

1 **Unveiling the critical role of opportunistic bacteria in post-wildfire**
2 **phosphorus cycling in tropical peat forests**

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19 **Abstract**

20 Although wildfires are known to significantly influence soil phosphorus (P)
21 dynamics in forest ecosystems, the mechanisms through which bacterial subgroups
22 with distinct life-history strategies respond to wildfire disturbances and regulate
23 inorganic phosphorus (Pi) remain poorly understood. To address this knowledge gap,
24 we collected soil samples from wildfire-affected tropical peat forests and oil palm
25 agroforests to investigate how bacterial subgroups regulate phosphorus cycling. Results
26 showed that although wildfire led to an overall decline in bacterial communities,
27 including reductions in diversity and absolute abundance, soil Pi content increased
28 significantly. This increase may be linked to shifts in bacterial life-history strategies.
29 Following the classification of the community into opportunistic (r-strategists),
30 sensitive, and tolerant (K-strategists) taxa, we observed that wildfire enhanced soil
31 moisture, which in turn significantly increased the relative abundance of opportunistic
32 taxa. Following wildfire, these r-strategists showed higher speciation rates, greater
33 dormancy potential, stronger dispersal limitation (favoring local habitat colonization),
34 and greater number of keystone species in networks. These traits provided opportunistic
35 taxa with a competitive advantage, enabling rapid proliferation after wildfires.
36 Additionally, they enhanced the expression of organic P-mineralization genes,
37 particularly *phnA*, *phnW*, and *phoN*, accelerating the release and accumulation of Pi.
38 This study for the first time revealed how bacteria with different life-history strategies
39 influence P cycling, particularly highlighting the role of opportunistic taxa. This finding
40 offers new insights into post-fire P dynamics driven by microbial communities in

41 tropical forests and peatlands.

42 **Keywords:** Wildfire; Tropical peatlands; Bacterial community; Life-history strategies;

43 Opportunistic taxa; Organic phosphorus mineralization; Inorganic phosphate

44 **1. Introduction**

45 Phosphorus is a critical nutrient that supports plant and microbial growth in
46 tropical ecosystems. It fundamental in life processes such as photosynthesis and
47 respiration (Lambers, 2022), and strongly affects biomass accumulation and
48 community assembly. However, P cycling in forest soil is increasingly threatened.
49 Climate change has driven a rise in the frequency of extreme wildfires, which have
50 increased by 2.2 times over the past two decades (Cunningham et al., 2024). The global
51 annual burned forest area is projected to increase by 29% by the end of the 21st century
52 (Senande-Rivera et al., 2022). Wildfires not only disturb surface soil ecosystems
53 directly but also contribute to climate change by releasing particulate aerosols and
54 greenhouse gases. In the tropical regions of Southeast Asia, many lowland, coastal
55 forest soils are composed of organic-rich peat, which is highly flammable. Fires on
56 Southeast Asian peatlands have resulted in significant greenhouse gas emissions (c. 0.1
57 Gt C eq yr⁻¹) (Hooijer et al., 2010). Yet there remains a significant gap in literature
58 related to processes which occur post fire. Specifically, fires may also trigger
59 biogeochemical decoupling of nutrient cycles (Shen et al., 2025). This characteristic
60 heightens the vulnerability of P cycling to wildfire disturbances. Understanding of
61 microbial assemblage structure and its role in mediating nutrient availability within
62 tropical lowland peatlands, remains virtually unexplored (Too et al., 2018). Further, the
63 impact of fire on microbial communities in tropical peats is completely unknown and
64 represents a major gap in our understanding of post-fire ecosystem function.
65 Investigating post-fire P dynamics is, therefore, crucial for assessing the stability of P

66 cycling in impacted tropical peatlands. This is a relevant phase in considerations of soil
67 processes under global change.

68 Bacterial communities play a central regulatory role in soil P cycling through key
69 biological processes such as solubilization, mineralization, and transport (Chu and Ma,
70 2024). However, forest wildfires have been shown to exert highly detrimental effects
71 on soil microbiota. One meta-analysis study revealed that wildfires reduced soil
72 bacterial biomass, abundance, and diversity to 90%, 96%, and 93% of the pre-fire levels,
73 respectively (Pressler et al., 2018). The primary cause is that high temperatures directly
74 inactivate enzymes and damage cellular structures, resulting in widespread mortality.
75 Additionally, wildfires diminish the complexity of soil bacterial co-occurrence
76 networks (Li et al., 2024; Yang et al., 2024). Such effects may indicate potentially
77 negative impacts on Pi enrichment, given that the diversity and network complexity of
78 bacterial communities are strongly associated with the multifunctionality of soil
79 ecosystems (Delgado-Baquerizo et al., 2016; Zhai et al., 2024; Xiao et al., 2025).
80 Notably, some studies suggest that soil Pi may increase following wildfires—a
81 phenomenon potentially more widespread than previously recognized (Shen et al., 2024;
82 Wu et al., 2024; Zhou et al., 2025). This increase has been commonly attributed to heat-
83 induced mineralization and short-term input from ash deposition. However, this
84 explanation overlooks the potential and sustained role of bacterial communities. For
85 example, wildfires may stimulate the expression of organic P-mineralization genes and
86 the secretion of phosphatases by bacteria, thereby facilitating the conversion of organic
87 P into plant-available Pi (Shen et al., 2025). As such, given the global importance of

88 peatlands, it is critical that the functional alteration of these systems associated with
89 anthropogenic disturbance, be fully understood. Nevertheless, the post-fire dynamics
90 of soil Pi in tropical peatlands remain unclear, and the underlying microbial
91 mechanisms require further investigation.

92 Soil moisture is recognized as a key driver of global soil biodiversity and
93 ecosystem functioning (Zhang et al., 2023) and is a fundamental driver of the
94 functionality of peatlands; with water table heights and associated moisture content of
95 peat soils controlling both carbon storage and sequestration rates (Evers et al., 2017).
96 Alterations in soil moisture, under limiting conditions especially, can profoundly
97 influence various ecological processes mediated by soil biota. Wildfires are intricately
98 connected to the hydrological cycle. In the immediate aftermath of a wildfire, surface
99 soil is often covered by a layer of highly porous ash that is rich in organic and inorganic
100 matter (Abrantes et al., 2025), the loss of vegetation reduces evapotranspiration
101 (Belongia et al., 2023), thereby increasing the proportion of precipitation that infiltrates
102 into the soil. However, in tropical regions, the greater exposure of the peat surface to
103 sunlight and heat, often increases evaporation rates and reducing moisture content
104 (Harrison et al., 2024). Fire can also change the physiochemical properties of peat,
105 promoting water repellence of burnt peat (Doerr et al., 2000) via alteration of soil
106 particle size (Balfour, 2015), increase subsidence and associated compaction, and an
107 overall reduced water storage capacity and infiltration rate (Wösten et al., 2006). In
108 contrast, peat fires can burn to significant depth, often removing >50cm of height from
109 the soil surface, exposing deeper reserves of recalcitrant peat material to oxidation.

110 During these events, the soil surface drops, driving it closer to the water table height,
111 thereby enhancing the soil moisture content of the peat post fire (Lupascu et al., 2020).
112 Consequently, the decline in soil moisture negatively influences P solubilization and
113 diffusion; while excess soil moisture (caused by short-term spikes or poor drainage)
114 can reduce soil aeration and create anaerobic conditions. Anaerobiosis alters bacterial
115 community composition and metabolic activity, thereby indirectly influencing P cycling
116 rates. Nevertheless, despite these observations, the extent to which wildfire-induced
117 alterations in soil moisture affect bacterial community structure and function, as well
118 as their subsequent roles in P cycling, remains insufficiently understood.

119 Understanding how soil bacteria respond to wildfire and how soil moisture
120 influences this response, can enhance predictions of degraded tropical peat recovery
121 and long-term impacts of both degraded and intact peatlands under global climate
122 change. Bacterial taxa can be classified according to life-history strategies to assess
123 their resistance to wildfire and capacity for post-disturbance recovery. These strategies
124 represent suites of functional traits that have evolved in response to environmental
125 disturbances and resource competition (e.g., resource allocation, reproduction, survival
126 mechanisms, and stress tolerance), and confer competitive advantages under specific
127 environmental conditions (Malik et al., 2020). For example, in highly variable moisture
128 conditions, communities may become enriched with stress-tolerant taxa showing
129 metabolic plasticity. In contrast, in environments with consistently high moisture,
130 bacteria tend to optimize resource utilization either through enhanced decomposition of
131 organic matter or by reducing metabolic rates to maintain competitive advantages

132 within their niches (Evans and Wallenstein, 2014). These contrasting adaptive strategies
133 often give rise to three distinct ecological groups within bacterial communities:
134 opportunistic, sensitive, and tolerant taxa (Coyle et al., 2017). As r-strategists,
135 opportunistic taxa flourish in response to resource pulses and demonstrate high growth
136 rates and metabolic versatility. Sensitive taxa depend on stable environments, exhibit
137 slow growth, and possess low tolerance to stress, which makes them potentially poorly
138 adapted to habitat disturbances. Tolerant taxa, typically with high biomass and
139 persistence, act as K-strategists and play critical role in maintaining community
140 stability and functional continuity during recovery (Rojas-Tirado et al., 2017; Lavallee
141 et al., 2024). Given their distinct functional traits, these three taxonomic groups may
142 assume different roles in regulating soil Pi levels. A key scientific question is whether
143 Pi enrichment after wildfires is primarily driven by a specific bacterial strategy. Do
144 these taxa possess sufficient functional advantages to sustain phosphorus cycling under
145 reduced community diversity? Investigating the role of bacteria with specific life-
146 history strategies in P cycling is important for predicting ecosystem recovery, and the
147 availability of Pi post fires. Specifically, understanding these changes has significance
148 towards vegetation recovery and land management decisions.

149 Here, we focus on tropical forest peatlands in Malaysia, where frequent wildfires
150 occur due to drainage of the peatlands and the reduced associated soil moisture,
151 seasonal drought, lightning strikes, and agricultural development (driving wildfires to
152 spread from slash and burn activities). We hypothesize that wildfire disturbance exerts
153 selective pressure on bacterial communities by altering soil moisture and nutrient

154 balance, preferentially enriching r-strategist taxa (e.g., opportunistic taxa), rather than
155 uniformly affecting the entire community structure. These bacteria, due to their high
156 growth rates and enhanced expression of organic P-mineralization genes, may drive the
157 accumulation of soil Pi after wildfire. The objectives of this study are: 1) to investigate
158 the effects of wildfire on soil physicochemical properties, bacterial taxa with different
159 life-history strategies, and P-cycling functional genes; and 2) to clarify the potential
160 mechanisms by which bacterial taxa with distinct life-history strategies influence soil
161 Pi levels post wildfire disturbance.

162 **2. Materials and methods**

163 **2.1. Study site and sampling**

164 The study sites were located in Southern Selangor, Malaysia (2.69 – 2.88 °N,
165 101.51 – 101.64 °E), within mosaic landscapes of remnant peat swamp forests
166 surrounding the Kuala Langat north and south peat swamp forest reserves. The
167 peatlands are characterized by acidic peat deposits, and the peat soils sit on marine clay.
168 The area experiences a tropical climate, with a mean annual temperature of 28°C and
169 precipitation exceeding 2,000 mm. Seasonal rainfall patterns are distinct: the dry season
170 (May – September) has an average monthly rainfall of 146 mm, while the wet seasons
171 (October – April) receive approximately 295 mm monthly. Agricultural expansion has
172 caused land conversion into four distinct ecosystems: young oil palm, mature oil palm,
173 secondary forests, and drained forests. The secondary forests represent relic primary
174 vegetation with mixed historical logging, though perimeter drainage and illegal
175 encroachment has meant none of the forest sites were pristine.

176 Wildfires in the area typically occur during the dry season. We conducted soil
177 sampling as soon as possible after the fire was extinguished during August 2020. During
178 fieldwork, we selected two land cover classes representing the dominant land uses of
179 the areas: secondary forest (F) and the mature oil palm (OP) plantation. At each site,
180 we established both burned and control plots (denoted: BF/BOP for burned, and
181 CF/COP for control, respectively). However the overwhelming differences between
182 post fire and unburnt, compared to variation within land cover class meant that soil
183 samples from F and OP land classes were combined into 'Control' and 'Burnt' for
184 downstream analyses. Sampling was conducted at 12 locations (see Fig. S1). At each
185 location, soil samples were collected at three depths (0 – 10 cm, 20 – 30 cm, and 40 –
186 50 cm) along the soil cores, covering the main oxic and anoxic zones. Four replicates
187 were taken per location, resulting in a total of 144 soil samples. All samples were kept
188 refrigerated and promptly transported to the laboratory for further analysis.

189 **2.2. Physicochemical analysis**

190 All samples were lyophilized. Soil moisture content was calculated from the
191 weight difference before and after drying. The dried soils were mixed with ultrapure
192 water, shaken thoroughly, and centrifuged at 6,800 g for 10 min. The resulting
193 supernatant was used for pH measurement. The supernatant was filtered through a 0.22
194 μm membrane before analyzing anion concentrations (PO_4^{3-} , NH_4^+ , and NO_3^-) using
195 anion chromatography (ICS-600, Thermo Fisher Scientific, Waltham, MA), PO_4^{3-} was
196 used as the indicator of soil Pi content (Li et al., 2025). Total nitrogen (TN), total
197 organic carbon (TOC), and C/N ratios were determined using an elemental analyzer

198 (Vario MACRO cube, Elementar, Germany). Dissolved organic matter (DOM) in the
199 supernatant was quantified with a three-dimensional fluorescence spectrometer (Fluoro
200 Max 4, HORIBA Jobin Yvon, Edison, NJ). Its composition was qualitatively assessed
201 using fluorescence indices: fluorescence index (FI), biological index (BIX), and
202 humification index (HIX) (Wang et al., 2017).

203 **2.3. DNA extraction, PCR amplification, and sequencing**

204 Total DNA was extracted from the soil samples using the DNeasy PowerSoil Kit
205 (12888-100; Qiagen, Hilden, Germany) following the manufacturer's protocol. The
206 V3–V4 region of the bacterial 16S rRNA gene was amplified using primers 338F/806R,
207 and sequenced on an Illumina MiSeq PE250 platform (Chen et al., 2025). Absolute
208 gene copy numbers of the 16S rRNA gene were quantified via real-time quantitative
209 PCR (qPCR) using the CFX96 system (Bio-Rad, USA). Each 20 μL qPCR reaction
210 contained 1 μL of template DNA, 10 μL of TB GreenTM Premix Ex Taq II (RR820A;
211 TaKaRa, Kyoto), 0.5 μL each of forward and reverse primers, 3.2 μL of 25 mM MgCl_2 ,
212 and 4.8 μL of RNase-free water. Thermocycling conditions were as follows: initial
213 denaturation at 95 °C for 30 s; 40 cycles of 95 °C for 5 s and 60 °C for 30 s; followed
214 by melting curve analysis to assess amplification specificity. Primer sequences and
215 detailed methods were provided in previous research (Tettamanti Boshier et al., 2020).

216 **2.4. Data processing**

217 Raw sequence data were processed in QIIME 2. Primer sequences were trimmed
218 from paired-end reads using Cutadapt, and DADA2 was used for quality filtering,

219 denoising, chimera removal, and generation of amplicon sequence variants (ASVs)
220 along with their representative sequences. Taxonomic assignment was performed
221 against the SILVA 138 16S rRNA gene reference database. To standardize sequencing
222 depth across samples, all datasets were rarefied to 16,335 sequences per sample.
223 Functional prediction of the bacterial communities was inferred using PICRUSt2, with
224 predicted gene families annotated against the Kyoto Encyclopedia of Genes and
225 Genomes (KEGG) database (Douglas et al., 2020). Specifically, we focused on the
226 relative abundances of genes associated with dormancy and P cycling.

227 **2.5. Statistical analysis**

228 We assessed differences between control and burned soils using t-tests. STAMP
229 analysis was employed to identify between-group differences in bacterial abundance at
230 the phylum level and genus level. To capture divergent bacterial life-history strategies
231 in response to wildfire, ASVs were categorized into three subgroups based on their
232 relative abundance shifts between burned and control soils: (1) opportunistic taxa,
233 defined as ASVs whose relative abundances significantly increased after burning; (2)
234 sensitive taxa, defined as ASVs whose abundances significantly decreased in burned
235 soils; and (3) tolerant taxa, defined as ASVs that showed no significant change in
236 abundance between treatments. Spearman's rank correlation was applied to evaluate the
237 relationships between the variables, and to evaluate the influence of environmental
238 drivers on bacterial subgroups. To further investigate the community assembly
239 mechanisms, we applied the iCAMP framework to quantify the relative contributions
240 of ecological assembly processes via the *iCAMP* package (Ning et al., 2020). Finally,

241 we used the Binary State Speciation and Extinction (BiSSE) model to estimate rates of
242 speciation, extinction, and state transitions of opportunistic and sensitive taxa (He et al.,
243 2023).

244 Ecological co-occurrence networks were constructed in R software (version 4.4.2)
245 using the Hmisc package to compute Spearman correlations among ASVs and igraph
246 to assemble the network from significant associations ($r > 0.06$, $p < 0.05$). Network
247 visualizations were performed using Gephi software (version 0.10.1). We then extracted
248 site-specific subnetworks for each sampling location and evaluated their topological
249 metrics in relation to soil moisture. Keystone species were identified using *Zi-Pi*
250 analysis based on within-module connectivity and among-module connectivity,
251 methodological details were described in previous research (Shi et al., 2016). ASVs
252 classified as Module hubs, Network hubs, or Connectors were collectively designated
253 as keystone species. To identify the bacterial genera most strongly associated with soil
254 PO_4^{3-} content, we employed a random forest regression model and ranked genera based
255 on their variable importance scores (Liao et al., 2024). Finally, we used structural
256 equation modeling (SEM) to assess the direct and indirect effects of soil moisture, the
257 three bacterial life-history strategy subgroups, and P-cycling gene abundances on soil
258 PO_4^{3-} content (Dai et al., 2020).

259 **3. Results**

260 **3.1. Wildfire increased soil moisture and PO_4^{3-} content**

261 Wildfire significantly increased average PO_4^{3-} levels from 6.92 mg/kg to 14.72
262 mg/kg and raised soil moisture from 62.75% to 70.40% (Fig. 1). A significant positive

263 correlation was observed between mean soil moisture and PO_4^{3-} (Fig. S2). NH_4^+
264 concentration also rose markedly, from 10.89 mg/kg to 90.22 mg/kg, and FI from
265 increased 1.34 to 1.46. In contrast, NO_3^- decreased from 26.90 mg/kg to 12.59 mg/kg,
266 TN dropped from 1.08% to 0.90%, TOC declined from 42.70% to 37.59%, and BIX
267 fell from 0.18 to 0.16. However, wildfire had no significant effect on pH, C/N ratio, or
268 HIX, which averaged 4.16, 41.17, and 15.89, respectively.

269 **Insert Fig. 1 in here**

270 **3.2. Wildfire reduced the overall bacterial diversity and abundance**

271 Bacterial Shannon diversity decreased significantly from 6.86 to 6.01 after the
272 wildfire, with concurrent reductions in niche breadth (from 61.33 to 36.91) and absolute
273 abundance (from 1.90×10^9 to 7.10×10^8 copies g^{-1} DW) (Fig. 2a). At the phylum level,
274 *Actinomycetota* (average relative abundance: 32.58%) and *Pseudomonadota* (28.70%)
275 were dominant (Fig. 2b). Wildfire significantly increased the relative abundance of
276 *Bacillota* alone, from 2.47% to 27.61%, while other phyla decreased to varying extents
277 (Fig. S4a).

278 At the genus level, *Acidothermus* (16.66%) and *Sulfobacillus* (6.55%) were
279 dominant (Fig. S3). *Sulfobacillus* and *Alicyclobacillus*, both belonging to *Bacillota*,
280 showed the largest relative abundance increases after wildfire, rising from 0.16% to
281 12.94% and from 0.44% to 6.92%, respectively. Conversely, wildfire significantly
282 decreased the relative abundances of *Acidothermus* (21.11% to 12.20%), Subgroup_2
283 (6.25% to 1.96%), and WPS-2 (3.08% to 0.53%) (Fig. S4b).

284 **Insert Fig. 2 in here**

285 **3.3. Bacterial subgroups with different life-history strategies showed distinct**
286 **responses to wildfire**

287 We identified bacterial taxa with distinct wildfire response strategies to explore
288 those potentially contributing to soil PO_4^{3-} enrichment, especially under varying soil
289 moisture. Opportunistic, sensitive, and tolerant taxa accounted for 17.04%, 24.25%,
290 and 58.71% of the community, respectively. Among opportunistic taxa, *Bacillota*
291 showed the highest relative abundance (8.65%). *Pseudomonadota* (10.02%) and
292 *Actinomycetota* (8.96%) were predominant phyla among the sensitive taxa, while
293 *Actinomycetota* (20.53%) and *Pseudomonadota* (14.81%) were dominated among the
294 tolerant taxa (Fig. 3a). Compared with sensitive taxa, opportunistic and tolerant taxa
295 had greater dormancy potential, which increased significantly after wildfire (Fig. 3b).
296 Soil moisture was positively correlated with opportunistic and tolerant taxa, but
297 negatively with sensitive taxa (Fig. 3c). In contrast, TN, NO_3^- , and BIX were positively
298 associated with sensitive taxa but negatively with opportunistic and tolerant taxa (Fig.
299 S5).

300 The iCAMP model showed that drift was the main driver of soil bacterial
301 community assembly, contributing 51.12% on average, followed by dispersal limitation
302 (28.45%) and homogeneous selection (15.41%). Wildfire increased the contribution of
303 dispersal limitation to the assembly of opportunistic taxa from 8.45% to 34.64%, while
304 reducing the influence of homogeneous selection and drift by 12.11% and 11.14%,
305 respectively. For sensitive taxa, wildfire increased the contribution of drift from 45.46%
306 to 75.31%, while reducing homogeneous selection and dispersal limitation by 11.99%

307 and 18.45%, respectively. In contrast, the assembly of tolerant taxa was minimally
308 affected by wildfire (Fig. 3d). According to BiSSE model estimates, the speciation rate
309 of opportunistic taxa was 1.7 times higher than that of sensitive taxa, while their
310 extinction rate was 2.6 times higher. Moreover, opportunistic taxa were predicted to
311 transition to sensitive taxa at a rate five times higher than the reverse (Fig. 3e).

312

Insert Fig. 3 in here

313 **3.4. Wildfire enhanced the network complexity and stability of opportunistic taxa**

314 Wildfire significantly increased the proportion of opportunistic taxa in the network
315 by 17.68%, while reducing those of sensitive and tolerant taxa by 13.23% and 4.45%,
316 respectively (Fig. 4a). Subnetworks were constructed for each bacterial subgroup based
317 on sampling locations (Fig. S6), and their topological features were correlated with soil
318 moisture. All topological features of the opportunistic taxa subnetworks were positively
319 correlated with soil moisture, while no significant correlations were observed for
320 sensitive or tolerant taxa (Fig. 4b).

321 The *Zi-Pi* analysis revealed the impact of wildfire on keystone species within the
322 bacterial networks (Fig. 4c), with their taxonomic identities and topological roles
323 detailed in Table S1. Notably, only the number of keystone species belonging to the
324 opportunistic taxa increased after wildfire, while those associated with sensitive and
325 tolerant taxa declined (Fig. 4d).

326

Insert Fig. 4 in here

327 **3.5. Opportunistic taxa promoted soil PO₄³⁻ enrichment**

328 Peat PO_4^{3-} content was significantly positively correlated with the relative
329 abundance of opportunistic taxa, but not with that of sensitive or tolerant taxa (Fig. 5).
330 This suggests that opportunistic taxa may promote soil PO_4^{3-} enrichment.

331 A random forest model further identified *Methylovirgula* (1.08%), *Lysinibacillus*
332 (0.06%), and *Candidatus Koribacter* (0.07%) as the key genus-level opportunistic taxa
333 significantly associated with PO_4^{3-} enrichment ($p < 0.05$) (Fig. S7, Table S2).

334 **Insert Fig. 5 in here**

335 Genes involved in organic P-mineralization (e.g., *phnA*, *phnW*, and *phnX*), P-
336 starvation response (*phoB*, *phoR*), inorganic P-dissolution (*ppx*), and P-transport (e.g.,
337 *pstABC*) were significantly more abundant in burned soil (Fig. 6a). These results
338 indicate that changes following the wildfire activated P-cycling genes. Furthermore, the
339 abundance of opportunistic taxa was significantly positively correlated with P-
340 starvation response genes (*phoB*, *phoR*), organic P-mineralization genes (*phnA*, *phnW*,
341 *phoN*, and *phnX*), and P-transport genes (*ugpA*, *ugpB*, *ugpE*, and *pstB*) (Fig. 6b). Only
342 three organic P-mineralization genes (*phnA*, *phnW*, and *phoN*) were significantly
343 positively correlated with soil PO_4^{3-} content (Fig. 6c). These results suggest that wildfire
344 enhanced the coupling between opportunistic taxa and organic P-mineralization genes,
345 promoting their expression and facilitating the sustained enrichment of soil PO_4^{3-} .

346 A conceptual model based on SEM illustrates the potential mechanism by which
347 wildfire promotes peat PO_4^{3-} enrichment (Fig. 7). Wildfire drops peat surface level,
348 bringing it closer to the water table and increased soil moisture, which subsequently
349 significantly promoted opportunistic taxa and suppressed sensitive taxa but had no

350 notable effect on tolerant taxa. Opportunistic taxa enhanced the expression of organic
351 P-mineralization genes (*phnA*, *phnW*, and *phoN*) by increasing their relative abundance,
352 thereby elevating soil PO_4^{3-} levels. In contrast, sensitive and tolerant taxa did not
353 contribute to PO_4^{3-} accumulation.

354 **Insert Fig. 6 in here**

355 **4. Discussion**

356 **4.1. The wildfire anomaly: overall decline in soil communities and Pi enrichment**

357 Wildfires represent a significant ecological threat to tropical peat forests. As
358 observed in this study, wildfire within the tropical peatland context significantly
359 reduced the Shannon diversity, niche breadth, and absolute abundance of soil bacterial
360 communities (Fig. 2), suggesting a substantial decline in the integrity of these microbial
361 communities following fire disturbance. Similar trends have also been reported in soils
362 from subtropical and coniferous forest ecosystems (Nelson et al., 2022; Shi et al., 2024).
363 Typically, a decrease in bacterial diversity is associated with a decline in ecosystem
364 functions (or nutrient availability) (Cardinale et al., 2012; Wagg et al., 2014; Wagg et
365 al., 2019) such as the reductions in TN and TOC observed after wildfire (Fig. 1). Given
366 that P is often a key limiting nutrient for forest productivity (Cui et al., 2025), it is likely
367 that soil Pi would decrease after wildfire. However, contrary to predictions derived from
368 the diversity-ecosystem function theory, our findings reveal that the loss of overall
369 bacterial diversity did not impair P mineralization processes; rather, it coincided with
370 an unexpected increase in Pi accumulation (Fig. 1). This paradox suggests that wildfire
371 does not uniformly inhibit all bacterial subgroups. In contrast, it functions as an

372 environmental filter that selectively eliminates sensitive taxa while promoting the
373 growth of opportunistic taxa with higher potential for organic P mineralization. The
374 relatively rapid P mineralization performed by these opportunistic taxa likely provides
375 critical Pi nutrients, which play a crucial role in supporting early vegetation recovery
376 after wildfire. Mineralization may also release P in soil drainage and thereby enhance
377 eutrophication effects in receiving water ways. Additionally, alteration to peat nutrient
378 status is likely to have implications for the significant GHG emissions associated with
379 degraded peatlands (Hooijer et al., 2012).

380 <https://bg.copernicus.org/articles/9/1053/2012/>

381 **Insert Fig. 7 in here**

382 **4.2. Establishment of opportunistic taxa dominance in peat forest soils under** 383 **wildfire disturbance**

384 In response to wildfire disturbance, forest soil microbial communities often shift
385 from K-strategist to r-strategist dominance, as the latter exhibit faster growth rates and
386 higher nutrient utilization efficiency (Qin et al., 2025). However, the underlying
387 mechanisms of this functional shift have remained poorly understood. Our study
388 extends current knowledge by providing detailed mechanistic insights: (i) As typical r-
389 strategists, opportunistic taxa exhibited speciation and extinction rates 1.7 and 2.6 times
390 higher, respectively, than those of sensitive taxa (Fig. 3e). This high turnover rate
391 enabled them to rapidly reproduce and diversify, allowing swift occupation of newly
392 created post-fire niches. (ii) Microorganisms can enter dormancy—a low-metabolic
393 state—to survive unpredictable environmental stress (Lennon and Jones, 2011). Under

394 wildfire disturbance, opportunistic taxa showed a higher potential for deploying
395 dormancy strategies than sensitive or tolerant taxa (Fig. 3a). For example, *Bacillota*, a
396 dominant phylum among opportunistic taxa, can form multi-layered protective
397 endospores to enhance stress resistance (Hashimi and Tocheva, 2024). (iii) The high
398 temperatures of wildfires can disrupt the compact structure of forest soil aggregates
399 (Jordán et al., 2011), increasing soil porosity and habitat fragmentation. This enhances
400 the role of dispersal limitation in driving the colonization of opportunistic taxa (Fig.
401 3d), enabling them to establish locally and thereby reduce direct competition with
402 tolerant taxa (Walters et al., 2022). (iv) Wildfire increased both the relative abundance
403 and number of keystone species of opportunistic taxa in co-occurrence networks, while
404 those of sensitive and tolerant taxa declined (Fig. 4). This suggests that opportunistic
405 taxa play more central functional roles in the restructured bacterial communities,
406 potentially accelerating short-term nutrient cycling in peat soils. These findings
407 elucidate the ecological mechanisms underlying the increased dominance of
408 opportunistic taxa after wildfire events.

409 We found that soil moisture in peat soils increased significantly after wildfire (Fig.
410 1). This was also observed in post-fire observations by Lupascu et al., 2020. The clear
411 drop in surface level associated with the frequently deep burn scars, contributed to
412 bringing the peat surface closer to the water table height, thereby enhancing peat
413 moisture content. Reduced plant transpiration and enhanced rainfall infiltration may
414 also have contributed to this. Similarly, a study of boreal shrub wetlands reported that
415 13 years post wildfire, soil moisture had increased at least 26% (Li et al., 2023),

416 indicating that fire-induced poor drainage may have long-lasting effects. Soil moisture
417 correlated significantly positively with the relative abundances of opportunistic and
418 tolerant taxa but negatively with sensitive taxa (Fig. 3c). Soil moisture was also
419 positively correlated with the topological features of opportunistic taxa in the co-
420 occurrence network (Fig. 4b). These findings collectively suggest that moisture is a key
421 driver linking fire disturbance to bacterial community responses in warm and humid
422 tropical peatlands (secondary forests and agricultural plantations). Adequate soil
423 moisture facilitates the dissolution and transport of nutrients and metabolites (Smith et
424 al., 2023), thereby promoting the rapid proliferation of r-strategist microbes. Moreover,
425 increased soil moisture can revive the dormant spores of opportunistic taxa (Imminger
426 et al., 2024), further enhancing their abundance. This aligns with r/K selection theory,
427 which suggests that increased moisture may create transient resource-rich conditions
428 favorable to r-strategists (Li et al., 2015). However, prolonged high-moisture conditions
429 may be unfavorable for some aerobic bacteria. For example, *Acidothermus*, an aerobic
430 genus, showed one of the most notable declines in relative abundance after wildfire
431 (Fig. S4b). Overall, increased soil moisture appears to serve as a critical “trigger” for
432 the rise of opportunistic taxa during the early stages of post-fire ecosystem recovery.

433 **4.3. Organic P-mineralization genes and Pi accumulation: driven by opportunistic** 434 **taxa**

435 The relative abundance of opportunistic taxa was positively correlated with both
436 organic P-mineralization gene abundance (*phnA*, *phnW*, and *phoN*) and soil Pi content
437 (Figs. 5, 6, 7). This highlights the crucial role of opportunistic taxa in the wildfire-

438 induced Pi enrichment. Previous global-scale studies have suggested that wildfires can
439 increase soil P through mechanisms such as enhanced mechanical weathering of rocks
440 and plant ash deposition (Zhou et al., 2025). The present study further reveals the
441 critical bacterial contribution: by increasing soil moisture, wildfire not only stimulated
442 the proliferation of opportunistic taxa but may also have enhanced phosphatase activity
443 (Margalef et al., 2021), thereby accelerating the mineralization of organophosphorus
444 compounds. In fact, the abundance of both opportunistic taxa and organic P-
445 mineralization genes (*phnA*, *phnW*, and *phoN*) increased in parallel (Figs. 6, 7),
446 indicating that wildfire strengthened the coupling between r-strategist microbes and P
447 mineralization functions. Thus, opportunistic taxa may promote Pi accumulation by
448 upregulating community-level organic P-mineralization genes. This coupling may be
449 transient and ecologically fragile because shifting soil moisture regimes (e.g., moisture
450 loss due to long-term declines in water-holding capacity of soil aggregates) could
451 disrupt this microbial process in tropical peat forests under climate change. These
452 results therefore reinforce the need to raise water tables and rewet degraded peatlands
453 to restore ecological functionality.

454 **5. Conclusions**

455 Our investigation of tropical peatland revealed that although wildfire reduced the
456 overall diversity and abundance of bacterial communities, they significantly increased
457 soil moisture and Pi level. This suggests that fire dose not uniformly suppress all
458 bacterial taxa. Based on life-history strategies, we classified bacteria into opportunistic,
459 sensitive, and tolerant taxa, and revealed the central role of opportunistic taxa. Wildfire

460 initially elevated soil moisture, which in turn enhanced the relative abundance of
461 opportunistic taxa while reducing that of sensitive ones. After the fire disturbance,
462 opportunistic taxa demonstrated increasing speciation rates, greater dormancy potential,
463 stronger dispersal limitation, and an increased number of keystone species in co-
464 occurrence networks. Meanwhile, wildfire intensified the coupling between these taxa
465 and key P-mineralization genes (*phnA*, *phnW*, and *phoN*), thereby promoting the
466 accumulation of Pi. This study, for the first time, uncovered a sequential ecological
467 mechanism—from environmental disturbance to community reorganization, functional
468 response, and ultimately nutrient cycling—highlighting the pivotal role of bacterial
469 communities in sustaining soil nutrient dynamics and ecosystem stability under
470 escalating wildfire frequency and global climate change.

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481 **CRedit authorship contribution statement**

482 **Pengfei Gao**: Conceptualization, Formal analysis, Visualization and Writing –
483 original draft. **Xiaoyu Cheng**: Methodology, Conceptualization, Funding acquisition,
484 and Writing – review and editing. **Wen Tian**: Investigation. **Xiaoyan Liu**:
485 Methodology. **Olli H Tuovinen**: Writing – review and editing. **Stephanie L. Evers**:
486 Investigation, Funding acquisition, Writing – review and editing. **Thomas E.L. Smith**:
487 Investigation, Funding acquisition. **Hongmei Wang**: Conceptualization, Funding
488 acquisition, Resources, and Writing – review and editing.

489 **Data accessibility**

490 The raw sequence data reported in this paper have been deposited in the Genome
491 Sequence Archive in National Genomics Data Center, China National Center for
492 Bioinformation/Beijing Institute of Genomics, Chinese Academy of Sciences, under
493 BioProject accession No. PRJCA043095 and are publicly accessible at
494 <https://ngdc.cncb.ac.cn/gsa>.

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714

715 **Figure captions**

716 **Fig. 1** Effects of wildfire on physicochemical parameters of soil samples collected from
717 Kuala Lumpur, Malaysia. Different letters (*a, b*) above the boxes indicate significant
718 differences in physicochemical parameters. Control, CF/COP; Burned, BF/BOP; FI,
719 fluorescence index; BIX, biological index; HIX, humification index.

720 **Fig. 2** Shannon diversity, niche breadth and absolute abundance of soil bacterial
721 communities in response to wildfire (a). Different letters above the boxes indicate
722 significant differences between groups. Bacterial community composition at the
723 phylum level in the control and burned soils (b).

724 **Fig. 3** Impacts of wildfire on bacterial subgroups with different life-history strategies.
725 Relative abundance of dominant phyla within the three bacterial subgroups (a). Effects
726 of wildfire on the relative abundance of sporulation genes in the three subgroups (b).
727 Different letters above the boxes indicate significant differences between groups.
728 Relationship between bacterial subgroup abundance and soil moisture (c), * $p < 0.05$,
729 ** $p < 0.01$, *** $p < 0.001$. Ecological assembly mechanisms of the three bacterial
730 subgroups in response to wildfire (d). Evolutionary traits of opportunistic and sensitive
731 taxa based on the BiSSE model estimates (e).

732 **Fig. 4** Changes in soil bacterial co-occurrence networks between control and burned
733 peat. Relative abundances of the three bacterial subgroups in the networks (a).
734 Relationships between soil moisture and network topological features for the three
735 bacterial subgroups (b), * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$. *Zi-Pi* analysis showing

736 wildfire effects on the keystone species in bacterial networks (c), and changes in the
737 keystone species numbers across the three subgroups (d).

738 **Fig. 5** Relationships between soil PO_4^{3-} content and the abundances of bacterial
739 subgroups, * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

740 **Fig. 6** P cycling processes and differences in P-cycling gene abundances between
741 control and burned soils (a), genes in red indicate significantly higher abundance in
742 burned soil, and genes in blue indicate significantly higher abundance in control soil.
743 The P-cycling functions corresponding to each gene are shown in Table S3.
744 Correlations between the abundance of opportunistic taxa and P-cycling genes involved
745 in P-starvation response, organic P-mineralization, and P-transport (b). Only
746 significantly correlated genes are shown. Correlations between P-cycling gene
747 abundance and soil PO_4^{3-} content (c). Only significantly correlated genes are shown. *
748 $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

749 **Fig. 7** SEM revealed causal relationships among soil moisture, three bacterial
750 subgroups, organic P-mineralization genes (*phnA*, *phnW* and *phoN*), and soil PO_4^{3-}
751 levels. Goodness of fit index is 0.767. Colors of bacteria in the central circle indicate
752 different life-history strategies: red (opportunistic), yellow (sensitive), and blue
753 (tolerant). Red lines indicate significant positive effects; black lines, significant
754 negative effects; and dashed lines, nonsignificant effects. * $p < 0.05$, ** $p < 0.01$, ***
755 $p < 0.001$.

756

757 **Supplementary materials**

758 **Fig. S1** Locations of soil sampling sites.

759 **Fig. S2** Spearman correlation between soil moisture and PO_4^{3-} content. *** $p < 0.001$.

760 **Fig. S3** Genus-level composition of bacterial communities in control and burned soils.

761 **Fig. S4** Effects of wildfire on the relative abundance of bacterial taxa at the phylum (a)
762 and genus (b) levels based on STAMP analysis. Taxa showing significant differences
763 between control and burned soils are highlighted.

764 **Fig. S5** Spearman correlation between environmental parameters and the three bacterial
765 subgroups (opportunistic, sensitive, and tolerant taxa). * $p < 0.05$, ** $p < 0.01$, *** $p <$
766 0.001 .

767 **Fig. S6** Subnetworks of the three bacterial subgroups (opportunistic, sensitive, and
768 tolerant taxa) constructed based on sampling locations.

769 **Fig. S7** Importance ranking of bacterial genera (belonging to the three life-history
770 strategy subgroups) in predicting soil PO_4^{3-} content based on random forest model. * p
771 < 0.05 , ** $p < 0.01$, *** $p < 0.001$.

772 **Table S1** Taxonomic information and topological roles of keystone species between
773 control and burned soils.

774 **Table S2** Relative abundance of genus-level bacteria affecting soil PO_4^{3-} -enrichment
775 in three survival-strategy bacterial subgroups.

776 **Table S3** Phosphorus cycling functions associated with P-cycling genes.