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An obesity-associated risk allele within the FTO gene affects human brain activity for areas important for emotion, impulse control and reward in response to food images.

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1 **Title**

2 An obesity-associated risk allele within the *FTO* gene affects brain activity for areas
3 important for emotion, impulse control, and reward in response to food images.

4

5 **Running Title**

6 FTO Associated Brain Activity

7

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40 **Number of:**

- 41 • Figures = 3
- 42 • Tables = 1
- 43 • Words:
 - 44 ○ Abstract = 197
 - 45 ○ Introduction = 565
 - 46 ○ Entire Manuscript (excluding references and figure legends) = 3,972

47

48 **Keywords:**

49 FTO, fMRI, SNP, obesity, food

50

51 **Conflict of Interest:**

52 The authors declare no conflicts of interest.

53

ABSTRACT

54 Understanding how genetics influences obesity, brain activity, and eating behavior will
55 add important insight for developing strategies for weight-loss treatment, as obesity may stem
56 from different causes and as individual feeding behavior may depend on genetic differences.
57 To this end, we examined how an obesity risk-allele for the *FTO* gene affects brain activity in
58 response to food images of different caloric content via fMRI. 30 participants homozygous
59 for the rs9939609 single nucleotide polymorphism were shown images of low- or high-calorie
60 food while brain activity was measured via fMRI. In a whole-brain analysis, we found that
61 people with the *FTO* risk-allele genotype (AA) had increased activity than the non-risk (TT)
62 genotype in the posterior cingulate, cuneus, precuneus, and putamen. Moreover, higher BMI
63 in the AA genotype was associated with reduced activity to food images in areas important for
64 emotion (cingulate cortex), but also in areas important for impulse control (frontal gyri and
65 lentiform nucleus). Lastly, we corroborate our findings with behavioral scales for the
66 behavioral inhibition and activation systems (BIS/BAS). Our results suggest that the two
67 genotypes are associated with differential neural processing of food images, which may
68 influence weight status through diminished impulse control and reward processing.

69

Keywords:

71 fMRI, FTO, SNP, BMI, food images, obesity

72

INTRODUCTION

73

74 Genetic differences may influence satiety and metabolism via effects in the brain
75 (Singh, 2014; Yeo, 2014). One such example is the influence of the *fat mass and obesity*
76 *associated (FTO)* gene on brain activity governing feeding behavior. Several different *FTO*
77 single nucleotide polymorphisms (SNPs) are associated with a higher body mass index (BMI)
78 (Sällman Almén et al., 2013; Scuteri et al., 2007a), and higher energy intake (Speakman,
79 2013). Current theory suspects that risk alleles of these SNPs may increase obesity via effects
80 in the brain to modulate feeding behavior and metabolism (Singh, 2014; Yeo, 2014).
81 Moreover, experiments in rodents show that changes in *FTO* expression levels in the
82 hypothalamus affect feeding behavior (Frederiksen, Skakkebaek, & Andersson, 2007;
83 Olszewski et al., 2009; Tung et al., 2010). However, whether the hyperphagia and subsequent
84 obesity induced in these experiments is due solely to metabolic changes or to manipulations
85 of the reward response to food is still controversial. Indeed, obesity may depend on
86 psychological characteristics, specifically those related to reward or self-control (Gerlach,
87 Herpertz, & Loeber, 2015). Personality scales for the Behavioral Inhibition System (BIS) and
88 Behavioral Activation System (BAS), which measure punishment and reward sensitivity
89 respectively, are two such tools which correlate with inactivity and poor diet (Carver &
90 White, 1994; Dietrich, Federbusch, Grellmann, Villringer, & Horstmann, 2014; Meule, 2013;
91 Voigt et al., 2009). Furthermore, some suggest obesity could be given a differential diagnosis
92 and treatment regimen based on this distinction between obesity caused by physiological
93 factors or by psychological effects (Yu et al., 2015). Thus, *FTO* could promote obesity
94 through metabolic effects, augmented reward signaling, or a combination of both.

95 Obesity is increasingly considered an addiction-type disorder with disruptions of the
96 reward pathway, and several fMRI studies have explored this topic (Batterink, Yokum, &
97 Stice, 2010; Brooks, Cedernaes, & Schiöth, 2013; Gearhardt AN et al., 2011; Goldstone et al.,
98 2009; Tomasi et al., 2015; Volkow, Wang, Tomasi, & Baler, 2013; Zhang et al., 2015). To
99 date, however, few fMRI studies have examined how genetic profile is associated with brain
100 responses to food in obesity. A recent study found that people with the *FTO* risk allele for
101 rs8050136 had reduced activity in the right prefrontal cortex while viewing food images in a
102 postprandial state, but not while fasting (Heni et al., 2014). Another study found that fasted
103 participants with the risk allele for rs9939609 had lower activations in areas important for
104 both reward and metabolism when shown food images, such as the hypothalamus, ventral
105 tegmental area and substantia nigra, posterior insula, globus pallidus, thalamus, and
106 hippocampus. They also showed that circulating ghrelin may affect brain areas important for
107 both metabolism and reward signaling depending on genotype (Karra et al., 2013). Notably,
108 their cohort of participants had a normal BMI with no obese participants. Thus, possible
109 interactions between genotype and BMI on brain activity remain unstudied.

110 Against this background, we explore for the first time the association between *FTO*
111 genotype, BMI, and neural responses to food images of either low- or high-calorie content.
112 In whole-brain analysis of BOLD responses to food images, we test whether the genotype
113 homozygous for the at-risk allele for rs9939609 (which is A) (Dina et al., 2007; Frayling et
114 al., 2007; Scuteri et al., 2007b) affects brain activity differently from the homozygous
115 genotype with the non-risk allele (T). We also examine within-group regressions between
116 BMI and BOLD responses to food images within each genotype and behavioral
117 characteristics of each genotype.

118

METHODS

119

120 *Participants*

121 Prior to any experimental procedures, all participants gave written informed consent to
122 the study which conformed to the Declaration of Helsinki and approved by the local ethics
123 committee. Participants were 30 right-handed, northern-European males, with a mean age of
124 26 ± 1 years, recruited locally in Uppsala, Sweden by advertisement. Genotyping of the *FTO*
125 single nucleotide polymorphism (SNP) rs9939609 was performed with a pre-designed
126 Taqman single-nucleotide polymorphism genotyping assay (Applied Biosystems, Foster City,
127 USA) and an ABI7900 genetic analyzer with SDS 2.2 software at the Uppsala Genome Center
128 (<http://www.genpat.uu.se/node462>). The genotype call rate was 97.8%. Only homozygous
129 participants were included in the study. There was a similar distribution of body mass index
130 (BMI) for each genotype of the rs9939609 SNP: 26.8 ± 1.2 , with a range of 13.1 kg/m^2 in the
131 AA genotype ($n = 13$), and 24.1 ± 0.7 , with a range of 9.7 kg/m^2 in the TT genotype ($n = 17$).
132 Hunger ratings were also assessed on a 1-10 scale with higher numbers indicating greater
133 feelings of hunger.

134

135

136 *Behavioral Questionnaires*

137 Clinical measures for punishment sensitivity and reward-seeking behavior were
138 acquired using the Behavioral Inhibition System (BIS) and Behavioral Activation System
139 (BAS) questionnaires (Carver & White, 1994). Both consisted of 24 items. Each item was
140 represented by a statement, where the participant indicated how much s/he agreed or

141 disagreed on a four-point scale. The BIS included only one scale, evaluating the reactions to
142 the anticipation of punishment and anxiety, while the BAS included three subscales: Drive,
143 which represents the pursuit of desired goals; Fun Seeking, which evaluates the desire for new
144 rewards and impulsivity; and Reward Responsiveness, which focuses on positive reactions
145 anticipating rewards.

146

147 *fMRI Paradigm and Image Acquisition*

148 Images of food were presented in the scanner using MRI-compatible goggles
149 (NordicNeuroLab, Bergen, Norway) attached to the headcoil. The food images were shown
150 at 3 second intervals and no images were repeated. Participants were instructed to imagine
151 what it felt like to eat the food presented. Blood oxygen-level dependent (BOLD) signals in
152 response to food stimuli were measured as participants were shown images of low-calorie
153 (LC) food, high-calorie (HC) food, or control (C) images in five cycles of the following block
154 design pattern: C, LC, C, HC. Each block contained 6 images. In total, the number of
155 measurements collected for each group was 54 for control, 30 for LC, and 30 for HC. Low
156 and high calorie food images were determined by caloric content and selected for familiarity
157 according to local palate, and were controlled for visual features (color, size, etc.). Caloric
158 content of the HC and LC images was confirmed by their perceived caloric content in focus
159 groups representative of the population to be studied. Example HC images were cakes, pies,
160 ice cream, candy, fried foods, and hamburgers. Example LC images were vegetables, fruits,
161 and salads without high-calorie dressing. Control images were a grey screen with a crosshair
162 in the center.

163 Participants were scanned at 08:00 following an overnight fast of at least 8 hours, i.e.,
164 were in a fasted state during testing. Structural and functional brain images were acquired

165 with a Philips 3-Tesla (Achieva, Philips Healthcare, Best, Netherlands) using a standard head
166 coil. 125 volumes were registered during the T2*-weighted echo-planar imaging (EPI)
167 sequence with whole brain coverage of 30 slices (slice thickness = 3 mm; 1 mm gap,
168 interleaved scan order, in-plane resolution: (3 mm x 3 mm), repetition time (TR) = 3 s; echo
169 time (TE) = 35 ms, flip angle = 90°).

170

171 *Preprocessing of fMRI data*

172 All preprocessing steps were performed using software package Statistical Parametric
173 Mapping (SPM, version 8, <http://www.fil.ion.ucl.ac.uk/spm/>), implemented in MATLAB
174 (version R2014a, 11 FEB 2014, 8.3.0.532, 64-bit). The images were realigned and estimated
175 to remove movement artefacts in the data. EPI images were further matched with the
176 structural image using coregistration. The anatomical image was segmented to strip away
177 unnecessary tissue in the images. Tissue probability maps were introduced in the
178 segmentation step to differentiate between gray matter, white matter and cerebrospinal fluid in
179 each individual. Volume was calculated based on the extracted tissue maps of each subject.
180 The gray matter and white matter volumes were added together to find the total brain volume
181 (TBV), to serve as a nuisance covariate in the analyses. Then, functional images were
182 normalized to fit the segmented anatomical image. Finally, images were smoothed using a
183 Gaussian function (8 mm full-width, half-maximum (FWHM)) to minimize noise and bias.

184

185 *Statistical Analysis*

186 All fMRI statistical analysis was performed using the same versions of SPM and
187 MATLAB listed in preprocessing steps. For all whole-brain results, a family wise error

188 (FWE) corrected significance level was set at $p < 0.05$ to correct for multiple testing. For the
189 main analysis of the imaging data, we tested a contrast between the neural responses to
190 images of high calorie (HC) foods to the neural response of images from low calorie (LC)
191 foods. This contrast was then tested using a between-groups t-test followed by directional
192 post-hoc comparisons as well as with a multiple regression analysis testing for interactions
193 between genotype and BMI, BIS, or BAS individually. The t-test used both BMI and total
194 brain volume (TBV, grey matter + white matter) as covariates of no interest, while the
195 multiple regression analysis used only TBV as a covariate of no interest.

196 Region of interest (ROI) analysis was performed by preparing masks of brain areas
197 based on previous MRI studies on *FTO* (de Groot et al., 2015; Karra et al., 2013). Bilateral
198 masks of such areas were produced using the Wake Forest University Pickatlas toolbox
199 (Maldjian, Laurienti, Kraft, & Burdette, 2003) within SPM. Small volume corrections were
200 performed on clusters trending towards significance using a 6 mm radius sphere over the
201 greatest FWE-corrected suprathreshold voxel.

202 For the within-group regression analyses, the same contrast of HC versus LC was used
203 and TBV served as a covariate of no interest. For both genotypes, positive or negative
204 correlations between BMI and BOLD signal were tested.

205 Other statistical tests, as well as the principle component analysis (PCA) were
206 performed with R statistical software (version 3.2.1, 64-bit) using the FactoMineR package
207 (Lê et al., 2008). Results for the PCA were considered significant if the percentage of inertia
208 summing from the two largest eigenvalues exceeded values listed in a significance table based
209 on 10,000 analyses with similar numbers of individuals and independent variables (Lê et al.,
210 2008).

211

RESULTS

212

213 **The obesity-associated *FTO* SNP rs9939609, is associated with increased activity in**
214 **response to food images.**

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First, we tested for a difference in the processing of food stimuli as measured by fMRI depending on genotype and/or body-mass index (BMI). We tested 30 participants (13 AA and 17 TT) in a fasted state. BMI for the AA genotype was significantly higher than the non-risk TT genotype (Student's t-test, $p < 0.05$), as expected from previous reports (Fredriksson et al., 2008; Sällman Almén et al., 2013). There was no significant difference in age between groups of differing genotype (Student's t-test, $p = 0.7$; Table 1). BOLD signals were measured as participants were shown images of low-calorie (LC) food, high-calorie (HC) food, or control images in a block design format. Patients were instructed to imagine the feeling of eating the food presented. We compared one contrast in our analysis: HC versus LC, which tests the neural response discriminating food images of different caloric contents.

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Between-group comparisons found three clusters with greater activity in the AA genotype compared to the TT genotype. (Figure 1A). Areas included the posterior cingulate cortex (PCC), cingulate gyrus, cuneus, and precuneus (Table 2). A multiple regression analysis found an interaction between genotype and BMI, post-hoc comparisons found significant clusters for the AA genotype while BMI was decreasing in the PCC, cingulate gyrus, middle occipital gyrus, and precuneus (Supplementary Table 1).

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Based on results from previous studies (de Groot et al., 2015; Karra et al., 2013), we performed a region-of-interest (ROI) analysis on areas putatively associated with *FTO* genotype: such as the insula, putamen, and nucleus accumbens. Within the t-test comparison between genotypes, a significant cluster showing greater activity in the AA genotype was

235 found in the putamen (Figure 1B) after performing a small-volumes correction using a 6 mm-
236 radius sphere over the lowest FWE-corrected p-value in the cluster (Table 2).

237

238 **Differential patterns of neural activation in each *FTO* genotype depending on body-mass**
239 **index.**

240 We then tested regressions of BMI and whole brain activity within each genotype. We
241 tested both positive and negative regressions in the HC versus LC contrast. Two regressions
242 yielded results which survived corrections for multiple testing. In addition, there was almost
243 no overlap between the two patterns of activity, with only a small area on the motor cortex
244 showing overlap of clusters from each genotype, neither of which survived corrections for
245 multiple testing (Figure 3A). In the AA genotype, there was a negative correlation between
246 caloric discrimination (HC versus LC) and BMI (Figure 3B) in the following brain areas:
247 frontal gyrus (medial and superior), lentiform nucleus, cerebellum (declive, uvula, and
248 pyramis), and cingulate gyrus (Table 3); i.e., overweight AA participants had weaker
249 responses to HC images than LC, while normal-weight AA participants had stronger
250 responses to HC images compared to LC. However, in the TT genotype, there was a positive
251 relationship between caloric discrimination and BMI (Figure 3C), but only in a cluster
252 containing the superior temporal gyrus (Table 4). Thus, the two genotypes have a divergence
253 in the neural processing of caloric discrimination to food based on BMI.

254

255 **Differential patterns of behavior for each *FTO* genotype depending on body-mass index.**

256 We next tested if behavioral questionnaires corroborated the findings from the imaging
257 experiments. We employed two questionnaires: the Behavioral Inhibition System (BIS) and

258 Behavioral Activation System (BAS) questionnaires, which measure punishment and reward
259 sensitivity, respectively (Carver & White, 1994). We choose these questionnaires as they
260 represented the main elements of personality that we wished to explore in this study. We then
261 performed a principle component analysis within each genotype using the BIS and the three
262 BAS subscales (Drive, Fun Seeking, and Reward Responsiveness) as variables of interest
263 with BMI as a quantitative supplementary variable. For both analyses, all the variables of
264 interest were well projected and the first two dimensions accounted for $\approx 80\%$ of the variability
265 (considered significant based on criteria listed in methods under statistical analysis
266 subheading, $81.4 > 80.0$ for the AA group and $79.2 > 76.5$ in the TT group). Moreover, the
267 variables of interest projected to the same quadrants except for the Drive and Fun Seeking
268 subscales, which were switched between the two different genotypes. Interestingly, the
269 supplementary variable, BMI, was projected in opposite quadrants relative to the BIS
270 depending on genotype. In the AA genotype, BMI correlated positively with the BAS Drive
271 subscale and negatively with BIS, while in the TT genotype, BMI correlated somewhat
272 positively with BIS and negatively with the BAS Fun Seeking subscale. Furthermore, we
273 followed up the association between the BIS and BMI using a multiple regression analysis
274 testing if BIS scores could be predicted by genotype, BMI, or their interaction. This model
275 found significance for the genotype ($p < 0.01$) and interaction terms ($p < 0.01$). Thus, the two
276 genotypes appear to have differing behavioral measures for impulsivity, punishment, and
277 reward depending on BMI.

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DISCUSSION

We examined whether an obesity-associated genotype affects the neural processing of food images with different caloric content and to what extent body-mass index (BMI) is an important factor. We tested 30 male participants homozygous for either the risk (A)- or non-risk (T) allele of the *fat mass and obesity associated (FTO)* SNP rs9939609 (13 in the AA group and 17 in the TT group). Participants were shown images of low-calorie (LC) food, high-calorie (HC) food, or control images while blood-oxygen-level dependent (BOLD) signals were obtained via fMRI. We found the AA genotype had increased brain activity compared to the TT genotype when viewing food images with different caloric contents, specifically in areas important for emotion (cingulate gyrus), memory, and self-image (cuneus and precuneus) and reward (putamen). Moreover, while both genotypes had a similar distribution of BMI, ranging between normal-weight to obese in each genotype, we found that BMI was associated with differential activity within each genotype – with comparatively more brain regions associated with BMI in the AA genotype than in the TT genotype. Thus, discrimination between HC and LC foods may be handled differently for each genotype depending on BMI. Next, we corroborate our findings in the imaging study with personality questionnaires examining behavioral characteristics related to impulsivity and reward-processing: namely the Behavioral Inhibition System (BIS) and Behavioral Activation System (BAS) scales. We found that the BIS as well as subscales of the BAS correlated with BMI oppositely in each genotype. Thus, obesity within each genotype may stem from different behavioral characteristics. Our results suggest that this polymorphism of the *FTO* gene affects the processing of visual food stimuli differently, possibly involving emotion, self-image, and reward processing.

302 In between-groups comparisons, as well as multiple regression analysis, we found
303 significant clusters of brain activity when testing a contrast for caloric discrimination (HC
304 food images opposed to LC food images). Specifically, we found increased neural activation
305 in the AA genotype compared to the TT genotype within the posterior cingulate cortex (PCC),
306 cingulate gyrus, cuneus and precuneus. The PCC is a well-connected and multifunctional
307 brain area associated with emotional processing, and a central node in the default mode
308 network (DMN): involved in arousal/awareness, balancing external and internal thought, and
309 emotion (Leech & Sharp, 2014). Decreased activity in the PCC is associated with low levels
310 of arousal (Fiset et al., 1999). The cuneus and precuneus are also implicated in obesity, as a
311 previous study showed that obese adolescents had less activation in the cuneus and precuneus
312 when viewing food versus non-food commercials (Gearhardt, Yokum, Stice, Harris, &
313 Brownell, 2014). Thus, there may be a difference in the salience of food images depending
314 on *FTO* genotype, and perhaps an increased salience of HC food images for people with the
315 AA genotype leads to an increase in food intake.

316 Participants with the AA genotype also had increased activity in the putamen compared
317 to TT participants in a region-of-interest analysis. The putamen has an important functional
318 role in reward processing (Delgado, 2007), and several imaging studies have shown increased
319 activity for this structure in obese participants (Boutelle et al., 2015; Jastreboff et al., 2014;
320 Zhang et al., 2015). Thus, in our study, finding increased activity in the putamen for the risk
321 allele (AA) is a supportive finding, as both are associated with obesity. This also further
322 implicates the involvement of the reward system in obesity with the *FTO* risk allele.

323 Other studies have examined participants of different genetic background for the *FTO*
324 gene using MRI techniques (de Groot et al., 2015; Heni et al., 2014; Karra et al., 2013), but no
325 study has yet included obesity as a factor. However, we only partially replicated previous
326 findings (Karra et al., 2013). For example, for their cohort in a fasted state, they reported

327 reduced activity in the AA versus TT genotype in the hypothalamus, globus pallidus,
328 thalamus, ventral tegmental area, and substantia nigra (Karra et al., 2013). This difference in
329 results is perhaps because the BMI in our cohort was much more variable, whereas
330 overweight and obese participants in their previous study were excluded. Furthermore,
331 others suggest that BMI should be regarded as a confounding factor (Cole et al., 2013).
332 However, the variability of BMI in our cohort allowed us to perform an analysis with
333 regressions between BMI and BOLD responses. BMI appears to be an important factor in
334 studies of this type, and our results suggest that the neural processing of food images between
335 genotypes may be differentially affected as BMI increases.

336 Next, we further explored the neural response to caloric discrimination of food images
337 based on BMI. We find that BMI augments the neural response to food images differently
338 within each genotype. As BMI decreased in the AA genotype, neural activity increased in the
339 superior and medial frontal gyrus, lentiform nucleus, cingulate gyrus, middle temporal gyrus,
340 and cerebellar areas: declive, uvula, and pyramis. Thus, the overweight AA participants have
341 less activity in these areas. And, in line with our results, a previous fMRI study also showed
342 reduced activation in the superior and middle frontal gyri as BMI increased (Batterink et al.,
343 2010). Thus, the AA genotype again corresponds with previously-established brain activity in
344 the obese/overweight. Our findings of altered activity among AA participants in cerebellar
345 areas are also pertinent, given the relatively recent association with the cerebellum to emotion
346 (Schienle & Scharmüller, 2013) and appetitive processes (Zhu & Wang, 2007). As for the TT
347 genotype, we found two areas where neural activity positively correlated with BMI: the
348 superior and middle temporal gyri. These affected areas are perhaps due to functional
349 relationships within the default mode network and the temporal lobe network, where
350 functional connectivity is augmented in the obese (Kullmann et al., 2012). From the above,
351 and similar to (Karra et al., 2013), we find a divergence in the processing of food images

352 between the two genotypes, and our results highlight the importance of BMI as a factor
353 mediating the effect of genotype on neural responses to images of food. Thus, obese as well
354 as normal-weight individuals may be expected to process food images differently depending
355 on their genotype.

356 Lastly, we corroborate our findings in the imaging study with personality measures
357 from the BIS and BAS scales, as obesity is associated with anomalies in both (Carver &
358 White, 1994; Dietrich et al., 2014; Meule, 2013; Voigt et al., 2009). Higher scores of the BIS
359 are indicative of increased sensitivity to punishment, and correlate positively with such
360 personality measures as harm avoidance, reward dependence, susceptibility to punishment,
361 negative affect, and anxiety, and correlate negatively with optimism and socialization.
362 Whereas higher scores of the BAS and its subscales, are indicative of greater reward
363 sensitivity, and correlate positively with extraversion, novelty seeking, and positive affect,
364 and correlate negatively with harm avoidance, and susceptibility to punishment (Carver &
365 White, 1994). The two genotypes had opposing relationships with BMI and the BIS score.
366 Specifically, BMI in the AA genotype correlated negatively with BIS, while BMI in the TT
367 genotype correlated positively with BIS. This suggests that the overweight/obese people with
368 the AA genotype have less inhibitory personality characteristics than the overweight/obese
369 with the TT genotype. Thus, the AA genotype in our cohort confirms previous reports
370 equating impulsivity with obesity/overeating (Meule, 2013) specifically in one study which
371 also found a negative correlation between BIS and BMI in males (Dietrich et al., 2014).
372 Subscales of the BAS also had different correlations with BMI in each genotype: the Drive
373 subscale correlated positively with BMI in the AA genotype, while the Fun Seeking subscale
374 correlated negatively in the TT genotype. The correlation between the Drive subscale and
375 BMI in the AA genotype corroborates with previous associations with BAS, arousal, and
376 overeating (Voigt et al., 2009) and also a previous fMRI report associating the Drive subscale

377 with neural activity in response to appetitive food images (Beaver et al., 2006). Thus, people
378 in the TT genotype may possess a reward processing that is optimal for maintaining or
379 decreasing weight, while the AA genotype has a more impulsive attitude and matches
380 traditional descriptions of obesity and unhealthy eating behavior. Indeed, the same *FTO*
381 variant modulated reward processing and avoidance learning (Sevgi et al., 2015). However,
382 we were unable to establish effects of BIS or BAS or interactions with genotype using the
383 brain-imaging data. Thus, while this finding is intriguing, we caution that it requires further
384 validation.

385 In summary, we show that a genotype homozygous for an *FTO* risk allele for obesity is
386 associated with increased neural response to HC versus LC food images in brain regions
387 associated with emotion, impulsivity, and reward compared to the low-risk for obesity TT
388 genotype, and that BMI is a mediating factor for each genotype. Our results suggest that
389 overweight/obese people with the AA genotype may be prone to unhealthy eating behavior
390 due to food images being less salient at evoking normal appetitive responses compared to
391 overweight/obese people with the TT genotype. Some limitations of this study are the use of
392 only males in the fMRI experiments, as well as the exclusion of the AT genotype. We
393 maintain that BMI is an important factor in fMRI research as well as in the relationship
394 between brain activity and genotype. In conclusion, our findings offer insight into the
395 relationship between *FTO*, obesity, and brain activity; and suggest that overweight/obese
396 populations have different attitudes and functional processing for food images depending on
397 genetic background.

398

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ACKNOWLEDGEMENTS

537 This study was supported by the Swedish Research Council and the Swedish Brain Research

538 foundation.

539

FIGURE LEGENDS

540

541 **Figure 1. Participants homozygous for the *FTO* risk-allele (rs9939609) have increased**
542 **brain activity in response to high- and low-calorie food images.** 13 participants
543 homozygous for the *FTO* risk allele (A) and 17 for the non-risk allele (T) were shown images
544 of either low-calorie (LC), high-calorie (HC) food, or control while brain activations were
545 measured via fMRI. A whole-brain, between-groups comparison tested the neural response to
546 foods of different caloric content (HC versus LC contrast) using total brain volume (TBV)
547 and body-mass index (BMI) as covariates of no interest. Greater activity was found in four
548 clusters for participants homozygous for the risk allele (A) compared to the normal allele (T).
549 **A)** Axial brain slices showing significant clusters. Exact MNI z-coordinate given below
550 slices in white numbers, and t-statistics indicated by color-coded scale bar (critical $t = 3.5$).
551 **B)** A region of interest analysis found a significant cluster within the putamen after a small-
552 volumes correction.

553

554 **Figure 2. Opposite patterns of activity in each *FTO* genotype depending on body-mass**
555 **index.** For each genotype, the caloric discrimination to food was tested in a contrast between
556 the BOLD response to high-calorie versus low-calorie food images in regressions with body-
557 mass index (BMI). A negative relationship was found between brain activity and BMI in
558 participants with the *FTO* risk genotype (AA), while a positive relationship was found in the
559 non-risk genotype (TT). Patterns of activity were mostly independent of each other, with only
560 a small area in motor cortex overlapping between the two. **A)** Color-coded activity for each
561 genotype (AA in magenta and TT in cyan) imposed on a 3D rendering of the brain.
562 Overlapped activity between genotypes shown as white. **B & C)** Threshold images of brain

563 activity for AA and TT genotype respectively. Axial brain images are shown with respective
564 MNI z-coordinated displayed below and t-statistics indicated by color-coded scale bar (critical
565 $t = 4.1$ for AA, and 3.8 for TT).

566

567 **Figure 3. The AA genotype displays a different balance of behavioral inhibition and**
568 **activation systems depending on body-mass index compared to the TT genotype.**

569 Participants completed the Behavioral Inhibition System (BIS) and Behavioral Activation
570 System Questionnaires (BAS) questionnaires. The BIS evaluates inhibitory behavior in the
571 anticipation of punishment and anxiety, while the BAS included three subscales: Drive, which
572 represents the pursuit of desired goals; Fun Seeking, which evaluates the desire for new
573 rewards and impulsivity; and Reward Responsiveness, which focuses on positive reactions
574 anticipating rewards. Principle component analysis was used to compare the relationships of
575 these variables along with BMI in the two different genotypes. The behavioral variables were
576 all well projected in each group. Body-mass index (BMI) was used as a supplementary
577 variable (not included in the analysis, but still plotted to evaluate which variables it correlated
578 with). Interestingly, for each genotype, BMI was projected in opposite quadrants relative to
579 the BIS. The subscales for the BAS were also differently projected relative to BMI for each
580 genotype. **A)** variables factor map for the AA genotype. BMI was negatively correlated with
581 BIS and positively correlated with the BAS Drive subscale. **B)** variables factor map for the
582 TT genotype. BMI was positively correlated with BIS and negatively correlated with the
583 BAS Fun Seeking subscale.