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McCard, M, McCard, N, Coughlan, NE, South, J, Kregting, L and Dick, JTA (2024) Functional response metrics explain and predict high but differing ecological impacts of juvenile and adult lionfish. Royal Society Open Science. 11.

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Cite this article: McCard M, McCard N, Coughlan NE, South J, Kregting L, Dick JTA. 2024 Functional response metrics explain and predict high but differing ecological impacts of juvenile and adult lionfish. *R. Soc. Open Sci.* **11**: 240855. https://doi.org/10.1098/rsos.240855

Received: 31 May 2024 Accepted: 22 July 2024

Subject Category:

Ecology, conservation, and global change biology

Subject Areas:

behaviour, ecology, environmental science

Keywords:

invasive, functional response ratio, Pterois volitans, impact prediction, Nephrops norvegicus

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Functional response metrics explain and predict high but differing ecological impacts of juvenile and adult lionfish

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Recent accumulation of evidence across taxa indicates that the ecological impacts of invasive alien species are predictable from their functional response (FR; e.g. the maximum feeding rate) and functional response ratio (FRR; the FR attack rate divided by handling time). Here, we experimentally derive these metrics to predict the ecological impacts of both juvenile and adult lionfish (Pterois volitans), one of the world's most damaging invaders, across representative and likely future prey types. Potentially prey-population destabilizing Type II FRs were exhibited by both life stages of lionfish towards four prey species: Artemia salina, Gammarus oceanicus, Palaemonetes varians and Nephrops norvegicus. FR magnitudes revealed ontogenetic shifts in lionfish impacts where juvenile lionfish displayed similar if not higher consumption rates than adult lionfish towards prey, apart from N. norvegicus, where adult consumption rate was considerably higher. Additionally, lionfish FRR values were very substantially higher than mean FRR values across known damaging invasive taxa. Thus, both life stages of lionfish are predicted to contribute to differing but high ecological impacts across prey communities, including commercially important species. With lionfish invasion ranges currently expanding across multiple regions globally, efforts to reduce lionfish numbers and population size structure, with provision of prey refugia through habitat complexity, might curtail their impacts. Nevertheless, the

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present study indicates that management programmes to support early detection and complete eradication of lionfish individuals when discovered in new regions are advised.

1. Introduction

Invasive species are predicted to continue to increase in number and severity of ecological impacts [1,2]. However, species-specific impacts are difficult to predict with surety and can vary considerably among invaded ecosystems due to a plethora of biotic and abiotic interactions (e.g. [3,4]). Also, invaders with highly similar morphological and behavioural traits, and even congeneric species, may differentially impact invaded regions (e.g. [5]). Thus, we require robust metrics that are both explanatory and predictive of ecological impact across habitats and trophic and taxonomic groups [6,7].

The red lionfish, *Pterois volitans*, is currently considered one of the most invasive and ecologically harmful marine fish worldwide [8], with populations spreading across the western Atlantic Ocean, Caribbean Sea [9] and Mediterranean Sea [10]. Both P. volitans and its sister species, Pterois miles, are also currently establishing themselves across the Mediterranean. Due to the significant impact of P. miles and the extensive invasion by P. volitans, we use P. volitans in this study as a proxy when referring to lionfish. As generalist and opportunistic predators, lionfish show dietary plasticity for prey items [11] and demonstrate clear negative impacts on some prey species [12]. Although adult lionfish are primarily piscivorous, they have also been recorded to predate on a variety of invertebrates (e.g. [13]). Most lionfish dietary studies have been completed on adult individuals due to ease of capture compared to juveniles (i.e. <15 cm; see [14-21]), and therefore juvenile lionfish predation is data limited; however, information available suggests a predominance of small crustaceans and larval fish [17,22,23]. As a consequence of culling programmes, selective removal of large individuals tends to occur in invaded areas, with the remaining population being largely composed of juveniles and smaller-sized individuals which are cryptic, fast moving and avoid culling activity by spear fishers [24,25]. Accordingly, a predictive assessment of lionfish invasion impacts thus requires dual examination of juvenile and adult feeding rates on representative and future likely prey communities.

Functional response (FR) metrics (described as Type I, Type II and Type III) have been used successfully to assess the ecological impact of current, future and emerging invasive species, through consideration of resource consumption (i.e. the amount of a resource utilized, such as prey) as a function of resource density [3,4,6]. The Type I response is a linear response, characteristic of filter feeders which are not constrained by handling times [26]; density-dependent Type II responses are characterized by a plateauing of consumption as prey density increases [27]; Type III responses are represented by a sigmoidal curve due to a reduction of consumption at low prey densities, often providing low-density refugia to rare prey species and sometimes related to learned predator avoidance behaviour in prey populations [28,29]. Deriving the type of FR, plus the attack rates, handling times and maximum feeding rates, has been successful across taxa in explaining and predicting invader impacts [4,6,30]. However, while high attack rates and low handling times predict high impact [7], predictions based on either parameter alone can be contradictory (e.g. when handling times are low, but attack rates are also low [31]). To resolve this, Cuthbert et al. [31] proposed a composite metric, the functional response ratio (FRR), that is, attack rate divided by handling time. The FRR has a clear pattern of high values predicting high ecological impact, where the worst invaders have an FRR mean of 83.36. This benchmarking allows comparison of newly derived FRRs, and hence likely ecological impact, in studies such as the present regarding lionfish (see [31]). FRR, however, is applicable for Type II and Type III comparison as Type I FR is devoid of a handling time.

We thus assessed the predatory impacts of both juvenile and adult *P. volitans* on four prey species by employing the FR metrics approach. The selected prey species were used to mimic a host of similar prey found across the invaded and future ranges of lionfish, namely the brine shrimp (*Artemia salina*), marine gammarid (*Gammarus oceanicus*), dwarf white shrimp (*Palaemonetes varians*) and finally Dublin Bay prawn (*Nephrops norvegicus*), as the lionfish range is currently expanding across the Mediterranean (albeit predominantly *P. miles*) into areas where commercially and ecologically valuable *N. norvegicus* are located [32]. Given that morphological and metabolic changes may affect diet, leading to variations in preferred prey types, as well as possible restricted ability of juvenile lionfish to predate larger prey due to the size relationship between predator and prey. We hypothesized that juvenile and adult lionfish will display FR metrics consistent with high ecological impact, with ontogenetic shifts in these metrics due to predator/prey size.

2. Material and methods

2.1. Animal collection and maintenance

Experiments were undertaken at Queen's University Marine Laboratory (QML), Portaferry, UK, between January and October 2019. Juvenile P. volitan lionfish (n = 8) and adult P. volitan lionfish (n = 8) were obtained from Seahorse Aquarium, Dublin. Juveniles had a total body length (mean \pm s.e.) of 102.80 ± 3.18 mm, with a pectoral fin diameter of 57.89 ± 4.80 mm, as measured across the widest point when elongated. Adults measured 305.51 ± 3.73 mm in length with a pectoral fin diameter of 265.51 ± 6.37 mm. Juveniles were kept in a holding tank (W: 32 cm × L: 152 cm × H: 45 cm, 218 l) with external filtration containing UV- and sand-filtered recirculated Strangford Lough seawater. Adult holding tanks separately employed the same filtration set-up, while two adults were housed per tank (W: 82 cm × L: 227 cm × H: 61 cm, 1130 l). Up to 10% of the tank water was changed daily, monitoring temperature, and conditions including pH and ammonium. Seawater was maintained at $25.0 \pm 1.0^{\circ}$ C using an aquarium heater under a natural light regime. Lionfish were maintained daily ad libitum on frozen anchovy to avoid predator learning behaviour of the experimental prey species. Feeding experiments were conducted within glass tanks (juveniles: W: 33 cm × L: 46 cm × H: 30 cm, 45 l; adults: W: 51 cm × L: 132 cm × H: 38 cm, 250 l) maintained at 25.0 ± 1.0 °C to ensure lionfish welfare. Experimental tanks were scaled to reflect the difference between juvenile and adult lionfish when pectoral fins were fully elongated during feeding trails, where adults were approximately five times the size of juvenile lionfish. All fish were acclimated in the experimental arenas for a 30 min period immediately prior to experimentation.

Brine shrimp (*A. salina*), marine gammarid (*G. oceanicus*), dwarf white shrimp (*P. varians*) and Dublin Bay prawn (*N. norvegicus*) were used as live prey. *Artemia salina* were obtained from Seahorse Aquariums, Dublin, Ireland, while *G. oceanicus* and *P. varians* were obtained from Grosvenor Tropicals, Lisburn, UK. *Nephrops norvegicus* were caught in fishing grounds off the western Irish Sea, by the FV Fulmar, an 11.33 m trawler using a SELTRA in single-rig configuration. Once samples were landed, they were immediately brought to QML. *Artemia salina, G. oceanicus* and *P. varians* were maintained under identical conditions to hose for the predators in separate holding tanks (W: 15 cm × L: 20 cm × H: 18 cm, 10 l), whereas *N. norvegicus* were housed in a dark outdoor holding tank (H: 94 cm × W: 142 cm × L: 211 cm, 2800 l), which included tunnels for refuge. All prey species were acclimated to lionfish maintenance temperature of $25.0 \pm 1.0^{\circ}$ C before being introduced to the testing tank. *Artemia salina, G. oceanicus*, and *P. varians* were kept at $22.0 \pm 1.0^{\circ}$ C, which was then raised to $25.0 \pm 1.0^{\circ}$ C 60 min before introduction. In contrast, *N. norvegicus* was initially kept at $18.0 \pm 1.0^{\circ}$ C and gradually exposed to a temperature change in the testing tank over 60 min until reaching $25.0 \pm 1.0^{\circ}$ C. Intraspecific prey size was standardized throughout all trials. Total length (mean \pm s.e.): *A. salina*, (6.2 \pm 0.8 mm); *G. oceanicus*, (10.7 \pm 0.9 mm); *P. varians*, (11.3 \pm 0.5 mm); and total carapace length for *N. norvegicus*, (20.1 \pm 3.1 mm).

Selected prey species mimic those that are commonly found in lionfish stomachs across their invaded and potential future regions and have been used in previous lionfish FR experiments using similar laboratory set-ups to the present study [13,17,30,33–36]. The present study represented the first comparative assessment of juvenile and adult lionfish, as well as the first assessment of lionfish impact on *N. norvegicus*, which are a valuable commercial fishery species across the United Kingdom and European Union that will likely be threatened by the expanding lionfish invasion [37–39]. Furthermore, *N. norvegicus* can be used as a proxy for juveniles of other large crustacean species such as the Caribbean spiny lobster (*Panulirus argus*), the European lobster (*Homarus gammarus*) and anomurans (squat lobsters).

2.2. Functional response procedure

Each prey species was separately supplied at 15 densities (2, 4, 6, 8, 12, 16, 20, 25, 30, 35, 40, 45, 50, 55, 60; experiment replication n = 8 per density for each of the four prey species) in a randomized pattern of both prey species and densities. This was achieved with the re-use of the available lionfish in the following manner: following the addition of the allotted prey to the experimental tanks that contained an individual predator, FR experiments were initiated. Lionfish were allowed to feed for 3 h before being removed for enumeration of prey consumed. In a one month period, there were eight experiment days, with all lionfish being used on each experiment day (adult n = 8; juvenile n = 8). This facilitated a 3 day recovery period between experiment days. Re-use of individuals was essential due to the limited number of lionfish available, hence the recovery period (see [40]). The entire experiment



Figure 1. FRs showing the order of highest to lowest consumption of prey with both juvenile (*a*) and adult (*b*) *P. volitans* towards all prey types: *A. salina* (red), *G. oceanicus* (orange), *P. varians* (blue) and *N. norvegicus* (green). Shaded areas are bootstrapped (*n* = 2000) 95% CIs.

was conducted over a 10 month period, with all lionfish being systematically exposed to all prey items at all densities, in a randomly allocated order. Controls consisted of one replicate of each prey type across all densities in the absence of lionfish predators.

2.3. Statistical analyses

Statistical analyses were undertaken using the 'frair' package in R [41]. Logistic regression was used to derive FR types based on analyses of proportional prey consumption across prey densities, with 'prey density' included as a continuous variable [42]. To model the FRs, data were fit using Rogers' random predator equation, as prey were not replaced once consumed [43]:

$$N_{\rm e} = N_0 (1 - \exp(a(N_{\rm e}h - T))).$$
(2.1)

Wherein N_e represents the amount of prey consumed, N_0 is initial prey density, *a* is the attack rate parameter, *h* is the handling time and *T* is the total time available. Data of prey eaten were then non-parametrically bootstrapped (*n* = 2000) to produce 95% confidence intervals (CIs) using initial maximum-likelihood estimates of *a* and *h*. The handling time parameter was used to determine maximum feeding rates (1/*h*) of lionfish across prey groups. Additionally, the FRR was calculated for each prey species using the parameter estimates of *a* and *h* derived from the FR curve from equation (2.1):

$$FRR = a/h.$$
(2.2)

3. Results

Across all control groups (i.e. no predator) for all prey species, survival of the prey exceeded 99% in the absence of lionfish; therefore, all mortality of prey in experimental groups was assumed to be due to predation by lionfish.

3.1. Functional responses

First-order terms were significantly negative as per Juliano [44], indicating Type II FRs by all lionfish towards all prey species (table 1; figure 1).

3.1.1. Juvenile lionfish

The attack rates and handling times of juvenile lionfish towards the four prey species resulted in FR magnitudes greatest for *A. salina*, lowest for *N. norvegicus* and intermediate for *G. oceanicus* and *P. varians* (table 1; figure 1). FRR values for juvenile lionfish were greatest for *A. salina* (564.29), followed

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Table 1. First-order terms from logistic regression of prey consumed, alongside rounded initial and bootstrapped (*n* = 2000; 95% Cls) FR parameters (attack rates, handling times, maximum feeding rates and FRBs), of juvenile and adult *P. volitans* with different prey types.

predator	prey	first-order term, <i>p</i>	FR type	a, 95% Cl	<i>h</i> , 95% Cl	maximum feeding rate (1/h)	FRR (<i>a/h</i>)
juvenile <i>P. volitans</i>	A. salina	-0.09, <0.001	_	7.11, 5.47–9.43	0.01, 0.01–0.02	79.37	564.29
juvenile <i>P. volitans</i>	G. oceanicus	-0.06, <0.001	=	4.44, 3.18–6.14	0.03, 0.02–0.03	35.71	158.54
juvenile <i>P. volitans</i>	P. varians	-0.08, <0.001	=	10.15, 7.22–14.55	0.03, 0.03–0.03	34.48	350.14
juvenile <i>P. volitans</i>	N. norvegicus	-0.06, <0.001	=	6.45, 3.85–11.84	0.11, 0.10–0.13	8.93	57.62
adult <i>P. volitans</i>	A. salina	-0.07, <0.001	=	6.42, 5.19–7.94	0.03, 0.03–0.03	37.04	237.74
adult <i>P. volitans</i>	G. oceanicus	-0.06, <0.001	=	5.27, 3.61–7.83	0.04, 0.04–0.05	23.26	122.61
adult <i>P. volitans</i>	P. varians	-0.07, <0.001	=	6.86, 5.11–10.07	0.03, 0.03–0.04	30.30	207.76
adult <i>P. volitans</i>	N. norvegicus	-0.08, <0.001	=	7.15, 5.60–9.27	0.02, 0.02–0.03	43.48	310.91

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by *P. varians* (350.14), *G. oceanicus* (158.54), and *N. norvegicus* (57.62: table 1). The first three FRR values are markedly higher than the mean FRR value of 83.36 that was found for highly damaging invaders across taxa (see [31]), by factors of 7, 4.5 and 2, predicting high ecological impacts of juvenile lionfish on prey with similar traits to *A. salina*, *P. varians* and *G. oceanicus*.

3.1.2. Adult lionfish

The attack rates and handling times of adult lionfish result in quite different FR magnitudes compared with juveniles, with adult FRs ordered greatest for *N. norvegicus* followed by *A. salina*, *P. varians* and *G. oceanicus* (table 1; figure 1). FRR values for adult lionfish were greatest for *N. norvegicus* (310.91), then *A. salina* (237.74), *P. varians* (207.76) and *G. oceanicus* (122.61; table 1). All these FRR values are substantially higher than the mean FRR value of 83.36 found by Cuthbert *et al.* [31] by factors of 3.8, 3, 2.6 and 1.5, predicting high ecological impacts of adult lionfish on such crustacean prey.

4. Discussion

The observed consumption patterns and FR metrics predict that both juvenile and adult lionfish can exert damaging impact on a range of crustacean prey where larger individuals show size-dependent preferences for larger prey items compared to the gape-limited juveniles [13,45,46]. These findings are in line with *in situ* stomach content analyses (e.g. [15,17,23]).

The Type II FRs by both juveniles and adults suggest potential destabilizing effects on invertebrate prey species populations, although lionfish may switch prey preference for species that are more abundant in the environment, which provides a low-density prey refuge, and complex habitat structure can mitigate impacts [29,36]. These mitigating drivers may explain the difference in field impact between the Bahamas (high) and Belize (low) [47,48]. Habitat structure can offer refuges for prey, reducing predator search success at low prey densities, leading to sigmoid Type III FRs [49,50]. In contrast, the absence of habitat structure and the effects of arena size often result in Type II FRs [51]. Our comparative laboratory study used standardized conditions for all organisms without additional habitat complexity. Despite this, FR analyses and related impact assessment metrics are highly predictive of *per capita* impacts and simple laboratory settings of FR analyses are predictive of actual field impacts [6,7,52]. While lionfish use the entire water column to hunt, benthic and pelagic prey can attempt escape in both the upward and downward direction, which adds additional complexity to *in situ* foraging by lionfish in nominal two-dimensional benthic and three-dimensional pelagic environments.

Further predictive confidence of high lionfish impact is their remarkable FRR values, that were up to seven times higher than the mean FRR across known damaging invasive taxa [31]. The benchmark FRR values of Cuthbert *et al.* [31] indicate that, overall, mean FRRs of 83.36 typify high-impact invaders, since attack rates are high and handling times are low. The FRR values found here for lionfish were distinctively high, indicating an ecologically damaging ability of lionfish to find, subdue, consume and digest prey, which is clearly commensurate with actual field impacts of lionfish.

Lionfish pose a threat to commercial crustacean fisheries and can disturb benthic food webs in both current and potential invasion areas [53,54]. Lionfish have been observed at varying depths in regions they have infiltrated: surpassing 100 m in the Bahamas [55], reaching depths of 250 m in Honduras [56] and descending as far as 304 m in Bermuda [56] where dense lionfish populations have been identified at specific locations, particularly at or below 60 m, in select Bermuda sites [57,58]. The potential for high consumption rates of N. norvegicus by adult lionfish is a cause for concern considering that lionfish range expansions will overlap with commercially important fishing grounds. While juveniles also fed on N. norvegicus, they did so in smaller numbers, which may be due to limitations in gape and their inability to efficiently predate the hard exoskeleton [46,59]. Nephrops norvegicus are generally a deep-water species which reside in mud-flat burrows at depths of 20-800 m; given that lionfish have been found at mesophotic depths this suggests that deep-water populations could be sustained on N. norvegicus [60]. Furthermore, high FRR values on P. varians indicates high potential for consumption of functional analogue species such as Pandalus montagui and Cragon crangon, both of which form a high percentage of diet for native fish predators [61]. The full and pernicious impacts of lionfish invasion may be further revealed if prey depletion leads to trophic cascades through loss of prey for native predators [62,63].

While *A. salina* and *P. varians* are generally pelagic with high mobility [64,65], *G. oceanicus* and *N. norvegicus* tend to be epibenthic with a relatively lower rate of mobility [66–68]. It appears that both lionfish life stages can exploit pelagic and epibenthic prey [69,70], with utilization of prey items being linked to body type [15,71], size [69,72] and digestibility [73,74] rather than mobility [74] for juvenile lionfish, while adults appeared to better utilize the largest prey and least mobile prey. Previous studies have shown lionfish may specialize on small prey species that are solitary, nocturnal and bottom dwelling [34,69]; however, the adult lionfish in this study showed a reduced consumption of small epibenthic species (*G. oceanicus*) compared with the larger *N. norvegicus* and the pelagic species. This may indicate some difficulty in consumption within the tank confines due to spatial limitations on manoeuvrability.

The current geographical spread and increased growth in abundance of lionfish in the Atlantic have made eradication impossible [75]. While the data in this study were collected in a laboratory setting, with prey being presented in isolation from other prey resources, this work provides a basis for estimates of consumption rates of both juvenile and adult lionfish on representative and future prey types. Further, current management strategies for lionfish populations rely on the removal of adults [75,76]; however, our data indicate that juvenile lionfish can have a greater impact on native prey species than adult lionfish. This increased predation pressure reduces prey availability for native predators. Therefore, populations composed of juveniles and adults will have wide ranging impacts across multiple prey species, potentially driven by ontogenetic shifts in functional morphology, since juveniles have traits associated with a mechanical advantage during prey capture, whereas adult morphology is more associated with locomotion and sustained swimming but a lower suction velocity [46]. Accordingly, management strategies will need to be developed to efficiently control all life stages, rather than adults alone. Finally, while culling might reduce lionfish impacts through reduced numbers of individual predators, imaginative strategies to alter FR metrics might also be employed; for example Type III FRs and lowered FRRs may result from increased habitat complexity such as artificial reefs. The present study demonstrates both juvenile and adult lionfish can have a considerable impact on prey populations if allowed to establish and persist in regions at risk of invasion. Ultimately, early detection and eradication remain the best, if least utilized, strategies for invasive species management.

Ethics. Ethical approval for work with lionfish was granted by the School of Biological Sciences Animal Research Ethics Committee, Queen's University Belfast.

Data accessibility. The data are available from the Dryad Digital Repository [77].

Declaration of Al use. We have not used AI-assisted technologies in creating this article.

Authors' contributions. M.M.: conceptualization, formal analysis, investigation, methodology, project administration, resources, visualization, writing—original draft, writing—review and editing; N.M.: methodology, writing—review and editing; N.E.C.: formal analysis, methodology, writing—review and editing; J.S.: formal analysis, supervision, writing—review and editing; J.T.A.D.: supervision, writing—review and editing.

All authors gave final approval for publication and agreed to be held accountable for the work performed therein.

Conflict of interest declaration. We declare we have no competing interests.

Funding. M.M. is part funded from G & M Williams fund, Queen's Marine Laboratory, Portaferry.

Acknowledgements. We extend thanks to Emma Healey for technical support. N.E.C. gratefully acknowledges support from the Irish Research Council (GOIPD/2022/861).

References

- Seebens H et al. 2021 Projecting the continental accumulation of alien species through to 2050. Glob. Chang. Biol. 27, 970–982. (doi:10.1111/ qcb.15333)
- IPBES. 2022 Summary for policymakers of the global assessment report on biodiversity and ecosystem services of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services, pp. 86–101.
- Dickey JWE et al. 2021 Breathing space: deoxygenation of aquatic environments can drive differential ecological impacts across biological invasion stages. Biol. Invasions 23, 2831–2847. (doi:10.1007/s10530-021-02542-3)
- Coughlan NE, Dickey JWE, Dick JTA, Médoc V, McCard M, Lacroix G, Fiorini S, Millot A, Cuthbert RN. 2022 When worlds collide: invader-driven benthic habitat complexity alters predatory impacts of invasive and native predatory fishes. *Sci. Total Environ.* 843, 156876. (doi:10.1016/j. scitotenv.2022.156876)
- Števove B, Kováč V. 2013 Do invasive bighead goby Neogobius kessleri and round goby N. melanostomus (Teleostei, Gobiidae) compete for food? Knowl. Manage. Aquat. Ecosyst. 410, 08. (doi:10.1051/kmae/2013064)

- 6. Dick JTA *et al.* 2014 Advancing impact prediction and hypothesis testing in invasion ecology using a comparative functional response approach. *Biol. Invasions* **16**, 735–753. (doi:10.1007/s10530-013-0550-8)
- 7. Dick JTA et al. 2017 Functional responses can unify invasion ecology. Biol. Invasions 19, 1667–1672. (doi:10.1007/s10530-016-1355-3)
- 8. Goodbody-Gringley G, Eddy C, Pitt J, Chequer A, Smith S. 2019 Ecological drivers of invasive lionfish (pterois volitans and pterois miles) distribution across mesophotic reefs in bermuda. *Front. Mar. Sci.* **6**, 1–12. (doi:10.3389/fmars.2019.00258)
- 9. Schofield P. 2009 Geographic extent and chronology of the invasion of non-native lionfish (*Pterois volitans* [Linnaeus 1758] and *P. miles* [Bennett 1828]) in the Western North Atlantic and Caribbean sea. *Aquatic Invasions* **4**, 473–479. (doi:10.3391/ai.2009.4.3.5)
- Bariche M, Torres M, Azzurro E. 2013 The presence of the invasive lionfish *Pterois miles* in the Mediterranean Sea. *Med. Mar. Sci.* 14, 292–294. (doi:10.12681/mms.428)
- 11. Côté IM, Green SJ, Hixon MA. 2013 Predatory fish invaders: insights from Indo-Pacific lionfish in the western Atlantic and Caribbean. *Biol. Conserv.* **164**, 50–61. (doi:10.1016/j.biocon.2013.04.014)
- Del Río L, Navarro-Martínez ZM, Ruiz-Abierno A, Chevalier-Monteagudo PP, Angulo-Valdes JA, Rodriguez-Viera L. 2022 Feeding ecology of invasive lionfish in the Punta Frances MPA, Cuba: insight into morphological features, diet and management. *PeerJ* 10, e14250. (doi:10.7717/ peerj.14250)
- 13. Morris JAJ, Akins JL. 2009 Feeding ecology of invasive lionfish (*Pterois volitans*) in the Bahamian archipelago. *Environ. Biol. Fishes* **86**, 389–398. (doi:10.1007/s10641-009-9538-8)
- 14. Dahl KA, Patterson WF. 2014 Habitat-specific density and diet of rapidly expanding invasive red lionfish, *Pterois volitans*, populations in the northern Gulf of Mexico. *PLoS One* **9**, e105852. (doi:10.1371/journal.pone.0105852)
- 15. Muñoz RC, Currin CA, Whitfield PE. 2011 Diet of invasive lionfish on hard bottom reefs of the southeast USA: insights from stomach contents and stable isotopes. *Mar. Ecol. Prog. Ser.* **432**, 181–193. (doi:10.3354/meps09154)
- 16. Mizrahi M, Chapman J, Gough C, Humber F, Anderson L. 2017 Management implications of the influence of biological variability of invasive lionfish diet in Belize. *Manag. Biol. Invasions* **8**, 61–70. (doi:10.3391/mbi.2017.8.1.06)
- 17. Dahl KA, Patterson WF, Robertson A, Ortmann AC. 2017 DNA barcoding significantly improves resolution of invasive lionfish diet in the northern Gulf of Mexico. *Biol. Invasions* **19**, 1917–1933. (doi:10.1007/s10530-017-1407-3)
- Zannaki K, Corsini-Foka M, Kampouris TE, Batjakas IE. 2019 First results on the diet of the invasive *Pterois miles* (Actinopterygii: Scorpaeniformes: Scorpaenidae) in the Hellenic waters. *Acta Ichthyol. Piscat.* 49, 311–317. (doi:10.3750/AIEP/02616)
- Acero PA, Bustos-Montes D, Pabón Quintero P, Polo-Silva CJ, Sanjuan Muñoz A. 2019 Feeding habits of *Pterois volitans*: a real threat to Caribbean coral reef biodiversity. In *Impacts of invasive species on coastal environments* (eds C Makowski, CW Finkl), pp. 269–314. UK: Springer. (doi:10. 1007/978-3-319-91382-7_8)
- 20. Batjakas IE, Evangelopoulos A, Giannou M, Pappou S, Papanikola E, Atsikvasi M, Poursanidis D, Gubili C. 2023 Lionfish diet composition at three study sites in the Aegean Sea: an invasive generalist? *Fishes* **8**, 314. (doi:10.3390/fishes8060314)
- 21. Samourdani A, Ketsilis-Rinis V, Koutsidi M, Lazaris A, Peristeraki P, Tzanatos E. 2024 Ecology and behaviour of the invasive lionfish *Pterois miles* colonizing coastal areas of the central Mediterranean. *Estuar. Coast. Shelf Sci.* **303**, 108796. (doi:10.1016/j.ecss.2024.108796)
- Cure K, Benkwitt CE, Kindinger TL, Pickering EA, Pusack TJ, McIlwain JL, Hixon MA. 2012 Comparative behavior of red lionfish *Pterois volitans* on native Pacific versus invaded Atlantic coral reefs. *Mar. Ecol. Prog. Ser.* 467, 181–192. (doi:10.3354/meps09942)
- 23. Eddy C, Pitt J, Morris J Jr, Smith S, Goodbody-Gringley G, Bernal D. 2016 Diet of invasive lionfish (*Pterois volitans* and *P. miles*) in Bermuda. *Mar. Ecol. Prog. Ser.* **558**, 193–206. (doi:10.3354/meps11838)
- 24. Morris JAJ, Sullivan CV, Govoni JJ. 2011 Oogenesis and spawn formation in the invasive lionfish, *Pterois miles* and *Pterois volitans*. Sci. Mar. **75**, 147–154. (doi:10.3989/scimar.2011.75n1147)
- Frazer TK, Jacoby CA, Edwards MA, Barry SC, Manfrino CM. 2012 Coping with the lionfish invasion: can targeted removals yield beneficial effects? *Rev. Fish. Sci.* 20, 185–191. (doi:10.1080/10641262.2012.700655)
- Hoxha T, Crookes S, Lejeusne C, Dick JTA, Chang X, Bouchemousse S, Cuthbert RN, MacIsaac HJ. 2018 Comparative feeding rates of native and invasive ascidians. *Mar. Pollut. Bull.* 135, 1067–1071. (doi:10.1016/j.marpolbul.2018.08.039)
- 27. Hassell MP. 1978 The dynamics of arthropod predator-prey systems. Princeton, NJ: Princeton University Press.
- Colton TF. 1987 Extending functional response models to include a second prey type: an experimental test. *Ecology* 68, 900–912. (doi:10.2307/1938361)
- 29. Kalinkat G, Rall BC, Uiterwaal SF, Uszko W. 2023 Empirical evidence of type III functional responses and why it remains rare. *Front. Ecol. Evol.* **11**, 1033818. (doi:10.3389/fevo.2023.1033818)
- South J, Dick JTA, McCard M, Barrios-O'Neill D, Anton A. 2017 Predicting predatory impact of juvenile invasive lionfish (*Pterois volitans*) on a crustacean prey using functional response analysis: effects of temperature, habitat complexity and light regimes. *Environ. Biol. Fishes* 100, 1155–1165. (doi:10.1007/s10641-017-0633-y)
- 31. Cuthbert RN, Dickey JWE, Coughlan NE, Joyce PWS, Dick JTA. 2019 The functional response ratio (FRR): advancing comparative metrics for predicting the ecological impacts of invasive alien species. *Biol. Invasions* **21**, 2543–2547. (doi:10.1007/s10530-019-02002-z)
- 32. Lolas A, Vafidis D. 2021 Population dynamics, fishery, and exploitation status of Norway lobster (*Nephrops norvegicus*) in eastern Mediterranean. *Water* **13**, 289. (doi:10.3390/w13030289)
- Ortiz M et al. 2015 Control strategy scenarios for the alien lionfish Pterois volitans in Chinchorro Bank (Mexican Caribbean): based on semiquantitative loop analysis. PLoS One 10, e0130261. (doi:10.1371/journal.pone.0130261)

- Layman CA, Allgeier JE. 2012 Characterizing trophic ecology of generalist consumers: a case study of the invasive lionfish in the Bahamas. *Mar. Ecol. Prog. Ser.* 448, 131–141. (doi:10.3354/meps09511)
- 35. Layman CA, Jud ZR, Nichols P. 2014 Lionfish alter benthic invertebrate assemblages in patch habitats of a subtropical estuary. *Mar. Biol.* **161**, 2179–2182. (doi:10.1007/s00227-014-2491-x)
- McCard M, South J, Cuthbert RN, Dickey JWE, McCard N, Dick JTA. 2021 Pushing the switch: functional responses and prey switching by invasive lionfish may mediate their ecological impact. *Biol. Invasions* 23, 2019–2032. (doi:10.1007/s10530-021-02487-7)
- Smith CJ, Papadopoulou KN. 2003 Burrow density and stock size fluctuations of Nephrops norvegicus in a semi-enclosed bay. ICES J. Mar. Sci. 60, 798–805. (doi:10.1016/S1054-3139(03)00065-1)
- Bell MC, Redant F, Tuck I. 2006 Nephrops species. In Lobsters: biology, management, aquaculture and fisheries (ed. BF Phillips), pp. 412–462. Oxford, UK: Wiley-Blackwell. (doi:10.1002/9780470995969.ch13)
- Nikolic N, Diméet J, Fifas S, Salaün M, Ravard D, Fauconnet L, Rochet M-J. 2015 Efficacy of selective devices in reducing discards in the Nephrops trawl fishery in the Bay of Biscay. ICES J. Mar. Sci. 72, 1869–1881. (doi:10.1093/icesjms/fsv036)
- 40. Alexander ME, Dick JTA, Weyl OLF, Robinson TB, Richardson DM. 2014 Existing and emerging high impact invasive species are characterized by higher functional responses than natives. *Biol. Lett.* **10**, 20130946. (doi:10.1098/rsbl.2013.0946)
- 41. R Core Development Team. 2018 R: a language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing.
- Pritchard DW, Paterson RA, Bovy HC, Barrios-O'Neill D. 2017 Frair: an R package for fitting and comparing consumer functional responses. *Methods Ecol. Evol.* 8, 1528–1534. (doi:10.1111/2041-210X.12784)
- 43. Rogers D. 1972 Random search and insect population models. J. Anim. Ecol. 41, 369. (doi:10.2307/3474)
- Juliano SA. 2001 Nonlinear curve fitting predation and functional response curves. In *Design and analysis of ecological experiments* (eds S Scheiner, J Gurevitch), pp. 178–196. Oxford, UK: Oxford University Press. (doi:10.1093/oso/9780195131871.003.0010)
- Mihalitsis M, Bellwood DR. 2017 A morphological and functional basis for maximum prey size in piscivorous fishes. *PLoS One* 12, e0184679. (doi: 10.1371/journal.pone.0184679)
- Rojas-Vélez S, Tavera J, Acero P. A. 2024 Is lionfish ontogenetic dietary change mediated by morphological functionality? *Hydrobiologia* 851, 447–456. (doi:10.1007/s10750-023-05254-y)
- Green SJ, Akins JL, Maljković A, Côté IM. 2012 Invasive lionfish drive Atlantic coral reef fish declines. PLoS One 7, e32596. (doi:10.1371/journal. pone.0032596)
- Hackerott S, Valdivia A, Cox CE, Silbiger NJ, Bruno JF. 2017 Invasive lionfish had no measurable effect on prey fish community structure across the Belizean barrier reef. *PeerJ* 5, e3270. (doi:10.7717/peerj.3270)
- Rickers S, Scheu S. 2005 Cannibalism in Pardosa palustris (Araneae, Lycosidae): effects of alternative prey, habitat structure, and density. Basic Appl. Ecol. 6, 471–478. (doi:10.1016/j.baae.2005.02.007)
- Birkhofer K, Gavish-Regev E, Endlweber K, Lubin YD, von Berg K, Wise DH, Scheu S. 2008 Cursorial spiders retard initial aphid population growth at low densities in winter wheat. *Bull. Entomol. Res.* 98, 249–255. (doi:10.1017/S0007485308006019)
- Vucic-Pestic O, Birkhofer K, Rall BC, Scheu S, Brose U. 2010 Habitat structure and prey aggregation determine the functional response in a soil predator-prey interaction. *Pedobiologia* 53, 307–312. (doi:10.1016/j.pedobi.2010.02.003)
- 52. Dickey JWE *et al.* 2018 Assessing the relative potential ecological impacts and invasion risks of emerging and future invasive alien species. *NeoBiota* **40**, 1–24. (doi:10.3897/neobiota.40.28519)
- Ballew N, Bacheler N, Kellison G et al. 2016 Invasive lionfish reduce native fish abundance on a regional scale. Sci. Rep. 6, 32169. (doi:10.1038/ srep32169)
- Savva I et al. 2020 They are here to stay: the biology and ecology of lionfish (*Pterois miles*) in the Mediterranean Sea. J. Fish Biol. 97, 148–162. (doi:10.1111/jfb.14340)
- Lesser MP, Slattery M. 2011 Phase shift to algal dominated communities at mesophotic depths associated with lionfish (*Pterois volitans*) invasion on a Bahamian coral reef. *Biol. Invasions* 13, 1855–1868. (doi:10.1007/s10530-011-0005-z)
- Gress E, Andradi-Brown DA, Woodall L, Schofield PJ, Stanley K, Rogers AD. 2017 Lionfish (*Pterois* spp.) invade the upper-bathyal zone in the western Atlantic. *PeerJ* 5, e3683. (doi:10.7717/peerj.3683)
- 57. Eddy C. 2016 An investigation of the biology and ecology of the invasive lionfish (*Pterois volitans* and *P. miles*) to explore their ecological impact and inform management in Bermuda's marine ecosystem. PhD dissertation, University of Massachusetts, Dartmouth, MA, USA.
- Andradi-Brown DA *et al.* 2017 Depth-dependent effects of culling: do mesophotic lionfish populations undermine current management? *R. Soc.* Open Sci. 4, 170027. (doi:10.1098/rsos.170027)
- Björnsson B, Álvaro Dongala Dombaxe M. 2004 Quality of Nephrops as food for Atlantic cod (Gadus morhua L.) with possible implications for fisheries management. ICES J. Mar. Sci. 61, 983–991. (doi:10.1016/j.icesjms.2004.06.017)
- 60. Andradi-Brown DA *et al.* 2017 Large-scale invasion of western Atlantic mesophotic reefs by lionfish potentially undermines culling-based management. *Biol. Invasions* **19**, 939–954. (doi:10.1007/s10530-016-1358-0)
- 61. Myers RA, Worm B. 2003 Rapid worldwide depletion of predatory fish communities. Nature 423, 280–283. (doi:10.1038/nature01610)
- 62. Moll RJ, Redilla KM, Mudumba T, Muneza AB, Gray SM, Abade L, Hayward MW, Millspaugh JJ, Montgomery RA. 2017 The many faces of fear: a synthesis of the methodological variation in characterizing predation risk. J. Anim. Ecol. **86**, 749–765. (doi:10.1111/1365-2656.12680)
- Palmer MS, Gaynor KM, Becker JA, Abraham JO, Mumma MA, Pringle RM. 2022 Dynamic landscapes of fear: understanding spatiotemporal risk. Trends Ecol. Evol. 37, 911–925. (doi:10.1016/j.tree.2022.06.007)

- 64. Ward-Booth K, Reiss M. 1988 Artemia salina: an easily cultured invertebrate ideally suited for ecological studies. J. Biol. Educ. 22, 247–251. (doi: 10.1080/00219266.1988.9654995)
- 65. Harding L, Jackson AL, Payne N. 2023 Energetic costs increase with faster heating in an aquatic ectotherm. *Conserv. Physiol.* **11**, coad042. (doi: 10.1093/conphys/coad042)
- Maynou F, Sarda` F. 2001 Influence of environmental factors on commercial trawl catches of Nephrops norvegicus (L.). ICES J. Mar. Sci. 58, 1318– 1325. (doi:10.1006/jmsc.2001.1091)
- 67. Navarro-Barranco C, Irazabal A, Moreira J. 2020 Demersal amphipod migrations: spatial patterns in marine shallow waters. J. Mar. Biol. Ass. 100, 239–249. (doi:10.1017/S002531542000003X)
- Coughlan M, Guerrini M, Creane S, O'Shea M, Ward SL, Van Landeghem KJJ, Murphy J, Doherty P. 2021 A new seabed mobility index for the Irish Sea: modelling seabed shear stress and classifying sediment mobilisation to help predict erosion, deposition, and sediment distribution. *Cont. Shelf Res.* 229, 104574. (doi:10.1016/j.csr.2021.104574)
- 69. Green SJ, Côté IM. 2014 Trait-based diet selection: prey behaviour and morphology predict vulnerability to predation in reef fish communities. J. Anim. Ecol. 83, 1451–1460. (doi:10.1111/1365-2656.12250)
- 70. Green SJ, Dilley ER, Benkwitt CE, Davis ACD, Ingeman KE, Kindinger TL, Tuttle LJ, Hixon MA. 2019 Trait-mediated foraging drives patterns of selective predation by native and invasive coral-reef fishes. *Ecosphere* **10**. (doi:10.1002/ecs2.2752)
- 71. McCleery C. 2011 A comparative study of the feeding ecology of invasive lionfish (*Pterois volitans*) in the Caribbean. J. Mar. Sci. 9, 38–43.
- 72. Kindinger TL, Anderson ER. 2016 Preferences of invasive lionfish and native grouper between congeneric prey fishes. *Mar. Ecol. Prog. Ser.* **558**, 247–253. (doi:10.3354/meps11833)
- 73. Steell SC, Van Leeuwen TE, Brownscombe JW, Cooke SJ, Eliason EJ. 2019 An appetite for invasion: digestive physiology, thermal performance and food intake in lionfish (*Pterois* spp.). J. Exp. Biol. **222**, jeb209437. (doi:10.1242/jeb.209437)
- 74. Trehern R, Garg A, Bigelow W, Hauptman H, Brooks A, Hawkes L, Van Leeuwen T. 2020 Low salinity negatively affects metabolic rate, food consumption, digestion and growth in invasive lionfish *Pterois* spp. *Mar. Ecol. Prog. Ser.* **644**, 157–171. (doi:10.3354/meps13340)
- 75. Johnston MW, Purkis SJ. 2015 A coordinated and sustained international strategy is required to turn the tide on the Atlantic lionfish invasion. *Mar. Ecol. Prog. Ser.* **533**, 219–235. (doi:10.3354/meps11399)
- 76. Barbour AB, Allen MS, Frazer TK, Sherman KD. 2011 Evaluating the potential efficacy of invasive lionfish (*Pterois volitans*) removals. *PLoS One* **6**, e19666. (doi:10.1371/journal.pone.0019666)
- 77. McCard M, McCard N, Coughlan NE, Kregting L, Dick JTA. 2023 Functional response metrics explain and predict high but differing ecological impacts of juvenile and adult lionfish. Dryad Digital Repository. (doi:10.5061/dryad.h70rxwdpq)