). The locations were thus split into Low litter - Low effort (N = 7), Low litter - High effort (N = 10), and High litter - High effort (N = 7) by combining the obtained classifications (Fig. 2B).

The redundancy analysis (RDA) was performed to evaluate the relationship of taxonomic group distribution with environmental features, anthropic pressures, and sampling sites. According to the cluster analysis, sites were classified into three groups, with arrows indicating the correlation between the parameters. Sites and organisms were exposed to different stressors (Fig. 3). In particular, variables related to river and seafloor litter are connected with the I quadrant. The distance from the coast is related to the II quadrant, while depth, urban areas, shipping, and harbours are linked to the III quadrant. Finally, the IV quadrant is related to fishing effort pressures. Sampling sites were spread across all four quadrants, with the low litter-low effort sites being concentrated exclusively in quadrant II. In contrast, the remaining two clusters appear to be distributed across multiple quadrants to some extent. The RDA species representation revealed a clear separation of ecological groups along a left-to-right gradient of the x-axis (RDA1). The bathypelagic and bathydemersal individuals were positioned on the left of this axis, while the sessile individuals were on the right side. In contrast, the distribution of benthic, demersal, and benthopelagic species was widespread.

According to the results of the indicator species analysis (Table 2), Moridae and Sergestidae are significantly related with low litter and low effort sampling sites, Epizoanthidae, Peristediidae, and Funiculinidae are strongly related with low litter and high effort sampling sites, and Ascidiidae and Luidiidae are related with high litter and high effort sampling sites.

4. Discussion

The results provided in this study offer a thorough examination of the spatial distribution and prevalence of marine organisms with various anthropogenic pressures in the central Tyrrhenian Sea. These findings hold critical significance for the enhancement of our comprehension of anthropogenic stressors' repercussions on marine ecosystems and the complex interconnections between environmental characteristics and the fauna inhabiting this area. In this study, eDNA data are used for the first time to relate the spatial distribution of species composition with the accumulation of seafloor litter and fishing activities at different sites. Using this method, it was possible to identify consistent differences in the faunal community composition under various levels of pressure.

The study included a comprehensive selection of 120 species identified in 24 sampling sites, which covered a wide spectrum of organisms, from sharks, rays, and bony fish to invertebrates, such as cnidaria hydroids, polychaetes, crustaceans, and echinoderms. Sampling sites presented a clear separation in terms of both litter abundance and fishing effort. The cluster analysis facilitated the discrimination of areas characterised by distinct levels of accumulation, which included both low and high-impact litter, and represented a fundamental starting point for understanding the spatial distribution patterns of marine litter. As observed in the map, high-impact sampling sites were near the Tiber river mouth. This is also supported by the RDA analysis, which showed a strong relationship between river and seafloor litter. Rivers are important sources of waste. Their runoff and discharged debris significantly influence seas and the surrounding seabed (Atwood et al., 2019; Campanale et al., 2020: Crosti et al., 2018: Noce et al., 2013: Poeta et al., 2016; Rech et al., 2014; Sbrana et al., 2020). The fishing effort followed a decreasing gradient from coast to offshore depending on both



Fig. 2. Results of the cluster analyses, which grouped eDNA sampling sites according to fishing effort (h) and seafloor litter (number of objects \cdot km⁻²) (A). Map of the combined classifications (B). *blue*: outgroup sites (control); *green*: low pressures; *red*: high pressures; *orange*: low litter – high effort.



Fig. 3. Results of the redundancy analysis showing the associations between environmental characteristics (distance from the coast, depth, distance from the river), anthropogenic pressures (urban areas, shipping, harbours, seafloor litter, and trawl effort), sampling sites (RDA - sites) and the distribution of the taxonomic groups found (RDA - species) in the central Tyrrhenian Sea. The sampling sites are enclosed in coloured polygons that belong to their respective cluster group. Centroids represent species based on their respective families (Refer to Table 1 for a list of their code numbers).

bathymetry and distance from the coast. The analysis indicated that most fishing activities took place close to the coast, while only a small number of vessels operated in deep waters off the coast influenced by the presence of specific habitats occupied by target species, e.g., the bathyal muddy sediments hosting the red prawns *Aristeomorpha foliacea* and *Aristeus antennatus* (Ardizzone et al., 2018). Indeed, coastal location facilitated cheaper and more accessible fishing resources, and legal restrictions limited access to deeper areas (i.e., Regulation (EU) 2019/1241 of the European Parliament and of the Council of June 20, 2019 on the conservation of fisheries resources and the protection of marine ecosystems through technical measures).

The combination of seafloor litter and fishing effort clusters led to the identification of three different groups (i.e., low litter - low effort, low litter - high effort, and high litter – high effort), which were influenced by the synergistic interplay between the hydrodynamic conditions and the Tiber river discharges. Notably, heavily impacted sampling sites (high litter – high effort) are constrained between the river mouth and the southward-descending current from the northern coast (Iacono

et al., 2021), while low litter – high effort sampling sites are influenced primarily by the fishing activity itself.

The Redundancy Analysis (RDA) results connected multiple environmental features and anthropogenic disturbances that mainly influenced the distribution of fauna on the bathyal seabed. Seafloor litter accumulation was found to be predominantly associated with invertebrates, such as hydroids, alcyonidiids, crinoids, and Loliginidae cephalopods, and with bony fishes (i.e., Cepolidae and Citharidae). These organisms differ considerably in motility modes: the first two being sessile, the others slow-moving, and the latter two are typically nektonic demersal animals. Based on the constraints of the animalsubstrate relationships, we hypothesise that these organisms benefit from the availability of additional substrates and increased habitat complexity provided by the accumulated litter, which promotes spatial heterogeneity and offers new settlement opportunities, particularly for sessile organisms (e.g., hydroids and alcyonidiids). This interpretation was documented by the numerous findings of such animals as a peculiar component of fouling, mainly in shallow waters (Angiolillo et al., 2015;

Table 2

Significant species resulting from Indicator Species Analysis for each cluster of sampling sites. Silhouettes are coloured based on ecological groups: bathype-lagic (light blue), sessile (purple), demersal (green), and benthic (orange). Significant codes: **: p-value <0.01; *: p-value <0.05

Low Litter - Low	stat	p-value						
Moridae	Mora moro		0.727	0.018*				
Segestidae	Eusergestes arcticus Parasergestes vigilax Robustosergia robusta	AND A	0.715	0.039*				
Low Litter - High Effort								
Epizoanthidae	Epizoanthus arenaceus		0.754	0.008**				
Peristediidae	Peristedion cataphractum		0.714	0.018*				
Funiculinidae	Funiculina quadrangularis	X.	0.678	0.013*				
High Litter - High Effort								
Ascidiidae	Ascidia spp.	8	0.728	0.009**				
Luidiidae	Luidia sarsi	\star	0.678	0.003**				

Giangrande et al., 2020; Mancini et al., 2021) and was confirmed in the present study for deep substrates.

The other stressors, such as depth, urban areas, shipping, harbours, and distance from the coast, mainly affected fish and cephalopods. In particular, these included bathypelagic fishes (e.g., Mictophydae, Sternoptychidae), and the cephalopods of the family Enoploteuthidae and Histioteuthidae, with a high capacity for movement, as well as bathyidemersal fish (e.g. some Chondrichthyes such as Dalatiidae, Lophidae and Trachirincidae), which inhabit deep seabeds and are fairly mobile The distribution of these organisms is primarily influenced by depth and strongly associated with the distance from the coast. This is especially true for deeper species, which are a significant part of the assemblages of the bathyal plain of the Mediterranean Sea (Carlucci et al., 2018; Follesa et al., 2011).

Finally, the indicator species analysis highlighted a significant correlation between the offshore sampling sites subjected to low litter – low effort and two bathypelagic families of Moridae and Sergestidae, which are highly motile swimming in deep water. These species are typical inhabitants of the deeper muddy seabed and so their observed distribution can be explained by both depth and distance from the coast.

These results showed that environmental features and anthropogenic disturbances have different effects in shaping marine community structure, depending on the life strategy traits and adaptability of the species. In addition, they suggest important implications for the assessment of trawling impacts. The stressors connected with the intensity of fishing effort were proved to play a pivotal role in shaping the faunal assemblage, which was mainly composed of typical demersal species, widely distributed over the seabed, from the edge of the continental shelf to the middle-bathyal slope. Many of these species belonged to fish families, like Bothidae, Caproidae, Triglidae, and Gobidae, that swim near the bottom to hunt their prey (Colloca et al., 2003; Relini et al., 2011). Specifically, the Peristediidae *Peristedion cataphractum* was identified as an indicator of low litter-high effort. This species uses its distinctive morphological feature, the rostrum, to dig in the mud and extract prey, and probably they benefit from the

resuspension of sediments by trawling to find their prev. Others were properly benthic species, such as the asteroids of the families Luidiidae and Astropectinidae, which exhibited their positive response to trawling disturbance. We hypothesise that trawling supplies them with the discarded bycatch or bait from the fishing gear, which perfectly matches their scavenger habit and carnivorous diet (Groenewold et al., 2000; Juan et al., 2007). Moreover, some sessile species, such as the anthozoan, Epizoanthidae and Funiculinidae, were significantly associated with the sites subjected to low litter - high effort. This can be explained by the characteristic species-specific response that Funiculina quadrangularis (Funiculinidae) has to withstand the fishing pressures (Pierdomenico et al., 2018). This species has a bulb, i.e., peduncle, that enters the sediment up to about 50 cm, allowing the colony to strongly anchor to the seafloor, but maintaining it to bend and lie flat temporarily when fishing gear passes over it (Lauria et al., 2017). As regards the other species Epizoanthus arenaceus (Epizoanthidae), we assume that it withstands the pressures of fishing effort due to its epibiotic habit on motile invertebrates (Reimer, 2023), like molluscs and hermit crabs, which allow it to move away from the impact along with its host species.

Two species were significantly associated with high impacts of seafloor litter and fishing effort: Ascidia spp. (Ascidiidae) and Luidia sarsi (Luidiidae). These species were typically found on the seafloor with high litter accumulation and high fishing effort due to their ecological preferences and behaviour. Ascidia species are frequently found in the biofouling of seabed litter (Ramalhosa et al., 2021; Subías-Baratau et al., 2022). As filter-feeding invertebrates, they tend to inhabit areas with high concentrations of organic matter and detritus, which are often accompanied by litter accumulation. Luidia sarsi, a species of starfish, is commonly found in seabeds with a high abundance of food (Koukouras and Kitsos 2010; Lejeune et al., 2023). Due to the abundance of potential food sources, including waste and decaying material, these species could be attracted to areas with high levels of littering and fishing activity. Based on the idea that non-random distributions of species co-occurrence are the fundamental components of ecological communities (Tulloch et al., 2018), the presence of different species in a given area may be influenced by both environmental requirements and biological interactions, such as competition, mutualism and predation. As a result, certain taxa coexist more frequently than others, while some coexist less frequently than expected by chance. Our results reveal significant non-random associations, indicating potential ecological relationships between taxonomic groups, particularly in varying environmental conditions. The indicator species analysis supports the hypothesis that most species exhibit specific adaptations to litter and trawling effort, based on their substrate interaction and motility patterns.

5. Conclusions

The bathyal zone in the central Tyrrhenian Sea off the Latium coast was affected by multiple environmental features and anthropogenic pressures. Our results showed that the species composition may be related to fishing effort and seafloor litter. The adaptive traits of species have played a significant role in explaining faunal assemblages, primarily related to their environment, mobility, and feeding habits. The accumulation of litter on the seafloor and fishing activity forces these species to exhibit shared and effective responses to disturbances. These responses typically involve the relocation of mobile species, the resistance of some sessile species, or the passive transport of epibiotic species. Conversely, some species thrive in the most affected areas, taking advantage of available food sources.

In conclusion, the results provide a comprehensive overview of the distribution of seafloor litter and fishing pressures, analysing their impacts on marine organisms in the central Tyrrhenian Sea. The methodologies employed, including metabarcoding, statistical analyses, and indicator species analysis, converge to understanding the complex relationships between environmental variables, anthropogenic factors,

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and community composition. This study not only improves our understanding of the response of marine ecosystems to litter accumulation and fishing pressures but also provides essential information for designing effective conservation and management strategies to mitigate their impact on marine organisms.

CRediT authorship contribution statement

Alice Sbrana: Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Resources, Validation, Visualization, Writing – original draft, Writing – review & editing. Giulia Maiello: Data curation, Investigation, Methodology, Visualization, Writing – original draft, Writing – review & editing. Maria Flavia Gravina: Investigation, Resources, Writing – original draft. Davide Cicala: Investigation. Simone Galli: Investigation, Resources, Validation, Writing – original draft. Matteo Stefani: Investigation, Methodology. Tommaso Russo: Conceptualization, Formal analysis, Methodology, Resources, Supervision, Writing – original draft, Writing – review & editing.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data availability

Data will be made available on request.

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.marenvres.2024.106415.

References

- Albonetti, L., Maiello, G., Cariani, A., Carpentieri, P., Ferrari, A., Sbrana, A., Shum, P., Talarico, L., Russo, T., Mariani, S., 2023. DNA metabarcoding of trawling bycatch reveals diversity and distribution patterns of sharks and rays in the central Tyrrhenian Sea. ICES (Int. Counc. Explor. Sea) J. Mar. Sci. 80, 664–674. https://doi. org/10.1093/ICESJMS/FSAD022.
- Angiolillo, M., Fortibuoni, T., 2020. Impacts of marine litter on mediterranean reef systems: from shallow to deep waters. Front. Mar. Sci. 7, 581966 https://doi.org/ 10.3389/FMARS.2020.581966/BIBTEX.
- Angiolillo, M., Lorenzo, B. di, Farcomeni, A., Bo, M., Bavestrello, G., Santangelo, G., Cau, Angelo, Mastascusa, V., Cau, Alessandro, Sacco, F., Canese, S., 2015. Distribution and assessment of marine debris in the deep Tyrrhenian Sea (NW Mediterranean Sea, Italy). Mar. Pollut. Bull. 92, 149–159. https://doi.org/10.1016/ j.marpolbul.2014.12.044.
- Angiolillo, M., Fortibuoni, T., Di Lorenzo, B., Tunesi, L., 2023. First baseline assessment of seafloor litter on Italian coralligenous assemblages (Mediterranean Sea) in accordance with the European Marine Strategy Framework Directive. Mar. Pollut. Bull. 187, 114597 https://doi.org/10.1016/J.MARPOLBUL.2023.114597.
 Ardizzone, G., Belluscio, A., Criscoli, A., 2018. Atlante degli habitat dei fondali marini
- del Lazio.
 Atwood, E.C., Falcieri, F.M., Piehl, S., Bochow, M., Matthies, M., Franke, J., Carniel, S., Sclavo, M., Laforsch, C., Siegert, F., 2019. Coastal accumulation of microplastic particles emitted from the Po River, Northern Italy: Comparing remote sensing and hydrodynamic modelling with in situ sample collections. Mar. Pollut. Bull. 138, 561–574. https://doi.org/10.1016/j.marpolbul.2018.11.045.
- Beauchard, O., Brind'Amour, A., Schratzberger, M., Laffargue, P., Hintzen, N.T., Somerfield, P.J., Piet, G., 2021. A generic approach to develop a trait-based indicator of trawling-induced disturbance. Mar. Ecol. Prog. Ser. 675, 35–52. https://doi.org/ 10.3354/MEPS13840.
- Bohmann, K., Evans, A., Gilbert, M.T.P., Carvalho, G.R., Creer, S., Knapp, M., Yu, D.W., de Bruyn, M., 2014. Environmental DNA for wildlife biology and biodiversity monitoring. Trends Ecol. Evol. 29, 358–367. https://doi.org/10.1016/J. TREE.2014.04.003.
- Boucher, J., Billard, G., 2020. The mediterranean: mare plasticum. International Union for Conservation of Nature and Natural Resources 78.
- Boucher, J., Billard, G., Simeone, E., Sousa, J., 2020. The marine plastic footprint. The marine plastic footprint. https://doi.org/10.2305/iucn.ch.2020.01.en.

- Boyer, F., Mercier, C., Bonin, A., Le Bras, Y., Taberlet, P., Coissac, E., 2016. obitools: a unix-inspired software package for DNA metabarcoding. Mol Ecol Resour 16, 176–182. https://doi.org/10.1111/1755-0998.12428.
- Bozzano, A., Sardaà, F., 2002. Fishery discard consumption rate and scavenging activity in the northwestern Mediterranean Sea. ICES (Int. Counc. Explor. Sea) J. Mar. Sci. 59, 15–28. https://doi.org/10.1006/JMSC.2001.1142.
- Brandt, A., Gutt, J., Hildebrandt, M., Pawlowski, J., Schwendner, J., Soltwedel, T., Thomsen, L., 2016. Cutting the umbilical: new technological perspectives in benthic deep-sea research. Journal of Marine Science and Engineering 2016 4 (36 4), 36. https://doi.org/10.3390/JMSE4020036.
- Campanale, C., Stock, F., Massarelli, C., Kochleus, C., Bagnuolo, G., Reifferscheid, G., Uricchio, V.F., 2020. Microplastics and their possible sources: the example of Ofanto river in southeast Italy. Environ. Pollut. 258, 113284 https://doi.org/10.1016/J. ENVPOL.2019.113284.
- Canals, M., Pham, C.K., Bergmann, M., Gutow, L., Hanke, G., van Sebille, E., Angiolillo, M., Buhl-Mortensen, L., Cau, A., Ioakeimidis, C., Kammann, U., Lundsten, L., Papatheodorou, G., Purser, A., Sanchez-Vidal, A., Schulz, M., Vinci, M., Chiba, S., Galgani, F., Langenkämper, D., Möller, T., Nattkemper, T.W., Ruiz, M., Suikkanen, S., Woodall, L., Fakiris, E., Molina Jack, M.E., Giorgetti, A., 2021. The quest for seafloor macrolitter: a critical review of background knowledge, current methods and future prospects. Environ. Res. Lett. https://doi.org/10.1088/1748-9326/abc6d4.
- Carlucci, R., Bandelj, V., Ricci, P., Capezzuto, F., Sion, L., Maiorano, P., Tursi, A., Solidoro, C., Libralato, S., 2018. Exploring spatio-temporal changes in the demersal and benthopelagic assemblages of the northwestern Ionian Sea (central Mediterranean Sea). Mar. Ecol. Prog. Ser. 598, 1–19. https://doi.org/10.3354/ MEPS12613.
- Cau, A., Sbrana, A., Franceschini, S., Fiorentino, F., Cristina Follesa, M., Galgani, F., Garofalo, G., Gerigny, O., Profeta, A., Rinelli, P., Sbrana, M., Russo, T., 2024. In press. What, where and when: spatial-temporal distribution of macro-litter on the seafloor of the western and central Mediterranean Sea. Environ. Pollut. 342, 123028 https://doi.org/10.2139/ssrn.4540467.
- Cesarini, G., Crosti, R., Secco, S., Gallitelli, L., Scalici, M., 2023. From city to sea: spatiotemporal dynamics of floating macrolitter in the Tiber River. Sci. Total Environ. 857, 159713 https://doi.org/10.1016/J.SCITOTENV.2022.159713.
- Cicala, D., Maiello, G., Fiorentino, F., Garofalo, G., Massi, D., Sbrana, A., Mariani, S., D'Alessandro, S., Stefani, M., Perrodin, L., Russo, T., 2024. Spatial analysis of demersal food webs through integration of eDNA metabarcoding with fishing activities. Front. Mar. Sci. 10, 1209093 https://doi.org/10.3389/ FMARS.2023.1209093/BIBTEX.
- Clark, M.R., Althaus, F., Schlacher, T.A., Williams, A., Bowden, D.A., Rowden, A.A., 2016. The impacts of deep-sea fisheries on benthic communities: a review. ICES (Int. Counc. Explor. Sea) J. Mar. Sci. 73, i51–i69. https://doi.org/10.1093/ICESJMS/ FSV123.
- Coll, M., Piroddi, C., Albouy, C., Ben Rais Lasram, F., Cheung, W.W.L., Christensen, V., Karpouzi, V.S., Guilhaumon, F., Mouillot, D., Paleczny, M., Palomares, M.L., Steenbeek, J., Trujillo, P., Watson, R., Pauly, D., 2012. The Mediterranean Sea under siege: spatial overlap between marine biodiversity, cumulative threats and marine reserves. Global Ecol. Biogeogr. 21, 465–480. https://doi.org/10.1111/j.1466-8238.2011.00697.x.
- Colloca, F., Cardinale, M., Belluscio, A., Ardizzone, G., 2003. Pattern of distribution and diversity of demersal assemblages in the central Mediterranean Sea. Estuar. Coast Shelf Sci. 56, 469–480. https://doi.org/10.1016/S0272-7714(02)00196-8.
- Consoli, P., Sinopoli, M., Deidun, A., Canese, S., Berti, C., Andaloro, F., Romeo, T., 2020. The impact of marine litter from fish aggregation devices on vulnerable marine benthic habitats of the central Mediterranean Sea. Mar. Pollut. Bull. 152, 110928 https://doi.org/10.1016/J.MARPOLBUL.2020.110928.
- Cózar, A., Sanz-Martín, M., Martí, E., González-Gordillo, J.I., Ubeda, B., gálvez, J.Á., Irigoien, X., Duarte, C.M., 2015. Plastic accumulation in the mediterranean sea. PLoS One 10, 1–12. https://doi.org/10.1371/journal.pone.0121762.
- Crosti, R., Arcangeli, A., Campana, I., Paraboschi, M., González-Fernández, D., 2018. 'Down to the river': amount, composition, and economic sector of litter entering the marine compartment, through the Tiber river in the Western Mediterranean Sea. Rendiconti Lincei 29, 859–866. https://doi.org/10.1007/s12210-018-0747-y.
- Davis, N.M., Proctor, D.M., Holmes, S.P., Relman, D.A., Callahan, B.J., 2018. Simple statistical identification and removal of contaminant sequences in marker-gene and metagenomics data. Microbiome 6 (1), 1–14. https://doi.org/10.1186/s40168-018-0605-2.
- De Cáceres, Miquel, Legendre, Pierre, Moretti, Marco, De Cáceres, M., Legendre, P., Moretti, M., 2010. Improving indicator species analysis by combining groups of sites. Oikos 119, 1674–1684. https://doi.org/10.1111/J.1600-0706.2010.18334.X.
- de Carvalho-Souza, G.F., Llope, M., Tinôco, M.S., Medeiros, D.V., Maia-Nogueira, R., Sampaio, C.L.S., 2018. Marine litter disrupts ecological processes in reef systems. Mar. Pollut. Bull. 133, 464–471. https://doi.org/10.1016/J. MARPOLBUL.2018.05.049.
- Deudero, S., Alomar, C., 2015. Mediterranean marine biodiversity under threat: reviewing influence of marine litter on species. Mar. Pollut. Bull. 98, 58–68. https:// doi.org/10.1016/j.marpolbul.2015.07.012.
- Downie, A.L., Piechaud, N., Howell, K., Barrio Froján, C., Sacau, M., Kenny, A., 2021. Reconstructing baselines: use of habitat suitability modelling to predict pre-fishing condition of a Vulnerable Marine Ecosystem. ICES (Int. Counc. Explor. Sea) J. Mar. Sci. 78, 2784–2796. https://doi.org/10.1093/ICESJMS/FSAB154.
- Eigaard, O.R., Bastardie, F., Hintzen, N.T., Buhl-Mortensen, L., Buhl-Mortensen, P., Catarino, R., Dinesen, G.E., Egekvist, J., Fock, H.O., Geitner, K., Gerritsen, H.D., González, M.M., Jonsson, P., Kavadas, S., Laffargue, P., Lundy, M., Gonzalez-Mirelis, G., Nielsen, J.R., Papadopoulou, N., Posen, P.E., Pulcinella, J., Russo, T.,

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Sala, A., Silva, C., Smith, C.J., Vanelslander, B., Rijnsdorp, A.D., 2017. The footprint of bottom trawling in European waters: distribution, intensity, and seabed integrity. ICES (Int. Counc. Explor. Sea) J. Mar. Sci. 74, 847–865. https://doi.org/10.1093/ ICESJMS/FSW194.

- Follesa, M.C., Porcu, C., Cabiddu, S., Mulas, A., Deiana, A.M., Cau, A., 2011. Deep-water fish assemblages in the central-western Mediterranean (south Sardinian deepwaters). J. Appl. Ichthyol. 27, 129–135. https://doi.org/10.1111/J.1439-0426.2010.01567.X.
- Fonseca, V.G., Davison, P.I., Creach, V., Stone, D., Bass, D., Tidbury, H.J., 2023. The application of eDNA for monitoring aquatic non-indigenous species: practical and policy considerations. Diversity 15, 631. https://doi.org/10.3390/D15050631/S1.
- Galgani, F., Pham, C.K., Claro, F., Consoli, P., 2018. Marine animal forests as useful indicators of entanglement by marine litter. Mar. Pollut. Bull. 135, 735–738. https:// doi.org/10.1016/J.MARPOLBUL.2018.08.004.
- Giangrande, A., Lezzi, M., Del Pasqua, M., Pierri, C., Longo, C., Gravina, M.F., 2020. Two cases study of fouling colonization patterns in the Mediterranean Sea in the perspective of integrated aquaculture systems. Aquac Rep 18, 100455. https://doi. org/10.1016/J.AQREP.2020.100455.
- Goulletquer, P., Gros, P., Boeuf, G., Weber, J., 2014. Biodiversity in the marine environment. Biodiversity in the Marine Environment. https://doi.org/10.1007/ 978-94-017-8566-2_1. Springer Netherlands.
- Groenewold, Stefan, Fonds, Mark, Groenewold, S., Fonds, M., 2000. Effects on benthic scavengers of discards and damaged benthos produced by the beam-trawl fishery in the southern North Sea. ICES (Int. Counc. Explor. Sea) J. Mar. Sci. 57, 1395–1406. https://doi.org/10.1006/jmsc.2000.0914.
- Halpern, B.S., Walbridge, S., Selkoe, K.A., Kappel, C.V., Micheli, F., D'Agrosa, C., Bruno, J.F., Casey, K.S., Ebert, C., Fox, H.E., Fujita, R., Heinemann, D., Lenihan, H.S., Madin, E.M.P., Perry, M.T., Selig, E.R., Spalding, M., Steneck, R., Watson, R., 2008. A global map of human impact on marine ecosystems. Science 319, 948–952. https://doi.org/10.1126/SCIENCE.1149345/SUPPL_FILE/HALPERN_SOM.PDF.
- Hardesty, B.D., Polidoro, B., Compa, M., Shim, W.J., Widianarko, B., Wilcox, C., 2019. Multiple approaches to assessing the risk posed by anthropogenic plastic debris. Mar. Pollut. Bull. 141, 188–193. https://doi.org/10.1016/J.MARPOLBUL.2019.02.017.
- Hiddink, J.G., Rijnsdorp, A.D., Piet, G., 2008. Can Bottom Trawling Disturbance Increase Food Production for a Commercial Fish Species?, pp. 1393–1401. https://doi.org/ 10.1139/F08-06465.
- Hiddink, J.G., Jennings, S., Sciberras, M., Szostek, C.L., Hughes, K.M., Ellis, N., Rijnsdorp, A.D., McConnaughey, R.A., Mazor, T., Hilborn, R., Collie, J.S., Pitcher, C. R., Amoroso, R.O., Parma, A.M., Suuronen, P., Kaiser, M.J., 2017. Global analysis of depletion and recovery of seabed biota after bottom trawling disturbance. Proc. Natl. Acad. Sci. U. S. A. 114, 8301–8306. https://doi.org/10.1073/PNAS.1618858114/ SUPPL FILE/PNAS.1618858114.SAPP.PDF.
- Hijmans, R.J., 2019. Introduction to the "Geosphere "Package, pp. 1–19. Version 1 . 5-10.
- Horton, A.A., Barnes, D.K.A., 2020. Microplastic pollution in a rapidly changing world: implications for remote and vulnerable marine ecosystems. Sci. Total Environ. 738, 140349 https://doi.org/10.1016/J.SCITOTENV.2020.140349.
- Iacono, R., Napolitano, E., Palma, M., Sannino, G., Passaro, S., Giorgio, A., Sarra, D., Badalamenti, F., Francocci, F., 2021. The Tyrrhenian Sea circulation: a review of recent work. Sustainability 2021 13. https://doi.org/10.3390/SU13116371, 6371 13, 6371.
- Inghilesi, R., Ottolenghi, L., Orasi, A., Pizzi, C., Bignami, F., Santoleri, R., 2008. Fate of river Tiber discharge investigated through numerical simulation and satellite monitoring. Ocean Sci. 8, 19. https://doi.org/10.5194/os-8-773-2012.
- Israels, A.Z., 1984. Redundancy analysis for qualitative variables. Psychometrika 49, 331–346. https://doi.org/10.1007/BF02306024/METRICS.
- Johnson, A.F., Gorelli, G., Gorelli, G., Hiddink, J.G., Hinz, H., 2015. Effects of bottom trawling on fish foraging and feeding. Proc. Biol. Sci. 282 https://doi.org/10.1098/ RSPB.2014.2336.
- Juan, S. de, Cartes, J.E., Demestre, M., 2007. Effects of commercial trawling activities in the diet of the flat fish Citharus linguatula (Osteichthyes: pleuronectiformes) and the starfish Astropecten irregularis (Echinodermata: asteroidea). J. Exp. Mar. Biol. Ecol. 349, 152–169. https://doi.org/10.1016/J.JEMBE.2007.05.003.
- Juhel, J.B., Marques, V., Utama, R.S., Vimono, I.B., Sugeha, H.Y., Kadarusman, K., Cochet, C., Dejean, T., Hoey, A., Mouillot, D., Hocdé, R., Pouyaud, L., 2022. Estimating the extended and hidden species diversity from environmental DNA in hyper-diverse regions. Ecography 2022, e06299. https://doi.org/10.1111/ ECOG.06299.
- Kaiser, M.J., Spencer, B.E., 1994. Fish scavenging behaviour in recently trawled areas. Mar. Ecol. Prog. Ser. 112, 41–49.
- Kaiser, M.J., Clarke, K.R., Hinz, H., Austen, M.C.V., Somerfield, P.J., Karakassis, I., 2006. Global analysis of response and recovery of benthic biota to fishing. Mar. Ecol. Prog. Ser. 311, 1–14. https://doi.org/10.3354/MEPS311001.
- Kemp, J., López-Baucells, A., Rocha, R., Wangensteen, O.S., Andriatafika, Z., Nair, A., Cabeza, M., 2019. Bats as potential suppressors of multiple agricultural pests: a case study from Madagascar. Agric. Ecosyst. Environ. 269, 88–96. https://doi.org/ 10.1016/j.agee.2018.09.027.
- Kipson, S., Fourt, M., Teixidó, N., Cebrian, E., Casas, E., Ballesteros, E., Zabala, M., Garrabou, J., 2011. Rapid biodiversity assessment and monitoring method for highly diverse benthic communities: a case study of mediterranean coralligenous outcrops. PLoS One 6, e27103. https://doi.org/10.1371/JOURNAL.PONE.0027103.
- Koukouras, A., Kitsos, M.-S., 2010. The biodiversity of the Mediterranean Sea: estimates, patterns, and threats, 2010 Echinoderms. pp.288–296. In: Coll, M., et al. (Eds.), PLoS ONE, p. 36, 5(8).

- Kühn, S., Bravo Rebolledo, E.L., van Franeker, J.A., 2015. Deleterious effects of litter on marine life. In: Marine Anthropogenic Litter. Springer International Publishing, pp. 75–116. https://doi.org/10.1007/978-3-319-16510-3_4.
- Lauria, V., Garofalo, G., Fiorentino, F., Massi, D., Milisenda, G., Piraino, S., Russo, T., Gristina, M., 2017. Species distribution models of two critically endangered deep-sea octocorals reveal fishing impacts on vulnerable marine ecosystems in central Mediterranean Sea. Scientific Reports 2017 7 (1 7), 1–14. https://doi.org/10.1038/ s41598-017-08386-z.
- Lejeune, B., Marcout, A., Kopp, D., Morandeau, F., Mehault, S., Mouchet, M.A., 2023. Assessing discard consumption dynamic in shallow coastal environment using underwater video. Fish. Res. 260, 106587 https://doi.org/10.1016/J. FISHRES.2022.106587.
- Mačić, V., Đorđević, N., Đurović, M., Petović, S., Russo, T., 2022. Improving knowledge of Funiculina quadrangularis and vulnerable marine ecosystems in the south Adriatic. Mediterr. Mar. Sci. 23, 805–816. https://doi.org/10.12681/mms.29898.
- Mahé, F., Rognes, T., Quince, C., de Vargas, C., Dunthorn, M., 2015. Swarmv 2: highlyscalable and high-resolution amplicon clustering. PeerJ 2015 (12), 1–12. https:// doi.org/10.7717/peerj.1420.
- Maiello, G., Talarico, L., Carpentieri, P., De Angelis, F., Franceschini, S., Harper, L.R., Neave, E.F., Rickards, O., Sbrana, A., Shum, P., Veltre, V., Mariani, S., Russo, T., 2022. Little samplers, big fleet: eDNA metabarcoding from commercial trawlers enhances ocean monitoring. Fish. Res. 249, 106259 https://doi.org/10.1016/J. FISHRES.2022.106259.
- Maiello, G., Talarico, L., Brodie, C., Carpentieri, P., Sbrana, A., Shum, P., Mariani, S., Russo, T., 2023. Net gain: low-cost, trawl-associated eDNA samplers' upscale ecological assessment of marine demersal communities. Environmental DNA 1–13. https://doi.org/10.1002/EDN3.389.
- Mallet, D., Pelletier, D., 2014. Underwater video techniques for observing coastal marine biodiversity: a review of sixty years of publications (1952–2012). Fish. Res. 154, 44–62. https://doi.org/10.1016/J.FISHRES.2014.01.019.
- Mancini, E., Miccoli, A., Piazzolla, D., Saraceni, P.R., Lezzi, M., Tiralongo, F., Bonifazi, A., Picchietti, S., Marcelli, M., 2021. Macrozoobenthic fauna associated with benthic marine litter (Northern Tyrrhenian Sea, Italy) and first report of two bryozoan species in Italian waters. Reg Stud Mar Sci 47, 101912. https://doi.org/ 10.1016/J.RSMA.2021.101912.
- Mangano, M.C., Bottari, T., Caridi, F., Porporato, E.M.D., Rinelli, P., Spanò, N., Johnson, M., Sarà, G., 2017. The effectiveness of fish feeding behaviour in mirroring trawling-induced patterns. Mar. Environ. Res. 131, 195–204. https://doi.org/ 10.1016/J.MARENYRES.2017.09.004.
- MEDITS working group, 2012. MEDITS. International Bottom Trawl Survey in the Mediterranean. Instruction Manual. Version 6. MEDITS-Handbook. Revision n.6, April 2012. MEDITS Working group. https://archimer.ifremer.fr/doc/00117/22783.
- Noce, T.La, Pagnotta, R., Pettine, M., Puddu, A., 2013. Coastal Water Pollution Around Tiber River Mouth – a Case Study, Mediterranean Coastal Pollution. IAWPR/ Pergamon Press Ltd. https://doi.org/10.1016/b978-0-08-026058-7.50020-0.
- Oberle, F.K.J., Storlazzi, C.D., Hanebuth, T.J.J., 2016. What a drag: quantifying the global impact of chronic bottom trawling on continental shelf sediment. J. Mar. Syst. 159, 109–119. https://doi.org/10.1016/J.JMARSYS.2015.12.007.
- Oksanen, J., et al.. vegan: Community Ecology Package. R package version 2.5-3. http s://CRAN.R-project.org/package=vegan.
- Pante, E., Bouhet, B.S., 2015. Making and using bathymetric maps in R with marmap. Pennino, M.G., Conesa, D., López-Quílez, A., Muñoz, F., Fernández, A., Bellido, J.M.,
- 2016. Fishery-dependent and -independent data lead to consistent estimations of essential habitats. ICES (Int. Counc. Explor. Sea) J. Mar. Sci. 73, 2302–2310. https://doi.org/10.1093/ICESJMS/FSW062.
- Pérès, J.M., Picard, J., 1964. Nouveau manuel de Bionomie benthique de la Mediterranee. Extrait du Recueil des Travaux de la Station Marine d'Endoume 31 (47), 137.
- Pierdomenico, M., Russo, T., Ambroso, S., Gori, A., Martorelli, E., D'Andrea, L., Gili, J. M., Chiocci, F.L., 2018. Effects of trawling activity on the bamboo-coral isidella elongata and the sea pen Funiculina quadrangularis along the gioia canyon (western mediterranean, southern Tyrrhenian Sea). Prog. Oceanogr. 169, 214–226. https:// doi.org/10.1016/J.POCEAN.2018.02.019.
- Pinochet, J., Urbina, M.A., Lagos, M.E., 2020. Marine invertebrate larvae love plastics: habitat selection and settlement on artificial substrates. Environ. Pollut. 257, 113571 https://doi.org/10.1016/J.ENVPOL.2019.113571.
- Poeta, G., Conti, L., Malavasi, M., Battisti, C., Acosta, A.T.R., 2016. Beach litter occurrence in sandy littorals: the potential role of urban areas, rivers and beach users in central Italy. Estuar. Coast Shelf Sci. 181, 231–237. https://doi.org/10.1016/j. ecss.2016.08.041.
- Póvoa, A.A., Skinner, L.F., de Araújo, F.V., 2021. Fouling organisms in marine litter (rafting on abiogenic substrates): a global review of literature. Mar. Pollut. Bull. 166, 112189 https://doi.org/10.1016/J.MARPOLBUL.2021.112189.
- Pusceddu, A., Bianchelli, S., Martín, J., Puig, P., Palanques, A., Masqué, P., Danovaro, R., 2014. Chronic and intensive bottom trawling impairs deep-sea biodiversity and ecosystem functioning. Proc. Natl. Acad. Sci. U. S. A. 111, 8861–8866. https://doi. org/10.1073/PNAS.1405454111/SUPPL_FILE/PNAS.201405454SI.PDF.
- Ramalhosa, P., Gestoso, I., Rocha, R.M., Lambert, G., Canning-Clode, J., 2021. Ascidian biodiversity in the shallow waters of the Madeira Archipelago: fouling studies on artificial substrates and new records. Reg Stud Mar Sci 43, 101672. https://doi.org/ 10.1016/J.RSMA.2021.101672.
- Rech, S., Macaya-Caquilpán, V., Pantoja, J.F., Rivadeneira, M.M., Jofre Madariaga, D., Thiel, M., 2014. Rivers as a source of marine litter - a study from the SE Pacific. Mar. Pollut. Bull. 82, 66–75. https://doi.org/10.1016/j.marpolbul.2014.03.019.

- Rech, S., Salmina, S., Borrell Pichs, Y.J., García-Vazquez, E., 2018. Dispersal of alien invasive species on anthropogenic litter from European mariculture areas. Mar. Pollut. Bull. 131, 10–16. https://doi.org/10.1016/J.MARPOLBUL.2018.03.038.
- Reimer, J.D., 2023. World List of Zoantharia. Epizoanthus Arenaceus (Delle Chiaje, 1841). Accessed through: World Register of Marine Species at: https://www.marinespecies.org/aphia.php?p=taxdetails&id=101023 on 2023-11-02.
- Relini, G., Sartor, P., Reale, B., Orsi Relini, L., Mannini, A., De Ranieri, S., Ardizzone, G. D., Belluscio, A., Serena, F., 2011. In: Cataudella, S., Spagnolo, M. (Eds.), GSA 9 44–51.
- Russo, T., D'Andrea, L., Parisi, A., Cataudella, S., 2014. VMSbase: an R-package for VMS and logbook data management and analysis in fisheries ecology. PLoS One 9, e100195. https://doi.org/10.1371/JOURNAL.PONE.0100195.
- Russo, T., D'Andrea, L., Parisi, A., Martinelli, M., Belardinelli, A., Boccoli, F., Cignini, I., Tordoni, M., Cataudella, S., 2016. Assessing the fishing footprint using data integrated from different tracking devices: issues and opportunities. Ecol. Indicat. 69, 818–827. https://doi.org/10.1016/J.ECOLIND.2016.04.043.
- Sandra, M., Devriese, L.I., Booth, A.M., De Witte, B., Everaert, G., Gago, J., Galgani, F., Langedock, K., Lusher, A., Maes, T., Pirlet, H., Russell, J., Pham, C.K., 2023. A systematic review of state-of-the-art technologies for monitoring plastic seafloor litter. J. Ocean Eng. Sci. https://doi.org/10.1016/J.JOES.2023.07.004.
- Sbrana, A., Valente, T., Scacco, U., Bianchi, J., Silvestri, C., Palazzo, L., de Lucia, G.A., Valerani, C., Ardizzone, G., Matiddi, M., 2020. Spatial variability and influence of biological parameters on microplastic ingestion by Boops boops (L.) along the Italian coasts (Western Mediterranean Sea). Environ. Pollut. 263, 114429 https://doi.org/ 10.1016/j.envpol.2020.114429.
- Sbrana, A., Cau, A., Cicala, D., Franceschini, S., Giarrizzo, T., Gravina, M.F., Ligas, A., Maiello, G., Matiddi, M., Parisi, A., Sartor, P., Sbrana, M., Scacco, U., Valente, T., Viva, C., Russo, T., 2022. Ask the shark: blackmouth catshark (Galeus melastomus) as a sentinel of plastic waste on the seabed. Mar. Biol. 169, 1–17. https://doi.org/ 10.1007/S00227-022-04084-1/FIGURES/6.

- Siegenthaler, A., Wangensteen, O.S., Soto, A.Z., Benvenuto, C., Corrigan, L., Mariani, S., 2019. Metabarcoding of shrimp stomach content: harnessing a natural sampler for fish biodiversity monitoring. Molecular Ecology Resources 19 (1), 206–220. https:// doi.org/10.1111/1755-0998.12956.
- Subías-Baratau, A., Sanchez-Vidal, A., Di Martino, E., Figuerola, B., 2022. Marine biofouling organisms on beached, buoyant and benthic plastic debris in the Catalan Sea. Mar. Pollut. Bull. 175, 113405 https://doi.org/10.1016/J. MARPOLBUL.2022.113405.
- Thomsen, P.F., Willerslev, E., 2015. Environmental DNA an emerging tool in conservation for monitoring past and present biodiversity. Biol. Conserv. 183, 4–18. https://doi.org/10.1016/J.BIOCON.2014.11.019.
- Tiano, J.C., van der Reijden, K.J., O'Flynn, S., Beauchard, O., van der Ree, S., van der Wees, J., Ysebaert, T., Soetaert, K., 2020. Experimental bottom trawling finds resilience in large-bodied infauna but vulnerability for epifauna and juveniles in the Frisian Front. Mar. Environ. Res. 159, 104964 https://doi.org/10.1016/J. MARENVRES.2020.104964.
- Tulloch, A.I.T., Chadès, I., Lindenmayer, D.B., 2018. Species co-occurrence analysis predicts management outcomes for multiple threats. Nature Ecology & Evolution 2018 2 (3 2), 465–474. https://doi.org/10.1038/s41559-017-0457-3.
- Wangensteen, O.S., Palacín, C., Guardiola, M., Turon, X., 2018. DNA metabarcoding of littoral hardbottom communities: high diversity and database gaps revealed by two molecular markers. PeerJ 2018 (5), e4705. https://doi.org/10.7717/peerj.470.
- Yoccoz, N.G., 2018. The Future of Environmental DNA in Ecology the Future of Environmental DNA in Ecology. https://doi.org/10.1111/j.1365-294X.2012.05505.
- Zettler, E.R., Mincer, T.J., Amaral-Zettler, L.A., 2013. Life in the "plastisphere": microbial communities on plastic marine debris. Environ. Sci. Technol. 47, 7137–7146. https://doi.org/10.1021/es401288x.