1	Cytokine responses to repeated, prolonged walking in lean versus
2	overweight/obese individuals
3	RUNNING TITLE: Cytokine responses to repeated exercise
4	
5	REBECCA J.H.M. VERHEGGEN, MD, MSC ¹
6	THIIS M.H. EIJSVOGELS, PHD ^{1,3}
7	MILÈNE CATOIRE, PHD ¹
8	RIENEKE TERINK, MSC ⁴
9	ROB RAMAKERS, BSC ¹
10	COEN C.W.G. BONGERS, MSC ¹
11	MARCO MENSINK, MD, PHD ⁴
12	AD R.M.M. HERMUS, MD, PHD^2
13	DICK H.J. THIJSSEN, PHD ^{1,3}
14	MARIA T.E. HOPMAN, MD, PHD ¹
15	Radboud university medical center, Radboud Institute for Health Sciences, Nijmegen, the Netherlands;
16	¹ Department of Physiology, ² Department of Internal Medicine, Division of Endocrinology; ³ Research
17	Institute for Sport and Exercise Sciences, Liverpool John Moores University, Liverpool, United
18	Kingdom; ⁴ Wageningen University, Division of Human Nutrition, Wageningen, the Netherlands
19	Total Word Count: 2987
20	Word Count Abstract: 228
21	Total Number of Figures: 1
22	Total Number of Tables: 1
23	Author for correspondence:
24	Rebecca Verheggen, MD, Dept. of Physiology (392), Radboud university medical center, Radboud
25	Institute for Health Sciences, PO Box 9101, 6500 HB Nijmegen, The Netherlands
26	e-mail: rebecca.verheggen@radboudumc.nl
27	Tel. (+31) (0)24 36 14906
28	Fax. (+31) (0)24 354 0535

30 Abstract **Objectives.** Obesity is characterized by a pro-inflammatory state, which plays a role in pathogenesis 31 32 of metabolic and cardiovascular disease. An exercise bout causes a transient increase in proinflammatory cytokines, whilst training has anti-inflammatory effects. No previous study examined 33 whether the exercise-induced increase in pro-inflammatory cytokines is altered with repeated 34 35 prolonged exercise bouts and whether this response differs between lean and overweight/obese 36 individuals. **Design.** Lean (n=25, BMI 22.9±1.5kg/m²) and age-/sex-matched overweight/obese (n=25; BMI 37 38 27.9±2.4kg/m²) individuals performed walking exercise for 30, 40 or 50 km per day on four 39 consecutive days (distances similar between groups). 40 **Methods**. Circulating cytokines (IL-6, IL-10, TNF- α , IL-1 β and IL-8) were examined at baseline and 41 <30 minutes after the finish of each exercise day. **Results**. At baseline, no differences in circulating cytokines were present between groups. In response 42 to prolonged exercise, all cytokines increased on Day 1 (IL-1β: P=0.02; other cytokines: P<0.001). IL-43 44 6 remained significantly elevated during the 4 exercise days, when compared to baseline. IL-10, TNF-45 α, IL-1β and IL-8 returned to baseline values from exercise day 2 (IL-10, IL-1β, IL-8) or exercise day 3 (TNF-α) onward. No significant differences were found between groups for all cytokines, except IL-46 8 (Time*Group Interaction P=0.02). 47 48 Conclusion. These data suggest the presence of early adaptive mechanisms in response to repeated prolonged walking, demonstrated by attenuated exercise-induced elevations in cytokines on 49 50 consecutive days that occurs similar in lean and overweight/obese individuals. 51 **Keywords:** obesity, inflammation, training, adaptive response 52

53

Introduction

In individuals with obesity, a chronic state of low grade-inflammation is present which is characterized by elevated circulating levels of cytokines.¹ This chronic inflammation is associated with the pathogenesis of cardiovascular and metabolic diseases, which are strongly associated with obesity.^{2, 3} Exercise training represents a potent non-pharmacological intervention with strong anti-inflammatory effects, leading to lower levels of circulating pro-inflammatory cytokines and increased expression of anti-inflammatory cytokines.⁴ Paradoxically, an acute exercise bout elicits a pro-inflammatory response, characterized by a transient rise of pro-inflammatory cytokines.^{5, 6} The response of cytokines to acute exercise seems dose-dependent, as higher cytokine levels are observed after exercise of higher intensity and/or longer duration.⁶ To support these observations, flu-like symptoms have been reported in relation to an exhaustive acute exercise bout, such as a marathon, which are accompanied by a (transient) rise in circulating cytokines.⁶ Even exercise bouts of lower intensity have shown to cause a rise in pro-inflammatory cytokines.⁵

The pro-inflammatory effects of acute exercise *versus* the anti-inflammatory effect of regular exercise training imply the presence of an adaptive mechanism. Repeated exposure to the pro-inflammatory effects of acute exercise may induce an adaptive response, leading to an attenuated exercise-induced release of cytokines, as was previously demonstrated for Interleukin-6 (IL-6) in trained cyclists performing repeated exercise bouts of prolonged duration and moderate intensity (~72% of maximal heart rate). In recent years an increasing number of voluntary exercise events, characterized by repeated prolonged exercise on consecutive days (e.g. walking, swimming, hiking, cycling), is organized. Since the release of cytokines in response to acute exercise seems to increase with longer duration and higher intensity, it is highly relevant to examine physiological responses of cytokines to repeated prolonged exercise during such events.

Obesity is characterized by the presence of low grade inflammation.³ Accordingly, the acute changes in cytokines in response to prolonged exercise may be affected in overweight individuals because of the presence of higher circulating cytokine levels in resting conditions.

Therefore, the aim of this study is to examine differences in the effect of repeated moderate-intensity prolonged exercise (i.e. prolonged walking 30, 40 or 50km on four consecutive days during the Nijmegen Four Day Marches, a voluntary walking event) on circulating cytokine levels (IL-6, IL-10, Tumor necrosis factor (TNF)-α, IL-1β, and IL-8) and between lean and overweight/obese individuals. We hypothesize that the presence of low-grade inflammation at baseline in overweight/obese subjects leads to exaggerated increases in pro-inflammatory cytokines in response to prolonged exercise when compared to lean individuals.

Methods

A total of 50 adult participants of the Nijmegen Four Days marches were included. Subjects were recruited form a cohort of participants in the Nijmegen 4 Day Marches that filled out a questionnaire as part of the Nijmegen Exercise Study. Subjects with a chronic inflammatory disease (e.g. inflammatory bowel disease, rheumatoid arthritis) and participants that used anti-inflammatory drugs (non-steroidal anti-inflammatory drugs, corticosteroids) were excluded from participation since these conditions can cause a change in circulating cytokines independent from overweight/obesity. All participants completed a distance of 30, 40 or 50 km per day on four consecutive days at a self-selected pace. Every participant was assigned to an individual distance (30, 40 or 50 km) and completed the same distance on the four consecutive exercise days. To answer our research question, subjects were allocated either to a lean (BMI <25 kg/m²) or overweight/obese (BMI ≥25 kg/m²) cohort. Furthermore, subjects were individually matched based on age, sex and walking distance and were selected for recruitment accordingly. Since exercise intensity is known to influence cytokine levels, participants were also matched based on exercise intensity, calculated based on individually recorded heart rate during the walking event. Written informed consent was obtained from all participants prior to the start of the study. This study was approved by the Medical Ethical Committee

of the Radboud University Nijmegen Medical Center, and was conducted in accordance with the declaration of Helsinki. Baseline data (subject characteristics and blood sample; day 0) were collected 1 or 2 days prior to the start of the event after a minimum resting period of 24 hours. During Day 1, exercise intensity was assessed with the use of a 2-channel chest band system (Polar Electro Oy, Kempele, Finland). At baseline, height and body weight (Seca 888 scale, Hamburg, Germany) were measured to calculate body mass index (BMI). Waist and abdominal circumference were measured with a measuring tape (Seca 201, Chino, USA) to calculate waist-to-hip ratio. A four-point skinfold thickness measurement (biceps, triceps, sub-scapular, supra-iliac) was obtained by a well-trained, experienced researcher to calculate the body fat percentage as previously described. Resting heart rate and blood pressure were measured in supine position, after a 5 minute rest period. Habitual daily energy intake, macronutrient and anti-oxidant intake were assessed with use of an online validated 180-item semi-quantitative Food Frequency Questionnaire (FFQ).⁹ The FFQ reference period was one month, and portion sizes were estimated using standard portions. Intake of total energy and nutrients was calculated using the Dutch Food Composition Database. 10 At baseline, physical activity levels were assessed with the use of the Short QUestionnaire to ASsess Health-enhancing physical activity (SQUASH), a validated tool to assess physical activity levels in the Dutch population.¹¹

129 130

131

132

133

107

108

109

110

111

112

113

114

115

116

117

118

119

120

121

122

123

124

125

126

127

128

Heart rate was measured with a 2-channel ECG chest band system (Polar Electro Oy, Kempele, Finland) at every 5 km point during Day 1. Exercise intensity was calculated for each measurement by dividing the mean heart rate during exercise by the maximal predicted heart rate (208-0.7*age). ¹² By calculating the mean of these percentages of maximal heart rate, the mean intensity for the exercise bout was recorded for each participant.

Venous blood was sampled at baseline (between 9.00 AM to 4.00 PM after a minimum resting period of 24 hours) and at each walking day within 30 minutes after completion of the exercise bout by venepuncture. Blood was centrifuged at 3000 RPM for 15 minutes and plasma was stored at -80°C until analysis. Cytokines (IL-6, IL-10, TNF- α , Il-1 β and IL-8) were simultaneously analyzed using the ultrasensitive MesoScale Discovery (MSD) QuickPlex SQ 120 Instrument with Multi-spot assay (Human Proinflammatory Panel 1, K15049D, MSD) according to the manufacturer's recommendations. The lower detection limit varied per plate and was 0.029–0.159 (IL-6), 0.025–0.051 (IL-8), 0.021–0.042 (IL-10), 0.008–0.061 (IL-1 β), and 0.034–0.079 (TNF- α) pg/ml. 34 of the 250 (<15%) samples for IL-1 β were below the lower detection limit. These samples were excluded from further analysis. The other cytokines were all above the detection limit.

Statistical Analysis. Data were checked for normality with use of the Shapiro-Wilk test and visual inspection of Q-Q plots. Baseline characteristics were normally distributed and therefore assessed with use of a one-way ANOVA. Cytokine data that was not normally distributed was transformed with use of square root transformation (IL-6 and TNF- α) or inverse transformation (IL-10). Cytokine data were analyzed using a time (exercise day) X group (lean vs. overweight) linear mixed model analysis. Post hoc analysis (Bonferroni) per group was performed when a significant effect was found. The level of statistical significance was defined at α =0.05. Data are presented as mean±SD, unless stated otherwise. The statistical analyses were conducted in SPSS 25 (Statistical Package for Social Sciences 25.0, SPSS Inc., Chicago, Illinois, USA)

Results

Subject characteristics are presented in Table 1. We found significant differences between the lean and overweight/obese subgroups for weight, BMI, body fat percentage and waist-hip-ratio, whilst no differences in age and sex distribution were present due to selective matching. Furthermore, the groups

reported comparable habitual physical activity levels, daily energy intake and intake of macronutrients and anti-oxidants (Table 1).

All subjects successfully completed the four exercise days. No group differences were present for exercise intensity and exercise duration (Table 1). At baseline, circulating levels of IL-6, IL-10, IL-8, IL-1β and TNF-α were not significantly different between the lean and overweight groups (Figure 1).

Repeated prolonged exercise resulted in a significant change of all cytokines (Figure 1). For all cytokines, except for IL-8 (interaction effect P=0.02), we found no differences in the post-exercise levels between lean and overweight/obese subjects (all P>0.05, Figure 1). Specifically, IL-6 showed a significant increase that remained elevated on all exercise days (P<0.001), with no differences between groups. In contrast, IL-10 increased significantly on exercise day 1(lean group: P = 0.005; overweight/obese group: P=0.003), but post-exercise levels were similarly declined to baseline on subsequent exercise days in both groups (interaction-effect P>0.05). For TNF- α , a significant effect of exercise was only present at exercise day 1 and 2 in the overweight/obese group (P<0.001 day 1, P = 0.02 day 2), whilst the lean group exhibited no change after exercise on any of the exercise days. IL-1 β was significantly higher on day 1 (P=0.04) in the overweight/obese group, whilst no post-exercise increases were found in the lean group. For IL-8 a significant Time*Group Interaction effect (P=0.02) was found. Both groups showed an increase in IL-8 on day 1 that returned to baseline on subsequent days. The lean group demonstrated a significantly larger decline resulting in below-baseline levels on day 4 (P=0.001). (Figure 1)

Discussion

This study presents the following findings. First, prolonged exercise induced an immediate increase in pro- and anti-inflammatory cytokines, and the magnitude of this response was not different between lean and overweight/obese individuals. The exercise-induced elevation in cytokine levels was

attenuated following exercise on consecutive days. Except for IL-8, no differences in cytokine responses between lean and overweight/obese individuals were found. Our data suggest the presence of early adaptive mechanisms in inflammatory cytokines in response to repeated prolonged exercise bouts performed on consecutive days, which did not markedly differ between lean and overweight/obese individuals.

In contrast to our hypothesis, no differences in plasma cytokines between the lean and overweight/obese group were present at baseline. In this study, relatively fit subjects were included since all subjects participated in a 4-day walking event. Previous work has shown that overweight and obese subjects with higher cardiorespiratory fitness levels, as a result of higher levels of physical activity, demonstrate lower levels of circulating pro-inflammatory cytokines, compared to unfit individuals.¹³ Furthermore, we included subjects with only modest obesity (range BMI: 25-32.9 kg/m²). Higher levels of BMI are significantly related to higher levels of inflammation.¹⁴ Last, the individuals in the overweight/obese cohort report similar caloric and macronutrient intake when compared to the individuals in the lean cohort, despite being overweight/obese. It can be hypothesized that the reported dietary intake of the overweight/obese cohort is relatively healthy because these are fit individuals who perform exercise on a regular basis. Therefore, the relatively high level of fitness, modest level of obesity and similar dietary intake when compared to lean controls in our study may explain the absence of differences in baseline levels of cytokines between the overweight/obese and lean group.

To our knowledge, this is the first human study that examined responses of different cytokines to repeated exercise bouts on subsequent days and whether these responses differ between lean and overweight/obese individuals. We found no differences between lean and overweight/obese individuals in responses of IL-6, IL-10, TNF- α and IL-1 β to repeated exercise. Exercise caused a subsequent rise in circulating IL-6 across the four consecutive exercise days in both groups. Of all known cytokines, IL-6 shows the largest response to exercise. This might explain why IL-6 plasma

levels remain elevated throughout the four-days of walking. Furthermore, previous work has shown that expression and circulating levels of IL-6 remain elevated at least 24 hours after cessation of an exercise bout, which might also have contributed to the persistent rise of circulating IL-6 in our study and why no group differences were found. Anti-inflammatory IL-10 showed a significant rise after the first exercise day. The release of IL-10 into the circulation is induced by the presence of IL-6, which was previously observed in both in vitro and in vivo work. This might explain the rise in IL-10 we observed after the first exercise day in both groups. However, IL-10 returns to baseline levels after the subsequent exercise days in both groups, despite the elevated levels of IL-6 on all 4 exercise days. It has been hypothesized previously that IL-6 levels have to reach a certain threshold to cause IL-10 production by leukocytes. Possibly this threshold was not reached on exercise day 2-4, since IL-6 levels are lower on exercise day 2-4 when compared to exercise day 1, which may explain the return to baseline of IL-10 levels from exercise day 2 onwards.

We observed a significant change in cytokines on day 1 (IL-8, TNF- α and IL-1 β) and day 2 (TNF- α in the overweight/obese cohort) when compared to baseline, that was no longer present on the consecutive exercise days. This suggests an attenuated acute response to exercise of pro-inflammatory cytokines (TNF- α , IL-8 and IL-1 β) after repeated bouts of prolonged exercise. In discordance with our hypothesis, we found no differences in this attenuation between lean and overweight/obese individuals, except for IL-8. Our time-effects results show a transient rise in IL-1 β on day 1 in the overweight/obese group, whilst IL-1 β in the lean group shows no change. The modest response of IL-1 β to exercise might relate to the presence of a persistent rise in IL-6. Previous work postulated that under influence of IL-6, the presence of IL-1receptor antagonist (IL1-ra) in the circulation is induced, which subsequently causes a decrease in IL-1 β by competitively binding to the IL-1receptor. The presence of elevated levels of IL-6, therefore, may contribute to the attenuated exercise-induced increase in IL-1 β in the overweight/obese group.

obesity and related to constitutes of the metabolic syndrome, such as waist circumference and insulin

resistance (i.e. HOMA-IR). ²⁰ The difference between the lean and overweight/obese group in IL-8 response to repeated prolonged exercise seems to be caused by the decrease in IL-8 in the lean cohort on exercise day 4 when IL-8 decreases below baseline. This attenuated response of IL-8 suggests the presence of early adaptations to repeated bouts of prolonged exercise. This is in line with previous work that found a decrease in IL-8 after exercise training, although the exercise stimulus in our study is different due to the prolonged duration.²¹

247

248

249

250

251

252

253

254

255

256

257

258

259

260

261

262

263

264

265

266

241

242

243

244

245

246

Based on our data, one may speculate that the shift from the pro-inflammatory effects of a single bout of prolonged exercise to the known anti-inflammatory effects of exercise training is mediated by a change in cytokine secretion in response to repeated prolonged exercise bouts. During acute prolonged exercise, cytokines are secreted from adipose tissue²² and skeletal muscle.²³ Exercise training is known to change gene expression in these tissues, which eventually results in altered secretion patterns of cytokines.²⁴⁻²⁷ Gene expression in skeletal muscle is altered during each prolonged exercise bout because of altered contractile activity,23 but is also believed to be influenced by the increased respiratory capacity in skeletal muscle that occurs by aerobic exercise training.²⁸ These adaptive responses, where responses to acute bouts of exercise relate to subsequent adaptation, have been referred to as hormesis: a biological process in which exposure to a low amount of a damaging factor leads to an adaptive beneficial effect in the organism.²⁹ Pro-inflammatory cytokines, i.e. the proinflammatory state which occurs during and after a single bout of exercise could be classified as a "hormesis stimulus", where the acute responses to exercise mediate an adaptive response contributing to health benefits when performed repeatedly.³⁰ The attenuated response of cytokines we observed in our study fits well in this hypothesis. This is further supported by a study that examined responses of IL-6 mRNA expression in skeletal muscle after a 3-h exercise protocol, before and after 10 weeks of exercise training in untrained men. A decrease in IL-6 mRNA expression levels in response to prolonged exercise from 76-fold (before training) to 8-fold (after the training period) was observed. 2727 Although it is important to emphasize that our design does not resemble the typical exercise training response, our data support the presence of an attenuated magnitude in exercise-induced changes in circulating cytokines when subjects repeat the same exercise stimulus on subsequent days.

Some limitations must be considered. Due to practical reasons, it was impossible to measure cytokines directly before the start of the walking exercise on the four consecutive days. Baseline levels were measured one or two days prior to the start of the walking event. Therefore, we were unable to assess potential adaptations in resting levels of cytokines (prior to each exercise bout). However, the primary goal of this study was to investigate differences between overweight and lean individuals in cytokine responses to repeated prolonged exercise bouts, which were therefore assessed immediately after cessation of such a bout. In our study, a prolonged exercise stimulus was used to examine cytokine responses to repeated exercise. Because of the duration of the exercise bouts $(8.6 \pm 2.1 \text{ hours})$ this design is not intended as a training study but rather as a model to examine physiological changes in response to repeated exercise stimuli.

Conclusion

This study demonstrated that prolonged exercise induces an immediate increase in pro- and antiinflammatory cytokines in lean and overweight/obese individuals while repeated bouts of prolonged
exercise lead to an attenuated exercise-induced cytokine response. Our data suggest that
overweight/obese subjects, when matched for sex, age and fitness, largely show comparable exerciseinduced changes in levels of cytokines across consecutive days of prolonged walking exercise.
Therefore, our data suggest the presence of early adaptive mechanisms in circulating cytokines in
response to repeated exercise bouts..

Practical Implications

• Cytokines are circulating factors that play a role in inflammation in the human body.

Inflammation contributes to the development of metabolic and cardiovascular disease. Our

293		study reveals that a prolonged walking exercise results in a rise in these cytokines that
294		attenuates when this exercise bout is repeated.
295	•	Our study demonstrates that both lean and overweight individuals largely show comparable
296		exercise-induced changes of cytokines across four days of repeated prolonged walking.
297	•	The attenuation of cytokine IL-8 occurs delayed in overweight individuals when compared to
298		lean controls.
299		
300		
301		
302		
303		
304		
305		
306		
307		
308		
309		
310		
311		
312		
313		
314		

References

- 1. Hotamisligil GS. Inflammation and metabolic disorders. *Nature*. 2006; 444(7121):860-867.
- 2. Eckel RH, Krauss RM. American Heart Association call to action: obesity as a major risk factor
- for coronary heart disease. AHA Nutrition Committee. Circulation. 1998; 97(21):2099-2100.
- 319 3. Kahn SE, Hull RL, Utzschneider KM. Mechanisms linking obesity to insulin resistance and type
- 320 2 diabetes. *Nature*. 2006; 444(7121):840-846.
- 321 4. Petersen AM, Pedersen BK. The anti-inflammatory effect of exercise. Journal of applied
- 322 physiology (Bethesda, Md. : 1985). 2005; 98(4):1154-1162.
- 323 5. Ostrowski K, Hermann C, Bangash A, Schjerling P, Nielsen JN, Pedersen BK. A trauma-like
- delevation of plasma cytokines in humans in response to treadmill running. The Journal of
- 325 *physiology.* 1998; 513 (Pt 3):889-894.
- 326 6. Suzuki K, Nakaji S, Yamada M, Totsuka M, Sato K, Sugawara K. Systemic inflammatory
- response to exhaustive exercise. Cytokine kinetics. Exercise immunology review. 2002; 8:6-
- 328 48.
- 329 7. Robson-Ansley P, Barwood M, Canavan J, et al. The effect of repeated endurance exercise on
- 330 IL-6 and sIL-6R and their relationship with sensations of fatigue at rest. Cytokine. 2009;
- 331 45(2):111-116.
- 332 8. Durnin JV, Womersley J. Body fat assessed from total body density and its estimation from
- 333 skinfold thickness: measurements on 481 men and women aged from 16 to 72 years. *The*
- 334 British journal of nutrition. 1974; 32(1):77-97.
- 335 9. Siebelink E, Geelen A, de Vries JH. Self-reported energy intake by FFQ compared with actual
- energy intake to maintain body weight in 516 adults. The British journal of nutrition. 2011;
- 337 106(2):274-281.
- 338 10. RIVM/Voedingscentrum. NEVO-tabel 2016. http://nevo-online.rivm.nl/.

339 11. Wendel-Vos GC, Schuit AJ, Saris WH, Kromhout D. Reproducibility and relative validity of the 340 short questionnaire to assess health-enhancing physical activity. Journal of clinical 341 epidemiology. 2003; 56(12):1163-1169. 12. Tanaka H, Monahan KD, Seals DR. Age-predicted maximal heart rate revisited. Journal of the 342 American College of Cardiology. 2001; 37(1):153-156. 343 344 13. Hong S, Dimitrov S, Pruitt C, Shaikh F, Beg N. Benefit of physical fitness against inflammation 345 in obesity: role of beta adrenergic receptors. Brain, behavior, and immunity. 2014; 39:113-120. 346 347 14. Festa A, D'Agostino R, Jr., Williams K, et al. The relation of body fat mass and distribution to markers of chronic inflammation. International journal of obesity and related metabolic 348 disorders: journal of the International Association for the Study of Obesity. 2001; 349 25(10):1407-1415. 350 351 Fischer CP. Interleukin-6 in acute exercise and training: what is the biological relevance? 15. 352 Exercise immunology review. 2006; 12:6-33. 353 16. Louis E, Raue U, Yang Y, Jemiolo B, Trappe S. Time course of proteolytic, cytokine, and 354 myostatin gene expression after acute exercise in human skeletal muscle. Journal of applied 355 physiology (Bethesda, Md.: 1985). 2007; 103(5):1744-1751. 356 17. Cullen T, Thomas AW, Webb R, Hughes MG. Interleukin-6 and associated cytokine responses 357 to an acute bout of high-intensity interval exercise: the effect of exercise intensity and 358 volume. Applied physiology, nutrition, and metabolism = Physiologie appliquee, nutrition et 359 metabolisme. 2016; 41(8):803-808. 360 18. Nieman DC, Henson DA, Davis JM, et al. Blood leukocyte mRNA expression for IL-10, IL-1Ra, 361 and IL-8, but not IL-6, increases after exercise. Journal of interferon & cytokine research: the official journal of the International Society for Interferon and Cytokine Research. 2006; 362

363

26(9):668-674.

- 19. Lancaster GI, Febbraio MA. The immunomodulating role of exercise in metabolic disease.
- 365 *Trends in immunology.* 2014; 35(6):262-269.
- 366 20. Kim CS, Park HS, Kawada T, et al. Circulating levels of MCP-1 and IL-8 are elevated in human
- obese subjects and associated with obesity-related parameters. *International journal of*
- 368 *obesity (2005).* 2006; 30(9):1347-1355.
- 369 21. Troseid M, Lappegard KT, Claudi T, et al. Exercise reduces plasma levels of the chemokines
- 370 MCP-1 and IL-8 in subjects with the metabolic syndrome. European heart journal. 2004;
- 371 25(4):349-355.
- 372 22. Keller C, Keller P, Marshal S, Pedersen BK. IL-6 gene expression in human adipose tissue in
- response to exercise--effect of carbohydrate ingestion. *The Journal of physiology.* 2003;
- 374 550(Pt 3):927-931.
- 375 23. Steensberg A, van Hall G, Osada T, Sacchetti M, Saltin B, Klarlund Pedersen B. Production of
- interleukin-6 in contracting human skeletal muscles can account for the exercise-induced
- increase in plasma interleukin-6. *The Journal of physiology.* 2000; 529 Pt 1:237-242.
- 378 24. Pedersen BK, Febbraio MA. Muscles, exercise and obesity: skeletal muscle as a secretory
- 379 organ. *Nature reviews. Endocrinology.* 2012; 8(8):457-465.
- 380 25. Stanford KI, Middelbeek RJ, Goodyear LJ. Exercise Effects on White Adipose Tissue: Beiging
- and Metabolic Adaptations. *Diabetes*. 2015; 64(7):2361-2368.
- 382 26. Bluher M, Williams CJ, Kloting N, et al. Gene expression of adiponectin receptors in human
- visceral and subcutaneous adipose tissue is related to insulin resistance and metabolic
- parameters and is altered in response to physical training. *Diabetes care.* 2007; 30(12):3110-
- 385 3115.
- 386 27. Fischer CP, Plomgaard P, Hansen AK, Pilegaard H, Saltin B, Pedersen BK. Endurance training
- reduces the contraction-induced interleukin-6 mRNA expression in human skeletal muscle.
- 388 American journal of physiology. Endocrinology and metabolism. 2004; 287(6):E1189-1194.

389	28.	Hawley JA, Hargreaves M, Joyner MJ, Zierath JR. Integrative biology of exercise. Cell. 201	
390		159(4):738-749.	
391	29.	Mattson MP. Hormesis defined. Ageing research reviews. 2008; 7(1):1-7.	
392	30.	Ost M, Coleman V, Kasch J, Klaus S. Regulation of myokine expression: Role of exercise and	
393		cellular stress. Free radical biology & medicine. 2016; 98:78-89.	
394			
395			

Table 1. Physiological characteristics of the study groups

Table 1. Physiological characteris	Lean subjects (n=25)	Overweight/Obese subjects (n=25)	P-value*
Baseline characteristics			
Age (years)	56.4 ± 14.4	58.4 ± 11.9	0.60
Male sex (%)	56%	56%	1.00
Weight (kg)	69.3 ± 7.7	84 ± 12.6	< 0.0001
Body mass index (kg/m²)	22.9 ± 1.5	27.9 ± 2.4	< 0.0001
Body fat percentage (%)	27.3 ± 6.6	33.5 ± 6.7	0.002
Waist-to-hip ratio	0.89 ± 0.1	0.95 ± 0.1	0.02
Systolic blood pressure (mmHg)	139 ± 21	142 ± 16	0.59
Diastolic blood pressure (mmHg)	86 ± 12	89 ± 9	0.92
Resting heart rate (bpm)	62 ± 8	63 ± 7	0.54
Daily physical activity levels			
Total SQUASH score	6342 ± 3974	7397 ± 4687	0.41
METmin/day	968 ± 522	1130 ± 629	0.32
Habitual dietary intake			
Caloric intake (kJ)	9592 ± 2516	9570 ± 3441	0.98
Total protein (g)	82 ± 21	87 ± 30	0.48
Total fat (g)	93 ± 34	89 ± 35	0.72
Saturated fat (g)	31 ± 13	31 ± 15	0.87
Total carbohydrates (g)	249 ± 64	244 ± 101	0.86
Fibre (g)	27 ± 7	25 ± 10	0.34
Dietary anti-oxidant intake			
Retinol (µg)	616 ± 369	655 ± 446	0.74
Vitamine E (mg)	16 ± 5	16 ± 7	0.71
Vitamine C (mg)	121 ± 54	115 ± 59	0.71
Exercise characteristics			
Exercise intensity (%HR _{max})	66 ± 5	69 ± 5	0.11
Exercise distance	5 15 5	5 15 5	- - -
Exercise duration day 1 (minutes)	510 ± 129	444 ± 167	0.12
Exercise duration day 2 (minutes)	534 ± 83	522 ± 98	0.64

Exercise duration day 3 (minutes)	508 ± 140	509 ± 114	0.97
Exercise duration day 4 (minutes)	565 ± 112	540 ± 124	0.46

*One-way ANOVA between lean and overweight subgroups

Figure Legends

Figure 1 Mean circulating cytokine levels of IL-6 (A); IL-10 (B); TNF α (C); IL-1 β (D) and IL-8 (E) at baseline and after each exercise day, with data being presented for lean subjects (\bigcirc) and overweight/obese subjects (\blacksquare). Error bars represent the standard error of the mean. * Significantly different from baseline in lean group (P <0.05); * Significantly different from baseline in overweight/obese group (P <0.05)

411 Figure 1

