

ANALGESIC, ANTI-INFLAMMATORY AND ANTIPYRETIC ACTIVITIES OF THE
AQUEOUS EXTRACT OF *GERANIUM CAROLINIANUM* L.

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Abstract

Background: *Geranium carolinianum* L. (Geraniaceae) is widely used for a variety of diseases including herpetic keratitis, eczema, rheumatism etc. However, there is lack of relevant scientific research.

Materials and Methods: GCE (125, 250, 500mg/kg body weight) was evaluated for its pharmacological properties by using the acetic acid-induced writhing test, the hot plate test and the fresh egg white-induced paw edema in rats. The dimethylbenzene-induced mouse inflammation model and the lipopolysaccharide (LPS)-induced rat fever model were employed and the acute toxicity of GCE was also assessed.

Results: The *Geranium carolinianum* aqueous extract (GCE) significantly inhibited the writhing responses in mice, increased reaction time of mice in the hot plate test, and suppressed the fresh egg white-induced paw edema in rats and the dimethylbenzene-induced ear edema in mice whilst attenuating LPS-induced fever in rats in a dose dependent manner. Furthermore, no deaths were observed when mice were orally administered GCE up to 14 g/kg body weight (approximately 553times of clinical dose).

Conclusions: GCE possesses analgesic, anti-inflammatory and antipyretic activities and is non-toxic at the doses used. The results of this study support the clinical use and effectiveness of *Geranium carolinianum* as an analgesic, anti-inflammatory and antipyretic agent in folk medicine.

Key words: Plant drug; Pain; Inflammation; Pyrexia; Toxicity; Animal

Introduction

Geranium carolinianum L. (Geraniaceae) is a commonly used herb in folk medicine for the treatment of herpetic keratitis, rheumatoid arthritis, neuralgia, disc prolapsed, gastrointestinal disorders, bacillary dysentery, febrile diseases etc. It has a wide distribution in North East China, East China, Central China and Sichuan Province (Han et al., 2001; Wu et al., 2008). One of the most important clinical applications of this herb is in the treatment of herpetic keratitis (Wang et al., 1995). In addition, *Geranium carolinianum* ointment is reported to be a good therapy for eczema and herpes zoster (Han et al., 2001). *Geranium carolinianum* has been recorded as a Chinese herb in the Chinese Pharmacopoeia owing to its exact curative effects.

The phytochemical study of *Geranium carolinianum* reveals the presence of tannins, flavones, flavonoids, volatile oils and organic acids (Lei et al., 2002), which have been extracted from *Geranium carolinianum* as well as most of the congeneric plants (He et al., 2011). This plant possesses a variety of pharmacological effects, such as antibacterial, antiviral, antioxidant, antitumor, hypoglycemic, liver protecting, and cough relieving properties (Li et al., 2008; Wu et al., 2011). The water extract from *Geranium carolinianum* contains 5–40% of corilagin, which is known to be effective for the treatment of gastroenteritis (Li et al., 2009).

As far as we are aware, there have been no scientific reports on the analgesic, antipyretic, and anti-inflammatory effects of this plant in animals. In this study, we report these activities of the aqueous extract of *Geranium carolinianum* which were evaluated in

mice and rats to substantiate and expand its clinical applications. In addition, the acute oral toxicity of the extract was also determined in mice.

Materials and methods

Plant material and extraction

The whole plant of *Geranium carolinianum* was collected in Great Khingan of Heilongjiang Province in September 2013 and the plant was identified by Professor Han-Ming Zhang, a pharmacognosist from the Department of Pharmacognosy, School of Pharmacy, Second Military Medical University (Shanghai, China). A voucher specimen of *Geranium carolinianum* was deposited and assigned the number YZ130905 in the herbarium of the Department of Pharmaceutical Botany, School of Pharmacy, Second Military Medical University (Shanghai, China). The dried plant (500g) was pulverized with amotor-driven grinder to prepare the extract which was then refluxed with distilled water (8L) three times for 0.5 h, 1 h, and 2h, respectively. The extract was then filtered, centrifugated and evaporated to dryness under reduced pressure in a rotary evaporator and the dry powder was obtained by spray drying and a yield of 10.13% was achieved.

Chromatographic Conditions

An Agilent 1100 HPLC system, equipped with a quaternary pump, an autosampler, a degasser, an automatic thermostatic column compartment, a DAD and an LC/MSD Trap XCT ESI mass spectrometer (Agilent Technologies, MA, USA), was used for separation. The separation was performed on a GS-120-5-C18-BIO chromatographic column (5 μ m, 250 \times 4.6 mm i.d.) with the column temperature set at 35°C. A linear gradient elution of A (0.1% formate acid water) and B (acetonitrile) was used with the gradient procedure as follows: 0 min, B 5%, to 60 min B 40% (v/v). The flow rate was 1.0 mL/min and the injection volume was 10 μ L. DAD was on and the target wavelength was simultaneously set at 210 nm. The split ratio to the mass spectrometer was 1:3. The acquisition parameters for negative ion mode were: collision gas, ultrahigh-purity helium (He), nebulizer gas (N₂), 35 psi, drying gas (N₂), 10 L/min, drying temperature, 350°C, HV, 3500 V, mass scan range, m/z 100-2200, target mass, 500 m/z, compound stability, 100%, trap drive level, 100%. All the data were analyzed by Chemstation software.

Animals

ICR mice (age range 8–10 weeks, weight range 20–25g), male Sprague-Dawley (SD) rats (age range 8–9 weeks, weight range 200–220 g) and male Wistar rats (age range 7–9 weeks, weight range 200–220 g), were obtained from the Experimental Animal Center of the Second Military Medical University (Shanghai, China), and were housed in a regulated environment (20 \pm 2°C), with a 12 h light/dark cycle (08:00–20:00, light). The animals were deprived of food for 12 h prior to the experiment, with water given *ad libitum* and each animal was used once in the experiment. All animal treatments were strictly in accordance with international ethical guidelines concerning the care and use of laboratory animals, and all the experiments were carried out under the approval of the Committee of Experimental Animal Administration of the University (No. LL130929). Each experimental group consisted of 10 animals.

Drugs and reagents

The following reagents and drugs were used: dimethylbenzene (AR), and acetic acid (AR) (Sinopharm Chemical Reagent, Shanghai, China), lipopolysaccharide (Sigma, St.

Louis, MO, USA), tramadol, indomethacin, dexamethasone, paracetamol (Chengdu Pharmaceutical Factory, Chengdu, China).

Treatment

GCE, indomethacin, dexamethasone, paracetamol, and tramadol were each suspended in 1% carboxymethylcellulose sodium (CMC-Na) solution respectively prior to administration. GCE at a concentration range of 125, 250, 500 mg/kg body weight was orally administered to the three groups of animals (n=10) respectively by intubation. The positive group of animals received indomethacin (5 mg/kg) in the writhing and paw edema tests, tramadol (30mg/kg) in the hot plate test, dexamethasone (5 mg/kg) in the ear edema test and paracetamol (100 mg/kg) in the antipyretic test, respectively. The control group of animals were given 1% CMC-Na solution and it established alongside the drug-treated groups, all of which received a volume of 10 mL/kg body weight irrespective of dose.

Analgesic test

The peripheral and central analgesic activity of GCE was investigated by monitoring test animals exposed to chemical and thermal stimuli respectively.

In the acetic acid-induced writhing test (Garcia et al., 2004), male mice were orally administered GCE or indomethacin (5mg/kg body weight) respectively, 1 h prior to intraperitoneal injection of acetic acid (1%, 10mL/kg body weight). The number of writhing reflexes was then counted for a period of 15 min.

In the hot plate test (Ma et al., 2011; Xu et al., 2002), the latency was recorded when female mice were screened on the hot plate at $55\pm0.5^{\circ}\text{C}$. Only mice that showed initial nociceptive responses between 5 and 30 sec were selected for this experiment. The posttreatment reaction time of each animal was recorded by observing either the licking of the hind paws or the jumping movements to avoid thermal pain after 30, 60, 90, and 120 min respectively of administration of GCE or tramadol (30 mg/kg body weight).

Anti-inflammatory test

The anti-inflammatory activity of GCE was measured against fresh egg white induced acute paw edema in rats (Xu et al., 2002) and dimethylbenzene induced ear edema in mice (Ma et al., 2011; Zheng et al., 2009).

In the rat paw edema test, male SD rats were orally premedicated with GCE or indomethacin (5 mg/kg body weight) 0.5h prior to subplantar injection of 0.1 mL freshly prepared 10% (v/v) egg white solution into the right hind paws of the rats. Paw volume was measured plethysmometrically using a paw edema detector (YLS-7A; Shandong Academy of Medical Science Device Station, Shandong, China) at 0, 0.5, 1, 2, 3, 4, and 6 h postinjection. The mean increase in paw volume was measured and inhibitory percentage was calculated. The edema rate of rats was calculated as follows:

$$\text{Edema rate (\%)} = (V_t - V_0) / V_0 \times 100\%$$

Where V_0 is the volume before fresh egg white injection (mL); V_t is the volume after t h of fresh egg white injection (mL).

In the dimethylbenzene-induced ear edema test, GCE or dexamethasone (5 mg/kg body weight) was orally administered 1h prior to local application of dimethylbenzene (40 μL /ear) to the right ear of mice. The ear swelling was measured by subtracting the weight of the left ear from that of the right post1 h of dimethylbenzene treatment as reported previously (Zheng et al., 2009).

Antipyretic test

The antipyretic activity of GCE was investigated according to the method previously reported (Ma et al., 2011). Male SD rats were randomly divided into five groups based on their basal temperature measured prior to the experiment. The temperature in external auditory meatus was measured in a temperature-controlled room (ambient temperature $20 \pm 2^{\circ}\text{C}$) using an electronic temperature recorder (DT-IDB, Shanghai Medical Instruments Factory, Shanghai). Rat fever can be induced by intraperitoneal injection of LPS (1 mg/kg). SD rats were pretreated with LPS post 0.5 h of oral administration of GCE or paracetamol (100 mg/kg body weight). The temperature was measured every 30 min from 9:00 hrs to 15:00 hrs.

Acute toxicity

Acute toxicity study was carried out as reported in the literature (Saha et al., 2013). ICR mice of either sex were tested by orally administering different doses of the extract by increasing or decreasing the dose according to the responses of the animals. The animals were fasted overnight and had access to water only prior to the experiment. The animals were administered GCE at a dose of 14 g/kg body weight and were test for any toxic signs or mortality for 7 days.

Statistical analysis

The data was analyzed using a SPSS 13.0 statistical package and data for multiple comparisons was performed by one-way ANOVA followed by Tukey's test. A value of $P < 0.05$ was considered statistically significant and all results are presented as mean \pm SD.

Results

Identification of compounds

The aqueous extract of *Geranium carolinianum* (GCE) was measured by high-performance liquid chromatography coupled with electrospray mass spectrometry (HPLC/ESI-MS) in negative-ion mode (Figure 1 a-g). Five compounds 1-5, with the retention time at 7.1 min, 19.2 min, 21.7 min, 29.8 min, and 30.5 min, were identified as gallic acid (1), brevifolin carboxylic acid (2), corilagin (3), ellagic acid (4), and hyperoside (5), on the basis of the observation of the pseudomolecular ion peak at m/z 168.8 $[\text{M-H}]^{-}$, m/z 632.9 $[\text{M-H}]^{-}$, and m/z 463.0 $[\text{M-H}]^{-}$ in HPLC/ESI-MS chromatogram, in accordance with the molecular weights of three compounds (gallic acid, coridagin, and hyperoside, Figure 1) occurring in *Geranium carolinianum*.

Effect of GCE on the writhing reflex of mice

As shown in Figure 2, the aqueous extract from *Geranium carolinianum* (GCE) significantly reduced the number of writhing responses in a dose dependent manner within 15 min of injection of acetic acid. At the higher dose of GCE the reduction in writing responses was much more noticeable and was even lower than that of the mice receiving indomethacin, suggesting that GCE has potent analgesic properties.

Effect of GCE in heat-stimulated mice

The analgesic activity of GCE was also markedly observed in the hot-plate test. It was found that GCE at doses of 250 and 500 mg/kg body weight prolonged the latency period of mice significantly when compared with the negative group of animals which received distilled water (Figure 3).

Effect of GCE on fresh egg white-induced paw edema in rats

As shown in Table 1, the subplantar injection of 0.1 mL of fresh egg white noticeably induced paw edema in rats, which had persisted for 6 h. The low dose of GCE (125 mg/kg body weight) evidently decreased the percentage of paw edema only after 2h and 3 h of treatment, and the middle dose (250 mg/kg body weight) significantly inhibited paw edema of rats after 1-4 h of treatment. However, both the high dose of GCE (500 mg/kg body weight) and indomethacin elicited the persistently inhibitory effect on edema formation after 0.5 - 6 h of treatment.

Effects of GCE on dimethylbenzene-induced ear edema in mice

Dimethylbenzene evidently increased ear edema in mice, GCE at the doses of 250 and 500 mg/kg significantly inhibited ear edema compared with the control group. These results reveal that GCE has significant activity against acute inflammation induced by dimethylbenzene (Figure 4).

Effects of GCE on fever in rats

As shown in Table 2, the temperature in rats was evidently elevated one hour postintraperitoneal injection of LPS and persisted for the following five hours. In contrast to the control group of animals given distilled water, the temperature in the rats receiving GCE significantly decreased in a time- and dose-dependent manner. These results indicate that oral administration of GCE inhibits pyrexia induced by LPS.

Acute toxicity

Although the mice were given GCE orally up to 14g/kg body weight which is equivalent to approximately 500 times of clinical dose (15 g/d herb each adult with 60 kg body weight), no mortality was observed during the assessment period (7 days). Thus, it can be concluded that oral administration of GCE at this dose is safe.

Discussion

It is well acknowledged that there is a growing need for safer and more effective drugs to treat pain, inflammation and fever. Due to the inevitable side effects of the drugs currently available there is strong interest in medicinal plants with a history of traditional use for the treatment of above diseases. This study was designed to evaluate the analgesic, anti-inflammatory and antipyretic effects of the aqueous extract of *Geranium carolinianum* (GCE) to give scientific authentication to the traditional claims. In the present study, GCE inhibited the writhing responses in mice in the acetic acid writhing reflex test, increased reaction time in mice in the hot-plate test, and reduced fresh egg white-induced paw edema in rats and dimethylbenzene-induced ear edema in mice. It further attenuated rat pyrexia induced by lipopolysaccharide (LPS), suggesting the favorable analgesic, anti-inflammatory and antipyretic activities of GCE. Additionally, the extract displayed no toxic effects at a dose of 14g/kg body weight supporting its safety in clinical use. These activities are associated with five main compounds contained in GCE, gallic acid, brevifolin carboxylic acid, corilagin, ellagic acid, and hyperoside (Daud et al., 2006; Gambari et al., 2012; Hsiang et al., 2013; Ku et al., 2014; Trevisan et al., 2014; Zhou et al., 2014).

The acetic acid writhing mice is a classic nonselective analgesic screening model. Acetic acid induces algesia by liberating endogenous substances such as prostaglandinsE₂ and many others that stimulate pain nerve endings (Liu et al., 2013). The results show that GCE significantly reduced the number of writhing responses in mice intraperitoneally

injected with acetic acid in a dose dependent manner. The analgesic effect of GCE may be due to inhibition of the local level of prostaglandins (Saha et al., 2013).

The hot-plate test method has been applied in analgesic investigations for several decades, which, when compared to the writhing test, can usually distinguish between central and peripheral effects (Srinivasan et al., 2003). The validity of this test has been shown even in the presence of substantial impairment of motor performance (Plummer et al., 1991). In this test, male animals are generally not used because they will lick their scrotums against thermal injury when they touch the heated plate. In this case, a false reaction time of licking the hind paws is often recorded.

Some peripheral analgesic drugs such as aspirin usually have few effects in the hot plated test (Zhu et al., 2011), while some other analgesic drugs such as ibuprofen and tramadol can reduce prostaglandin synthesis via central inhibition of cyclooxygenase, or bind to specific opioid receptors in the central nervous system, exhibiting both peripheral and central analgesic activities (Björkman, 1994). It is clearly demonstrated in our study that GCE at the doses of 250 and 500mg/kg body weight significantly prolonged the latency period of mice in the hot plate test, suggesting its centrally analgesic effect.

The rat paw edema induced by fresh egg white is suitable for evaluating anti-inflammatory agents and is used frequently to assess the anti-edematous effect of natural products (Du et al., 2013). It is well known that prostaglandins play an important role in pain progress in chemical nociception models (Santos et al., 1998) and are the target of action of commonly used anti-inflammatory drugs. GCE at the dose of 250 and 500 mg/kg body weight markedly reduced egg white-induced paw edema in rats, which lasted for 6 h, indicating that the extract suppresses the release or actions of some of inflammatory mediators, and possibly reduces the release of prostaglandins. The anti-inflammatory activity of the extract was further evidenced by the dimethylbenzene-induced ear edema test, in which GCE at the doses of 250 and 500mg/kg body weight also inhibited ear edema of mice markedly. In the process of inflammation, activated neutrophils release mediators such as platelet-activating factor and lysozyme, which can lead to vasodilatation and increase vascular permeability (Saeed et al., 2010).

LPS activates prostaglandin E_2 (PGE₂)-synthesizing enzymes in the lung and liver within minutes of systemic administration followed by PGE₂ which originates peripherally and is delivered to the brain by albumin or by locally influenced nerve fibers such as hepatic vagal fibers. Hepatic fibers contain PG receptors of the EP3 type causing pyrexia (Dogan et al., 2004). In our study, LPS-induced fever was employed to investigate the antipyretic activity of GCE and the results presented in Table 2 indicate that GCE possesses significant antipyretic action.

In addition, the acute toxicity study revealed that GCE did not cause any toxic effects as judged by the lack of fatalities in mice at a dose of 14 g/kg body weight (approximately 553 times of clinical dose), thus displaying good safety.

In conclusion, our study demonstrates that GCE exerts favorable analgesic, anti-inflammatory, and antipyretic activities possibly by inhibition of the central synthesis of prostaglandins, which are involved in five identified compounds contained in this extract. This validates the claim by traditional medicine practitioners that GCE can be used to treat rheumatic arthralgia and febrile diseases. However, detailed studies are needed to elucidate

the mechanisms of action responsible for the analgesic, anti-inflammatory, and antipyretic activities exhibited by the aqueous extract of *Geranium carolinianum*.

Conflict of interest statement

The authors report no declarations of interest.

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Table 1 Effects of GCE on egg white-induced paw edema in rats

Group	Animal (N)	Dose (mg/kg)	Percentage of paw edema (%)	
			0.5 h	1 h
Control	10	1% CMC-Na	66.7±7.7	69.7±7.4
IMC	10	5	50.8±6.3**	48.5±5.9**
GCE	10	125	64.4±8.6	62.5±9.0
GCE	10	250	60.9±8.0	59.2±8.2*
GCE	10	500	54.1±7.7**	40.5±6.9**

Percentage of paw edema (%)			
2 h	3 h	4 h	6h
68.8±8.9	51.5±6.7	38.2±5.6	26.7±4.5
50.2±5.3**	44.8±4.9**	22.6±4.4**	13.0±3.6**
51.6±6.1**	41.4±5.0**	33.3±5.3	27.0±10.9
45.1±6.2**	39.3±6.3**	32.2±5.6*	22.6±4.3
36.1±5.6**	34.1±5.3**	20.5±3.0**	11.8±2.5**

Table 1 Effects of *Geranium Carolinianum* extract (GCE) on fresh egg white-induced paw edema in rats. When 0.1 ml of freshly prepared 10% (v/v) egg white solution was injected subplantarily into the right hind paws of male rats, acute inflammation was induced. GCE and indomethacin (IMC) were orally premedicated. Data are presented as mean ± SD, n= 10. * $P < 0.05$, ** $P < 0.01$, significance versus control.

Table 2 Effects of GCE on temperature in external auditory meatus of rats

Group	Animal (N)	Dose (mg/kg)	Temperature (°C)				
			0 h	0.5 h	1 h	1.5h	2h
Control	10	1% CMC-Na	35.9±0.3	35.9±0.4	37.0±0.4	37.8±0.4	37.4±0.2
PRC	10	100	36.0±0.3	35.4±0.3*	35.7±0.4**	35.9±0.3**	35.7±0.3**
GCE	10	125	35.9±0.2	35.6±0.4	36.6±0.4	37.2±0.4*	36.8±0.5*
GCE	10	250	36.0±0.3	35.6±0.3	35.7±0.3**	36.7±0.4**	36.5±0.5**
GCE	10	500	36.1±0.2	35.7±0.4	36.0±0.5**	36.7±0.4**	36.4±0.4**

Temperature (°C)							
2.5h	3 h	3.5h	4 h	4.5h	5h	5.5h	6h
37.3±0.3	37.1±0.4	37.0±0.4	37.3±0.4	37.3±0.4	37.5±0.4	36.9±0.3	36.8±0.3
35.4±0.4**	35.6±0.3**	35.8±0.4**	35.9±0.4**	36.0±0.4**	35.5±0.4**	35.9±0.3**	35.9±0.4**
36.7±0.4**	36.9±0.5**	36.9±0.4	37.1±0.3	36.8±0.3	37.2±0.4	37.0±0.5	36.4±0.4
36.5±0.5**	36.6±0.4*	36.7±0.3	36.7±0.3**	36.2±0.5**	36.8±0.4**	36.5±0.4*	36.1±0.4**
36.3±0.4**	36.7±0.3*	36.4±0.5*	36.6±0.4**	36.4±0.5**	37.0±0.2*	36.6±0.3	36.1±0.4**

Table 2 Effects of *Geranium Carolinianum* extract (GCE) on temperature in external auditory meatus of rats. The rats were injected with lipopolysaccharides (LPS) after 0.5 h of the oral administration of GCE or paracetamol (PRC). The temperature was measured every 30 min from 09:00 hrs to 15:00 hrs. Data are presented as mean ± SD, n = 10. * $P < 0.05$, ** $P < 0.01$, significance versus control.

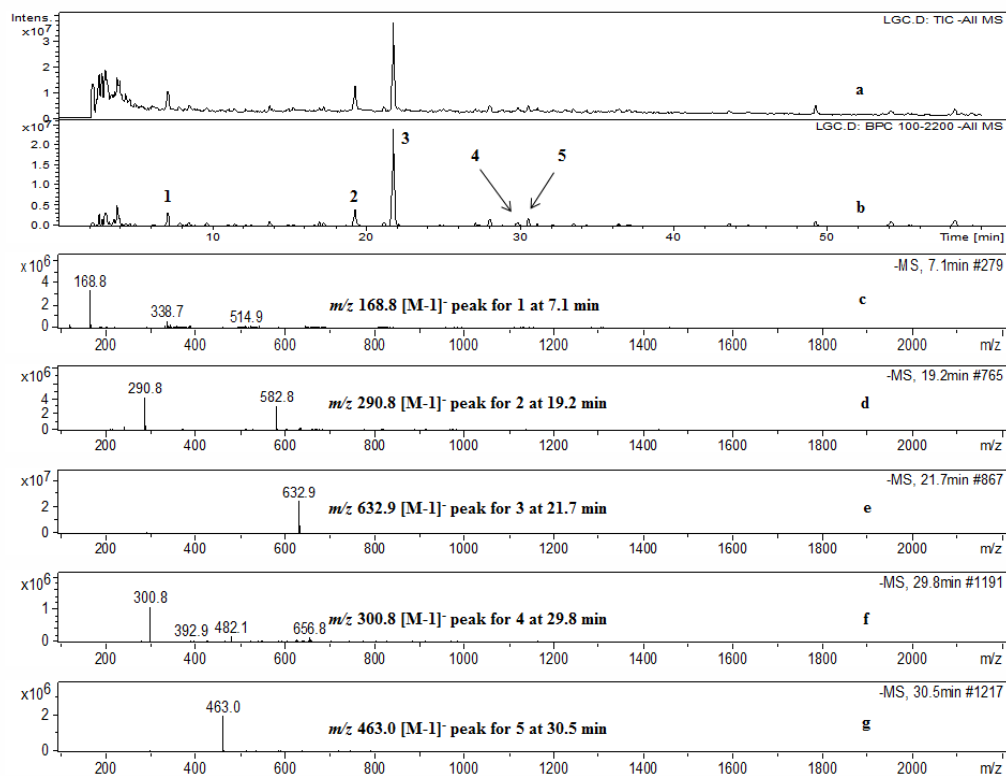


Figure 1 HPLC/ESI-MS chromatogram in negative mode of the extract of *Geranium Carolinianum* (a: total ion chromatogram b: base peak chromatogram c: ESI-MS spectra of $[M-1]^-$ ion of compound **1** (retention time: 7.1 min) d: ESI-MS spectra of $[M-1]^-$ ion of compound **2** (retention time: 19.2 min) e: ESI-MS spectra of $[M-1]^-$ ion of compound **3** (retention time: 21.7 min) f: ESI-MS spectra of $[M-1]^-$ ion of compound **4** (retention time: 29.8 min) g: ESI-MS spectra of $[M-1]^-$ ion of compound **5** (retention time: 30.5 min)).

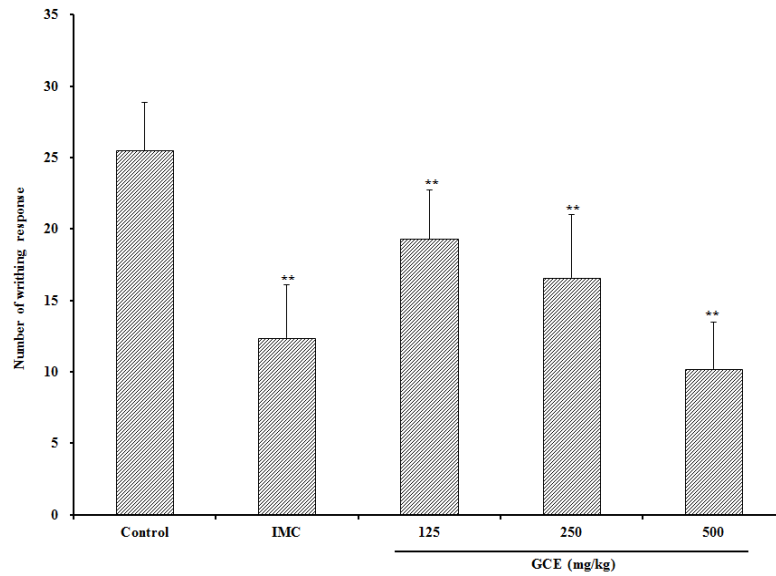


Figure 2 Effects of *Geranium Carolinianum* extract (GCE) on writhing reflex of mice in the writhing test. When mice were given acetic acid (1%, 10 mL/kg body weight) intraperitoneally, the writhing number was counted immediately for 15 min. The experiment was repeated twice. IMC is the abbreviation for indomethacin. Data are presented as mean \pm SD, n = 10. **P < 0.01, significance versus control.

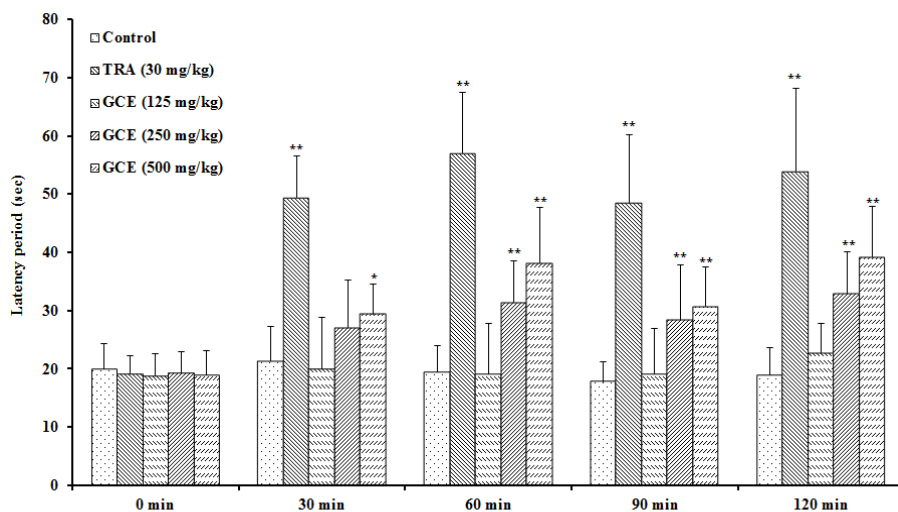


Figure 3 Effects of *Geranium Carolinianum* extract (GCE) on heat-stimulated mice in the hot-plate test. The latency period was recorded when female mice were put on the hot plate maintained at $55 \pm 0.5^{\circ}\text{C}$. TRA is the abbreviation for tramadol. Data are presented as mean \pm SD, $n = 10$. * $P < 0.05$, ** $P < 0.01$, significance versus control.

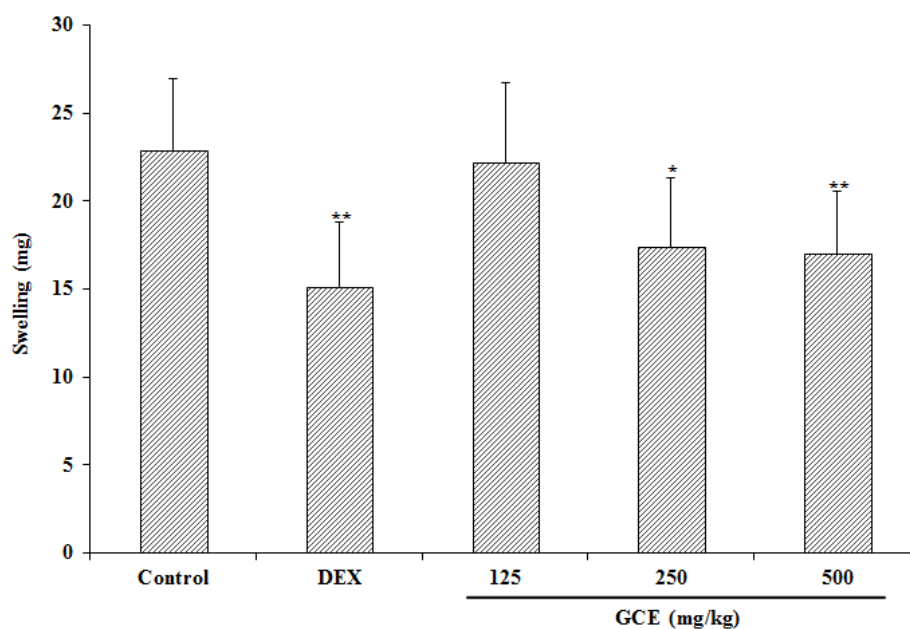


Figure 4 Influence of *Geranium Carolinianum* extract (GCE) on dimethylbenzene-induced ear edema in mice. Mice were given GCE or dexamethasone (DEX) orally prior to 1 h of topical application of dimethylbenzene (40 μ l/ear) to the right ears of mice. Data are presented as mean \pm SD, n = 10. *P < 0.05, **P < 0.01, significance versus control.